

Estimation of Freely-Dissolved Concentrations of Polychlorinated Biphenyls, 2,3,7,8-Substituted Congeners and Homologs of Polychlorinated Dibenzo-*p*-dioxins and Dibenzofurans in Water for Development of Total Maximum Daily Loadings for the Bluestone River Watershed, Virginia and West Virginia

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U.S. Department of the Interior U.S. Geological Survey

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By Robert W. Gale

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Contents

Abstract	1
Introduction	1
Experimental Method	2
SPMD Preparation	2
SPMD Performance Reference Compounds (PRCs)	2
Native PCB and PCDD/DF Fortification	3
Field Collection	3
Initial Sample Processing	3
Procedural Internal Standard Fortification and Dialytic Recovery of Contaminants	3
Sample Preparation	4
Secondary Reactive Column Cleanup	4
High Performance Size-Exclusion Chromatographic (HP-SEC) Fractionation	4
High Performance Porous Graphitic Carbon Chromatographic (HP-PGC) Fractionation	4
Low Pressure Basic Alumina Chromatographic (LP-BAC) Fractionation	4
Congener-Specific Polychlorinated Biphenyl Analysis	4
Polychlorinated Dibenzo- <i>p</i> -Dioxin and Dibenzofuran Analysis	5
Secondary Sample Preparation—Additional QC Samples	5
Confirmatory Congener-Specific Polychlorinated Biphenyl Analysis	5
Quality Assurance and Quality Control	6
Results and Discussion	6
SPMD Accumulation of Selected Congeners of Polychlorinated Biphenyls	6
SPMD Accumulation of 2,3,7,8-Substituted Congeners and Homologs of Polychlorinated Dibenzo- <i>p</i> -Dioxins and Dibenzofurans	
SPMD Release of Performance Reference Compounds (PRCs)	
Calculation of Estimated Concentrations of PCBs, PCDDs, and PCDFs in Water	
Concentrations of Selected Congeners of Polychlorinated Biphenyls in Water	11
Concentrations of 2,3,7,8-Substituted Congeners and Homologs of Polychlorinated	
Dibenzo-p-Dioxins and Dibenzofurans in Water	
Acknowledgments	13
References Cited	13
Tables	15

Tables

1.	SPMD samples deployed and collected from the Bluestone River watershed, Tazewell County, Virginia, and Mercer County, West Virginia (Waterbody VAS-N36/N37R)	7
2.	Amounts of total polychlorinated biphenyls (PCBs), total polychlorinated dibenzo- <i>p</i> -dio <i>x</i> ins (PCDDs) and total polychlorinated dibenzofurans (PCDFs) per semipermeable membrane device (SPMD) composite sample for the Bluestone River watershed sample set, fall 2005	8
3.	Percentages of performance reference compounds (PRCs) remaining in semipermeable membrane device (SPMD) composite samples from the Bluestone River watershed sample set, fall 2005	10
4.	Average a _{OPRC} intercept values of performance reference compounds (PRCs), maximum and minimum volumes of water sampled, and estimated uncertainties in sampling rates for semipermeable membrane device composite samples from the Bluestone River watershed sample set, fall 2005	12
5.	Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005	16
6.	Estimated total concentrations in water of polychlorinated biphenyls (PCBs), polychlorinated dibenzo- <i>p</i> -dio <i>x</i> ins (PCDDs) and polychlorinated dibenzofurans (PCDFs) from the Bluestone River watershed sample set, fall 2005	13
7.	Estimated concentrations of 2,3,7,8-substituted polychlorinated dibenzo- <i>p</i> -dio <i>x</i> ins (PCDDs) and dibenzofurans (PCDFs), and homologs of dioxins and furans in water from the Bluestone River watershed sample set, fall 2005	24

Conversion Factors and Datum

Multiply	Ву	To obtain
	Length	
micrometer (µm)	0.00003937	inch (in.)
centimeter (cm)	0.3937	inch (in.)
meter (m)	3.281	foot (ft)
	Area	
square meter (m ²)	10.76	square foot (ft ²)
square centimeter (cm ²)	0.1550	square inch (ft ²)
	Volume	
microliter (μL)	0.000002642	gallon (gal)
milliliter (mL)	0.0002642	gallon (gal)
liter (L)	0.2642	gallon (gal)
cubic meter (m ³)	264.2	gallon (gal)
	Flow rate	
liter per day (L·d ⁻¹)	0.2642	gallon per day (gal·d ⁻¹)
	Mass	
femtogram (fg)	3.527 x 10 ⁻¹⁷	ounce, avoirdupois (oz)
picogram (pg)	3.527 x 10 ⁻¹⁴	ounce, avoirdupois (oz)
nanogram (ng)	3.527 x 10 ⁻¹¹	ounce, avoirdupois (oz)
microgram (µg)	3.527 x 10 ⁻⁸	ounce, avoirdupois (oz)
milligram (mg)	3.527 x 10 ⁻⁵	ounce, avoirdupois (oz)
gram (g)	0.03527	ounce, avoirdupois (oz)
kilogram (kg)	2.205	pound avoirdupois (lb)
	Density	
gram per cubic centimeter (g/cm ³)	62.4220	pound per cubic foot (lb/ft ³)
	Concentration	
nanogram per gram (ng·g ⁻¹)	=	part per billion (ppb; 109)
milligram per liter (mg·L ⁻¹)	=	part per million (ppm; 106)
microgram per liter ($\mu g \cdot L^{\cdot 1}$)	=	part per billion (ppb; 109)
nanogram per liter (ng·L ⁻¹)	=	part per trillion (pptr; 10 ¹²)
picogram per liter (pg·L ⁻¹)	=	part per quadrillion (ppq; 1015)
nanogram per milliliter (ng·mL ⁻¹)	=	part per billion (ppb; 109)

Temperature in degrees Celsius (°C) may be converted to degrees Fahrenheit (°F) as follows:

°F=(1.8×°C)+32

Concentrations of chemical constituents in water are given in milligrams per liter ($mg\cdot L^{-1}$). micrograms per liter ($\mu g\cdot L^{-1}$), nanograms per liter ($ng\cdot L^{-1}$), picograms per liter ($pg\cdot L^{-1}$), or femtograms per liter ($fg\cdot L^{-1}$).

Estimation of Freely-Dissolved Concentrations of Polychlorinated Biphenyls, 2,3,7,8-Substituted Congeners and Homologs of Polychlorinated Dibenzo-*p*-dioxins and Dibenzofurans in Water for Development of Total Maximum Daily Loadings for the Bluestone River Watershed, Virginia and West Virginia

By Robert W. Gale

Abstract

The Commonwealth of Virginia Department of Environmental Quality, working closely with the State of West Virginia Department of Environmental Protection and the U.S. Environmental Protection Agency is undertaking a polychlorinated biphenyl source assessment study for the Bluestone River watershed. The study area extends from the Bluefield area of Virginia and West Virginia, targets the Bluestone River and tributaries suspected of contributing to polychlorinated biphenyl, polychlorinated dibenzo-*p*-dioxin and dibenzofuran contamination, and includes sites near confluences of Big Branch, Brush Fork, and Beaver Pond Creek.

The objectives of this study were to gather information about the concentrations, patterns, and distribution of these contaminants at specific study sites to expand current knowledge about polychlorinated biphenyl impacts and to identify potential new sources of contamination. Semipermeable membrane devices were used to integratively accumulate the dissolved fraction of the contaminants at each site. Performance reference compounds were added prior to deployment and used to determine site-specific sampling rates, enabling estimations of time-weighted average water concentrations during the deployed period.

Minimum estimated concentrations of polychlorinated biphenyl congeners in water were about 1 picogram per liter per congener, and total concentrations at study sites ranged from 130 to 18,000 picograms per liter. The lowest concentration was 130 picograms per liter, about threefold greater than total hypothetical concentrations from background levels in field blanks. Polychlorinated biphenyl concentrations in water fell into three groups of sites: low (130–350 picogram per liter); medium (640–3,500 picogram per liter; and high (11,000–18,000 picogram per liter). Concentrations at the high sites, Beacon Cave and Beaverpond Branch at the Resurgence, were about four- to sixfold higher than concentrations estimated for the medium group of sites.

Minimum estimated concentrations of polychlorinated dibenzo-*p*-dioxin and dibenzofuran congeners in water were about 0.2 to 1 femtograms per liter. Estimated total concentrations of 2,3,7,8-substituted congeners in water at study sites ranged from less than 1 to 22,000 femtograms per liter and less than 1 to 2,300 femtograms per liter for polychlorinated dibenzo-*p*-dioxin and dibenzofuran congeners, respectively. Total concentrations of 2,3,7,8-substituted congeners in water were comprised largely of octachlorodibenzo-*p*-dioxin and dibenzofuran, with less than 10 percent of the total contributed by concentrations of other congeners, mainly 2,3,7,8-hep-tachlorodibenzo-*p*-dioxin and dibenzofuran.

Of special interest for this study was 2,3,7,8-tetrachlorodibenzo-*p*-dioxin with a regulatory surface water-quality criterion of 1,200 femtograms per liter. Estimated concentrations in water ranged from 0.5 to 41 femtograms per liter. Concentrations in water were less than 5 femtograms per liter at all study sites, except the Bluefield Westside Sewage Treatment Plan, with an estimated concentration of 41 femtograms per liter.

Estimated total concentrations of homologs of polychlorinated dibenzo-*p*-dioxins and dibenzofurans in water at the study sites ranged from 3,200 to 36,000 femtograms per liter and 210–4,800 femtograms per liter, respectively. Again, homologs of polychlorinated dibenzo-*p*-dioxins and dibenzofurans in water were comprised largely of octachlorodibenzo*p*-dioxin and dibenzofuran.

Introduction

The Commonwealth of Virginia Department of Environmental Quality (VADEQ), working closely with the State of West Virginia Department of Environmental Protection (WVDEP) and the U.S. Environmental Protection Agency (USEPA) is undertaking a polychlorinated biphenyl (PCB) source assessment study for the Bluestone River watershed.

This watershed is located in the coalfields of Virginia and West Virginia, where historically, over 300 mining industry related facilities existed, many of which employed PCB-containing oils. Many of these operations have [since] closed. Some facilities were located in the remote valleys and ridges of the watershed, and other, smaller facilities, were home-based operations. The goals of the proposed source assessment are to ensure that no site is actively contributing PCBs to the watershed and to provide a baseline for the PCB Total Maximum Daily Load (TMDL) study required by the Clean Water Act. In the United States, commercially used technical mixtures of PCBs are commonly referred to as Aroclors® (Mosanto, St. Louis, Mo.) with a naming convention using a four-digit number denoting, first, the ring system (12 for biphenyl) and then the average percent chlorination (for example, 42 percent) (Frame and others, 1996); PCBs are referred to as Aroclors® in many State regulations. Currently, the calculated total PCB surface water-quality criteria (SWQC), based on the weight percent of each homolog group of the technical PCBs are 1,700 $pg\cdot L^{-1}$ and 47 $pg\cdot L^{-1}$ for the Commonwealth of Virginia and for West Virginia, respectively (Virginia State Water Control Board, 1997; Virginia Water Quality Standards, 2006; West Virginia Environmental Quality Board, 2000).

The Bluestone River watershed study is complicated by several factors (Lott and Newman, 2005). Most PCB misuse and improper disposal occurred decades ago and over a wide area. The karst geology of the region moves water from the surface through underground pathways near known or suspected PCB contaminated sites. Because of differential loadings of PCBs, differing exposure scenarios, and different target organisms, the transport mechanisms and bioaccumulation potential for PCBs are different in each watershed and even within a sub-watershed. The watershed crosses the Virginia– West Virginia State Line; tributaries from industrial areas in West Virginia enter Virginia and ultimately the Bluestone exits Virginia into West Virginia, thus requiring a cooperative assessment and TMDL approach to be developed by the Commonwealth of Virginia and the State of West Virginia.

Existing records of PCB related activities of State and Federal agencies and previous PCB sampling schemes, PCB environmental data, and reports of activity, cleanups, and site closures in the watershed have been compiled by the VADEQ. Potential PCB sources were identified by personal contacts and results of survey forms to gain information about historic PCB usage in the watershed; leading to locations of several contaminated sites.

The objectives of this study were to gather information about the concentrations, patterns, and distribution of selected congeners of PCBs, polychlorinated dibenzo-*p*-dioxins (PCDDs), and polychlorinated dibenzofurans (PCDFs) at specific study sites to expand current knowledge about PCB impacts and to identify potential new sources of contamination. Semipermeable membrane devices (SPMDs) were used to integratively accumulate the dissolved fraction of PCBs and PCDDs/DFs at each site. Performance reference compounds were added to the SPMDs prior to deployment and used to determine site-specific sampling rates of SPMDs, enabling estimations of time-weighted average (TWA) water concentrations of PCBs and PCDDs/DFs during the deployed period. This information will be used by the VADEQ, WVDEP, and USEPA to compare TWA concentrations with the SWQC for each analyte to determine those analytes potentially exceeding the proposed TMDLs being developed for the Bluestone River watershed.

Experimental Method

SPMD Preparation

Standard SPMDs (1 m x 2.5 cm x ~70–80 µm thickness, 1.00 mL 2,3-bis[[(E)-octadec-9-enoyl]oxy]propyl (E)-octadec-9-enoate (triolein), each) were deployed and composited for this work. Performance reference compound (PRC) loss rates equivalent to sampling rates of $2-4 \text{ L} \cdot \text{d}^{-1}$ were used to predict sampling rates for this study and the compositing required to attain these detection limits. The use of six standard SPMDs per site was calculated to provide sufficient sampling, once composited, to achieve the lowest quantification limit required; approximately tenfold less than targeted quantification SWQC for 2,3,7,8-TCDD of 1.2 pg-L⁻¹.

Semipermeable membrane devices were prepared for the VADEQ for deployment at 15 selected sites in the Virginia portion of the Bluestone River watershed, two of these sites were replicated (n=2) for a total of 17 sampling points. Three field blank SPMD composite samples, three fortified field blank SPMD composite samples, one fabrication blank SPMD composite sample, and one fortified field blank composite sample also were prepared. This required preparation and addition of PRCs to 150 standard SPMDs. Semipermeable membrane device composites also were prepared for the WVDEP for deployment at three selected sites in the West Virginia portion of the Bluestone River watershed along with three field blank samples. The number and configuration of the devices necessary to achieve the specified detection limits was the same as for VADEQ, six standard SPMDs per composite. No fortified field blank SPMD composite samples or other quality control (QC) samples were prepared for the WVDEP deployment. This required the additional preparation and addition of PRCs to 36 standard SPMDs.

SPMD Performance Reference Compounds (PRCs)

Suitable PRCs for the target analytes were selected for calibration (Huckins and others, 1999, 2002, 2006). The

previous deployment used only perdeuterated polycyclic aromatic hydrocarbons (PAH-*d*) (Lott and Newman, 2005), which were not considered completely acceptable as PRCs in this study because of their potential for photolytic degradation and dissimilarity to the analytes; rather, selected PCB congeners (PCB-014, PCB-029, and PCB-050) not found at any appreciable level in commercial Aroclors® were used here. However, the perdeuterated polycyclic aromatic hydrocarbons (acenaphthylene-*d*, acenaphthene-*d*, fluorene-*d*, phenanthrene-*d*, and pyrene-*d*) were included here as PRCs for comparative purposes.

A volume of 200 mL of purified triolein was required for this sample set, allowing enough residual triolein for anticipated SPMD losses and for preparing various QC samples. The 200 mL triolein was fortified with a 1,000 ng of each PCB and 200 μ g of each PAH-*d* by addition of isooctane solutions of the PRC-PCBs and the PRC-PAH-*d* to triolein and removal of the residual isooctane by rotary-evaporation. The PRC-PCBs were added at 5.0 ng·SPMD⁻¹ and the PRC-PAH-*d* were added at 1.0 μ g·SPMD⁻¹; or 30.0 ng and 6.00 μ g per composite, respectively.

Native PCB and PCDD/DF Fortification

Composites of six SPMDs, representing the one fortified fabrication blank and the three fortified field blanks were fortified with a mixture of native PCBs and PCDDs/DFs at levels near the quantification limits to determine the recovery efficiencies of the overall method. The native PCBs were added as a 1:1:1:1 mixture of Aroclors® 1242:1248:1254:1260. Current estimates of total PCB method background concentrations (as the sum of selected congeners) are about 7.5 ng·SPMD⁻¹, or 45 ng per composite. Therefore, 1,200 ng of Aroclor®1:1:1:1 was added, to allow quantification above potential background levels of PCBs. The PCDDs/DFs were added as a native standard solution of the tetra- through octa- chlorinated 2,3,7,8substituted congeners. There are no current estimates of PCDD/ DF method background concentrations for SPMDs; however, the 1997 Columbia River Study (McCarthy and Gale, 1999) reported levels for 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) of about 1-3 pg·SPMD⁻¹, which would be equivalent to about 6-20 pg in procedural blanks and in field blanks. Therefore, 240 pg of 2,3,7,8-TCDD were added; and because the native PCDD/ DF standard solution contains levels of congeners that increase with degree of chlorination, the amounts of individual PCDD and PCDF congeners added were: Cl₄-240 pg; Cl_{5.7}-1,200 pg; and Cl_o-2,400 pg, respectively.

Field Collection

The extent of the Bluestone River watershed to be sampled was in the Bluefield area of Virginia and West Virginia, targeting the Bluestone River and tributaries suspected of contributing to PCB and PCDD and PCDF contamination; including sites near confluences of Big Branch, Brush Fork, and Beaver Pond Creek. Fifteen deployment sites were selected by the VADEQ and three additional sites selected by the WVDEP. Site selection was based on considerations of appropriate site availability for SPMD deployment and specific stream conditions at each site. The design of the study required SPMD deployments at points where lateral mixing in the flow regime of each site was optimized, providing the most representative sampling of the stream at both high and low flow periods. The coordinates for each sampling point were documented by a geographic positioning system. Locations of these points were also described relative to existing landmarks, and all pertinent information concerning target analytes was compiled by the VADEQ as part of this study.

The deployment and collection of SPMDs for this project were accomplished by the VADEQ, WVDEP, and USGS personnel. These field personnel developed a sampling plan in accordance with USGS, VADEQ, and USEPA requirements (Standard Operating Procedures Manual, 2003; Water Quality Monitoring Consolidated Guidance, 2004). All deployed samples consisted of six standard SPMDs per site. Samples were transported to and from the site in precleaned sealed cans placed on ice. Field blanks were shipped, transported, and handled in the field in a manner consistent with the actual samples (for example, cans were opened and SPMDs were handled). All sampling equipment was cleaned prior to sampling and cleaned after each use. All samples were stored at -20 °C in sealed cans placed in coolers, and were handled under chain of custody. The samples were shipped overnight to the USGS Columbia Environmental Research Center, where they were logged, and stored at -20 °C.

Initial Sample Processing

Processing of SPMD samples consisted of inspection of the devices; all observations were noted and discussed with the principal investigator. Any punctures or leaking seals were resealed and documented. The exterior surface of the each SPMD was cleaned to remove any adhering particulates and the biofilm.

Procedural Internal Standard Fortification and Dialytic Recovery of Contaminants

The Procedural Internal Standard (PIS) solution was prepared by combining stock solutions to prepare 12.00 mL of a fortification solution containing PCB-155 (2,2',4,4',6,6'hexachlorobiphenyl) and PCB-204 (2,2',3,4,4',5,6,6'-octachlorobiphenyl) at 133 pg· μ L⁻¹ and ¹³C 2,3,7,8- substituted PCDDs/DFs at concentrations ranging from 3.33–6.67 pg· μ L⁻¹. Each SPMD in each composite sample (and in each field blank, fortified field blank, fabrication blank, and fortified fabrication blank) was opened, fortified with one-sixth (50 μ L) of the total specified amount of procedural internal standards,

and resealed as was performed for the native PCB and PCDD/ DF fortification in the previous section.

After fortification with PIS or PIS and native analytes and resealing, each SPMD was individually dialyzed according to established procedures. Hexane dialysis through the low density polyethylene membrane recovers small molecular weight target analytes (<1,000 Da) from the triolein reservoir and the membrane itself with greater than 99 percent efficiency. All dialysates from the individual SPMDs comprising one composited sample were combined, and concentrated in volume to $6.0 \text{ mL} (1.0 \text{ SPMD-equivalents} \cdot \text{mL}^{-1}, \text{ or SPMD-eq} \cdot \text{mL}^{-1}).$

Sample Preparation

Secondary Reactive Column Cleanup

Interfering anthropogenic and biogenic compounds in dialysates were removed by reaction in a layered sulfuric acidtreated silica gel/potassium hydroxide treated silica gel/silica gel column (SASG/KS/SG). The secondary reactive cleanup columns were calibrated for the collection of the appropriate target analyte fraction (c-PCBs/PCDDs/DFs) using a dichloromethane/hexane solvent scheme. The collected fractions were solvent-exchanged to dichloromethane and concentrated in volume to 1.00 mL.

High Performance Size-Exclusion Chromatographic (HP-SEC) Fractionation

HP-SEC was used to separate the analytes from residual higher molecular weight compounds in the reactive cleanup fractions. Higher molecular weight compounds are only slightly retained and elute prior to most target analytes; also, any elemental sulfur, a problematic interference during analysis, is more highly retained than the analytes, and elutes after the analyte fraction is collected. The HP-SEC was calibrated for the collection of the appropriate analyte fraction (c-PCBs/ PCDDs/DFs). The reactive cleanup fractions were then fractionated by HP-SEC at the rate of six SPMD-equivalents per HP-SEC cycle. The collected HP-SEC fractions were solventexchanged to isooctane and concentrated in volume to 1.0 mL.

High Performance Porous Graphitic Carbon Chromatographic (HP-PGC) Fractionation

Separation of the PCDDs and PCDFs from PCBs and other polyhalogenated hydrocarbons was necessary to remove potentially interfering chemicals that typically may be present at concentrations several orders of magnitude greater than those of the dioxins and furans, and to thereby achieve the required detection limits. The extended-*pi* systems of planar aromatic chemicals (PCDDs/DFs, and non-*ortho*-substituted PCBs) interact very strongly and very specifically with *pi*- system in the graphitic planes of the porous graphitic carbon, providing a highly selective and efficient fractionation mechanism. The HP-PGC was calibrated for the collection of the appropriate analytes: (fraction 1) weakly retained mono*ortho* – tetra-*ortho*-substituted PCBs (c-PCBs); (fraction 2) non-ortho-substituted PCBs (not collected); and (fraction 3) PCDDs/DFs. The collected c-PCB fractions were concentrated, solvent-exchanged to isooctane, and concentrated in volume to 6.0 SPMD-eq·mL⁻¹ for dual-column gas chromatography with electron-capture detection (GC-ECD) analysis. The collected PCDD/DF fractions were solvent exchanged to isooctane and concentrated in volume to about 0.5 mL for further fractionation by low pressure basic alumina chromatography.

Low Pressure Basic Alumina Chromatographic (LP-BAC) Fractionation

Residual PCBs and other polyhalogenated hydrocarbons that potentially interfere with the high-resolution mass-spectrometric analysis for PCDDs and PCDFs were minimized by chromatographic separation of the resulting HP-PGC PCDD/ DF fractions by LP-BAC. Most polyhalogenated hydrocarbon interferences are only slightly retained and elute prior to most analytes; also, any remaining biogenic materials may be more highly retained than the analytes, and elute after the analyte fraction is collected. The LP-BAC columns were calibrated for the collection of the appropriate analyte fraction (PCDDs/DFs) using a dichloromethane/cyclohexane solvent scheme. The collected LP-BAC fractions were concentrated in volume to about 0.5 mL.

Congener-Specific Polychlorinated Biphenyl Analysis

Instrumental internal standards (IIS) were added to each c-PCB fraction and the final volume adjusted to 6.0 SPMDeq·mL⁻¹. Individual PCB congeners were measured in sample fractions by dual-column GC-ECD. Analyses were performed using cool on-column capillary injection onto retention gaps connected to a 60 m x 0.25 mm x 0.25-µm film thickness 5 percent phenyl-, 95 percent methylsilicone capillary column, and a 60 m x 0.25 mm x 0.25-µm film thickness 50 percent phenyl-, 50 percent methylsilicone capillary column, respectively. Potential peaks for PCB congeners were matched and identified on one or both GC capillary columns with individual congener standards comprised of two IIS (PCB-030, -207), two PIS (PCB-155, -204), 128 PCB congeners found in Aroclors® at levels greater then 0.05 weight percent, and 16 PCB congeners found at less than this level. The capillary GC-ECD data were collected, archived in digital form, and processed using chromatography data-system software. Up to nine levels of calibration for each individual congener were used to quantify the targeted congeners. The calibration curve covers a concentration range of 0.01–0.03 to 100 ng·mL⁻¹ of each congener.

Instrumental internal standards were added to each PCDD/DF fraction and the final volume adjusted to about 25-50 µL. PCDFs and PCDDs were determined by gas chromatography with high-resolution mass spectrometric detection (GC-HRMS) by monitoring five sequential mass windows of selected ions for the Cl₄₋₈ PCDFs and PCDDs during the chromatographic separation. Analyses were performed using heated direct on-column injection onto a retention gap connected to a 30 m x 0.15 mm x 0.1-µm film thickness 5 percent phenyl-, 95 percent methylsilicone capillary column. The analytical column was directly interfaced to the MS. The HRMS system was tuned to a mass resolution of 10,000 and calibrated using perfluorotetradecahydrophenanthrene, and mass windows were established for five ion groups and monitored sequentially during the temperature program. Within each mass window, the two most abundant ions were measured for positive identification and quantification of each analyte; additional ions were monitored for any responses from potentially interfering chemicals (for example, Cl₅₀ polychlorinated diphenyl ethers, Cl_{5.7} polychlorinated terphenyls, Cl_{6.7} polychlorinated naphthalenes, and others), for Cl_{3 8} dibenzothiophenes, and Cl_{3 &} phenanthrenes and anthracenes. Isotope dilution calibration curves describing the response of each native congener (from 0.1 to 1,000 pg· μ L⁻¹) to that of a labeled procedural internal standard congener were used.

Window switching times were established using a window-defining PCDF/PCDD standard mixture; relative retention times were then established for polychlorinated dibenzothiophenes and other compounds of interest. Chromatographic columns were selected and temperature programmed on the basis that they must resolve 2,3,7,8-TCDD from 1,2,3,7/1,2,3,8-TCDD (and from 1,2,3,4-TCDD) by a resolution factor of at least 0.5. Column performance was verified by analyzing standards of individual components, and observing the chromatographic resolution of the TCDDs, hexachlorodibenzo-*p*-dioxins and dibenzofurans (HxCDDs and HxCDFs). Similarly, relative retention times for all other congeners of interest were evaluated with respect to labeled analogs.

Calibration curves describing the response of each native congener to that of a labeled procedural internal standard congener were used directly in the calculations, and their ranges of values were determined in the calibration procedure. Each calibration curve was matched to the range of analyte responses in the sample set.

Secondary Sample Preparation—Additional QC Samples

Because of the very low concentrations of PCBs that were anticipated in these samples, the cleanliness of all equipment was verified prior to any application of the extracts to a preparation step. The automated, multi-sample fractionation systems, HP-SEC and HP-PGC, were of special concern because they could be likely sources of sample carryover in preparation work. To minimize the potential for this to occur, the following checks were performed and the fractions analyzed prior to sample preparation; also, QC samples were added to the sample set to demonstrate the [low] levels of carryover throughout the preparation scheme.

The HP-SEC and the HP-PGC systems were demonstrated to be free of PCB background. All injector lines were thoroughly cleaned or replaced as necessary. Multiple solvent injection cycles were processed to equilibrate each system. Then, triplicate solvent blanks were collected and analyzed for PCBs. Each system was considered acceptably clean if levels of PCBs are less than 50 ng (total) with no single c-PCB contributing greater than 1 ng; otherwise, the systems were recleaned and further blanks evaluated.

Three separate procedural blanks were incorporated into the secondary portion of the sample preparation scheme to demonstrate the cleanliness of the post-dialysis part of the overall sample preparation plan. These procedural blanks were in addition to the procedural blanks previously prepared for the field and dialysis portions of this study. These procedural blanks were prepared and interspersed throughout the sample set, and then processed through the entire secondary scheme: secondary reactive cleanup, HP-SEC, HP-PGC, and LP-BAC for the PCDD/DF fractions. In addition, individual blanks were prepared for each discrete step of the scheme: a) secondary reactive cleanup, b) HP-SEC, c) HP-PGC, and d) LP-BAC; these blanks were used to assess and document the relative contributions of background PCBs (and PCDDs/DFs) from each step of the preparation scheme.

Confirmatory Congener-Specific Polychlorinated Biphenyl Analysis

Sample chromatography by dual-column GC-ECD exhibited high levels of interferences by coeluting peaks, which limited quantification of PCB-014, PCB-029, and PCB-050, the polychlorinated biphenyl congeners selected for use as PRCs in this study. All sample and field blank extracts were reanalyzed by gas chromatography with quadrupole mass spectrometric detection (GC-QMS) using both full-scan and selected-ion recording (SIR) for quantification of these congeners. Polychlorinated biphenyl congeners were determined by GC-QMS by monitoring selected ions for the Cl₁₋₁₀ PCBs during the chromatographic separation. Analyses were performed using cool on-column injection onto a retention gap connected to a 40 m x 0.18 mm x 0.1-µm film thickness 5 percent phenyl-, 95 percent methylsilicone capillary column. The analytical column was directly interfaced to the MS. The MS system was tuned to a mass resolution of 1,000 and calibrated using perfluorotributylamine, and the mass window was established for one ion group encompassing the two most abundant Cl₁₁₀ PCBs and monitored during the temperature program. Within

the mass window, the two most abundant ions were measured for positive identification and quantification of each analyte. Internal standard calibration curves describing the response of each congener to that of a procedural internal standard congener were used. Additional full-scan mass spectrometric analyses were conducted to confirm suspected interferences.

Quality Assurance and Quality Control

An operational QC system of checks was used to control and assess the quality of measurements. This system of checks was managed by a quality assurance (QA) system that ensures that defined standards of quality are being met (at stated levels of confidence). The accuracy and precision of environmental methods were assessed by utilizing appropriate checks for sample preparation and instrumental analysis. The appropriate QC sample types were selected on the basis of applicability to the objectives of this study and incorporated into the analysis plans.

Replicates, fortified field blanks, field blanks, fortified fabrication blanks, and fabrication blanks were prepared during the manufacture of SPMDs for deployment. The fortified field blanks and the fortified fabrication blanks were fortified with native analytes during fabrication. In addition to these QC samples, three procedural blanks were prepared to evaluate laboratory background levels of an analyte or background interferences with an analyte signal from the preparation method; and two fortification verification solutions were prepared to confirm analyte fortification levels.

Results and Discussion

Semipermeable membrane devices for the VADEQ and WVDEP parts of this study were prepared together and the samplers were shipped to VADEQ on September 13, 2005. Sampling began on September 15, 2005, for all VADEQ sites; October 1, 2005, for WVDEP Sites 1 and 2, Beacon Cave; and October 5, 2005, for WVDEP Site 3, Morris Spring. All deployed samples consisted of six standard SPMDs per site. Samples were transported to and from the site in precleaned sealed cans placed on ice. Field blanks were shipped, transported, and handled in the field in a manner consistent with the actual samples. All sampling equipment was cleaned prior to sampling and cleaned after each use. Most VADEQ samples were collected on November 1, 2005, with VADEQ Site-11, Cross-tributary to Beaverpond Branch collected on November 14, 2005. WVDEP samples for Sites 1 and 2, Beacon Cave, were collected on October 29, 2005; WVDEP Site-3, Morris Spring was collected on November 14, 2005. All sampling devices were retrieved without losses. All samples were stored at -20 °C in sealed cans placed in coolers, and were handled under chain of custody. Samplers were shipped overnight and received at the Columbia Environmental Research Center on November 15, 2005. The samples were inventoried, checked,

and stored at -20°C. The SPMD samples deployed and collected from the Bluestone River watershed, Tazewell County, Virginia, and Mercer County, West Virginia (Waterbody VAS-N36/N37R) are listed in table 1.

The SPMDs from VADEQ and WVDEP were treated as one set throughout sample preparation, analysis, and data reduction. Initial sample preparation entailed fortifying all samples with procedural internal standards, sample dialysis, compositing and concentration, along with preparation of quality control samples. Secondary sample preparation involved enrichment and purification of the SPMD dialysates through a series of cleanup and fractionation steps: secondary reactive cleanup, HP-SEC, HP-PGC, and an additional LP-BAC step for the PCDD/DF fractions from HP-PGC.

The instrumental analysis plan initially required only two analyses: congener-specific PCB analysis by dual-column GC-ECD, and 2,3,7,8-PCDD/DF/homolog analysis by GC-HRMS. However, additional perdeuterated PAH PRCs were incorporated into the study for comparison with the c-PCB PRCs, and required a separate analysis. Significant chromatographic interferences with the c-PCB PRC results required further GC-QMS analyses to obtain more reliable PRC information. In addition to unidentified chromatographic interferences, high levels of OCPs and PBDEs prompted semiquantitative analysis of the pesticides in these samples.

SPMD Accumulation of Selected Congeners of Polychlorinated Biphenyls

Amounts of total PCBs, PCDDs, and PCDFs accumulated by SPMDs at study sites are summarized in table 2. Amounts of PCB congeners in the composite samples were adjusted for recoveries of procedural internal standards and then corrected for background by subtraction of average field blank levels of PCBs. Low molecular weight congeners (PCB-001-PCB-035) are typically adjusted using PCB-029; however, in this study PCB-029 was used as a PRC and therefore its use as a PIS was precluded and these congeners were adjusted using PCB-155. A reporting limit of 0.03 ng per 6-SPMD composite was established for PCB congeners, with the exception of PCB-001, 002, and 003, wherein, a reporting limit of 0.3 ng was established. Congeners with coeluting chromatographic interferences were flagged, omitted from the final tabulations, and were not included in the summations for total amounts of PCBs, or in the calculation of PCB concentrations in water. Concentrations for chromatographically unresolved PCB congeners were calculated from the 5 percent phenyl and the 50 percent phenyl column information. Concentrations were averaged for PCB congeners resolved on both the 5 percent phenyl and the 50 percent phenyl columns. High levels of four organochlorine pesticides were determined to coelute, each with a specific PCB congener: PCB-063-heptachlor epoxide; PCB-090-cis-chlordane; PCB-093-trans-nonachlor; and PCB-196-methoxychlor. Therefore, these values were omitted. High levels of tetrabrominated diphenyl ether congener 047

Table 1.SPMD samples deployed and collected from the Bluestone River watershed, Tazewell County, Virginia, and Mercer County,
West Virginia (Waterbody VAS-N36/N37R).

Columbia Environmental		Station field			Deployn	nent	Collect	ion
Research Center sample name	State	identification number	Station name	Latitude, Iongitude	Date	Time	Date	Time
			WVDEP 2005 Study	_				
1. Site #1	WV	WVDEP-1	Beacon Cave # 1		10/1/2005	12:00	10/29/2005	11:00
2. Site #1 Field Blank	WV	WVDEP-FB-1	Beacon Cave # 1					
3. Site #2	WV	WVDEP-2	Beacon Cave # 2		10/1/2005	14:00	10/29/2005	12:15
4. Site #2 Field Blank	WV	WVDEP-FB-2	Beacon Cave # 2					
5. Site #3	WV	WVDEP-3	Morris Spring		10/5/2005	n/a	11/14/2005	13:15
6. Site #3 Field Blank	WV	WVDEP-FB-3	Morris Spring					
			VADEQ RARE 2005 Study					
1. Site #1	WV	9-LRR000.03	Laurel Fork Confluence	37° 18.89'N, 81° 19.60'W	9/15/2005	17:45	11/1/2005	16:00
2. Site #2	WV	9-ABB000.02, Replicate 1	Abbs Valley River	37° 18.41'N, 81° 18.88'W	9/15/2005	18:30	11/1/2005	15:30
3. Site #3	WV	9-ABB000.02, Replicate 2	Abbs Valley River	37° 18.41'N, 81° 18.88'W	9/15/2005	18:45	11/1/2005	15:40
4. Field Blank #3	WV	9-ABB000.02	Abbs Valley River					
5. Fortified Field Blank #3	WV	9-ABB000.02	Abbs Valley River					
5. Site #4	VA	9-BST022.43	Bluestone River below Yards	37° 18.17'N, 81° 17.84'W	9/15/2005	16:15	11/1/2005	15:00
7. Site #5	VA	9-BFK000.02	Brush Fork	37° 16.58'N, 81° 18.30'W	9/15/2005	14:45	11/1/2005	14:30
3. Site #6	WV	9-BFK003.14	Brush Fork, below Armory	37° 17.30'N, 81° 15.77'W	9/15/2005	14:51	11/1/2005	16:30
9. Site #7	WV	9-XRD000.43	Neal's Hollow	37° 16.11'N, 81° 16.45'W	9/15/2005	19:45	11/1/2005	17:00
10. Site #8	VA	VA0025054-001	Bluefield Westside STP	37° 15.64'N, 81° 16.92'W	9/15/2005	14:15	11/1/2005	14:10
1. Site #9	VA	9-BPB000.02	Beaverpond Creek	37° 15.31'N, 81° 16.55'W	9/15/2005	7:45	11/1/2005	13:40
12. Site #10	VA	9-WHI000.03	Whitley Creek	37° 15.14'N, 81° 16.25'W	9/15/2005	13:15	11/1/2005	12:05
13. Site #11	WV	9-XEF000.10	Cross-tributary to Beaverpond Branch	37° 14.84'N, 81° 14.41'W	9/15/2005	11:45	11/1/2005	14:23
14. Site #12	VA	9-BPB001.51, Replicate 1	Beaverpond Branch below Bluefield City Park	37° 14.65'N, 81° 15.46'W	9/15/2005	12:30	11/1/2005	11:30
15. Site #13	VA	9-BPB001.51, Replicate 2	Beaverpond Branch below Bluefield City Park	37° 14.65'N, 81° 15.46'W	9/15/2005	12:45	11/1/2005	11:40
16. Field Blank #2	VA	9-BPB001.51	Beaverpond Branch below Bluefield City Park					
17. Fortified Field Blank #2	VA	9-BPB001.51	Beaverpond Branch below Bluefield City Park					
18. Site #14	WV	9-BPB000.17	Beaverpond Branch at resur- gence	37° 14.48'N, 81° 14.07'W	9/15/2005	10:45	11/1/2005	10:30
19. Site #15	VA	9-BST025.23	Bluestone River above Bea- verpond Branch	37° 15.30'N, 81° 16.57'W	9/15/2005	8:05	11/1/2005	13:50
20. Site #16	VA	9-WVC000.05	Wright Valley Creek	37° 14.95'N, 81° 17.29'W	9/15/2005	10:00	11/1/2005	13:20
21. Site #17	VA	9-BST028.90	Bluestone River below water treatment plant	37° 14.38'N, 81° 16.66'W	9/15/2005	8:50	11/1/2005	9:55
22. Field Blank #1	VA	9-BST028.90	Bluestone River below water treatment plant					
23. Fortified Field Blank #1	VA	9-BST028.90	Bluestone River below water treatment plant					

Table 2. Amounts of total polychlorinated biphenyls (PCBs), total polychlorinated dibenzo-*p*-dioxins (PCDDs) and total polychlorinated dibenzofurans (PCDFs) per semipermeable membrane device (SPMD) composite sample for the Bluestone River watershed sample set, fall 2005.

[Total PCBs are reported in ng per composite, and total PCDDs and total PCDFs are reported in pg per composite]

	Field identification				
Sample name	number	Field sample label	Total PCBs	Total PCDDs	Total PCDFs
		VADEQ			
Site #1	9-LRR000.03	Laurel Fork Confluence	530	700	380
Site #2	9-ABB000.02, Replicate 1	Abbs Valley River	140	340	110
Site #3	9-ABB000.02, Replicate 2	Abbs Valley River	130	370	130
Site #4	9-BST022.43	Bluestone River below Yards	1,400	2,200	690
Site #5	9-BFK000.02	Brush Fork	2,500	670	300
Site #6	9-BFK003.14	Brush Fork, below Armory	190	580	260
Site #7	9-XRD000.43	Neal's Hollow	280	980	450
Site #8	VA0025054-001	Bluefield Westside STP	1,300	1,900	740
Site #9	9-BPB000.02	Beaverpond Creek	1,300	2,700	920
Site #10	9-WHI000.03	Whitley Creek	1,000	1,000	920
Site #11	9-XEF000.10	Cross-tributary to Beaverpond Branch	1,500	6,600	1,500
Site #12	9-BPB001.51, Replicate 1	Beaverpond Branch below Bluefield City Park	1,500	2,500	470
Site #13	9-BPB001.51, Replicate 2	Beaverpond Branch below Bluefield City Park	2,100	3,100	530
Site #14	9-BPB000.17	Beaverpond Branch at resurgence	15,000	1,800	550
Site #15	9-BST025.23	Bluestone River above Beaverpond Branch	780	1,000	370
Site #16	9-WVC000.05	Wright Valley Creek	290	750	400
Site #17	9-BST028.90	Bluestone River below water treatment plant	140	810	150
		WVDEP			
Site #1	WVDEP-1	Beacon Cave # 1	20,000	1,600	630
Site #2	WVDEP-2	Beacon Cave # 2	11,000	1,400	460
Site #3	WVDEP-3	Morris Spring	550	335	140

coeluted with PCB-180, which also was omitted. Unidentified chemicals were found to coelute with three additional PCB congeners (PCB-003, PCB-025, and PCB-101), again, these values were omitted.

Total amounts of PCBs in field samples ranged from 130 to 20,000 ng, much greater than field blank amounts (40–53 ng). Three groups of sites are apparent: low (130–530 ng); medium (550–2,500 ng); and high (11,000–20,000 ng). Amounts of PCBs determined in the three high sites: Beacon Cave #1 and #2, and Beaverpond Branch at resurgence #14, are about seven- to tenfold higher than the amounts determined for the medium group of sites.

Procedural internal standard recoveries for all samples ranged from 62 to 125 percent for PCB-155 and from 71 to 111 percent for PCB-204, excluding VADEQ Site-10, Whitley Creek, and WVDEP Site-3, Morris Spring. Whitley Creek recoveries ranged from 18 to 24 percent as a result of sample preparatory losses. Morris Spring recoveries were 175 percent and 140 percent, respectively; no explanation for this discrepancy was found.

As anticipated, background amounts of PCB congeners in quality-control samples incrementally increased for each additional sample preparation step in the sample processing scheme. The PIS solution used for recovery determination had amounts of only five congeners greater than the reporting limits. Four of these congeners (PCB-001, 006, 100, and 197) ranged from the reporting limits to less than threefold greater than these limits.

Values for all congeners in preparatory blanks for each cleanup or fractionation step were less than or very close to the reporting limits, with total amounts of PCBs ranging from 2.6 to 5.0 ng. Average total amounts of PCBs in procedural blanks were 11 ng, and average total amounts of PCBs in dialysis processing blanks were 21 ng. The contribution from the fabrication blank (materials for a six-SPMD composite plus all sample preparation) was 54 ng of total PCBs. The total amounts of PCBs in all six field blanks ranged from 40 to 53 ng (average 44 ng).

Analytical precision for the native PCB fortification solution (Aroclor® 1242:1248:1254:1260, 1,000 ng nominal amount fortified) averaged 6.6 percent, and the total PCB recovery for the solution was 870 ± 10 ng; the measured amounts of the congeners were used in the subsequent recovery calculations for fortified samples. Recoveries of individual congeners ranged from 53 to 118 percent and averaged 102 ± 21 percent. Recoveries of individual congeners in the fortified field blank samples ranged from 41 to 173 percent and averaged 100 ± 16 percent. It should be noted that the mixed Aroclor® standard used for fortification had very low concentrations of some PCB congeners, which biased the reported recovery ranges for analytes near the reporting limit; the majority of congeners, with amounts several-fold greater than the reporting limits, were quantitatively recovered.

Precision, calculated as relative difference between site replicates, generally was excellent. Relative differences varied more for those replicates with greater total amounts of PCBs (24 percent, VADEQ Site #12 and #13 samples) than for the replicates with lesser total amounts of PCBs (5 percent, VADEQ Site #2 and #3 samples).

SPMD Accumulation of 2,3,7,8-Substituted Congeners and Homologs of Polychlorinated Dibenzo-*p*-Dioxins and Dibenzofurans

Total amounts of PCDDs and of PCDFs for the Bluestone River watershed sample set are summarized in table 2. Amounts of 2,3,7,8-substituted congeners and homologs of PCDDs and PCDFs in the six-SPMD composite samples from the Bluestone River watershed (inherently adjusted for recoveries of PIS) were corrected for background by subtraction of average field blank levels of PCDDs and PCDFs.

The site groupings for PCDDs and PCDFs do not parallel those for PCBs; the three very high-level PCB sites corresponded to sites with only mid-range total amounts of PCDDs and PCDFs. Total amounts of PCDDs and PCDFs were found at very high levels at only one site: VADEQ Site-11, Crosstributary to Beaverpond Branch. At this site PCDDs were twofold greater than average, and PCDFs were about 50 percent greater than the next highest group of sites (VADEQ Site-9, Beaverpond Creek; VADEQ Site-10, Whitley Creek; and VADEQ Sites-12 and 13, Beaverpond Branch below Bluefield City Park). The remaining sites exhibited a gradual increase in amounts of PCDDs and PCDFs with no significant site groupings into low, medium, or high levels of PCDDs/DFs.

Of the 2,3,7,8-substituted congeners, octachlorodibenzofuran (OCDF) and 1,2,3,4,6,7,8-heptachlorodibenzofuran (HpCDF) contributed largely to total amounts of PCDFs; only OCDD contributed significantly to the total amount of PCDDs. Amounts of homologs (summed amounts of 2,3,7,8-substituted plus non-2,3,7,8-substituted congeners) were correlated differently with total amounts of PCDDs or PCDFs than were the 2,3,7,8-substituted congeners. Homolog amounts of TCDFs contributed most significantly to the total amount of PCDFs. All PCDF homolog groups were similarly correlated with total amounts of PCDFs. Amounts of TCDD and OCDD homologs contributed largely and about equally to the total amount of PCDDs, and as with the PCDFs, all PCDD homolog groups were similarly correlated with total amounts of PCDDs. The TCDF homologs were comprised of complex mixtures of TCDF congeners, the profiles of which did not change significantly among sites. The TCDD homolog series were found to be very different in composition compared with the TCDF homolog series. Of special note was that one specific non-2,3,7,8-substituted congener of PCDDs contributed to the majority of TCDD homologs and to total PCDDs. This congener has been tentatively identified as either 1,2,3,7-TCDD or 1,2,3,8-TCDD, members of an unresolved chromatographic pair on the 5 percent phenyl-, 95 percent methylsilicone capillary column.

Procedural internal standard recoveries for all samples ranged from 48 to 114 percent (average 70 ± 19 percent), excluding VADEQ Site-10, Whitley Creek because of sample preparatory losses (average recovery 14 percent). The analytical isotopic dilution method used here, inherently corrected for these preparatory losses. No inconsistencies in recoveries for the WVDEP Site-3 sample were found, indicating that only the post HP-PGC c-PCB fraction of the sample was compromised.

Preparatory blanks for the HP-PGC and LP-BAC steps were less than or very close to the reporting limits for all congeners, with total amounts of PCDDs and total PCDFs ranging from 2 to 4 pg. Average total amounts of PCDDs and PCDFs in procedural blanks ranged from 4 to 7 pg, and average total amounts of PCDDs and PCDFs in dialysis processing blanks ranged from 6 to 11 pg. The total amounts of PCDDs and PCDFs contributed from the fabrication blank were 24 pg and 14 pg, respectively. The average total amounts of PCDDs and PCDFs in VADEQ field blanks ranged from 14 to 17 pg and from 9 to 12 pg, and average total amounts in WVDEP field blanks ranged from 13 to 17 pg and from 4 to 5 pg, respectively. Average recoveries of native PCDD and PCDF congeners in the fortified field blank samples ranged from 69 to 104 percent.

Analytical precision for the native PCDD/DF fortification solution (fortified at 8,640 pg PCDDs and 12,240 pg PCDFs) averaged 3 percent. Recoveries of total amounts of PCDDs and PCDFs for the solution were $8,300 \pm 100$ pg and $12,000 \pm 100$ pg, respectively. Individual congener recoveries ranged from 93 to 100 percent (average 95 ± 3 percent) for PCDDs and from 86 to 105 percent (average 97 ± 5 percent) for PCDFs. These measured amounts of the congeners were used in the subsequent recovery calculations for fortified samples. Individual congener recoveries in the fortified fabrication blank sample ranged from 102 to 108 percent; however, recoveries of individual congeners in the fortified field blank samples ranged from 56 to 104 percent. Recoveries of all PCDD congeners in these samples were consistently 97–105 percent.

Precision of PCDD/DF accumulation between site replicates was generally excellent with differences of less than 20 percent for between total amounts of PCDDs and PCDFs in both replicate samples. As discussed for replication between sites for total amounts of PCBs, relative differences between replicate samples were found to be inversely related to the total amounts of PCDDs and PCDFs.

SPMD Release of Performance Reference Compounds (PRCs)

Initial amounts of PRCs in the six-SPMD composite samples before exposure were defined as the average of amounts determined in PRC-fortified field blank samples associated with VADEQ or with WVDEP sites (samplers not undergoing release of PRCs by environmental exposure). Overall average values from both sets of field blanks were not used because of small yet significant differences in averaged values between each set of sites. Final amounts of PRCs in SPMDs were defined as the amounts determined in field samples after exposure.

The c-PCB PRC amounts were adjusted for recoveries of PIS PCB-155. The PAH-*d* PRCs were quantified after dialysis and were not adjusted for recovery; however, values for the PAH-*d* PRCs were included in this study to enable comparisons of PCB and PAH-*d* release rates. Amounts of c-PCB and PAH-*d* PRCs in the associated blank samples were less than the reporting limits, therefore, neither the c-PCB nor the PAH-*d* PRC values were corrected for background. The dual-column GC-ECD values indicated positive biases for the c-PCB PRCs resulting from interfering compounds, therefore, subsequent SIR GC-QMS confirmations of the c-PCB PRCs were performed. Percentages of PRCs remaining in field samples at the end of exposures were determined by comparing the final amounts of PRCs measured in the samples to the initial amounts of PRC and are summarized in table 3.

Amounts of PRCs remaining in SPMDs after exposure ranged from <0.1 percent (not detectable) to much greater than 100 percent. Significant interferences in the quantification of some PRCs in some samples were indicated by recoveries of amounts in SPMDs greater than those initially fortified. Interferences in quantifying c-PCB PRCs are most evident in samples from Site-5 (Brush Fork), Site-14 (Beaverpond Branch at resurgence), and WVDEP-1 and 2 (Beacon Cave #1 and #2). These samples contained high total amounts of PCBs (2,500-20,000 ng), including amounts of PCB-014, PCB-029, and PCB-050 that, when accumulated in SPMDs, approached those amounts added initially as PRCs for monitoring release. No coeluting or interfering congeners for the confirmatory SIR GC-QMS analysis were noted. However, finding [of] high amounts of PCB congeners previously not considered to be present in the environment at significant levels, suggests that additional caution must be employed for the selection of candidate PRCs and for the selection of subsequent analytical methods for PRCs, as environmental levels of these compounds or analytical interferences with these compounds were much more prevalent than first assumed.

 Table 3.
 Percentages of performance reference compounds (PRCs) remaining in semipermeable membrane device (SPMD) composite samples from the Bluestone River watershed sample set, fall 2005.

	Performance Reference Compounds								
Sample name	Acenaphthylene-d	Acenaphthene-d	Fluorene- <i>d</i>	Phenanthrene- <i>d</i>	PCB-014	Pyrene-d	PCB-029	PCB-050	
			VADEQ						
Site #1	22	< 0.1	20	32	72	72	88	94	
Site #2	21	39	32	27	85	45	86	88	
Site #3	5.5	29	32	40	80	61	87	86	
Site #4	4.1	19	35	31	74	86	85	84	
Site #5	29	46	28	30	90	78	100	100	
Site #6	23	49	37	98	67	86	79	80	
Site #7	12	< 0.1	25	23	64	65	76	80	
Site #8	32	8.6	33	70	99	96	97	100	
Site #9	18	11	37	40	60	51	75	89	
Site #10	14	< 0.1	83	50	33	56	49	57	
Site #11	18	14	41	41	40	100	55	83	
Site #12	18	6.5	49	61	48	100	63	78	
Site #13	19	5.4	27	65	42	130	58	75	
Site #14	12	1.1	32	12	87	55	110	180	
Site #15	6.8	1.1	25	41	58	86	72	79	
Site #16	14	1.1	27	18	54	65	66	71	
Site #17	9.6	9.7	30	36	59	77	70	72	
			WVDEP						
Site #1	13	20	29	24	91	75	120	220	
Site #2	16	14	40	30	92	78	110	160	
Site #3	19	23	50	27	14	100	31	50	

Calculation of Estimated Concentrations of PCBs, PCDDs, and PCDFs in Water

Concentrations of PCBs, PCDDs, and PCDFs in water were estimated using the PRC-adjusted empirical uptake rate model incorporated into a spreadsheet calculator program (Huckins and others, 2006). Because of the availability of c-PCB and PAH-d PRC information, all PRC values were investigated for use in calculating estimates of the concentrations of target compounds in water. The final PRC amounts for PCB-014, PCB-029, and PCB-050 were determined by GC-QMS and were corrected for PCB-PIS recovery; final PRC amounts for acenaphthylene-d, acenaphthene-d, fluorene-d, phenanthrene-d, and pyrene-d were determined by gas chromatography with flame ionization detection of dialysates and were not corrected for recovery. Initial PRC amounts were determined from the average of SPMD field blank samples associated with VADEQ or with WVDEP sites using the methods just described. Overall average values from both sets of field blanks were not used because of differences between each set of sites. Final PRC amounts were determined in each SPMD field sample. PRC intercepts (a_{OPRC}) within each site and among sites were calculated using PRC-adjusted empirical uptake rate model and represent a measure of the sampling rate. The a_{OPRC} values were reviewed; data were omitted for PRCs at sites in which the final PRC amounts were not detectable (<0.01 µg for PAH-d) and in which the final PRC amounts were greater than the initial amounts (interferences or accumulation of PRC compounds from the site). Comparison of a_{0PRC} values among and between all sites was used to establish an acceptable range of PRC ratios (final amount/initial amount) for selecting a_{OPRC} values and for omitting outliers prior to averaging values at each site. Calculations provided similar values of a_{0PRC} over a broad range of releases, from PRC releases of only a few percent to those that were nearly quantitative. After data review, accepted values of a_{OPRC} were averaged for use and are summarized in table 4.

The total volumes of water sampled ranged from 40 to 2,300 L. At each site, sampling rates were maximum for $Cl_{2.3}$ PCBs, which ranged from 690 to 2,300 L of water sampled, and decreased about 17-fold to 40–140 L for $Cl_{7.8}$ PCDDs/ DFs. Across sites, sampling rates for the same analytes varied by only about three- to fourfold. This indicated that average water flows encountered by the deployed SPMDs were somewhat similar, possibly because the deployment structures had a greater effect on mediating flow to the SPMD surfaces more than differences in bulk streamflows outside the structures had in directly affecting flow at the SPMD surfaces.

The uncertainty factor for estimating concentrations of compounds in water from SPMD accumulation using PRC release data was calculated from the antilog of the standard deviation of the log-based a_{oPRC} values (table 4). Using these estimates of uncertainty, the uncertainties of most estimated concentrations in water were less than about ± twofold, with the exception of Sites-12 and 13 (Beaverpond Branch below

Bluefield City Park) which were about twofold and Site WVDEP-3 (Morris Spring) which was about threefold.

Concentrations of Selected Congeners of Polychlorinated Biphenyls in Water

Amounts of PCB congeners in SPMDs were adjusted for recoveries of PIS and were corrected for background amounts of c-PCBs in the associated field blank samples prior to calculation of the estimated concentrations in water. Specific PCB congeners with interferences from high levels of OCPs or PBDEs were omitted from the concentration calculations, as were congeners with unidentified co-eluting compounds. For PCB congeners less than the reporting limit, the reporting limit value was used in the calculation of the concentration of the congener in water. This resulted in minimum concentrations of congeners in water of about 1 pg·L⁻¹. Congener values less than the reporting limit were not included in total PCB concentrations.

Estimated concentrations of PCB congeners in water and estimated total concentrations of PCBs in water, calculated as the sum of all detectable concentrations of PCB congeners in water, are presented in table 5, at the end of this report. Estimated total concentrations of PCBs at study sites ranged from 130 to 18,000 pg·L⁻¹ (table 6) and were found to be linearly related to the relative amounts of PCBs accumulated by SPMDs.

The lowest estimated PCB concentration in water (130 $pg \cdot L^{-1}$) is almost threefold greater than total hypothetical concentrations of field blank amounts of PCBs (about 50 $pg \cdot L^{-1}$). As discussed previously for amounts of PCBs, three groups of sites are apparent for PCB concentrations in water: low (130–350 $pg \cdot L^{-1}$); medium (640–3,500 $pg \cdot L^{-1}$); and high (11,000–18,000 $pg \cdot L^{-1}$). Concentrations of PCBs in water estimated at the three high sites: Beacon Cave #1 and #2, and Beaverpond Branch at resurgence #14, are about four- to sixfold higher than concentrations estimated for the medium group of sites.

Concentrations of 2,3,7,8-Substituted Congeners and Homologs of Polychlorinated Dibenzo-*p*-Dioxins and Dibenzofurans in Water

Amounts of 2,3,7,8-substituted congeners and homologs of PCDDs and PCDFs in the six-SPMD composite samples from the Bluestone River watershed were inherently adjusted for recoveries of PIS by the isotopic dilution mass spectrometric technique used, and were corrected for background amounts of c-PCBs in the associated field blank samples prior to calculation of the estimated concentrations in water. Concentrations of PCDD and PCDF congeners with levels less than the reporting limit were derived from the reporting limit of the congeners in water. This resulted in minimum estimated concentrations of congeners in water of about 0.2

Table 4. Average a_{OPRC} intercept values of performance reference compounds (PRCs), maximum and minimum volumes of water sampled, and estimated uncertainties in sampling rates for semipermeable membrane device composite samples from the Bluestone River watershed sample set, fall 2005.

[Volumes of water for PCB are reported in L and are calculated for log K_{ow} 5.3 (maximum) and log K_{ow} 7.8 (minimum). Volumes of water for PCDDs and PCDFs are reported in L and are calculated for log K_{ow} 6.8 (maximum) and log K_{ow} 9.8 (minimum). Relative uncertainties are reported as multipliers/divisors of estimated concentrations]

		Polychlorina	ted biphenyls	-	dibenzo- <i>p</i> -dioxins nzofurans	Deletine
Sample name	a _{oprc}	Maximum	Minimum	Maximum	Minimum	Relative uncertainty
			VADEQ			
Site #1	-3.51	1,200	250	730	71	1.29
Site #2	-3.54	1,100	240	680	66	1.67
Site #3	-3.54	1,100	230	680	66	1.33
Site #4	-3.55	1,100	230	660	64	1.27
Site #5	-3.65	860	180	520	51	1.56
Site #6	-3.58	1,000	220	620	61	1.51
Site #7	-3.38	1,600	340	980	95	1.13
Site #8	-3.75	690	150	420	41	1.67
Site #9	-3.44	1,400	300	870	84	1.55
Site #10	-3.37	1,700	350	1,000	98	1.24
Site #11	-3.39	1,600	340	970	95	1.85
Site #12	-3.46	1,400	290	830	81	2.00
Site #13	-3.39	1,600	330	960	94	2.10
Site #14	-3.44	1,400	300	860	84	1.83
Site #15	-3.45	1,400	290	840	82	1.59
Site #16	-3.31	1,900	400	1,200	110	1.35
Site #17	-3.38	1,600	340	980	85	1.42
			WVDEP			
Site #1	-3.24	1,300	280	800	78	1.15
Site #2	-3.29	1,200	250	720	70	1.24
Site #3	-3.16	2,300	480	1,400	136	2.70

to about 1 fg·L⁻¹. Qualifiers for PCDD and PCDF congeners (or homologs) not meeting the quality-control criteria were propagated throughout the calculations and remain associated with their respective congener concentration in water. For analytes exceeding ± 15 percent tolerance in the ratio of the primary and secondary ions used for quantification, the lesser value is reported as the quantification limit and denoted LQ; for analytes not detected in the sample, the specified detection limit is reported and denoted ND. Analytes with recovery of the associated ¹³C-labelled surrogate exceeding the quality control limits set by the method were reported and denoted LR. Total concentrations of PCDDs and PCDFs were derived from the summation of detectable concentrations of each homolog group; therefore, congeners that were undetectable or unquantifiable were not included in concentrations in water of either the homologs or the total PCDDs or PCDFs. Estimated concentrations of 2,3,7,8-substituted congeners, homologs, and total PCDDs and PCDFs in water, calculated as the sum of all detectable concentrations of congeners in water (table 7, at the end of this report). Estimated concentrations of 2,3,7,8-substituted congeners in water at study sites ranged from <1 to 22,000 fg·L⁻¹ and <1 to 2,300 fg·L⁻¹ for PCDDs and PCDFs, respectively. Total concentrations of 2,3,7,8-substituted congeners in water were comprised largely of OCDD and OCDF, with less than 10 percent of the total contributed by concentrations of other, mainly 2,3,7,8-heptachloro-congeners.

Of special interest for this study was 2,3,7,8-TCDD, the only PCDD/DF congener with a regulatory SWQC value (1.2 pg·L⁻¹ or 1,200 fg·L⁻¹; Virginia Water Quality Standards, 2006). Estimated concentrations of 2,3,7,8-TCDD in water ranged from 0.5 to 41 fg·L⁻¹, as calculated from quantifiable amounts of 2,3,7,8-TCDD in SPMDs at the study sites. Concentrations of 2,3,7,8-TCDD in water were <5 fg·L⁻¹ at all study sites, except Site #8 (Bluefield Westside STP) with an estimated concentration of 2,3,7,8-TCDD in water of 41 fg·L⁻¹. Concentrations of 2,3,7,8-sustituted PCDD and PCDF congeners in water followed the same trends as previously described for amounts of 2,3,7,8-sustituted PCDD and PCDF congeners in SPMDs. **Table 6**. Estimated total concentrations in water of polychlorinated biphenyls (PCBs), polychlorinated dibenzo*p*-dioxins (PCDDs) and polychlorinated dibenzofurans (PCDFs) from the Bluestone River watershed sample set, fall 2005.

Field identification	Field cample label	PCBe	PCDDe	PCDFs					
number	•	1003	1 0003	TODIS					
VAUEU									
9-LRR000.03	Laurel Fork Confluence		4,900	980					
9-ABB000.02,	Abbs Valley River	180	3,200	210					
Replicate 1									
9-ABB000.02,	Abbs Valley River	180	3,700	270					
Replicate 2									
9-BST022.43	Bluestone River below Yards	1,800	10,000	1,500					
9-BFK000.02	Brush Fork	3,500	9,300	820					
9-BFK003.14	Brush Fork, below Armory	260	6,900	630					
9-XRD000.43	Neal's Hollow	260	7,200	720					
VA0025054-001	Bluefield Westside STP	2,300	11,000	2,300					
9-BPB000.02	Beaverpond Creek	1,300	13,000	1,900					
9-WHI000.03	Whitley Creek	910	7,100	1,600					
9-XEF000.10	Cross-tributary to Beaverpond Branch	1,300	36,000	4,800					
9-BPB001.51,	Beaverpond Branch below Bluefield City Park	1,500	15,000	1,000					
Replicate 1									
9-BPB001.51,	Beaverpond Branch below Bluefield City Park	1,700	16,000	940					
Replicate 2									
9-BPB000.17	Beaverpond Branch at resurgence	13,000	17,000	2,000					
9-BST025.23	Bluestone River above Beaverpond Branch	800	9,100	850					
9-WVC000.05	Wright Valley Creek	260	5,100	770					
9-BST028.90	Bluestone River below water treatment plant	130	6,500	230					
	WVDEP								
WVDEP-1	Beacon Cave #1	18,000	17,000	3,500					
WVDEP-2	Beacon Cave #2	11,000	17,000	3,000					
WVDEP-3	Morris Spring	350	1,600	160					
	number 9-LRR000.03 9-ABB000.02, Replicate 1 9-ABB000.02, Replicate 2 9-BST022.43 9-BFK000.02 9-BFK003.14 9-XRD000.43 VA0025054-001 9-BPB000.02 9-WHI000.03 9-XEF000.10 9-BPB001.51, Replicate 1 9-BPB001.51, Replicate 2 9-BPB001.71 9-BST025.23 9-WVC000.05 9-BST028.90	numberField sample labelVADEQ9-LRR000.03Laurel Fork Confluence9-ABB000.02,Abbs Valley RiverReplicate 19-ABB000.02,Abbs Valley RiverReplicate 29-BST022.43Bluestone River below Yards9-BFK000.02Brush Fork9-BFK003.14Brush Fork, below Armory9-XRD00.43Neal's HollowVA0025054-001Bluefield Westside STP9-BPB000.02Beaverpond Creek9-WH100.03Whitley Creek9-XEF000.10Cross-tributary to Beaverpond Branch9-BPB001.51,Beaverpond Branch below Bluefield City Park Replicate 19-BPB001.51,Beaverpond Branch below Bluefield City Park Replicate 29-BPB001.51,Beaverpond Branch below Bluefield City Park Replicate 19-BPB000.17Beaverpond Branch at resurgence9-BFT025.23Bluestone River above Beaverpond Branch9-WVC000.05Wright Valley Creek9-BST028.90Bluestone River above Beaverpond Branch9-WVDEP-1Beacon Cave #1WVDEP-2Beacon Cave #2	number Field sample label PCBs VADEQ VADEQ 9-LRR000.03 Laurel Fork Confluence 640 9-ABB000.02, Abbs Valley River 180 Replicate 1 - - 9-ABB000.02, Abbs Valley River 180 Replicate 2 - - 9-BST022.43 Bluestone River below Yards 1,800 9-BFK000.02 Brush Fork, below Armory 260 9-BFK000.02 Brush Fork, below Armory 260 9-XRD000.43 Neal's Hollow 260 VA0025054-001 Bluefield Westside STP 2,300 9-BPB00.02 Beaverpond Creek 1,300 9-WH1000.03 Whitley Creek 910 9-XEF000.10 Cross-tributary to Beaverpond Branch 1,500 Replicate 1 - - 9-BPB001.51, Beaverpond Branch below Bluefield City Park 1,700 Replicate 2 - - - 9-BPB000.17 Beaverpond Branch at resurgence 13,000 9-BST025.23 Bluestone Ri	number Field sample label PCBs PCDDs VADEQ VADEQ 9-LRR000.03 Laurel Fork Confluence 640 4,900 9-ABB000.02, Abbs Valley River 180 3,200 Replicate 1					

[Total PCBs are reported in pg $\cdot L^{-1}$, and total PCDDs and total PCDFs are reported in fg $\cdot L^{-1}$]

Estimated total concentrations of PCDDs and PCDFs in water at the study sites ranged from 3,200 to 36,000 fg·L⁻¹ and 210–4,800 fg·L⁻¹, respectively. Total PCDD and PCDF concentrations were calculated as the sum of homolog concentrations in water. Homologs of PCDDs and PCDFs in water were comprised largely of OCDD and OCDF.

Generally, homolog contributions decreased from octa- > hepta- > tetra- > penta- > hexa-chlorodibenzo-*p*dioxin and from octa- > tetra- > penta- > hepta- > hexachlorodibenzofuran. SPMDs accumulated significantly greater amounts of Cl_{4-5} homologs comprising non-2,3,7,8substituted congeners than those comprising only 2,3,7,8substituted congeners. The lesser chlorinated congeners contributed very much to the overall PCDD and PCDF concentrations in water.

Acknowledgments

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Tables

Table 5. Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005.

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

	RARE Study Field Samples—Fall 2005									
Sample name	Site #1 SPMDs	Site #2 SPMDs	Site #3 SPMDs	Site #4 SPMDs	Site #5 SPMDs	Site #6 SPMDs				
Field indentification	9-LRR000.03	9-ABB000.02, Rep 1	9-ABB000.02, Rep 2	9-BST022.43	9-BFK000.02	9-BFK003.14				
Field sample label	Laurel Fork Confluence	Abbs Valley River	Abbs Valley River	Bluestone River below Yards	Brush Fork	Brush Fork, below Armory				
PCB congener(s)	l									
001	3.9	9.5	2.6	5.0	15	7.5				
002	< 1	< 1	< 1	< 1	< 1	< 1				
004 005	4.9 0.06	4.2 0.07	3.0 0.1	19 0.2	250 0.3	4.6 0.1				
005	1.4	0.07	1.0	3.9	22	1.3				
007	< 1	< 1	< 1	2.5	3.4	< 1				
008	1.6	0.6	1.2	9.1	160	1.5				
)09)10	0.3 0.8	0.2 1.2	0.2 1.1	1.9 1.8	5.4 7.9	0.1 0.9				
012,013	5.3	1.2	1.0	20	4.3	1.8				
015	1.4	0.04	0.2	5.7	73	1.8				
016	4.0	0.5	0.7	15	71	2.0				
)17)18	7.5 12	0.8 1.2	0.8 1.4	26 41	170 170	3.5 6.6				
)19	3.9	0.4	0.4	19	94	2.4				
020	0.5	< 1	< 1	5.1	9.4	0.3				
)22	3.4	0.5	0.4	9.5	67	1.5				
024,027 026	1.9 1.6	< 1 0.2	< 1 0.1	6.3 6.5	41 39	0.7 0.9				
028	9.3	1.0	1.0	35	270	5.0				
031	7.1	4.0	3.5	49	200	4.7				
032	3.8	0.03	< 1	18	140	2.0				
033	8.5	4.4	3.6	31	27	4.3				
034 035	< 1 0.9	< 1 < 1	< 1 < 1	0.2 1.2	1.7 3.8	0.10 0.5				
)37,059	1.4	0.09	0.2	5.6	25	0.8				
040	2.8	0.2	0.1	9.4	35	1.2				
041	2.3	0.7	0.7	5.1	17	1.0				
042 044	3.7 17	0.3 1.2	0.2 1.3	12 46	46 150	1.5 5.8				
045	4.2	1.2	0.8	12	49	1.6				
046	1.1	< 1	< 1	7.4	19	0.7				
047	9.9	2.5	2.4	46	68	4.0				
048	2.4	0.1	0.2	6.7	35 110	1.1				
049 051	12 1.3	1.7 0.1	1.6 0.09	32 2.5	110	3.9 0.4				
052	37	18	17	94	160	14				
053	7.7	1.6	1.2	21	51	3.0				
054	6.6	5.3	4.9	15	4.7	3.0				
)56,060)64	2.8 7.6	0.9 1.0	0.8 0.9	15 20	83 82	0.7 2.9				
)66	7.3	1.5	1.7	33	110	3.8				
)67	1.1	0.2	0.4	1.2	5.2	0.1				
069	0.6	0.2	0.1	1.3	0.4	0.04				
070 071	14 4.4	2.7 0.3	2.4 0.6	36 13	85 53	5.5 1.5				
073	0.3	0.1	0.09	0.9	< 1	0.1				
)74	5.3 4.9	1.1	1.2 1.1	13 56	61	2.3				
)75		1.2	1.1	56	19	2.3 5.5 3.0				
)77)81,117	8.6 1.7	1.4 0.3	1.9 0.3	21 2.7	11 1.8	0.6				
082	5.4	0.6	1.3	11	6.7	1.8				
)83	2.6	0.6	0.6	6.2	3.0	0.9				
084	12	1.9	2.1	29	13	3.7				
)85)87	2.3	1.1 2.8	0.7 3.3	< 1 26	0.5 13	< 1 1.6				
)91	6.8	1.2	1.4	14	7.1	2.1				
92	7.4	2.0	2.3	19	6.5	2.1 2.2				
95	30	5.4	5.7	79	26	9.4				
)97	11	2.0	2.1	21	12	2.8 4.4				
)99 100	18 < 1	3.9 < 1	4.0 < 1	32 0.1	16 2.0	4.4 0.3				
103	1.3	2.1	1.8	3.4	0.6	0.8				
104	0.5	0.2	0.1	1.7	1.2	0.1				
05	10	2.5	2.7	17	9.4	2.4				
109 110	2.9 40	0.8 9.4	0.8 9.8	4.4 94	2.6 31	0.9 11				
110	40	0.1	0.1	0.9	0.4	0.06				
115 118	1.6 27	0.4 7.3	0.5 7.9	1.5 48	1.2 22	0.6 7.2				

Table 5. Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005.—Continued

	RARE Study Field Samples—Fall 2005								
Sample name	Site #1 SPMDs	Site #2 SPMDs	Site #3 SPMDs	Site #4 SPMDs	Site #5 SPMDs	Site #6 SPMDs			
Field indentification	9-LRR000.03	9-ABB000.02, Rep 1	9-ABB000.02, Rep 2	9-BST022.43	9-BFK000.02	9-BFK003.14			
Field sample label	Laurel Fork Confluence	Abbs Valley River	Abbs Valley River	Bluestone River below Yards	Brush Fork	Brush Fork, below Armory			
19	2.5	1.6	1.3	5.8	2.0	0.8			
22	2.3 0.9	0.4 0.1	0.3 0.2	12 1.7	1.5 0.6	0.5 0.2			
23 24	1.7	0.1	0.2	3.3	2.0	0.2			
28	7.1	2.0	2.2	13	3.4	2.3			
29	2.0	0.3	0.5	3.8	0.8	0.5			
30	3.1	0.8	0.8	5.9	1.6	1.0			
31	0.7	0.1	0.1	1.3	0.4	0.2			
32	< 1	< 1	< 1	25	6.4	4.9			
34	3.1	0.8	1.0	8.8	1.6	1.2			
35	4.5	1.2	1.3	9.4	2.1	1.3			
36	4.1	1.2	1.3	15	3.2	2.1			
37	8.8	1.1	0.5	3.5	1.0	0.5			
38	< 1	7.8	8.4	50	15	8.1			
41 44	4.8 1.4	1.5 0.6	1.5 0.6	13 3.8	3.1 0.8	1.8 0.5			
46	5.1	1.9	1.9	13	4.1	2.4			
47	1.4	0.6	0.6	2.2	0.9	0.8			
49	22	5.9	6.7	61	14	8.8			
51	5.5	1.3	1.9	21	5.4	2.8			
53	27	10.0	10	67	22	9.7			
54	0.6	< 1	0.09	1.1	0.6	0.2			
56	3.2	0.8	0.8	3.6	1.8	0.9			
57	1.1	0.2	0.2	2.5	0.5	0.4			
58	4.1	1.1	1.3	7.7	2.6	1.4			
63	7.6	2.5	2.7	18	5.6	3.3			
64	3.3	1.1	1.3	5.8	1.4	0.8			
65	< 1	< 1	< 1	< 1	< 1	< 1			
67 70	1.8	0.4	0.7	4.2	1.4	0.7			
70 71	4.3 1.8	1.3 0.8	1.3 1.0	11 4.8	4.2 1.3	2.8 1.3			
72	0.8	0.8	0.2	2.8	0.9	0.5			
73	< 1	< 1	< 1	0.1	< 1	< 1			
74	5.3	1.6	1.6	17	4.3	3.2			
75	1.1	0.2	0.1	0.8	0.2	0.1			
76	1.0	0.1	0.1	1.6	0.8	0.2			
77	3.0	0.8	1.0	9.4	2.5	1.9			
78	1.5	0.6	0.6	4.7	1.5	1.1			
79	2.2	0.6	0.5	8.5	2.2	1.3			
83	3.3	1.2	1.2	9.4	3.2	2.2			
85	0.5	0.1	0.2	2.2	0.5	0.4			
87	7.6	2.4	2.7	21	7.6	4.8			
89	< 1	< 1	0.1	0.3	< 1	< 1			
90 91	0.9	0.2	0.2	3.1	1.0 < 1	0.5			
93	0.2 1.9	< 1 0.5	< 1 0.2	0.5 4.5	< 1 1.0	0.1 0.3			
94	2.1	0.5	0.2	4.6	3.3	1.4			
95	0.8	0.5	0.5	2.5	1.0	0.6			
97	< 1	< 1	< 1	0.09	< 1	< 1			
99	3.1	0.8	0.8	6.0	2.9	1.8			
00	0.8	0.4	0.9	3.8	1.1	0.2			
01	1.0	0.8	0.8	0.9	2.2	1.5			
02	1.2	0.6	1.1	2.6	1.1	0.7			
03	2.4	0.6	0.9	5.0	2.6	1.6			
05	0.7	< 1	< 1	0.8	0.2	0.2			
06	5.2	1.6	1.6	3.7	3.1	1.7			
08	2.0	0.9	0.9	1.2	0.8	0.7			
09	4.4	1.3	1.7	1.8	1.1	0.9			
im of c-PCB water	640	180	180	1,800	3,500	260			
concentrations									

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

Table 5. Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005.—Continued

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

		RARE Study Field Samples—Fall 2005						
Sample name	Site #7 SPMDs	Site #8 SPMDs	Site #9 SPMDs	Site #10 SPMDs	Site #11 SPMDs	Site #12 SPMDs		
Field identification	9-XRD000.43	VA0025054-001	9-BPB000.02	9-WHI000.03	9-XEF000.10	9-BPB001.51, Rep 1		
Field sample label	Neal's Hollow	Bluefield Westside STP (g)	Beaverpond Creek	Whitley Creek	Cross-tributary to Beaverpond Branch	Beaverpond Branch below Bluefield City Park		
PCB congener(s)	Near 3 Honow	on (g/	Deaverpoila oreek	Wintey oreek	Deuverpond Branch			
	2.4			<i>(</i>)				
001 002	2.4 < 1	5.6 < 1	< 1 < 1	6.1 0.9	< 1 < 1	0.4 < 1		
004	1.6	210	12	11	5.1	8.6		
005 006	0.2 0.9	4.3 11	0.09 2.1	0.3 4.5	0.3 1.0	0.6 0.8		
007	0.3	5.2	0.3	0.4	0.9	0.3		
008	1.8	41	3.6	3.9	3.5	2.9		
009 010	0.3 0.3	10 26	0.6 6.3	0.5 1.7	1.6 1.0	0.6 11		
012,013	2.7	82	14	4.2	60	22		
015	1.1 1.3	30 37	4.7	4.1 6.5	6.2 6.0	6.3 19		
016 017	1.5	38	11 9.2	9.7	6.2	5.9		
018	3.6	74	16	16	12	13		
019 020	1.7 0.2	33 21	32 0.9	11 0.9	7.5 2.0	59 1.7		
022	1.6	19	5.7	4.6	6.4	7.3		
024,027	0.4 0.8	15 13	12 3.2	3.3 2.2	3.8 2.5	25 3.9		
026 028	4.9	45	5.2 16	14	20	24		
031	6.7	110	16	10	16	16		
032 033	0.8 6.3	16 94	26 15	7.9 10	4.7 44	54 19		
034	0.3	0.8	< 1	< 1	0.7	0.5		
035	0.3	3.5	1.8	1.0	1.6	3.3		
037,059 040	0.8 1.0	2.9 6.9	11 13	4.4 6.7	4.4 21	20 20		
041	0.9	9.2	5.4	3.0	9.7	7.4		
042 044	1.3 4.7	7.7 41	14 46	6.7 20	4.4 100	27 95		
045	2.1	11	24	12	14	48		
046	0.5	9.8	9.0	4.7	3.2	14		
047 048	3.4 1.0	100 8.8	28 4.4	14 2.5	60 2.3	34 7.0		
049	3.9	25	35	16	9.1	66		
051 052	0.3 20	4.1 110	5.8 100	4.3 46	2.2 150	7.8 150		
053	3.0	76	29	16	39	46		
054	5.5	25	28	12	100	38		
056,060 064	2.4 2.9	16 19	14 26	6.4 11	7.9 6.4	26 51		
066	4.4	24	38	11	10	42		
067 069	0.2 0.08	1.5 4.4	1.7 1.3	0.7 0.5	1.9 5.6	2.5 2.0		
070	5.9	47	20	15	13	20		
071 073	1.1 0.3	11 4.8	16 0.7	7.6 0.3	5.0 1.8	28 0.9		
073	2.3	14	10	4.8	6.0	16		
075	8.8	88	16	9.3	22 18 2.8	12		
077 081,117	3.6 0.5	0.2 4.5	14 1.6	14 1.2	18 2.8	12 13 2.6		
082	1.6	8.8	6.9	5.5	5.0	7.4		
083 084	0.9 3.2	6.0	3.9	3.3	6.5	4.3		
085	<1	25 15	18 < 1	16 0.03	11 < 1	15 0.1		
087	3.4	23	8.8	11	8.4	12 7.7		
091 092	1.8 2.8	8.0 14	9.1 11	8.7 9.5	5.2 11	7.7 7.6		
095	8.7	56	44	44	29	33		
097 099	2.9 4.1	21 20	13 17	10 16	11	11 15		
100	4.1 0.07	< 1	< 1	0.03	13 0.9	<1		
103	0.8	3.6	6.0	4.8	32	11		
104 105	0.2 3.0	1.8 26	2.5 8.6	1.0 6.2	0.7 8.5	1.7 8.1		
109	0.8	6.0	2.7	2.2	4.7	3.3		
110	11	90	50	40	34	40		

Table 5. Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005.—Continued

RARE Study Field Samples—Fall 2005 Sample name Site #7 SPMDs Site #8 SPMDs Site #9 SPMDs Site #10 SPMDs Site #11 SPMDs Site #12 SPMDs **Field identification** 9-XRD000.43 VA0025054-001 9-BPB000.02 9-WHI000.03 9-XEF000.10 9-BPB001.51, Rep 1 **Beaverpond Branch Bluefield Westside Cross-tributary to** below Bluefield Field sample label **Beaverpond Branch City Park Neal's Hollow** STP (g) **Beaverpond Creek** Whitley Creek 0.5 114 0.7 0.3 0.5 0.3 0.1 0.10 1.5 1.5 0.5 1.4 1.4 115 118 8.5 54 24 18 24 20 119 0.9 6.7 4.8 2.8 16 3.3 122 0.09 26 0.7 1.3 1.4 5.1 2.4 123 0.2 12 05 1.1 11 0.9 1.8 124 3.4 1.0 1.7 1.4 2.2 9.1 128 6.6 5.9 4.4 4.1 129 0.5 2.7 2.1 1.7 1.6 1.3 130 1.1 3.8 3.2 2.8 2.3 1.8 0.8 131 0.2 0.9 0.7 0.3 0.3 3.8 132 20 15 14 8.5 7.6 0.9 4.5 134 7.1 4.2 2.1 1.4 135 1.6 < 1 6.8 4.6 < 1 < 1 136 1.7 30 7.5 11 13 8.2 137 0.6 2.3 1.9 1.7 1.7 1.3 138 7.9 31 26 26 17 15 7.7 6.7 141 1.8 7.8 5.7 4.1 0.5 1.9 2.9 2.0 144 2.5 2.4 2.5 5.4 7.2 7.5 5.0 4.0 146 147 0.5 0.9 2.3 2.5 2.0 3.2 149 8.3 26 34 37 22 17 12 34 6.0 18 14 35 10 151 2.6 14 44 10 153 25 0.2 1.0 154 1.7 1.0 1.1 0.3 156 1.6 2.9 4.0 2.5 1.6 1.2 157 0.7 1.02.6 0.8 1.22.0158 1.3 47 43 4.0 34 2.6 10 10 56 163 33 11 6.6 2.0 0.8 3.2 164 3.8 3.4 1.7 0.1 165 < 1 < 1 < 1 < 1 < 1 2.5 2.8 3.6 2.3 167 1.4 2.1 170 1.8 4.5 7.5 11 4.4 3.2 171 0.7 1.6 3.7 3.8 1.6 1.0 2.0 172 0.5 0.7 2.0 1.1 0.7 0.3 173 < 1 < 1 0.1 0.08 < 1 2.2 7.2 5.0 174 11 15 8.0 175 0.09 1.0 0.6 0.8 0.7 1.6 176 0.1 0.5 1.1 1.4 1.2 0.4 2.7 1.8 177 1.3 3.1 6.2 7.9 4.0 2.5 3.4 178 0.73.9 3.8 2.7 179 0.9 4.2 2.8 6.5 6.2 3.0 5.5 4.9 183 2.3 8.8 9.0 0.3 0.2 1.5 1.2 0.6 185 1.9 187 3.7 5.8 15 18 10 7.2 0.2 0.08 189 < 1 0.1 0.4 < 1 1.0 2.2 190 0.3 0.8 2.0 1.1 0.6 0.07 0.4 0.2 1.3 191 0.2 < 1 1.2 2.7 193 0.2 1.8 3.5 194 0.7 3.0 2.9 5.7 4.6 1.6 195 0.2 < 1 2.1 2.2 2.4 1.5 197 < 1 0.1 0.3 0.1 < 1 199 1.3 1.7 4.8 2.6 4.6 3.4 0.9 200 0.6 0.9 0.2 1.0 5.6 201 7.6 2.5 0.4 1.3 1.0 6.7 202 0.4 < 1 1.8 2.2 4.3 1.4 203 0.8 1.8 3.4 3.7 2.6 1.7 0.9 205 0.3 0.3 1.5 0.8 1.6 206 19 2.8 3.0 11 2.7 42 0.9 1.5 208 1.0 0.4 0.5 1.4 209 0.3 0.3 1.7 7.3 4.5 2.5 Sum of c-PCB water 260 2,300 1,300 910 1,300 1,500

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

concentrations

Table 5. Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005.—Continued

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

	RARE Study Field Samples—Fall 2005								
Sample name	Site #13 SPMDs	Site #14 SPMDs	Site #15 SPMDs	Site #16 SPMDs	Site #17 SPMDs				
Field identification	9-BPB001.51, Rep 2	9-BPB000.17	9-BST025.23	9-WVC000.05	9-BST028.90				
	Beaverpond Branch		Bluestone River						
	below Bluefield	Beaverpond Branch	above	Wright Valley	Bluestone River below				
Field sample label	City Park	at resurgence (h)	Beaverpond Branch	Creek	water treatment plant				
PCB congener(s)									
001 002	0.8	< 1	0.9	< 1	< 1				
002 004	< 1 10	< 1 41	< 1 9.3	< 1 1.9	< 1 0.5				
005 006	0.2 1.0	1.2 3.1	0.10 1.0	0.08 0.3	0.08 0.6				
007	0.3	0.2	0.09	< 1	< 1				
008 009	3.0 0.7	8.9 0.7	3.9 0.4	0.9 0.4	0.2 0.08				
010	12	150	0.7	0.4	0.08				
012,013 015	24 7.6	6.6 35	2.3 5.9	1.4 0.5	1.4 0.2				
016	23	340	4.3	0.8	0.6				
017 018	6.6 15	23 58	24 10	1.6 2.1	0.8 1.2				
019	71	820	9.3	1.1	0.5				
020 022	2.1 9.0	16 73	0.4 2.4	0.06 0.6	0.05 0.4				
024,027	31	360	4.0	0.3	0.4				
026 028	4.6 28	24 120	1.4 11	0.3 2.0	0.1 1.3				
031	21	110	11	1.6	2.7				
032 033	68 21	1,200 9.9	10 4.7	0.7 1.5	0.2 2.4				
034	0.6	6.5	0.05	< 1	< 1				
035 037,059	4.4 24	63 250	1.4 1.3	0.06 0.4	< 1 0.2				
040	28	280	2.3	0.6	0.3				
041 042	9.0 33	49 380	1.4 5.5	0.5 0.7	0.4 0.4				
044	110	1,300	20	2.8	1.6				
045 046	56 16	570 170	3.3 2.8	1.0 0.3	0.5 < 1				
047	43	330	30	3.8	3.6				
048 049	8.4 85	77 940	1.8 23	0.3 2.2	0.2 1.4				
051	9.7	120	8.4	0.6	0.3				
052 053	190 56	1,400 500	38 19	7.8 1.8	5.9 1.6				
054	47	25	4.8	2.2	2.4				
056,060 064	31 65	320 700	4.0 6.9	0.3 1.6	0.6 1.0				
066	49	470	12	1.8	1.6				
067 069	2.8 2.4	9.5 4.7	0.7 0.3	< 1 0.1	< 1 0.08				
070 071	23 35	120 430	18 7.9	3.0 0.7	2.3 0.2				
073	1.1	4.7	0.6	0.07	0.2				
074 075	19 14	170 30	5.9 17	1.1 2.4	1.0 15				
077	13	50 52 8.4	14	3.7	2.0				
081,117 082	2.8 8.1	8.4 43	2.4 5.1	0.5 1.3	0.4 0.8				
083	4.5	18	3.2	0.7	0.5				
084 085	17 0.8	120 20	12 < 1	2.6 < 1	1.3 < 1				
087	11	89	15	3.0	1.8				
091 092	8.3 10.0	53 38	7.8 9.3	1.7 2.3	0.9 1.3				
095	38	210	30	8.1	3.6				
097 099	12 16	74 92	12 19	2.3 3.2	1.4 2.2				
100	< 1	16	0.5	0.3	0.2				
103 104	13 1.9	15 4.5	2.5 0.5	1.4 0.1	1.5 0.02				
105	8.7	39	11	1.9	1.4				
109 110	3.8 43	8.1 200	2.9 43	0.6 8.8	0.6 5.0				
114	0.4	2.6 3.2	0.5	0.09	0.09				
115	1.9	3.2	1.2	0.08	0.09				

Table 5. Estimated concentrations of selected congeners of polychlorinated biphenyls (PCBs) in water from the Bluestone River watershed sample set, fall 2005.—Continued

	[KAKE 5	tudy Field Samples—Fal	1 2005	
Sample name	Site #13 SPMDs	Site #14 SPMDs	Site #15 SPMDs	Site #16 SPMDs	Site #17 SPMDs
Field identification	9-BPB001.51, Rep 2	9-BPB000.17	9-BST025.23	9-WVC000.05	9-BST028.90
	Beaverpond Branch		Bluestone River		
Field sample label	below Bluefield City Park	Beaverpond Branch at resurgence (h)	above Beaverpond Branch	Wright Valley Creek	Bluestone River below water treatment plant
10		0.6	22	5.0	1.2
18 19	22 8.7	86 21	33 2.5	5.8 0.7	4.3 0.8
22	2.1	3.6	0.8	0.2	0.5
23	1.3	2.6	0.5	0.2	0.1
24	1.5	6.0	2.0	0.4	0.2
28	4.3	4.9	6.1	1.7	1.0
29	1.3	1.3	1.9	0.5	0.2
30	1.9	2.0	2.6	1.0	0.4
31 32	0.4 7.7	0.6 10	0.6 10	0.2 3.5	0.08 2.3
34	1.7	2.5	3.1	1.2	0.6
35	0.4	9.1	4.3	2.8	0.5
36	6.4	4.8	4.7	2.8	1.1
37	1.5	1.8	2.0	0.6	0.2
38	15	17	24	9.6	4.2
41	4.3	4.4	5.4	3.1	0.9
44	1.8	1.9	1.3	0.8	0.3
46	4.1 2.3	4.6	5.5 1.7	3.6	1.3
47 49	2.5 17	3.1 21	23	0.7 16	0.4 4.4
51	6.4	6.7	7.7	7.0	1.7
53	20	22	28	17	5.7
54	1.0	1.1	0.5	0.2	0.1
56	2.3	1.9	3.6	0.8	0.4
57	2.0	0.6	1.3	0.2	0.1
58	2.6	2.9	4.2	1.4	0.6
63	5.6	5.9	8.8	5.2	1.8
64 65	2.0 < 1	2.2	2.8	1.6	0.5
65 67	< 1 2.5	< 1 1.6	< 1 2.1	< 1 0.6	< 1 0.3
70	3.3	1.0	5.8	5.4	1.1
71	1.0	0.8	2.4	2.3	0.5
72	0.8	0.4	0.7	1.6	0.2
73	< 1	< 1	0.07	0.1	< 1
74	4.9	2.5	7.1	8.9	1.6
75	0.7	1.5	0.3	0.5	< 1
76	0.3	0.1	0.6	1.0	0.1
77 78	2.6 1.7	1.5 1.1	4.5 2.4	5.4 3.3	1.1 0.7
79	2.6	1.6	3.6	5.1	0.9
83	3.8	3.6	5.0	5.0	1.3
85	0.6	0.3	0.9	1.3	0.1
87	6.8	3.5	11	14	2.8
89	< 1	< 1	0.1	0.1	< 1
90	0.7	0.4	1.4	1.6	0.3
91	0.4	1.3	0.4	0.3	< 1
93 04	1.7	1.0	1.5	1.7	0.4
94 95	1.6 1.7	0.5 0.6	2.6 1.1	2.8 1.4	0.7 0.4
93 97	<1	< 1	0.09	0.1	< 1
99	2.4	1.3	3.2	3.8	0.9
00	0.5	0.3	0.2	0.4	0.06
01	3.0	7.3	1.1	1.0	0.8
02	1.7	1.7	1.2	1.6	0.4
03	1.6	0.9	2.6	3.1	0.6
05	1.1	1.5	0.6	0.7	0.1
06	2.9	1.9	2.0	1.7	0.8
08 09	0.9 3.1	0.7 1.0	0.7 0.3	0.5 0.3	0.2 < 1
~~	5.1	1.0	0.5	0.5	1
um of a PCB water	1,700	13,000	800	260	130
um of c-PCB water	1,700	15,000	000	200	150

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

concentrations

Table 5. Estimated concentrations of selected congeners of polychlorinatedbiphenyls (PCBs) in water from the Bluestone River watershed sample set, fall2005.—Continued

[Concentrations of PCBs are reported in pg·L⁻¹ water. PCB congener numbering is follows Frame and others, 1996]

	VVVDEP	Study Field Samples—	Fall 2005
Sample name	Site #1 SPMDs	Site #2 SPMDs	Site #3 SPMDs (*)
Field identification	WVDEP-1	WVDEP-2	WVDEP-3
Field sample label	Beacon Cave #1 (h)	Beacon Cave #2 (h)	Morris Spring
PCB congener(s)		· · · · · · · · · · · · · · · · · · ·	
01	0.4	0.8	< 1
02 04	< 1 72	< 1 47	< 1 0.5
05	0.8	0.2	0.5
06	5.3	2.9	0.4
07	0.7 13	0.3 7.0	0.6
08 09	1.4	0.7	< 1 0.4
10	240	160	0.2
12,013	6.4	4.8	5.5
15 16	44 490	29 330	1.6 1.1
17	490	22	<1
18	80	49	0.6
19	1,200	790	1.3
20 22	29 110	15 64	0.1 0.3
24,027	530	320	0.5
26	36	20	0.04
28 31	160	98 55	1.3
32	110 2,000	1,200	24 0.8
33	12	8.2	12
34	12	6.3	0.1
35 37,059	110 340	66 200	2.6 1.2
40	390	230	1.2
41	75	42	2.1
42	540	310	1.0
44 45	1,600 770	990 470	1.5 4.0
46	230	140	2.2
47	440	240	8.2
48 49	120 1,300	63 740	< 1 1.6
51	1,500	110	0.8
52	1,600	1,000	60
53	690	430	9.2
54 56,060	31 440	27 260	21 0.8
64	930	550	1.4
56	630	360	1.4
67 69	18 6.3	9.9 3.7	0.4 0.2
70	120	72	0.2
71	620	360	1.7
73 74	5.3 210	3.5 130	0.5 0.2
75	39	22	1.5
77	53	32	4.1
81,117 82	8.4	5.8	0.3
82 83	53 22	33 14	1.7 1.8
84	150	92	7.4
85	26	19	< 1
87 91	110 64	65 40	1.9 3.8
92	64 41	40 27	3.8 2.0
95	260	160	18
97	90	55	2.0
99 00	99 17	62 11	5.3 0.04
00	13	9.8	4.4
04	1.1	1.8	0.6
05	43	28 5.7	1.1
09 10	8.6 230	5.7 140	1.6 13
14	2.0	1.4	0.1
15	7.8	2.2	0.6

Table 5.Estimated concentrations of selected congeners of polychlorinatedbiphenyls (PCBs) in water from the Bluestone River watershed sample set, fall2005.—Continued

	WVDEP	Study Field Samples—	Fall 2005
Sample name	Site #1 SPMDs	Site #2 SPMDs	Site #3 SPMDs (*)
Field identification	WVDEP-1	WVDEP-2	WVDEP-3
Field sample label	Beacon Cave #1 (h)	Beacon Cave #2 (h)	Morris Spring
18	83	57	3.0
.19	22	16	5.5
22	4.1	2.6	0.4
23 24	3.4 6.2	2.2 3.9	0.4 0.4
28	3.3	2.6	2.5
29	1.1	0.7	0.7
30	1.6	1.1	1.4
31 32	0.5 8.9	0.3 7.4	0.3 6.1
34	2.1	1.6	1.4
35	6.0	4.8	<1
36	5.0	4.2	5.6
37	1.4	0.9	0.7
38	12 3.3	8.9	5.9
41 44	5.5 1.0	2.6 0.8	1.7 1.0
46	3.2	2.6	2.0
47	2.4	1.4	1.0
49	16	11	13
51	5.9 18	4.1	4.0
53 54	1.3	13 0.9	6.8 0.2
56	1.2	1.2	0.7
.57	0.3	0.4	0.7
58	2.2	1.7	1.1
63 64	4.3 1.7	3.2 1.2	2.9 1.5
65	< 1	<1.2	< 1
.67	0.9	0.8	1.0
70	1.1	0.8	2.3
71	0.5	0.4	0.9
72 73	0.2 < 1	0.2 < 1	0.6 0.04
.74	1.8	1.3	3.4
75	0.5	0.6	0.3
76	0.1	0.09	0.3
77 78	1.1 0.8	0.7 0.6	1.9 1.1
78 79	1.4	0.0	1.1
83	2.5	1.2	2.4
85	0.2	0.1	0.4
87	2.8	2.1	4.3
89 90	< 1 0.2	< 1 0.2	< 1 0.5
91	< 1	< 1	0.5
93	0.7	0.2	0.5
94	0.5	0.3	0.9
95 07	0.5	0.3	0.9
97 99	< 1 0.7	< 1 0.6	< 1 1.2
00	0.4	0.0	0.2
01	3.3	2.9	0.7
02	0.7	0.4	0.4
03	0.5	0.3	0.8
05 06	1.6 0.3	1.4 0.3	0.4 0.8
08	< 1	< 1	0.8
09	< 1	< 1	< 1
um of c-PCB water	18,000	11,000	350

[Concentrations of PCBs are reported in $pg \cdot L^{-1}$ water. PCB congener numbering is follows Frame and others, 1996]

Estimated concentrations of 2,3,7,8-substituted polychlorinated dibenzo-p-dioxins (PCDDs) and dibenzofurans (PCDFs), and homologs of dioxins and furans in water from the Bluestone River watershed sample set, fall 2005. Table 7.

[Concentrations of PCDDs and PCDFs are reported in fg·L⁻¹ water; ND, not detected at specified detection limit; LR, denotes recovery of 13C-labelled surrogate below 25% but above 10%: relative stan-dard deviation estimated as about 25%; LQ, ion ratio outside of ±15% tolerances: analyte is either less than the method quantification limit or is subject to interference]

			RA	RARE Study Field Sample—Fall 2005	ample-	-Fall 2005					
Sample name	Site #1 SPMDs	Site #2 SPMDs		Site #3 SPMDs		Site #4 SPMDs		Site #5 SPMDs		Site #6 SPMDs	
Field identification	9-LRR000.03	9-ABB000.02, Rep 1	7-6	9-ABB000.02, Rep 2		9-BST022.43		9-BFK000.02		9-BFK003.14	
Field sample label	Laurel Fork Conflence	Abbs Valley River	AI	Abbs Valley River	8	Bluestone River below Yards	-	Brush Fork		Brush Fork, below Armory	
Dioxins/Furans	_										
PCDDs 2,3,7,8-Tetrachloro TOTAL TETRA-PCDDs	1.3 430	0.6 110	LQ	0.6 93	LQ	4.6 2,500	LQ	4.3 150		3.3 78	
1,2,3,7,8-Pentachloro TOTAL PENTA-PCDDs	11 330	2.2 100	LQ	4.5 82		37 1,700		16 290	LQ	8.1 160	
1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8,9-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDDs	16 44 24 470	4.5 10 7.9 160	LQ	4.5 12 110	ГО	27 94 810		19 49 450		15 43 400	
1,2,3,4,6,7,8-Heptachloro TOTAL HEPTA-PCDDs	610 1,100	220 560		270 610		860 1,800		$\begin{smallmatrix}710\\1,500\end{smallmatrix}$		620 1,200	
Octachloro	2,600	2,300		2,800		3,700		6,900		5,100	
TOTAL PCDDs	4,900	3,200		3,700		10,000		9,300		6,900	
PCDFs 2,3,7,8-Tetrachloro TOTAL TETRA-PCDFs	8.0 280	5.1 110		5.5 120		22 620		11 350		7.7 250	
1,2,3,7,8-Pentachloro 2,3,4,7,8-Pentachloro TOTAL PENTA-PCDFs	3.6 9.4 150	1.2 2.1 45		1.9 3.6 50		7.3 34 390		4.3 14 160		2.6 11 150	
1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 2,3,4,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDFs	6.1 4.7 8.3 0.3 ND	0.6 0.6 0.3 0.3 26	ND LLQ	1.8 1.8 0.3 30	ND	11 8.6 14 0.3 200	ND	5.0 5.0 8.8 0.4	Ŋ	3.6 3.9 6.5 0.3 100	ŊŊ
1,2,3,4,6,7,8-Heptachloro 1,2,3,4,7,8,9-Heptachloro TOTAL HEPTA-PCDFs	62 2.2 150	7.1 0.5 18		13 0.5 30	LQ	83 1.9 160	LQ	42 1.2 91		33 0.5 72	
Octachloro	280	10		42		180		95		60	
TOTAL PCDFs	980	210		270		1,500		820		630	

				RARE Sti	July Field S	RARE Study Field Samples—Fall 2005	2005				
Sample name	Site #7 SPMDs	-	Site #8 SPMDs	Site #9 SPMDs		Site #10 SPMDs		Site #11 SPMDs		Site #12 SPMDs	
Field identification	9-XRD000.43		VA0025054-001	9-BPB000.02		9-WH1000.03		9-XEF000.10	-6	9-BPB001.51, Rep	-
international contractions in the contraction of th	Mool's Hollow		Bluefield Westside STD (2)	Beaverpond	-	door O welsid		Cross-tributary to Beaverpond Bronch	-	Beaver- pond Branch below Bluefield Cist. Boot	
Dioxins/Furans			oir (9)	CIEGN				DIallel			
PCDDs 2,3,7,8-Tetrachloro <i>TOTAL TETRA-PCDDs</i>	4.2 180		41 3,600	2.1 1,900		1.4 250	LQ, LR LR	5.0 3,300		1.7 1,500	
1,2,3,7,8-Pentachloro TOTAL PENTA-PCDDs	12 180	LQ	630 4,400	37 570		30 290	LR LR	58 940		19 800	
1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDDs	20 45 450 450		13 50 470	47 120 81 1,100		44 100 920	LQ, LR LR LQ, LR LR	79 230 150 1,700		32 72 58 1,100	
1,2,3,4,6,7,8-Heptachloro <i>TOTAL HEPTA-PCDDs</i>	$690 \\ 1,500$		430 850	1,600 3,200		1,300 2,300	LR	4,800 8,200		1,300 3,400	
Octachloro	4,900		1,400	6,700		3,400	LR	22,000		8,200	
TOTAL PCDDs	7,200		11,000	13,000		7,100		36,000		15,000	
PCDFs 2,3,7,8-Tetrachloro <i>TOTAL TETRA-PCDFs</i>	14 280		84 1,200	23 620		19 500	LR LR	19 500		9.7 320	
1,2,3,7,8-Pentachloro 2,3,4,7,8-Pentachloro TOTAL PENTA-PCDFs	2.8 13 160		3.8 19 800	6.8 24 340		$\begin{array}{c} 7.0\\18\\310\end{array}$	LR LR	8.2 30 390		3.3 13 170	
1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 2,3,4,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDFs	3.1 4.2 7.3 0.2 100	QN	2.9 3.3 7.1 LQ 0.5 ND 130	9.1 11 18 0.2 260	QN	7.3 10 18 0.2 20	LR LR LR ND, LR LR	21 23 0.2 560	ND	5.4 6.9 5.1 140	
1,2,3,4,6,7,8-Heptachloro 1,2,3,4,7,8,9-Heptachloro TOTAL HEPTA-PCDFs	36 1.3 86		43 0.8 83	130 5.6 260		120 5.1 240	LR LQ, LR LR	430 22 1,000		74 2.3 150	
Octachloro	100		95	420		370	LR	2,300		220	
TOTAL PCDFs	720		2,300	1,900		1,600		4,800		1,000	

Table 7. Estimated concentrations of 2,3,7,8-substituted polychlorinated dibenzo-*p*-dioxins (PCDDs) and dibenzofurans (PCDFs), and homologs of dioxins and furans in water from the Bluestone River watershed sample set fall 2005—Continued

25

Estimated concentrations of 2,3,7,8-substituted polychlorinated dibenzo-p-dioxins (PCDDs) and dibenzofurans (PCDFs), and homologs of dioxins and furans in water from the Bluestone River watershed sample set, fall 2005.—Continued Table 7.

[Concentrations of PCDDs and PCDFs are reported in fg·L⁻¹ water; ND, not detected at specified detection limit; LR, denotes recovery of 13C-labelled surrogate below 25% but above 10%: relative standard deviation estimated as about 25%; LQ, ion ratio outside of $\pm 15\%$ tolerances: analyte is either less than the method quantification limit or is subject to interference]

			R	ARE Stud	RARE Study Field Samples—Fall 2005	-Fall 20()5			
	Sample name	Site #13 SPMDs	Site #14 SPMDs		Site #15 SPMDs		Site #16 SPMDs		Site #17 SPMDs	
	Field identification	9-BPB001.51, Rep 2	9-BPB000.17		9-BST025.23		9-WVC000.05		9-BST028.90	
		Beaverpond Branch	Beaver-		Bluestone River above				Bluestone River below	
	Field sample label	below Bluefield City Park	pond Branch at resurgence (h)		Branch Branch		wright valley Creek		water treatment plant	
	Dioxins/Furans									
PCDDs	2,3,7,8-Tetrachloro TOTAL TETRA-PCDDs	1.3	0.9 36		2.7 180		1.8 62		2.5 71	
	1,2,3,7,8-Pentachloro TOTAL PENTA-PCDDs	19 800	29 210	ГQ	18 210		11 170		6.8 83	
	1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro 70TAL HEXA-PCDDs	27 61 1,100	46 120 85 880		28 80 53 640		19 58 30 450		12 26 320	LQ
	1,2,3,4,6,7,8-Heptachloro TOTAL HEPTA-PCDDs	1,300 3,600	2,000 3,700		1,000 2,100		710 1,500		540 1,200	
	Octachloro	8,300	12,000		6,000		3,000		4,800	
	TOTAL PCDDs	16,000	17,000		9,100		5,100		6,500	
PCDFs	2,3,7,8-Tetrachloro TOTAL TETRA-PCDFs	11 350	7.6 230		12 210		6.5 140		4.6 81	
	1,2,3,7,8-Pentachloro 2,3,4,7,8-Pentachloro TOTAL PENTA-PCDFs	3.1 11 160	2.1 3.6 70		3.9 12 130		3.0 8.6 110		1.2 3.3 45	
	1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 2,3,4,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDFs	4.4 6.1 0.2 120 N	6.6 7.3 4.7 0.2 220	QN	5.8 6.0 5.5 160	ND	3.5 4.4 0.2 120	ND QU ND CT	1.0 1.2 0.2 2 2	LQ
	1,2,3,4,6,7,8-Heptachloro 1,2,3,4,7,8,9-Heptachloro TOTAL HEPTA-PCDFs	60 2.0 120	190 13 490		73 2.3 160	ГО	64 1.7 170		15 0.3 32	ND
	Octachloro	200	1,000		200		230		27	LQ
	TOTAL PCDFs	940	2,000		850		770		230	

Table 7. Estimated concentrations of 2,3,7,8-substituted polychlorinated dibenzo-*p*-dioxins (PCDDs) and dibenzofurans (PCDFs), and homologs of dioxins and furans in water from the Bluestone River watershed sample set, fall 2005.—Continued [Concentrations of PCDDs and PCDFs are reported in fg·L⁻¹ water; ND, not detected at specified detection limit; LR, denotes recovery of 13C-labelled surrogate below 25% but above 10%: relative standard deviation estimated as about 25%; LQ, ion ratio outside of $\pm 15\%$ tolerances: analyte is either less than the method quantification limit or is subject to interference]

		M	/DEP Stud	WVDEP Study Field Samples—Fall 2005	all 2005-		
	Sample name	Site #1 SPMDs		Site #2 SPMDs	Site	Site #3 SPMDs (*)	
	Field identification	WVDEP-1		WVDEP-2		WVDEP-3	
	Field sample label	Beacon Cave #1 (h)	Be	Beacon Cave #2 (h)	≥	Morris Spring	
	Dioxins/Furans						
PCDDs	2,3,7,8-Tetrachloro TOTAL TETRA-PCDDs	0.7 15		0.5 11		1.8 52	
	1,2,3,7,8-Pentachloro TOTAL PENTA-PCDDs	31 130		23 110		24 180	
	1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDDs	50 140 820		39 110 650		2.8 11 7.3 130	LQ LQ
	1,2,3,4,6,7,8-Heptachloro TOTAL HEPTA-PCDDs	2,300 3,800		2,000 3,300		160 420	
	Octachloro	12,000		12,000		780	
	TOTAL PCDDs	17,000		17,000		1,600	
PCDFs	2,3,7,8-Tetrachloro TOTAL TETRA-PCDFs	1.9 59	LQ	1.3		1.9 56	
	1,2,3,7,8-Pentachloro 2,3,4,7,8-Pentachloro <i>TOTAL PENTA-PCDFs</i>	1.6 4.7 59		0.9 2.9 41		0.6 2.8 36	
	1,2,3,4,7,8-Hexachloro 1,2,3,6,7,8-Hexachloro 2,3,4,6,7,8-Hexachloro 1,2,3,7,8,9-Hexachloro TOTAL HEXA-PCDFs	10 9.9 15 380		8.2 12.9 0.3 280	ND	0.7 0.9 0.1 28 28	ND LQ
	1,2,3,4,6,7,8-Heptachloro 1,2,3,4,7,8,9-Heptachloro TOTAL HEPTA-PCDFs	290 20 930		250 17 760		9.5 0.2 20	QN
	Octachloro	2,100		1,900		22	
	TOTAL PCDFs	3,500		3,000		160	

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For more information concerning this publication, contact: Director U.S. Geological Survey Columbia Environmental Research Center 4200 New Haven Road Columbia, MO 65201 (573) 875–5399

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