Canada vario: 10

# Revised Robust Summaries for C.I. Acid Yellow 23

CAS No. 1934-21-0

# **Consortium Registration Number**

Submitted to the EPA under the HPV Challenge Program by:
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# **List of Member Companies**

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Noveon, Inc.

**Sensient Colors, Inc.** 

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## **Robust Summaries**

## for C.I. Acid Yellow 23

The evaluation of the quality of the following data uses a systematic approach described by Klimisch [Klimisch *et al.*, 1996]. Based on criteria relating to international testing standards for categorizing data reliability, four reliability categories have been established. The following categories are:

Reliability code 1. Reliable without restrictions
Reliable with restrictions

• Reliability code 3. Not reliable

• Reliability code 4. Not assignable

#### 1 CHEMICAL AND PHYSICAL PROPERTIES

#### 1.1 MELTING POINT

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for substance FD&C Yellow 5

Method/guideline Calculated

**GLP** 

Year

**Remarks for Test Conditions** 

Melting Point 350 °C

Decomposition

**Sublimation** 

#### **Remarks for Results**

#### **Conclusion Remarks**

References

Remarks for General Remarks	
Data Qualities Reliabilities	Reliability code 4. Not assignable.
Remarks for Data Reliability	Code 4. Calculated.

MPBPVPWIN EPI Suite (2000) US Environmental Protection Agency.

#### 1.2 BOILING POINT

CAS Numerical 1934-21-0

Substance Name	C.I. Acid Yellow 23	

Remarks for Substance FD&C Yellow 5

Method/guideline Calculated

GLP

Year

**Remarks for Test Conditions** 

**Boiling Point** 870 °C

**Pressure** 

**Pressure Unit** 

**Decomposition** 

**Remarks for Results** 

**Conclusion Remarks** 

**Remarks for General** 

Remarks

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References MPBPVPWIN EPI Suite (2000) US Environmental Protection

Agency.

#### 1.3 VAPOR PRESSURE

**CAS Numerical** 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for substance FD&C Yellow 5

Method/guideline Calculated/Mean of Antoine & Grain

**GLP** No

Year

**Remarks for Test Conditions** 

Vapor Pressure 7.43 X 10-22 mm Hg

Temperature 25 °C

**Decomposition** 

**Remarks for Results** 

**Conclusion Remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

**References** MPBPVPWIN EPI Suite (2000) US Environmental Protection

Agency.

#### 1.4 N-OCTANOL/WATER PARTITION COEFFICIENTS

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for substance FD&C Yellow No. 5

Method/guideline Calculated

GLP

Year

**Remarks for Test Conditions** 

**Log Pow** -10.17

**Temperature** 

**Remarks for Results** 

**Conclusion Remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

**References** KOWWIN EPI Suite (2000) US Environmental Protection

Agency.

#### 1.5 WATER SOLUBILITY

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance Purity not given

Method/guideline Not given

**GLP** Ambiguous

**Year** 1991

Remarks for Test Conditions Not given

Value (mg/L) at temperature 38,000 mg/ml at 2 °C; 200,000 mg/ml at 25 °C; 200,000 mg/ml

at 60 °C

**Description of Solubility** Not given

pH value and concentration

at temp

pKa value at 25 Celsius

**Remarks for Results** 

**Conclusion Remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4.Only secondary literature (review, tables, books, etc.).

**References** Marmion D.M. (1991) Handbook of U.S. Colorants: Foods,

Drugs, and Cosmetics and Medical Devices. 3rd Ed. New York,

John Wiley & Sons, Inc.

#### 2 ENVIRONMENTAL FATE AND PATHWAYS

#### 2.1 PHOTODEGRADATION

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance Data are for structurally related substance 2-

Naphthalenesulfonic acid, 6-hydroxy-5-[(2-methoxy-5-methyl-4-

sulfophenyl)azo]-, disodium salt (FD&C Red 40)

Method/guideline Not given

Test Type Experimental

**GLP** Ambiguous

**Year** 1991

Light Source 15-watt General Electric germicidal lamps

Light Spectrum (nm) Ultraviolet

**Relative Intensity** 

**Spectrum of Substance** 

Remarks for Test Conditions The concentration of the dye solution was measured before

and after the photolysis using the Hewlett-Packard 8452A diode-array UV/Visible Spectrophotometer. Red 40 was prepared in an initial concentration of 5 mg/l. In the first part of the study, photolysis experiments were conducted using two 15-W (30 Watts total) General Electric germicidal lamps as the ultraviolet light source. The distance between the light source and the reaction vessels was approximately 2.5 cm. Both direct photolysis and indirect photolysis experiments were conducted.

The indirect photolysis experiment used acetone as the sensitizer for indirect photodegradation.

Concentration of Substance 5 mg/L

**Temperature** 

**Direct photolysis** 7% degradation after 50 minutes

Halflife t1/2

Degradation % after

Quantum yield

**Indirect photolysis** 99% degradation after 20 minutes

Sensitizer acetone

Concentration of sensitizer 5 mg/L

Rate constant

**Degradation %after** 

**Breakdown products** 

Remarks field for results

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

**Remarks for Data Reliability** Code 2. Basic data given: comparable to guidelines/standards.

**References** Pasin B. and Rickabaugh J. (1991) Destruction of Azo Dyes by

Sensitized Photolysis. Hazard. Ind. Wastes, 359-367.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow 5

Method/guideline Calculation

Test Type AOPWIN

GLP

Year

**Light Source** 

**Light Spectrum (nm)** 

**Relative Intensity** 

**Spectrum of Substance** 

**Remarks for Test Conditions** 

**Concentration of Substance** 

**Temperature** 

**Direct photolysis** 

Halflife t1/2 3.5 hours

Degradation % after

Quantum yield

Indirect photolysis

Sensitizer

**Concentration of sensitizer** 

Rate constant

**Degradation %after** 

**Breakdown products** 

Remarks field for results

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References AOPWIN EPI Suite (2000) US Environmental Protection

Agency.

## 2.2 BIODEGRADATION

CAS Numerical	1934-21-0	
Substance Name	C.I. Acid Yellow 23	
Remarks for Substance	Data are for structurally related sulfonic acid C.I. Acid Red No. 9( benzenesulfonic acid 5-chloro-2-[(2-hydroxyl-1-naphthenyl)azo]-4-methyl, barium salt (CAS No-5160-02-1); Assay, 90%	
Method	OECD 301C	
Test Type		
GLP	Ambiguous	
Year	1992	
Contact time (units)	28 days	
Innoculum	Activated sludge	
Remarks for Test Conditions	Standard OECD 301C guideline study	
Results	Not biodegradable	

#### Classification

Remarks fields for results	In Zahn-Wellens test, after 21 days, 33% was loss with 10% absorbed on the sludge.
Conclusion remarks	
Data Qualities Reliabilities	Reliability code 1. Reliable without restriction.
Remarks for Data Reliability	Code 1. Comparable to guideline study.
References	OECD SIDS (1999) 9 <sup>th</sup> SIAM for D&C Red No. 9
CAS Numerical	1934-21-0
Substance Name	C.I. Acid Yellow 23
Remarks for Substance	Data are for structurally related sulfonic acid C.I. Acid Red No. 14.
Method	Not given
Test Type	
GLP	Ambiguous
Year	1993
Contact time (units)	24 hour
Innoculum	Activated sludge
Remarks for Test Conditions	Screened raw wastewater was used as the influent in three pilot scale activated sludge biological treatment systems. Each water soluble dye was tested at doses of 1 mg/L for low spike systems and 5 mg/L for high spike systems of influent flow. Before the data collection, dye analytical recovery studies were conducted by dosing the purified dye compound into organic free water, influent wastewater, and mixed liquor. These studies were run in duplicate and each recovery study was repeated at least once to ensure that the dye compound could be extracted. Purified dye standards were analytically prepared from the commercial dye product by repeated recrystallization.  The INF, primary effluent (PE), and ASE were filtered and the filtrate was passed through a column packed with resin. The filter paper and resin were soaked in an ammonia acetonitrile solution and then Soxhlet extracted with ammonia-acetonitrile. The extract was concentrated and brought up to 50 mL volume with a methanol/dimethylformamide solution. The mixed liquor samples were separated into two components, the filtrate or
	soluble fraction (SOL) and the residue (RES) fraction. The SOL fraction was processed similar to these samples but he resin adsorption step was omitted. All extracted samples were analyzed by HPLC with and ultraviolet-visible detector. Total suspended solids analyses were also performed on the INF, PE, ML, and ASE samples.

All systems were operated for at least three times the solids retention time to ensure acclimation prior to initiation of data collection. All samples were 24 hour composites made up of 6 grab samples collected every 4 hour and stored at 4 °C.

Percent recovery as measured: Organic Free Water: 101% at 1 mg/L and 90% at 5 mg/L; Wastewater: 98% at 1mg/L and 97% at 5 mg/L; Mixed Liquor: 88% at 1mg/L and 92% at 5 mg/L Mass Balance Data Summary: Low spike: 116% recovered, 1%

adsorbed; High spike: 148% recovered, less than 1%

adsorbed.

Classification

Results

Remarks fields for results Since the majority of the test substance was recovered, the

authors assumed that this compound was not biodegraded. The authors based this assumption on preliminary data indicating little or no problems in recovering the compounds from the sample matrix. Additionally the results also indicate that the material was not adsorbed. The authors attributed the high sulfonic acid substitution on the test substance as the reason why the material was not removed by the microbial cells

or cell byproducts and subject to aerobic biodegradation.

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

**Remarks for Data Reliability** Code 1. Comparable to guideline study.

**References** Shaul G.M., Holdsworth T.J., Dempsey C.R., and Dostal K.A.

(1990) Fate of water soluble azo dyes in the activated sludge

process. Chemosphere 22, p107-119.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow 5

Method

Test Type Calculated

**GLP** 

Year

Contact time (units)

Innoculum

**Remarks for Test Conditions** 

Results

Classification Not readily biodegradable

Remarks fields for results

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

**References** BIOWIN EPI Suite (2000) US Environmental Protection

Agency.

#### 2.3 FUGACITY

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5

Model Conditions 25 C, 100,000 lbs.

Test Type Environmental Equilibrium Partitioning Model

Method Mackay

Model Used (title, version,

date)

EQC V 2.70 Level III

**Input parameters** MW, log Kow, water solubility, MP & VP

Year

**Remarks for Test Conditions** 

**Media** Air

absorption coefficient

**Desorption** 

Volatility

Model data and results

**Estimated Distribution and** 

**Media Concentration** 

Remarks

3.05E-13%

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References ECOSAR EPI Suite (2000) US Environmental Protection

Agency. Level III. Fugacity.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5

Model Conditions 25 C, 100,000 lbs.

Test Type Environmental Equilibrium Partitioning Model

Method Mackay

Model Used (title, version,

date)

EQC V 2.70 Level III

**Input parameters** MW, log Kow, water solubility, MP & VP

Year

**Remarks for Test Conditions** 

Media Water

absorption coefficient

Desorption

Volatility

Model data and results

**Estimated Distribution and** 

**Media Concentration** 

Remarks

51.8%

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References ECOSAR EPI Suite (2000) US Environmental Protection

Agency. Level III. Fugacity.

**CAS Numerical** 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5

Model Conditions 25 C, 100,000 lbs.

Test Type Environmental Equilibrium Partitioning Model

Method Mackay

Model Used (title, version,

date)

EQC V 2.70 Level III

**Input parameters** MW, log Kow, water solubility, MP & VP

Year

**Remarks for Test Conditions** 

Media Soil

absorption coefficient

Desorption

Volatility

Model data and results

**Estimated Distribution and** 

**Media Concentration** 

**Remarks** 

48.1%

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References ECOSAR EPI Suite (2000) US Environmental Protection

Agency. Level III. Fugacity.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5

Model Conditions 25 C, 100,000 lbs.

Test Type Environmental Equilibrium Partitioning Model

Method Mackay

Model Used (title, version,

date)

EQC V 2.70 Level III

**Input parameters** MW, log Kow, water solubility, MP & VP

Year

**Remarks for Test Conditions** 

Media Sediment

absorption coefficient

Desorption

Volatility

Model data and results

Estimated Distribution and Media Concentration

Remarks

0.0981%

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References ECOSAR EPI Suite (2000) US Environmental Protection

Agency. Level III. Fugacity.

#### 3 ECOTOXICITY

#### 3.1 Acute Toxicity to Fish

Substance Name C.I. Acid Yellow 23

CAS No. 1934-21-0

Remarks for Substance Data are for structurally related azo dye, benzenesulfonic acid 5-chloro-2-[(2-hydroxyl-1-naphthenyl)azo]-4-methyl, barium salt (CAS No-5160-02-1); Assay, 90%

**Test Type** Experimental (semi-static) Method 84/449/EEC

GLP Yes Year 1982

**Species/Strain/Supplier** Fish (Oryzias latipes) (Orange killifish)

**Exposure Period** 96 hour

**Remarks for Test Condition** A group of 10 fish were exposed to 5 nominal concentrations.

Two controls, DMSO(0.5 mg/L) and lab water were used

**Endpoint value** 96-hr LC50 = >420 mg/L

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

Remarks for Data Reliability Code 1. Guideline study.

**Reference** Hoechst AG (1992). Unveroeffentlichte Untersuchung

(82.0250).

Substance Name C.I. Acid Yellow 23

**CAS No.** 1934-21-0

Remarks for Substance Data are for structurally related azo dye, benzenesulfonic acid

5-chloro-2-[(2-hydroxyl-1-naphthenyl)azo]-4-methyl, barium salt

(CAS No-5160-02-1); Assay, 90%

**Test Type** Experimental (static) Method 84/449/EEC

GLP Yes Year 1982

Species/Strain/Supplier Fish (Brachydanio rerio)

**Exposure Period** 96 hour

**Remarks for Test Condition** A group of 10 fish were exposed to 5 nominal concentrations.

Two controls, DMSO(0.5 mg/L) and lab water were used

Endpoint value	96-hr LC50 = >500 mg/L
----------------	------------------------

Data Qualities Reliabilities	Reliability code 1. Reliable without restriction.
	,
Remarks for Data Reliability	Code 1. Guideline study.
Reference	Hoechst AG (1992). Unveroeffentlichte Untersuchung (82.0250).
Substance Name	C.I. Acid Yellow 23
CAS No.	1934-21-0
Remarks for Substance	Data are for structurally related azo dye, D&C Red No. 7, 2-naphthalenecarboxylic acid, [(4-methyl-2-sulfophenyl)azo], calcium salt acid (CAS No-5281-04-9); Assay, 87%
Test Type GLP	Experimental (flow-through) Japanese Industrial Standard (JIS K 0102-1986) Yes
Year	1992
Species/Strain/Supplier	Fish (Oryzias latipes) (Orange killifish)
Exposure Period	96 hour
Remarks for Test Condition	NA
Endpoint value	48-hr LC50 = 50 mg/L
Data Qualities Reliabilities	Reliability code 2. Reliable with restriction.
Remarks for Data Reliability	Code 1. Comparable to guideline study.
Reference	MITI, Japan (1992).
Substance Name	C.I. Acid Yellow 23
CAS No.	1934-21-0
Remarks for Substance	Data are for structurally related azo dye, D&C Red No. 7, 2-naphthalenecarboxylic acid, [(4-methyl-2-sulfophenyl)azo], calcium salt acid (CAS No-5281-04-9); Assay, 87%
Test Type	Experimental (OECD Guideline 203-semi-static-open system)
GLP	Ambiguous
Year	Not given
Species/Strain/Supplier	Fish (Oryzias latipes) (Orange killifish)
Exposure Period	96 hour

Remarks for Test Condition A group of 10 fish were exposed to 5 nominal concentrations of

17.1 to180 mg/L. Two controls, DMSO(0.5 mg/L) and lab water

were used

**Endpoint value** 96-hr LC50 = 33 mg/L (95% C.I., 11-98 mg/L)

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

**Remarks for Data Reliability** Code 1. Comparable to guideline study.

Reference EA, Japan (1992).

Substance Name C.I. Acid Yellow 23

**CAS No.** 1934-21-0

Remarks for Substance Data are for structurally related azo dye, benzenesulfonic acid

5-chloro-2-[(2-hydroxyl-1-naphthenyl)azo]-4-methyl, barium salt

(CAS No-5160-02-1); Assay, 90%

**Test Type** Experimental (semi-static) Method 84/449/EEC

**GLP** Yes

**Year** 1982

**Species/Strain/Supplier** Fish (Oryzias latipes) (Orange killifish)

**Exposure Period** 96 hour

**Remarks for Test Condition** A group of 10 fish were exposed to 5 nominal concentrations.

Two controls, DMSO(0.5 mg/L) and lab water were used

Endpoint value 96-hr LC50 = >500 mg/L

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

Remarks for Data Reliability Code 1. Guideline study.

**Reference** Hoechst AG (1992). Unveroeffentlichte Untersuchung

(82.0250).

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow 5

Method/guideline ECOSAR

Test Type Calculated

**GLP** 

Year

Species/Strain/Supplier

Analytical monitoring

Exposure period (unit) 96 hour

Remarks for Test Conditions Input parameters: Molecular weight, Water solubility, 200,000

mg/L at 25 °C

Observations on precipitation

Nominal concentrations as

mg/L

Measured concentrations as

mg/L Unit

**Endpoint value** LC50 = 1.14 E+14 mg/L

Reference substances (if

used)

Remarks fields for results

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References ECOSAR EPI Suite (2000) U.S. Environmental Protection

Agency (Nabholz V. and G. Cash, 1998).

#### 3.2 Acute Toxicity to Aquatic Invertebrates

Substance Name C.I. Acid Yellow 23

CAS No. 1934-21-0

Remarks for Substance Data are for structurally related azo dye, benzenesulfonic acid 5-chloro-2-[(2-hydroxyl-1-naphthenyl)azo]-4-methyl, barium salt (CAS No-5160-02-1); Assay, 90%

Test Type Experimental OECD 202

**GLP** Yes

**Year** 1992

Species/Strain/Supplier Dapnid (Daphnia magna)

**Exposure Period** 48 hour

Remarks for Test Condition Saturated solution of test material was used

Endpoint value 48-hr EC50 = >2 mg/L

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

Remarks for Data Reliability Code 1. Guideline study.

Reference Hoechst AG (1993). Unveroeffentlichte Untersuchung

(93.0358).

Substance Name C.I. Acid Yellow 23

**CAS No.** 1934-21-0

Remarks for Substance Data are for structurally related azo dye, D&C Red No. 7, 2-

naphthalenecarboxylic acid, [(4-methyl-2-sulfophenyl)azo],

calcium salt acid (CAS No-5281-04-9); Assay, 87%

Test Type Experimental (static) OECD 202 Guideline Study

**GLP** No

**Year** 1984

Species/Strain/Supplier Daphnid (Daphnia magna)

**Exposure Period** 24 hour

Remarks for Test Condition 20 daphnids( 4 replicates, 5 organisms per plate) were exposed

to 5 nominal concentrations of 90-940 mg/L. Control was

DMSO;DCO40=9:1 (100 mg/L) and lab water.

**Endpoint value** 24-hr EC50 = 280 mg/L (95% C.I.=150-490 mg/L)

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

**Remarks for Data Reliability** Code 1. Comparable to guideline study.

Reference EA, Japan (1992).

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow 5

Method/guideline ECOSAR

Test Type Calculated

GLP

Year

Analytical procedures

Species/Strain Daphnia magna

Test details 48 hours

Input parameters: Water solubility, 200,000 mg/L at 25 °C; **Remarks for Test Conditions** 

Molecular weight 556.34

Nominal concentrations as

mq/L

Measured concentrations as

ma/L Unit

EC50, EL50, LC0, at 24,48

**Biological observations** 

EC50 = 5.25 E+13 mg/L

**Control response** satisfactory? Appropriate statistical

evaluations?

Remarks fields for results

**Conclusion remarks** 

Reliability code 4. Not assignable. **Data Qualities Reliabilities** 

Remarks for Data Reliability Code 4. Calculated.

ECOSAR EPI Suite (2000) U.S. Environmental Protection References

Agency (Nabholz V. and G. Cash, 1998).

#### 3.3 Acute Toxicity to Aquatic Plants

**CAS Numerical** 1934-21-0

C.I. Acid Yellow 23 **Substance Name** The test substance was an unidentified sulfonic acid substituted **Remarks for Substance** azo dye.

Method/guideline

**Test Type** Experimental

**GLP Ambiguous** 

Year 1996 Species/Strain/Supplier Green algae, Selenatrum capricornutum

**Endpoint basis** 

Exposure period (duration) 96 hour

**Analytical monitoring** 

Remarks for Test Conditions Algal chronic toxicity test were performed according the method

of EPA, 1988. Three replicates were performed for each dye at a nominal concentration of 1 mg/l for the active colorant. One ml of dye stock solution was added to 50 mg/l of algal assay medium in 125 ml Erlenmeyer flasks. *S. capricornutum* in continuous culture provided the initial innoculum (10,000 algal cells/ml). The cells were incubated in the solution for 96 hours. The diluent and negative control were algal assay medium. AAM was prepared by adding 1 ml from each of five stock solutions to 900 ml of deionized water. After spiking, the total volume was brought to 1 liter with deionized water. Population growth was used to establish potential toxicity. If the dye inhibited algal growth by more than 50% of that of the negative controls, a definitive test using several dilutions of the dye was performed to allow for determination of an EC50 concentration.

Nominal concentrations as mg/L

Measured concentrations as

mg/L Unit

Endpoint value Average yield: 36.6% with 95% C.I. (34.9-38.4).

NOEC, LOEC or NOEL, LOEL

**Biological observations** 26.4% stimulation of population growth compared to control.

Control response satisfactory?

**Appropriate statistical** 

evaluations?

Remarks fields for results

Yes, Dunnett's test

Not statistically significant.

Conclusion remarks

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

Remarks for Data Reliability Code 1. Comparable to guideline study.

Yes

**References** Greene J. C. and Baughman G.L. (1996) Effects of 46 dyes on

population-growth of fresh-water green-alga selenastrum-capricornutum. Textile Chemist And Colorist, 28, 23-30.

Green J.D. et al. (1988) Protocols for short term toxicity screening of hazardous waste sites. Report to EPA 600/3-88-029. U.S. Environmental Protection Agency. Corvallis, Oregon.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow 5

Method/guideline ECOSAR

Test Type Calculated

**GLP** 

Year

Species/Strain/Supplier Green algae

**Endpoint basis** 

Exposure period (duration) 96 hour

**Analytical monitoring** 

Remarks for Test Conditions Input parameters: Water solubility - 200,000 mg/L at 25 °C;

Molecular weight 556.34

Nominal concentrations as

mg/L

Measured concentrations as

mg/L Unit

**Endpoint value** EC50 = 1.63 E+13 mg/L

NOEC, LOEC or NOEL, LOEL

**Biological observations** 

Control response satisfactory? Appropriate statistical

evaluations?

Remarks fields for results

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4. Calculated.

References ECOSAR EPI Suite (2000) US Environmental Protection

Agency (Nabholz V. and G. Cash, 1998).

#### **4 HUMAN HEALTH TOXICITY**

#### 4.1 ACUTE TOXICITY

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance Not given

Method/guideline Not given

**Test Type** Acute Toxicity LD50

**GLP** No

**Year** 1957

Species/Strain Rat

Sex Not reported

# of animals per sex per

dose

Not given

Vehicle Not given

Route of administration Intraperitoneal

Remarks for test conditions

Value LD50 or LC50 with

confidence limits

Number of deaths at each

dose level

Remarks for results

2,000 mg/kg bw

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4.Only secondary literature (review, tables, books, etc.).

**References** Deutsche Forschungsgemeinschaft, Bad Godesberg, Federal

Republic of Germany, Farbstoff Kommission (1957) Mitteilung

6.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance Not given

Method/guideline Not given

**Test Type** Acute Toxicity LD50

**GLP** No

**Year** 1957

Species/Strain Rat

Sex Not reported

# of animals per sex per

dose

Not given

Vehicle Not given

Route of administration Intravenous

Remarks for test conditions

Value LD50 or LC50 with

confidence limits

Number of deaths at each

dose level

Remarks for results

1,000 mg/kg bw

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

**Remarks for Data Reliability** Code 4.Only secondary literature (review, tables, books, etc.).

**References** Deutsche Forschungsgemeinschaft, Bad Godesberg, Federal

Republic of Germany, Farbstoff Kommission (1957) Mitteilung

6.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance Not given

Method/guideline Not given

**Test Type** Acute Toxicity LD50

**GLP** No

**Year** 1964

Species/Strain Mice

Sex Not reported

# of animals per sex per

dose

Not given

Vehicle 1% gum arabic

Route of administration Oral

**Remarks for test conditions** 

Value LD50 or LC50 with

confidence limits

Number of deaths at each

dose level

Remarks for results

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 4. Not assignable.

Remarks for Data Reliability Code 4.Only secondary literature (review, tables, books, etc.).

References National Institute of Hygienic Sciences of Japan. Unpublished

data submitted to WHO, 1964 cited in ILSI report on FD&C

Yellow 5 6/2/83.

12,750 mg/kg bw

#### 4.2 GENETIC TOXICITY

#### 4.2.1 In vitro Genotoxicity

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

**Remarks for Substance** FD&C Yellow No. 5; Purity not given

**Method/guideline** Ames plate incorporation and liquid pre-incubation

**Test Type** Reverse mutation

System of Testing Bacterial

**GLP** Ambiguous

**Year** 1981

**Species/Strain** Salmonella typhimurium TA1535, TA 1537, TA1538, TA98,

TA100

Metabolic Activation With and without Rat liver microsome fraction S9 from Aroclor

induced rats

**Doses/concentration levels** 0.005- 5.0 mg/plate

#### **Statistical Methods**

Not given

#### Remarks for test conditions

The Salmonella typhimurium teste strains were grown in nutrient broth shaken for 14 hr at 37 deg C. The revertant colonies were counted by using a hand held tally. DMSO was the solvent except sterile distilled water was the solvent when sulfanilic acid was used. Liver homogenates were prepared from male Sprague Dawley rats stimulated with Aroclor 1254 (500 mg/kg intraperitoneally 5 days before sacrifice). The S9 mix added in samples of 0.5 ml per plate contained 3 mg of protein determined by the method of Lowry et al. Reverse mutation tests were carried out using *S. typhimurium* strains TA1535, TA 1537, TA1538, TA98, TA100. Plate incorporation tests were conducted according to Ames et al., with the Andrews et al. modifications. Duplicates were performed at each of the six concentrations of the test substance. Mutagenic compounds were assayed using duplicate plates. A substance was considered positive when the number of revertants above background was at least twice the value of the historical control mean or twice the value of the current control mean, whichever was greater and a dose response curve could be generated.

Positive controls without metabolic activation were sodium azide (TA1535 and TA100), 9-aminoacridine (TA97 and TA1535), and 4-nitro-o-phenylenediamine (TA98). The positive controls were sodium azide, 9-aminoacridine, 2-nitrofluorene, and 2-

aminoanthracene.

Result

Negative

Positive control Results	(-S9/+S9)
--------------------------	-----------

Cmpd Amt per plate	TA1538	TA98	TA100
None	9/24	11/35	100/87
DMSO .1 ml	11/19	21/27	124/99
Sodium azide 0.5 ug	13/20	11/23	1165/96
2-Nitrofluorene 5 ug	728/239	578/171	1586/525
2-Aminoanthracene 2.5	ug 15/882	22/799	90/2593

Cytotoxic concentration

5.0 mg/plate for plate-incorporation, and .5 mg/ml for pre-

incubation test.

Genotoxic effects

Negative

Appropriate statistical

evaluations?

None given

Remarks for results

Negative

Conclusion remarks

The test substance was negative in the AMES assa:v for reverse

mutation using Salmonella typhimurium TA1535, TA 1537,

TA1538, TA98, TA100.

**Data Qualities Reliabilities** 

Reliability code 1. Reliable without restriction.

Remarks for Data Reliability

Code 1. Guideline study.

References

Chung K.T., Fulk G.E., & Andrews A.W. (1981) Mutagenicity

testing of some commonly used dyes. Applied and

Environmental Microbiology 42, 641-648.

**CAS Numerical** 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5; Purity not given

Method/guideline Ames

**Test Type** Reverse mutation

System of Testing Bacterial

GLP No

**Year** 1979

Species/Strain Salmonella typhimurium TA1535, TA 1537, TA98, TA100

Metabolic Activation Rat liver microsome fraction S9 from Aroclor induced rats

Doses/concentration levels 10-250 mg/plate

Statistical Methods Not given

Remarks for test conditions 
The test substance was dissolved in DMSO. The test was

considered positive if 2 fold increase in revertants was observed. Positive controls included 9-aminoacridine; 2-aminoflourine; N-

methyl-N-nitrosoguanidine.

Result Negative

Cytotoxic concentration Not given

Genotoxic effects Negative

Appropriate statistical

evaluations?

None given

Remarks for results Negative

**Conclusion remarks** No evidence of genotoxicity was reported.

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

Remarks for Data Reliability Code 2. Basic data given: comparable to guidelines/standards.

References Muzzall J.M. and Cook W.I. (1979) Mutagenicity test of dyes

used in cosmetics with the Salmonella/mammalian microsome

test. Mutations Research 67, 1-8.a.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5; Purity not given

Method/guideline Ames

Test Type Reverse mutation

System of Testing Bacterial

GLP Ambiguous

**Year** 1984

Species/Strain Salmonella typhimurium TA1535, TA 1537, TA98, TA100, TA92,

TA94

Metabolic Activation Rat liver microsome fraction S9 from Aroclor induced rats

**Doses/concentration levels** Up to 5.0 mg/ml

Statistical Methods Not given

**Remarks for test conditions** Reverse mutation tests were carried out using *S. typhimurium* 

strains TA92, TA1535, TA100, TA1537, TA94 and TA98. Cells cultured overnight were pre-incubated with the test substance and the S-9 mix for twenty minutes at 37 degrees Celsius prior to plating. Duplicates were performed at each of the six concentrations of the test substance. The number of revertant colonies were counted following incubation for two days. Negative controls were either untreated plates or solvent.

Positive results were determined if the number of colonies found was twice the number in the control. If the test was positive and a dose response relationship was not detected, additional experiments at different doses or induced mutation frequency

assays were performed.

Result Negative

**Cytotoxic concentration** 5.0 mg/ml was the highest non-cytotoxic dose used in the

experiment.

Genotoxic effects Negative

Appropriate statistical

evaluations?

Remarks for results

None given

Negative

**Conclusion remarks** The test substance was negative in the AMES assay for reverse

mutation using Salmonella typhimurium TA1535, TA 1537,

TA98, TA100, TA92, TA94,

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

Remarks for Data Reliability Code 2. Basic data given: comparable to guidelines/standards.

References Ishidate, M., Sofuni, T., Yoshikawa, K., Hauashi, M., Nohmi, T.,

Sawada, M. and Matsuoka. (1984). Primary Mutagenicity Screening of Food Additives Currently Used in Japan. Fd.

Chem. Toxic. 22(8) 623-636.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

**Remarks for Substance** FD&C Yellow No. 5; Purity not given

Method/guideline Chromosomal aberration test was carried out using a Chinese

hamster fibroblast cell line, CHL. The cells were exposed to 3

different doses for 24 and 48 hours. No metabolic activation

system was applied.

Test Type Chromosomal aberration test

**System of Testing** Chinese hamster fibroblast cell line CHL.

**GLP** Ambiguous

**Year** 1984

**Species/Strain** Chinese hamster fibroblast cell line CHL.

Metabolic Activation None

Doses/concentration levels up to 2.5 mg/ml

Statistical Methods Not available

Remarks for test conditions

Chromosomal aberration tests were carried out using the Chinese hamster fibroblast line. Cells were exposed to the test substance at three different doses for 24 and 48 hour. No metabolic activation was employed. The maximum dose used for each test substance was found in a preliminary test to determine the dose required for 50% cell-growth inhibition. Colcemid at a final concentration of 0.2 ug/ml was added to the culture two hours prior to cell harvesting. The cells were prepared for viewing on slides. One hundred visible metaphases were observed under the microscope and the incidence of polyploid cells and structural chromosomal aberrations (including chromosome and chromatid gaps, breaks, exchanges, ring formations, fragmentations and others) were recorded. Negative controls included untreated cells and solvent treated cells. The incidence of aberrations in the negative controls was generally less than 3.0%. The results were considered negative if less than 4.9%, equivocal if between 5.0-9.9%, and positive if more than 10%. If dose response relationships were not observed, additional experiments were carried out at similar dose levels.

The maximum dose for positive results represents the dose at which the maximum effect was obtained.

For quantitative evaluation of the clastogenic potential, the D20 was calculated, which is the dose (mg/ml) at which structural aberrations (including gaps) were detected in 20% of the metaphases observed. In addition, the TR value was calculated, which indicates the frequency of cells with exchange-type aberrations per unit dose (mg/ml). These values are relatively high for chemicals that show carcinogenic potential in animals. The test substance was shown to be positive (23% total incidence of cells with aberrations) in chromosomal aberration test at 48 hours. TR value was 3.5 and D20 = 1.8. Weakly positive at 24 hour (11.0%, total incidence of cells with aberrations) The results were considered positive if the total incidence of cells with aberrations (including gaps) was 10.0% or more. Two percent (2%) reported as polyploid.

Result

Cytotoxic concentration Not given

Genotoxic effects Positive

Appropriate statistical

evaluations?

None given

Remarks for results

Positive

**Conclusion remarks**C.I. Acid Yellow 23 tested positive in the chromosomal

aberration test using Chinese hamster fibroblasts.

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

Remarks for Data Reliability Code 2. Acceptable, well-documented publication/study report

which meets basic scientific principles.

**References** Ishidate, M., Sofuni, T., Yoshikawa, K., Hauashi, M., Nohmi, T.,

Sawada, M. and Matsuoka. (1984) Primary Mutagenicity Screening of Food Additives Currently Used in Japan. Fd.

Chem. Toxic. 22(8) 623-636.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5; 94% purity

Method/guideline Wiliams, 1977

Test Type Unscheduled DNA Synthesis

System of Testing Rat hepatocytes

**GLP** Ambiguous

**Year** 1985

Species/Strain Rat/Sprague-Dawley

Metabolic Activation None

**Doses/concentration levels** 2 X 10-3

2 X 10-4 2 X 10-5 2 X 10-6

Statistical Methods None given

Remarks for test conditions Hepatocytes from rats were isolated and cultured according to

the two step in situ liver perfusion model (Malansky and Williams, 1982). Viable hepatocytes (2 X 10+5) were seeded in wells and incubated for 4 hours with [H3]-thymidine (10 uCi/ml) and the test substance (prepared in either DMSO or water) according to a procedure similar to Williams, 1977. Control incubations were conducted with and without DMSO. The authors state that DMSO had no effect on DNA repair. Two

experiments were conducted.

DNA repair was quantified by the autoradiographic

determination of incorporated [3H]-thymidine. Net nuclear grains

(NNG) were determined by counting the number of grains in each nuclei and subtracting the average number of grains present in the three equal size adjacent cytoplasmic areas. Average NNG counts of 5 or more were assumed to constitute a positive response, because these differed from the control response by greater than 2 standard deviations. In the negative controls, NNG counts ranged from -0.6- to -2.8 and from -0.9 to -2.1 for no solvent and 1% DMSO incubations, respectively. The proportion of cells with greater than or equal to 5 NNG was less than or equal to 8.1% for all control incubations. Therefore NNG below zero were considered negative responses.

Concentrations of dyes producing 90% or greater detachment of the hepatocytes from the coverslips were assumed to be toxic

and not counted.

The positive control was Solvent Yellow 3 (o-aminoazotoluene).

Result Experiment 1

Conc Avg NNG % >5NNG 2 X 10-3 -1.7 (+/-2.6) 5 2 X 10-4 -2.4 (+/-3.3) 5 2 X 10-5 -2.4 (+/-3.2) 2 2 X 10-6 -2.0 (+/-2.8) 3

Experiment 2

Conc Avg NNG % greater than 5NNG

2 X 10-3 -2.2 (+/-Greater than 2 X 10-3

Genotoxic effects Negative

Appropriate statistical

Cytotoxic concentration

evaluations?

Remarks for results

None given

Negative

Conclusion remarks C.I. Acid Yellow 23 did not induce unscheduled DNA synthesis

in an in vitro assay using rat hepatocytes isolated from the livers

of Sprague-Dawley rats.

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

**Remarks for Data Reliability** Code 2. Basic data given: comparable to guidelines/standards.

References Kornbrust D. and Barfknecht T. (1985) Testing Dyes in HPC/DR

systems. Enviromental Mutagenesis 7, 101-120.

## 4.2.2 In vivo Genotoxicity

CAS Numerical 1934
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0710 Humonou.	
Substance Name	C.I. Acid Yellow 23
Remarks for Substance	Data are for structurally related substance FD&C Yellow No. 6
Method/guideline	Rodent Micronucleus Test
Test Type	Rodent Micronucleus
GLP	Ambiguous
Year	1991
Species/Strain	Rat/PVG
Sex	Male
Route of administration	Oral-Gavage
Doses/concentration levels	10 ml/kg bw
Exposure period	Single dose
Remarks for test conditions	Male PVG rats received a single oral dose of 500, or 1000 mg/kg of the test substance. Bone marrow samples were taken at 24 and 48 hours later.
Effect on mitotic index or PCE/NCE ratio by dose level and sex	
Genotoxic effects	No significant increase in the frequency of micronucleated polychromatic erythrocytes at either time point and in either species was reported. Additionally, there was reported increase in the % PE (polychromatic erthyrocytes).
NOEL (C)/ LOEL (C)	(1.5)
Appropriate statistical evaluations?	Yes.
Remarks for results	No effects.
Conclusion remarks	
Data Qualities Reliabilities	Reliability code 2. Reliable with restriction.
Remarks for Data Reliability	Code 2. Acceptable, well-documented publication/study report which meets basic scientific principles.
References	Westmoreland C. and Gatehouse D.G. (1991) The differential clastogenicity of Solvent Yellow 14 and FD & C Yellow No. 6 in vivo in the rodent micronucleus test (observations on species and tissue specificity). Carcinogenesis 12 (8), 1403-8.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance 94% purity

Method/guideline Mirsalis and Butterworth, 1980

Test Type Unscheduled DNA Synthesis

**GLP** Ambiguous

**Year** 1985

Species/Strain Rat/Sprague Dawley

**Sex** Male

Route of administration Oral-Gavage

**Doses/concentration levels** 500 mg/kg bw

**Exposure period** 2 hr; 15 hr

Remarks for test conditions

Six to eight male Sprague-Dawley rats weighing 200-300 g were administered 500 mg acid yellow 23/kg bw *via* gavage. The control animal was administered corn oil only. Animals were killed at two time points, 2 hours and 15 hours. If negative results were obtained at time point 1 and time point 2, the *in vivo* testing was terminated and considered to be negative. If the initial test at time point 1 yielded a positive response, the test substance was retested at that time point. If another positive response was observed, the test was considered positive. Time points are the time the test substance was administered prior to the start of liver perfusion and isolation of hepatocytes.

Hepatocytes from rats were isolated and cultured according to the two step *in sit*u liver perfusion model (Malansky and Williams, 1982). Viable hepatocytes (2 X 10+5) were seeded in wells and incubated for 4 hours with [H3]-thymidine (10 uCi/ml) and the test substance (prepared in either DMSO or water) according to a procedure similar to Williams, 1977. Control incubations were conducted with and without DMSO. The authors state that DMSO had no effect on DNA repair.

DNA repair was quantified by the autoradiographic determination of incorporated [3H]-thymidine. Net nuclear grains (NNG) were determined by counting the number of grains in each nuclei and subtracting the average number of grains present in the three equal size adjacent cytoplasmic areas. Average NNG counts of 5 or more were assumed to constitute a positive response, because these differed from the control response by greater than 2 standard deviations. In the negative controls, NNG counts ranged from -0.6- to -2.8 and from -0.9 to -2.1 for no solvent and 1% DMSO incubations, respectively. The proportion of cells with greater than or equal

to 5 NNG was less than or equal to 8.1% for all control incubations. Therefore NNG below zero were considered negative responses. Concentrations of dyes producing 90% or greater detachment of the hepatocytes from the coverslips were assumed to be toxic and not counted.

The positive control was Solvent Yellow 3 (o-aminoazotoluene). Experiment 1

Effect on mitotic index or PCE/NCE ratio by dose level and sex

Dose (mg/kg bw) Time Avg NNG % >5NNG

500 2 hr -2.6 (+/-3.7) 2

> 15 hr -1.3 (+/-2.6)

Genotoxic effects Negative

Greater than 500 mg/kg bw NOEL (C)/ LOEL (C)

Appropriate statistical

evaluations?

None given

Remarks for results Negative

C.I. Acid Yellow 23 did not induce unscheduled DNA synthesis Conclusion remarks

in an invivo assay using rat hepatocytes isolated from the livers

of Sprague Dawley rats.

**Data Qualities Reliabilities** Reliability code 2. Reliable with restriction.

Code 2. Basic data given: comparable to guidelines/standards. Remarks for Data Reliability

References Kornbrust D. and Barfknecht T. (1985) Testing Dyes in

HPC/DR systems. Environmental Mutagenesis 7, 101-120.

#### 4.3 REPEATED DOSE TOXICITY

**CAS Numerical** 1934-21-0

C.I. Acid Yellow 23 **Substance Name** FD&C Yellow 5; 90% purity; 10% intermediates or volatile Remarks for Substance

matter

Chronic Toxicity/Carcinogenicity Study Method/guideline

**GLP** Yes

Year 1988

Species/Strain Rat/Charles River CD Sex Male and Female

Route of administration

0, 0.1, 1.0, or 2.0% (original study) 0, 5.0% (high dose study) **Doses/concentration levels** 

**Exposure period** 113 weeks (males) or 114 weeks (females) (original study); 122 weeks (males) or 125 weeks (females) high-dose study

Frequency of treatment Daily

**Oral-Diet** 

**Control Group** Yes, 2 concurrent controls (original study); 1 concurrent control (high-dose study)

Post exposure observation period Remarks for test conditions

In the *in utero* phase, groups of rats (60/sex/group) were administered 0, 0, 0.1, 1.0 or 2.0% FD & C Yellow No. 5 in the diet daily for approximately 2 months prior to mating. In the high-dose study, 60/sex/group received 0 or 5% FD&C Yellow 5 for approximately 2 months prior to mating. The 3 controls groups received the basal diet only. A maximum of 2 rats/sex/litter were randomly selected for the chronic phase of the study. There were 70/sex/group at the initiation of the chronic phase and these offspring were exposed to the same dietary levels as their parents.

Animals were housed individually and fed the test diet ad libitum. Clinical observations were recorded twice daily with at least 5 hours between observations. Detailed physical examinations and palpation for masses were performed weekly. Body weights and food consumption were determined weekly for the first fourteen weeks, bi-weekly for the next 12 weeks and every 4 weeks thereafter until the end of the study. The intake of the test substance was determined from body weight, food consumption and dietary concentration. Hematology tests, including hemoglobin, hematocrit, erythrocyte and total and differential leukocyte counts, and erythrocyte morphology, were conducted on ten randomly selected animals at months 3, 6, 12, 18 and 24 of the study. Necropsies were conducted on all animals dying prior to study termination, killed in a moribund condition or killed on schedule. Histological examinations were conducted on all animals from both control groups, the highest dose group (2.0 or 5.0%) from each study and also on 10 rats randomly selected from each group for an interim sacrifice at 12 months. Histology was also performed on any animal with gross lesions or masses.

Tissues examined included adrenal glands, aorta, blood smear, brain, cecum, colon, duodenum, epididymus or uterus, esophagus, eyes, femur including marrow, tissue masses, gallbladder, heart, ileum, jejunum, duodenum, kidneys, liver, lungs and bronchi, mammary gland, nerves (sciatic), ovaries, lymph nodes, pancreas, parathyroids, pituitary gland, prostrate, rectum, skin, spleen, seminal vesicles, skeletal muscle, testes with epididymides, stomach, thymus, thyroid gland including parathyroid, trachea, urinary bladder, uterus,

NOAEL(NOEL)

5.0 % (Males: 2641 mg/kg/d and Females: 3348 mg/kg/day)

LOAEL(LOEL)

Not determined

Actual dose received by dose level and sex Toxic response/effects by dose level

Males: 48, 491, 984 or 2641 mg/kg/day; Females: 58, 589, 1225 or 3348 mg/kg/day

In utero

There were no compound-related effects on fertility, gestation, parturition, lactation, pup survival through weaning or number of live and still-born pups. Slight decreases in body weight (4-5%) and slight increases in food consumption were noted in the F0 rats treated at dietary level of 5.0%. Two F0 female controls rats died during the *in utero* phase of the original study and one male and one female from the control and 5.0% group, respectively, died during the *in utero* phases of the high-dose study. There were no compound-related effects on pup survival.

In the F1 generation, a yellow tint was reported at all intake levels above 0.1%. At the 1.0% dietary level, group mean body weights at termination for both sexes were lower than the control animals, but the difference was only statistically significant for the females. In the high dose study (5.0% dietary level), group mean body weights were significantly lower in both sexes at termination. Food consumption was similar for control and treated animals at the 0.01, 1 or 2% dietary levels, but was slightly higher at the 5% level in the high-dose study, although not statistically significant. Hematological, clinical chemistry and urinalysis parameters did not differ significantly from the controls. Necropsies at one year did not reveal any treatment-related gross or microscopic changes.

At study termination, no treatment-related effects were reported on survival. No treatment-related changes were reported at gross necropsy. Histological evaluation revealed a variety of lesions, including neoplasms, present at similar incidences in control and treated animals. The authors considered the lesions to be spontaneous and not related to administration of the test material.

Appropriate statistical evaluations?

Remarks for results

Conclusion remarks

Yes, F-test, Anova

The decrease in mean body weight at the 5.0% treatment level was not considered toxicologically significant give the non-

nutritive character of FD & C Yellow No. 5.

The NOAEL of 5.0% providing an average daily intake of 2641 mg/kg/d and 3348 mg/kg/d for male and female rats,

respectively, under the conditions of this study.
Reliability code 1. Reliable without restriction.

**Data Qualities Reliabilities** 

•

Remarks for Data Reliability

Code 1. Comparable to guideline study.

References

Borzelleca J. and Hallagan J. (1988a) A chronic toxicity/carcinogenicity study of FD & C Yellow No. 5 (Tartazine) in rats. Fd Chem Toxic 26, 179-187.

CAS Numerical 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow 5; 90% purity; 10% intermediates or volatile

matter

Method/guideline Chronic Toxicity/Carcinogenicity Study

GLP Yes

**Year** 1988

Species/Strain Mice/Charles River CD-1

**Sex** Male and Female

Route of administration Oral-Diet

**Doses/concentration levels** 0, 0.5, 1.5, or 5.0%

**Exposure period** 104 weeks

Frequency of treatment Daily

Control Group Yes

Post exposure observation period

Remarks for test conditions

Groups of sixty male and sixty female mice each were administered 0, 0, 0.5, 1.5 or 5.0% FD & C Yellow No. 5 in the diet daily for 104 weeks. Animals were housed individually and fed the test diet ad libitum. Clinical observations were recorded twice daily, detailed physical examinations and palpations for masses were performed weekly. Body weights and food consumption were determined weekly for the first fourteen weeks, bi-weekly for weeks 16-26 and monthly from week 26 until the end of the study. The intake of the test substance was determined from body weight, food consumption and dietary concentration. Hematology tests, including hemoglobin, hematocrit, erythrocyte and total and differential leukocyte counts, and erythrocyte morphology, were conducted on randomly selected animals at months 3, 6, 12, 18 and 24 of the study. Necropsies were conducted on all animals dying prior to study termination, killed in a moribund condition or killed on schedule. Histological examinations were conducted on all animals from both control groups, the highest dose group (5.0%) and any animals with gross lesions or masses.

Tissues examined included adrenal glands, brain, cecum, colon, duodenum, epididymus or uterus, esophagus, eyes, femur including marrow, tissue masses, gallbladder, heart, ileum, jejunum, kidneys, liver, lungs and bronchi, mammary gland, nerves (sciatic), ovaries, lymph nodes, pancreas, parathyroids, pituitary gland, prostrate, rectum, skin, spleen, seminal vesicles, skeletal muscle, testes, stomach, thymus,

thyroid gland including parathyroid, trachea, and urinary

bladder.

**NOAEL(NOEL)** 5.0 % (8103 mg/kg/day)

LOAEL(LOEL) Not determined

Actual dose received by dose level and sex Toxic response/effects by dose level M: 714, 2173 or 8103; F: 870, 2662 or 9735 mg/kg/day

Physical observations included hair loss, lacrimation, nasal discharge, staining of hair in the anogenital region and soft stools. None of these observations was attributed to administration of the test substance. Discolored urine and feces was reported at all treatment levels within one week of the study initiation. Mean body weights of both sexes were slightly lower than controls at the 5.0% treatment group for a number of sampling intervals, and male mice at the 1.5% treatment group were lower than controls for a number of sampling intervals. These differences were significantly lower in some intervals. Mean food consumption was significantly increased in male mice at the 5.0% treatment level. No statistically significant differences were reported for any of the hematological parameters. Common neoplastic, inflammatory, and degenerative lesions were reported amongst treated and control animals but were not considered to be treatment related.

Appropriate statistical

evaluations?

Remarks for results

Yes, F-test, Anova

The decrease in mean body weight at the 5.0% treatment level was not considered toxicologically significant give the non-

nutritive character of FD & C Yellow No. 5.

**Conclusion remarks** The NOAEL of 5.0% providing an average daily intake of 8103

or 9753 mg/kg/d was established for male and female mice

under the conditions of this study.

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

Remarks for Data Reliability Code 1. Comparable to guideline study.

**References** Borzelleca J. and Hallagan J. (1988b) A chronic

toxicity/carcinogenicity study of FD & C Yellow No. 5 (Tartazine) in mice. Fd Chem Toxic 26, 189-194.

#### 4.4 DEVELOPMENTAL TOXICITY

**CAS Numerical** 1934-21-0

Substance Name C.I. Acid Yellow 23

Remarks for Substance FD&C Yellow No. 5; 92.7% purity

Method/guideline FDA Teratology Study

**Test Type** 

**GLP** Yes

**Year** 1990

Species/Strain Rat/Osborne-Mendel (FDA strain)

Sex Female

Route of administration Oral-Gavage

**Duration of test** 19 days

**Doses/concentration levels** 0, 60, 100, 200, 400, 600 or 1000 mg/kg bw/day

Exposure period 19 days

Frequency of treatment Daily

Control Group and treatment Yes

Remarks for test conditions Female Osborne-Mendel (FDA strain) rats (40-41 per group)

were administered FD & C Yellow No. 5 via gavage at dose levels of 0, 60, 100, 200, 400, 600 or 1000 mg/kg bw/day for the first 19 days of gestation. On day 19, the animals were examined for gross abnormalities followed by euthanization. Caesarean sections were performed. The uterus was examined for presence and position of resorption sites and fetuses, number of corpora lutea and implantation sites. All live fetuses were promptly weighed, sexed, and examined. Crown-rump lengths were measured. Fetuses were divided and assigned to

skeletal or soft tissue examination. Greater than 1000 mg/kg bw/day

NOAEL(NOEL) maternal

toxicity

LOAEL(LOEL) maternal

toxicity

NOAEL (NOEL)

developmental toxicity

LOAEL (LOEL)

developmental toxicity
Actual dose received by

dose level and sex Maternal data with dose

level

Not determined

Greater than 1000 mg/kg bw/day

Not determined

0, 60, 100, 200, 400, 600 or 1000 mg/kg bw/day

No unusual behavior or external findings were reported. One female at the 60 mg/kg bw/day dose level died on gestation day 13 due to gavage difficulties. The mean daily food consumption of rats administered the 1000 mg/kg bw/day dose level was

significantly greater than the controls. Initial body weight and maternal weight gain during gestation did not significantly differ between treated animals and controls. Pregnancy rate was

similar among all groups.

**Fetal data with dose level**No dose related findings were reported on fetal viability or fetal

development. The incidence of sternebral variations was similar

for all groups.

Appropriate statistical

evaluations?

Yes, ANOVA, Fisher's Exact Test, t-test.

Remarks for results The authors commented that the significant increase in food

consumption observed in the highest dose group without a corresponding effect on body weight indicated an effect on food

utilization.

**Conclusion remarks** The authors concluded that FD&C Yellow No. 5 was not

developmentally toxic or teratogenic under the conditions of the study. The NOAEL's for maternal and fetal toxicity were greater

than 1000 mg/kg bw/day.

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

Remarks for Data Reliability Code 1. Guideline study.

References Collins T., Black T.N., Brown L.H., and Bulhack P. (1990) Study

of the teratogenic potential of FD & C Yellow No. 5 when given

by gavage to rats. Fd. Chem. Toxic. Vol 28, pp 821-827.

## 4.5 REPRODUCTIVE TOXICITY

CAS Numerical 1934-21-0

Remarks for Substance FD&C Yellow 5: 90% purity: 10% intermediates or volatile

C.I. Acid Yellow 23

"."

matter

Method/guideline Lifetime Toxicity/Carcinogenicity study

**Test Type** 

**Substance Name** 

**GLP** Ambiguous

**Year** 1988

Species/Strain Rats/Charles River CD

Sex Male and Female

Route of administration Oral-Diet

**Duration of test** 114 weeks

**Doses/concentration levels** 0, 0.1, 1.0, or 2.0% (original study) 0, 5.0% (high dose study)

**Premating Exposure period** 

for males

Premating Exposure period

for females

Frequency of treatment Daily

Control Group and treatment

Yes.

2 months

2 months

Remarks for test conditions

In the *in utero* phase, groups of rats (60/sex/group) were administered 0, 0, 0.1, 1.0 or 2.0% FD & C Yellow No. 5 in the diet daily for approximately 2 months prior to mating. In the high-dose study, 60/sex/group received 0 or 5% FD&C Yellow 5 for approximately 2 months prior to mating. The 3 controls groups received the basal diet only. A maximum of 2 rats/sex/litter were randomly selected for the chronic phase of the study. There were 70/sex/group at the initiation of the chronic phase and these offspring were exposed to the same dietary levels as their parents.

Animals were housed individually and fed the test diet ad libitum. Clinical observations were recorded twice daily with at least 5 hours between observations. Detailed physical examinations and palpation for masses were performed weekly. Body weights and food consumption were determined weekly for the first fourteen weeks, bi-weekly for the next 12 weeks and every 4 weeks thereafter until the end of the study. The intake of the test substance was determined from body weight, food consumption and dietary concentration. Hematology tests, including hemoglobin, hematocrit, erythrocyte and total and differential leukocyte counts, and erythrocyte morphology, were conducted on ten randomly selected animals at months 3, 6, 12, 18 and 24 of the study. Necropsies were conducted on all animals dying prior to study termination, killed in a moribund condition or killed on schedule. Histological examinations were conducted on all animals from both control groups, the highest dose group (2.0 or 5.0%) from each study and also on 10 rats randomly selected from each group for an interim sacrifice at 12 months. Histology was also performed on any animal with gross lesions or masses.

Tissues examined included adrenal glands, aorta, blood smear, brain, cecum, colon, duodenum, epididymus or uterus, esophagus, eyes, femur including marrow, tissue masses, gallbladder, heart, ileum, jejunum, duodenum, kidneys, liver, lungs and bronchi, mammary gland, nerves (sciatic), ovaries, lymph nodes, pancreas, parathyroids, pituitary gland, prostrate, rectum, skin, spleen, seminal vesicles, skeletal muscle, testes with epididymides, stomach, thymus, thyroid gland including parathyroid, trachea, urinary bladder, uterus.

5.0 % (Males: 2641 mg/kg/d and Females: 3348 mg/kg/day)

NOAEL(NOEL)

#### LOAEL(LOEL)

Not determined

Actual dose received by dose level and sex Parental data and F1 as appropriate Males: 48, 491, 984 or 2641 mg/kg/day Females: 58, 589, 1225 or 3348 mg/kg/d

In utero

There were no compound-related effects on fertility, gestation, parturition, lactation, pup survival through weaning or number of live and still-born pups. Slight decreases in body weight (4-5%) and slight increases in food consumption were noted in the F0 rats treated at dietary level of 5.0%. Two F0 female controls rats died during the in utero phase of the original study and one male and one female from the control and 5.0% group, respectively, died during the in utero phases of the high-dose study. There were no compound-related effects on pup survival.

Offspring toxicity F1 and F2

In the F1 generation, a yellow tint was reported at all intake levels above 0.1%. At the 1.0% dietary level, group mean body weights at termination for both sexes were lower than the control animals, but the difference was only statistically significant for the females. In the high dose study (5.0% dietary level), group mean body weights were significantly lower in both sexes at termination. Food consumption was similar for control and treated animals at the 0.01, 1 or 2% dietary levels, but was slightly higher at the 5% level in the high-dose study, although not statistically significant. Hematological, clinical chemistry and urinalysis parameters did not differ significantly from the controls. Necropsies at one year did not reveal any treatment-related gross or microscopic changes.

At study termination, no treatment-related effects were reported on survival. No treatment-related changes were reported at gross necropsy. Histological evaluation revealed a variety of lesions, including neoplasms, present at similar incidences in control and treated animals. The authors considered the lesions to be spontaneous and not related to administration of the test material.

Appropriate statistical evaluations?
Remarks for results

Yes, F-test, Anova

The decrease in mean body weight at the 5.0% treatment level was not considered toxicologically significant give the non-nutritive character of FD & C Yellow No. 5.

**Conclusion remarks** 

**Data Qualities Reliabilities** Reliability code 1. Reliable without restriction.

Remarks for Data Reliability Code 1. Comparable to guideline study.

References

Borzelleca J. and Hallagan J. (1988a) A chronic toxicity/carcinogenicity study of FD & C Yellow No. 5 (Tartazine) in rats. Fd Chem Toxic 26, 179-187.