

**A Study of Non-covalent Protein-protein Complexes  
Under Native Condition by Matrix-assisted Laser  
Desorption/Ionization**

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## Major References of MALDI to Study Non-covalent Complexes

F. Hillenkamp, 1990, Streptavidin, a tetramer, nicotinic acid in 10% ethanol in water

F. Hillenkamp, 1995, Omp F porin, a trimer, ferulic acid in THF

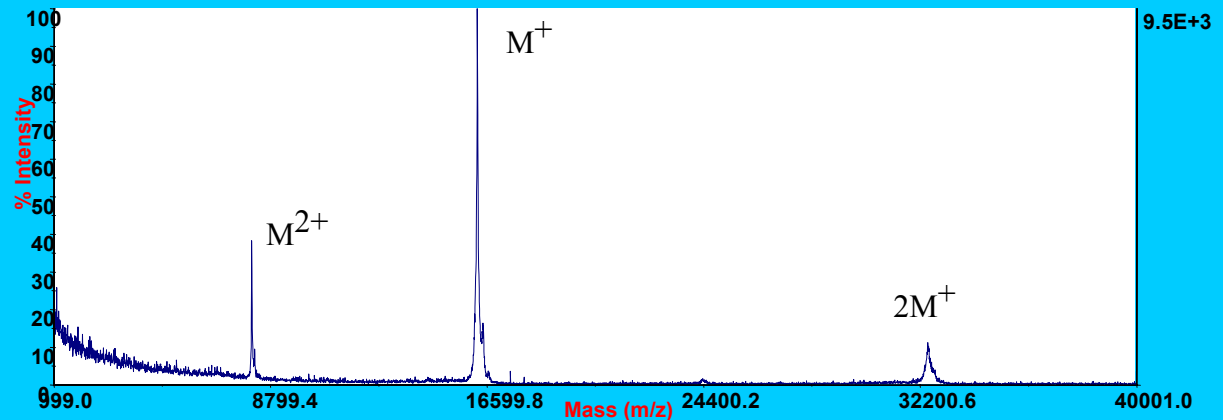
A. S. Woods and R. J. Cotter, 1995, enzyme and substrate complexes, sinapinic acid in ethanol-1M ammonium citrate at near pH 7.

M. Przybylski et al, 1996, RNase S consisting of S-protein and S-peptide using ATT with 10 mM ammonium acetate at pH5.5 .

F. Hillenkamp, 1997, Streptavidin, a tetramer, using DHAP in THF, ethanol, THAP in THF and ACN-TFA.

# Problems

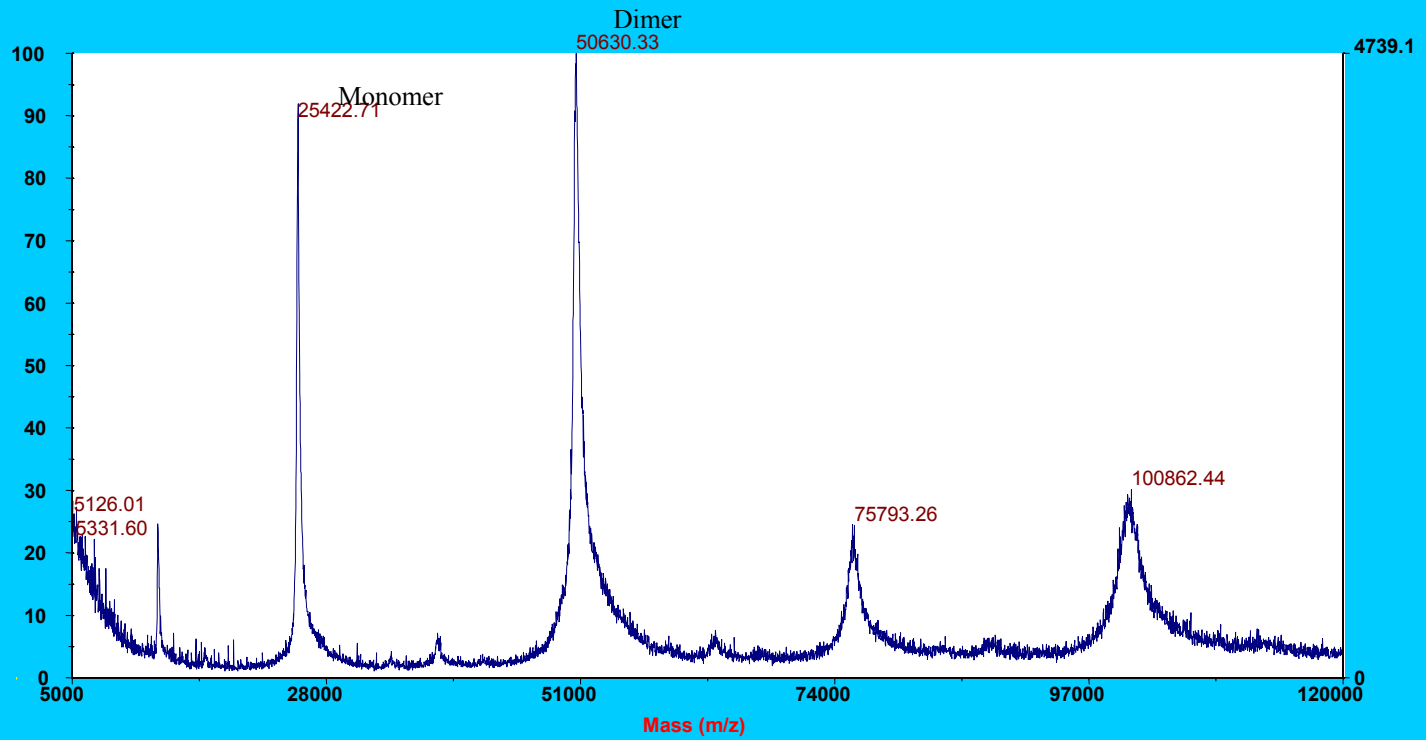
- The solution-phase acidity of these matrix substances and some typical additives (e.g the hydrophobicity of the organic (co)solvents and TFA) leads to dissociation of the noncovalent assemblies in most cases.
- It has been observed occasionally that an abundant signal of intact subunit assemblies could be obtained for the first laser shot on a not-yet-irradiated sample spot exclusively.
- Not very much is known about the gas-phase stability of molecular ions from noncovalent biomolecular complexes formed under MALDI-MS conditions.
- A further complication in MALDI-MS is the fact that nonspecific, noncovalent homo- and heteroligomeric aggregates, called cluster ions, are frequently observed.



# Possible Solutions

- Eliminate the organic co-solvent and TFA.
- Other new neutral matrices; use acidic matrices, then adjust the pH to neutral with base such as ammonium hydroxide.
- Compounds tested: 4-bromo-2,6-dimethylaniline, 2-pyridine carboxylic acid, 2-pyrazine-carboxylic acid, 2-quinolinecarboxylic acid, 2-isoquinolinecarboxylic acid, and other compounds.
- Sinapinic acid with ammonium citrate adjusted to pH 7 with ammonium hydroxide was found to be effective matrix for the detection of non-covalent protein complexes.

# Oligomeric States of Proteins



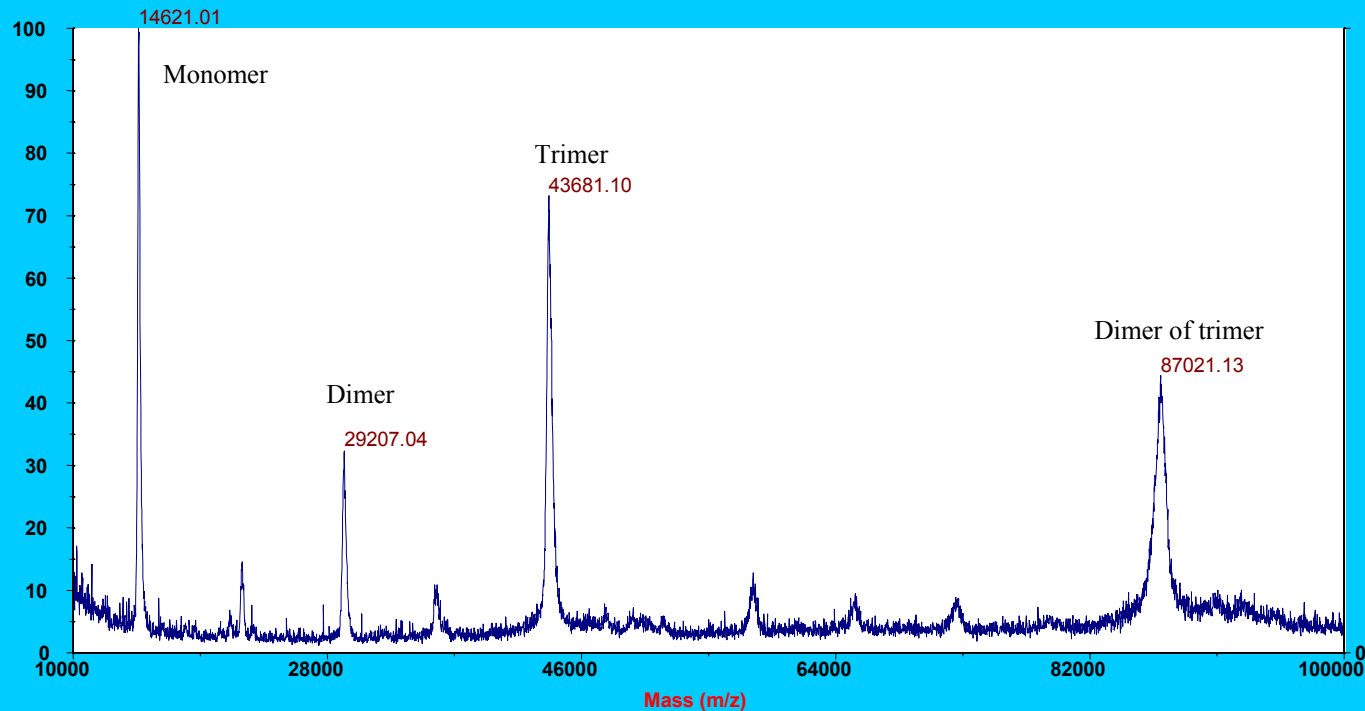
MALDI Spectrum of PhzD

# A Protein Trimer HI0719

Protein HI0719 belongs to a family of proteins are widely distributed in bacteria, archaea, plants and eukaryote.

HI0719 is a homotrimer by light scattering measurement and was proved as a trimer by solution NMR study.

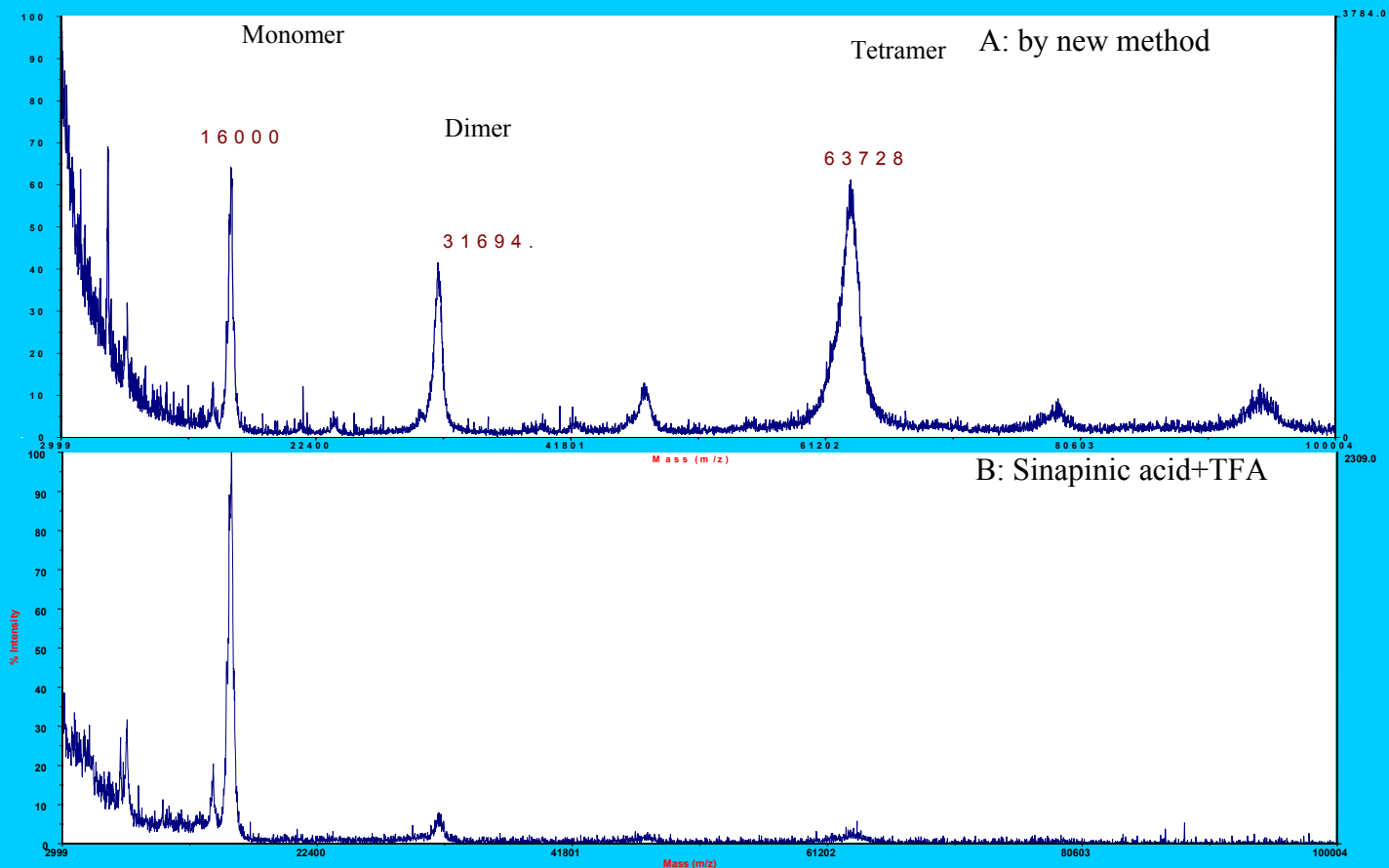
This protein is known as a trimer in solid state by X-ray crystallography.



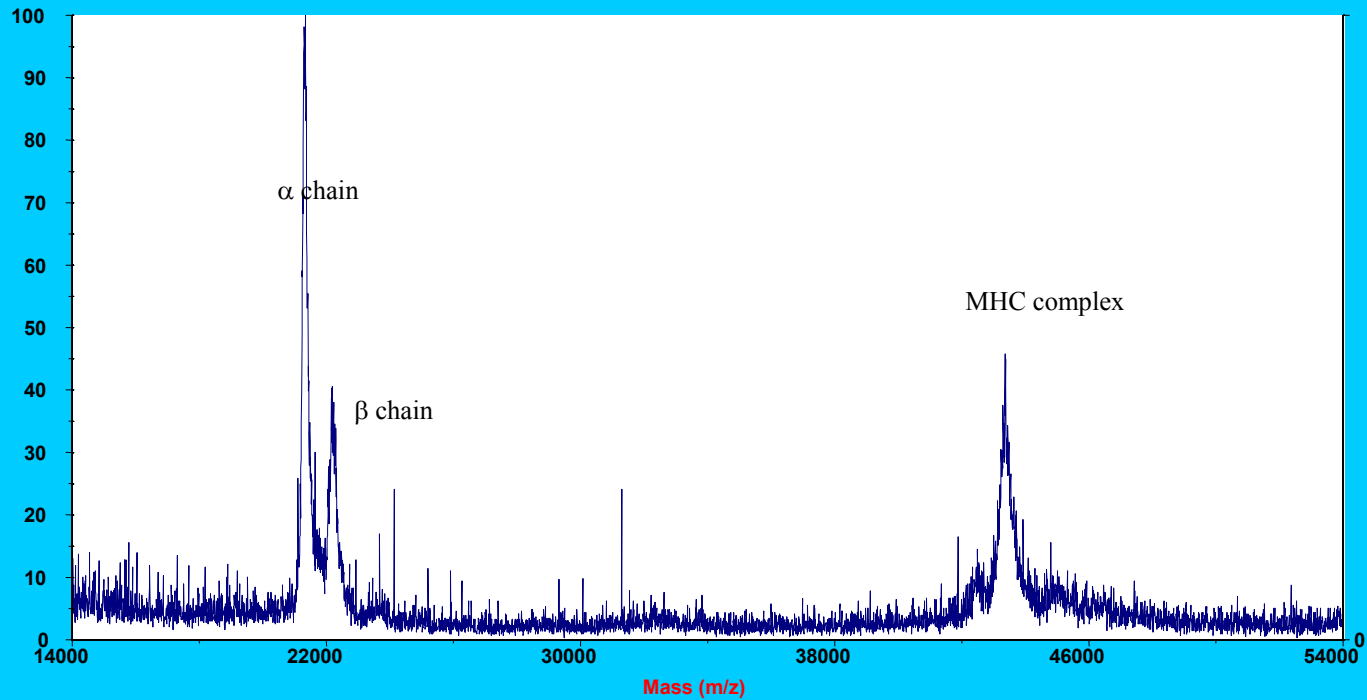
MALDI Spectrum of Protein HI0719

# Comparison of New Method with the Traditional Method

Avidin is a glycoprotein found in egg white whose active form is a tetramer composed of identical subunits. The tetramer has been detected by MALDI.



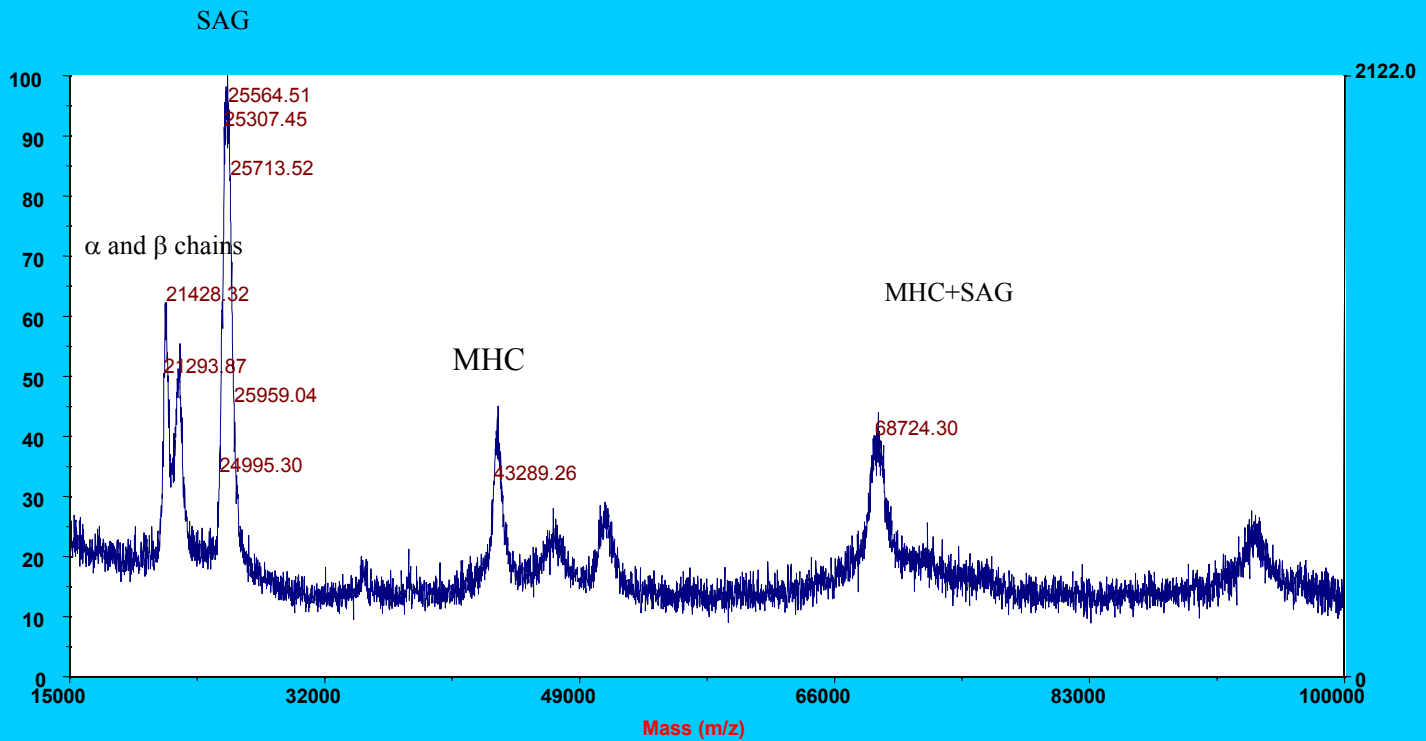
## Class II Major Histocompatibility Complex



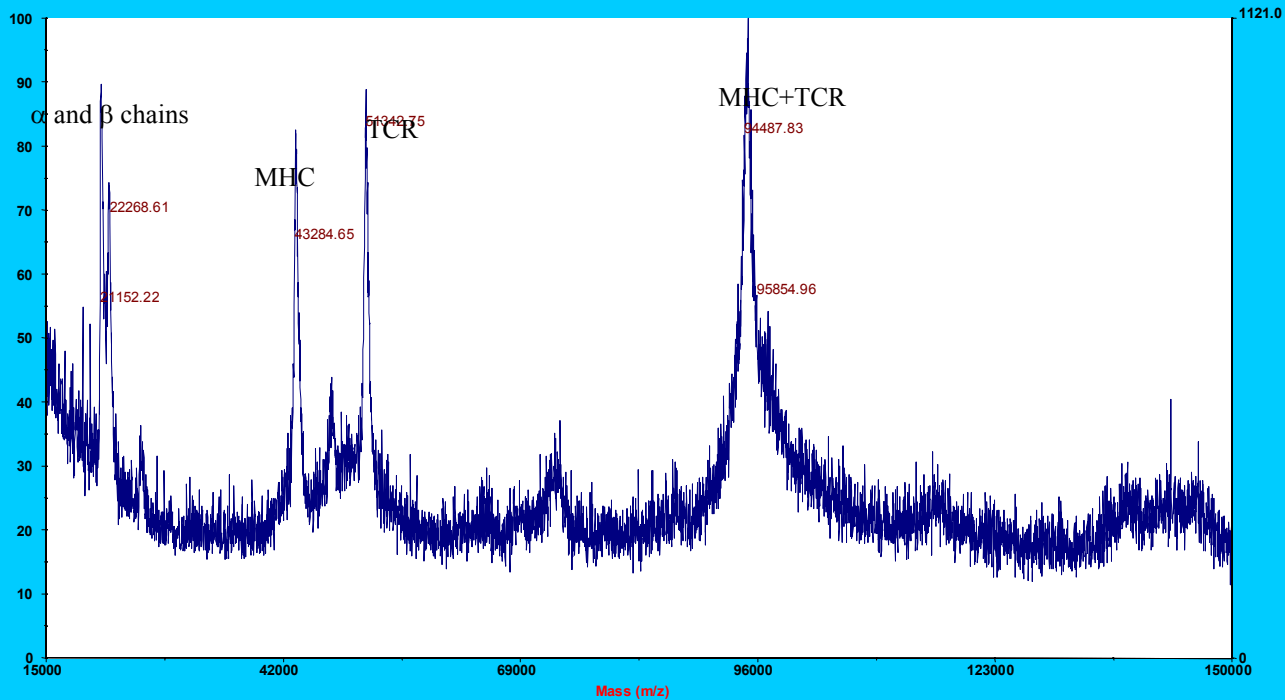
MALDI Spectrum of a Class II Major Histocompatibility Complex



# The Complex of Superantigen (SAG) with Peptide-bound Major Histocompatibility (MHC) Molecule

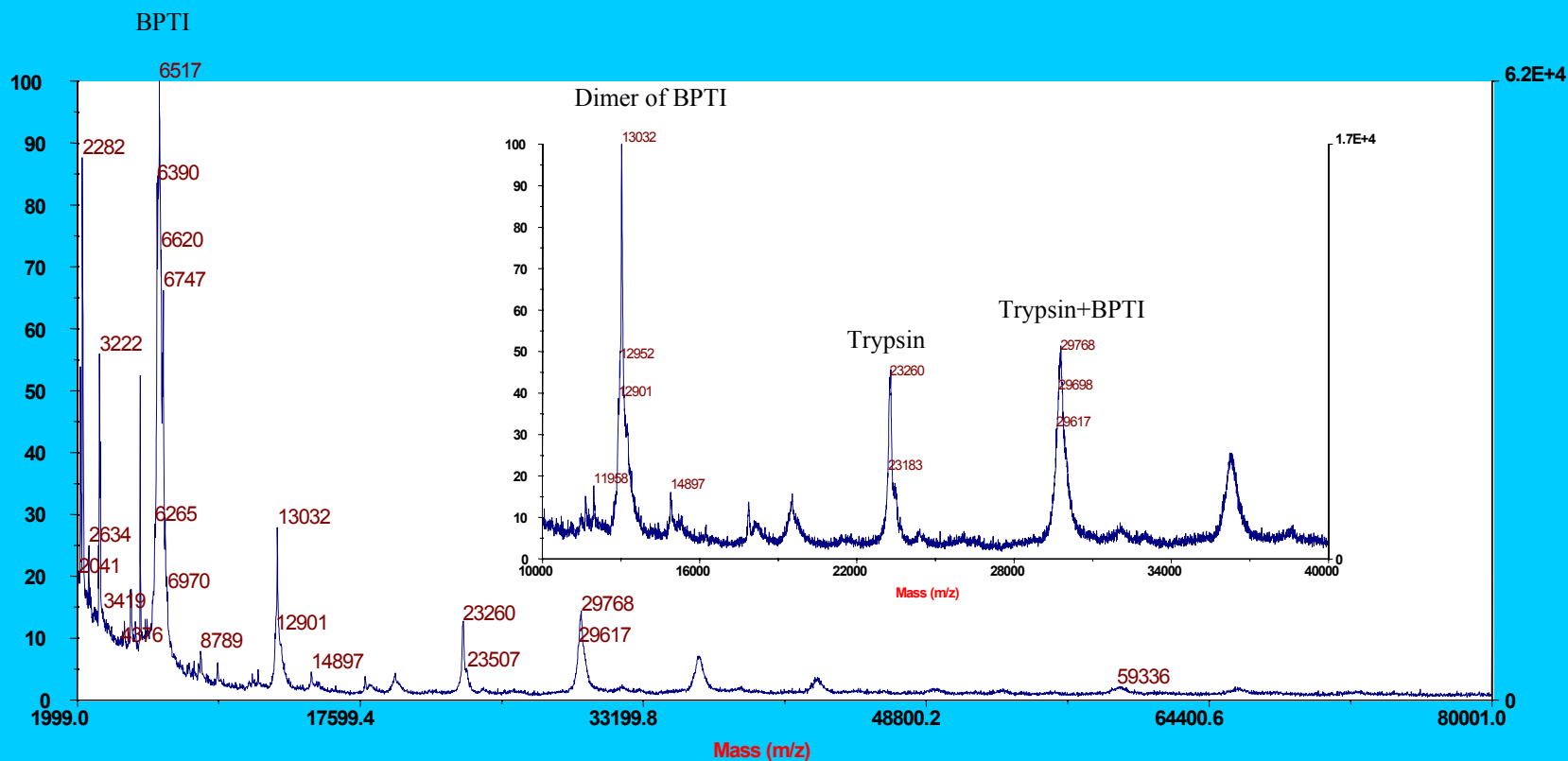


# The Complex of Peptide-linked T Cell Receptor (TCR) and MHC Molecule



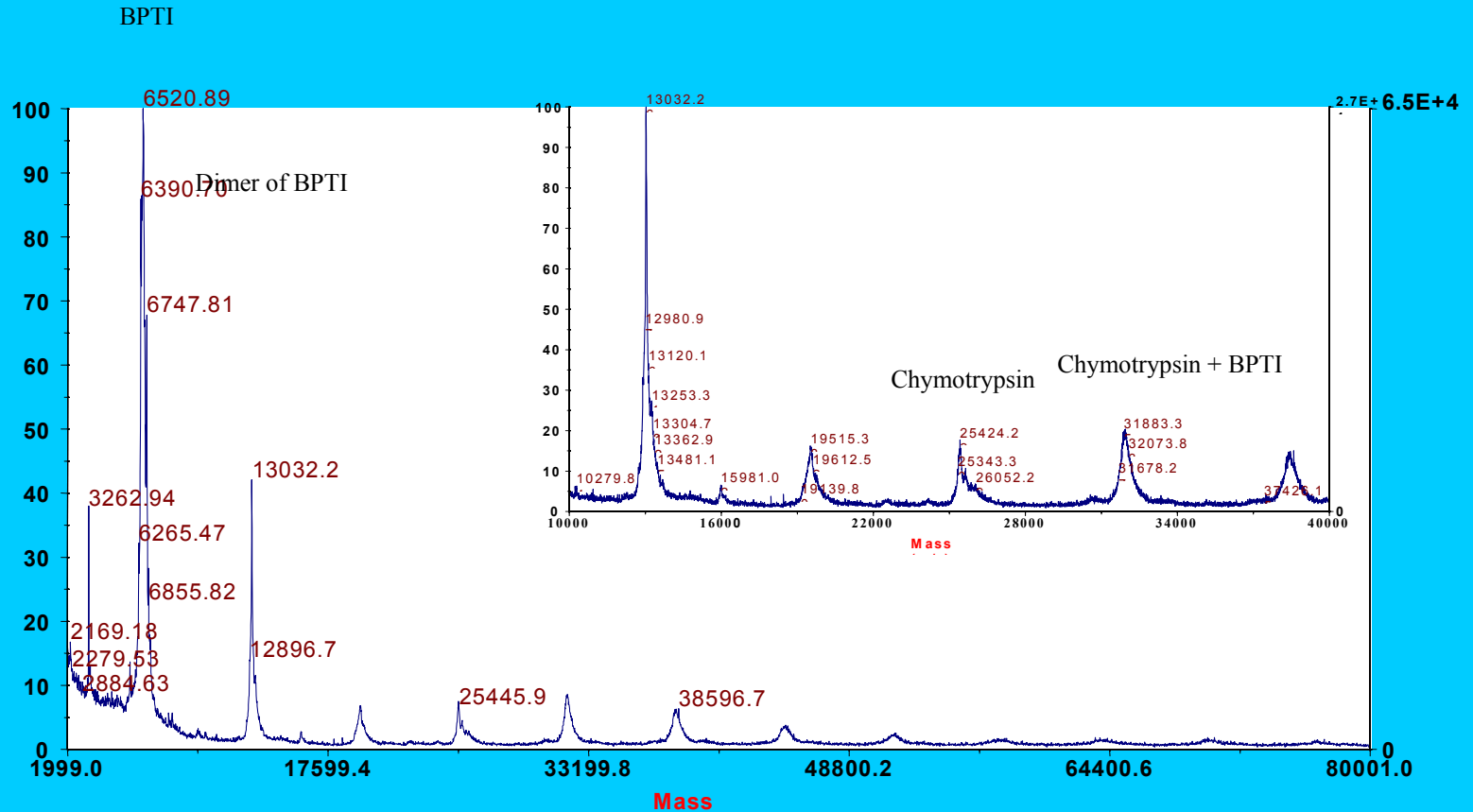
MALDI Spectrum of the Complex of Peptide-linked T Cell Receptor (TCR) and MHC Molecule

# Detection of Enzyme Inhibitor Complexes



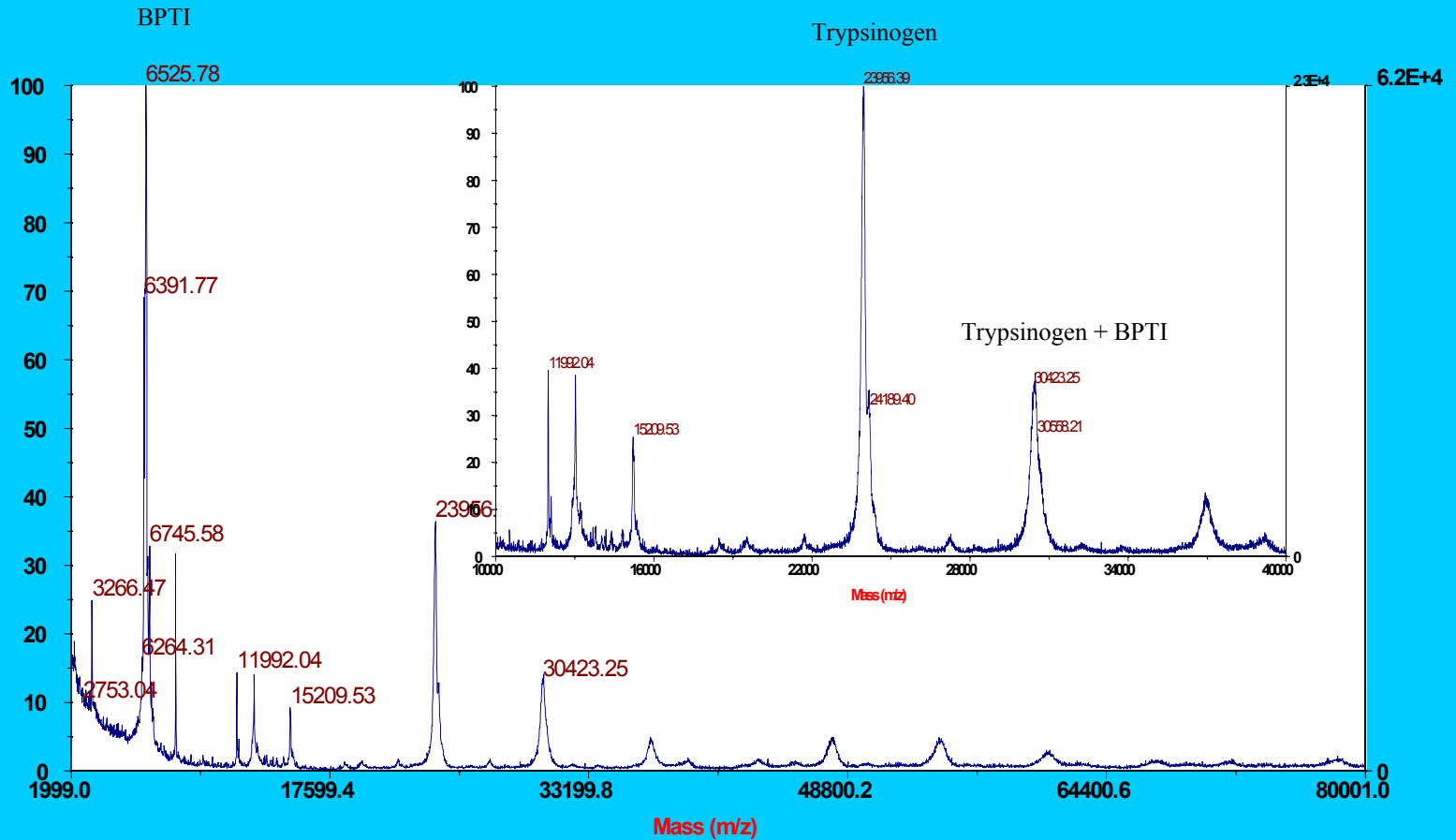
MALDI Spectra of Trypsin (6.3  $\mu$ M) with Bovine Pancreatic Trypsin Inhibitor (BPTI) (25  $\mu$ M)

# Complex of Chymotrypsin with BPTI



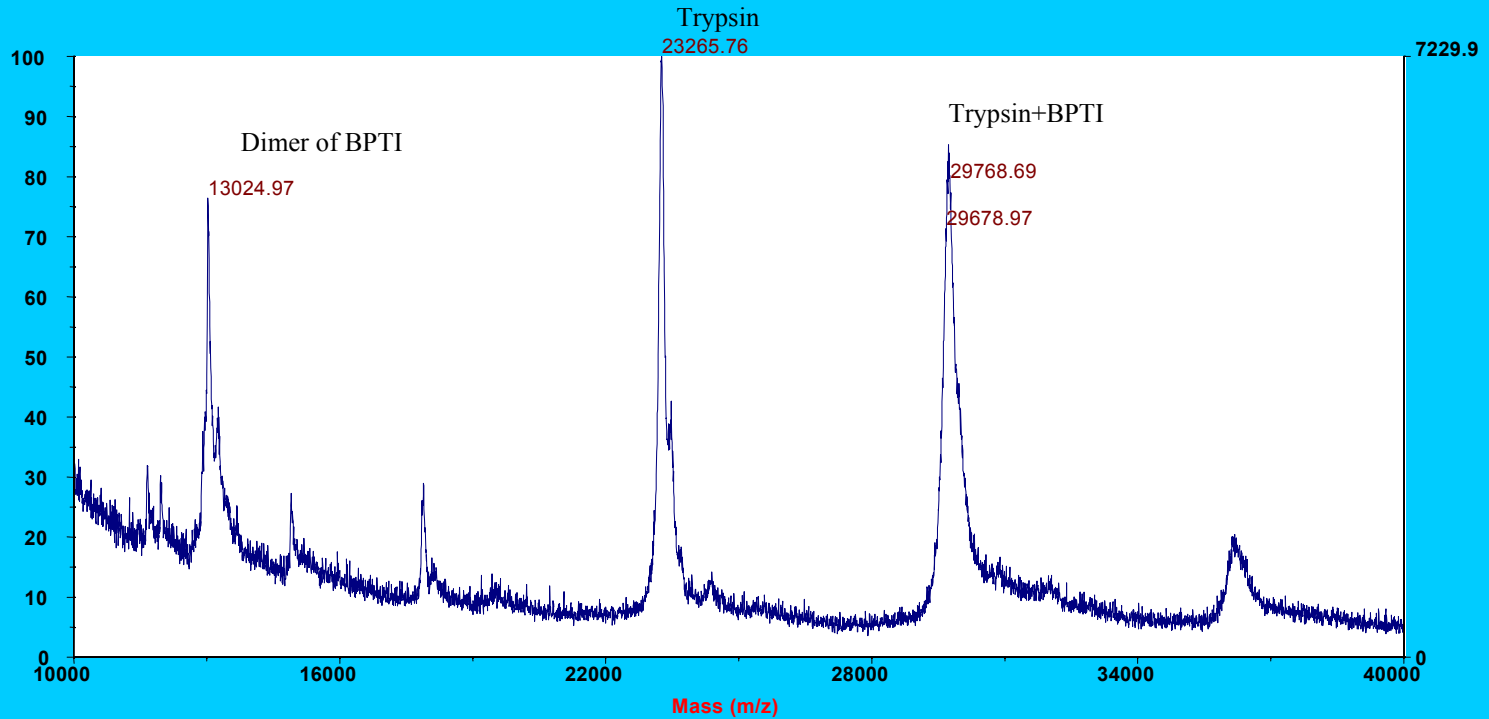
MALDI Spectrum of Chymotrypsin (6.3  $\mu$ M) With BPTI (25  $\mu$ M)

# Complex of Trypsinogen with BPTI



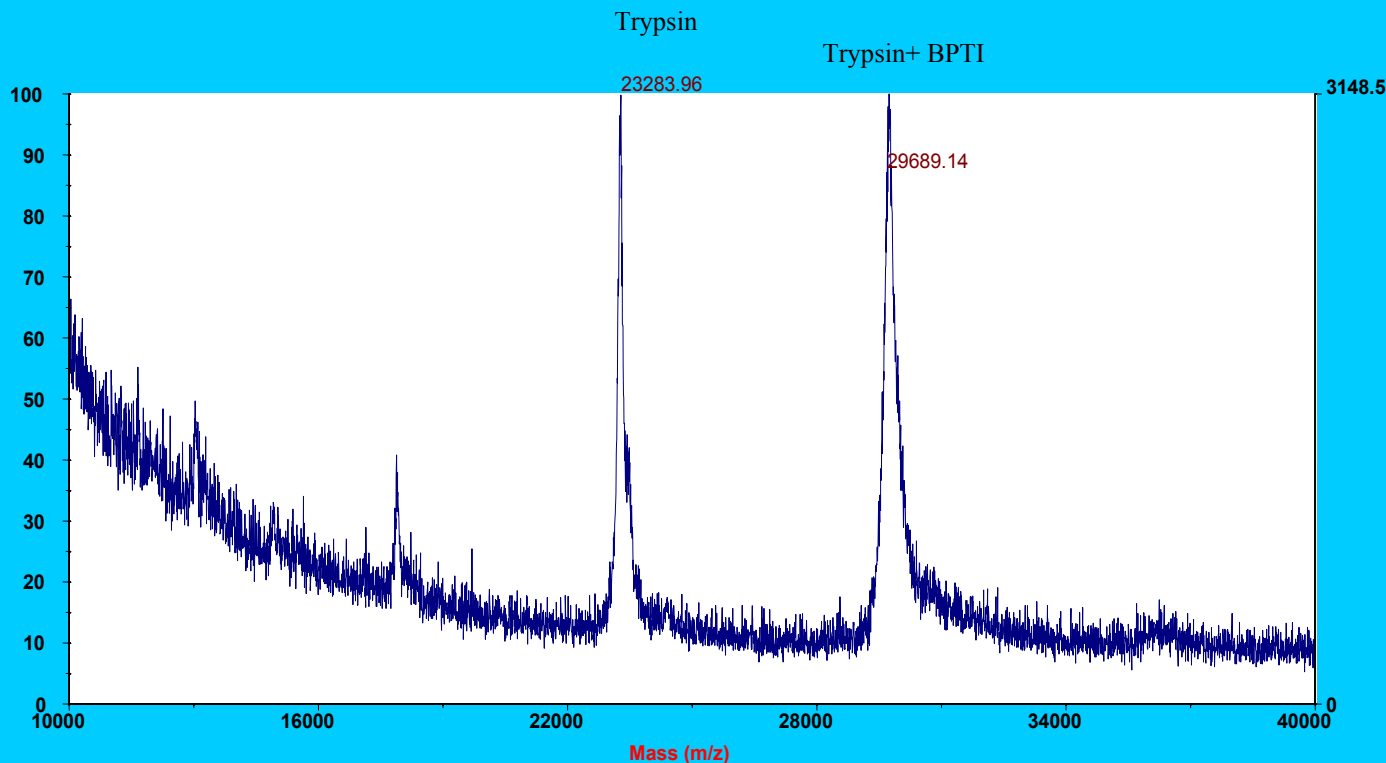
MALDI Spectrum of Trypsinogen (6 uM) With BPTI (25 uM)

## Dilution Study of Trypsin with Bovine Pancreatic Trypsin Inhibitor (BPTI) Complex



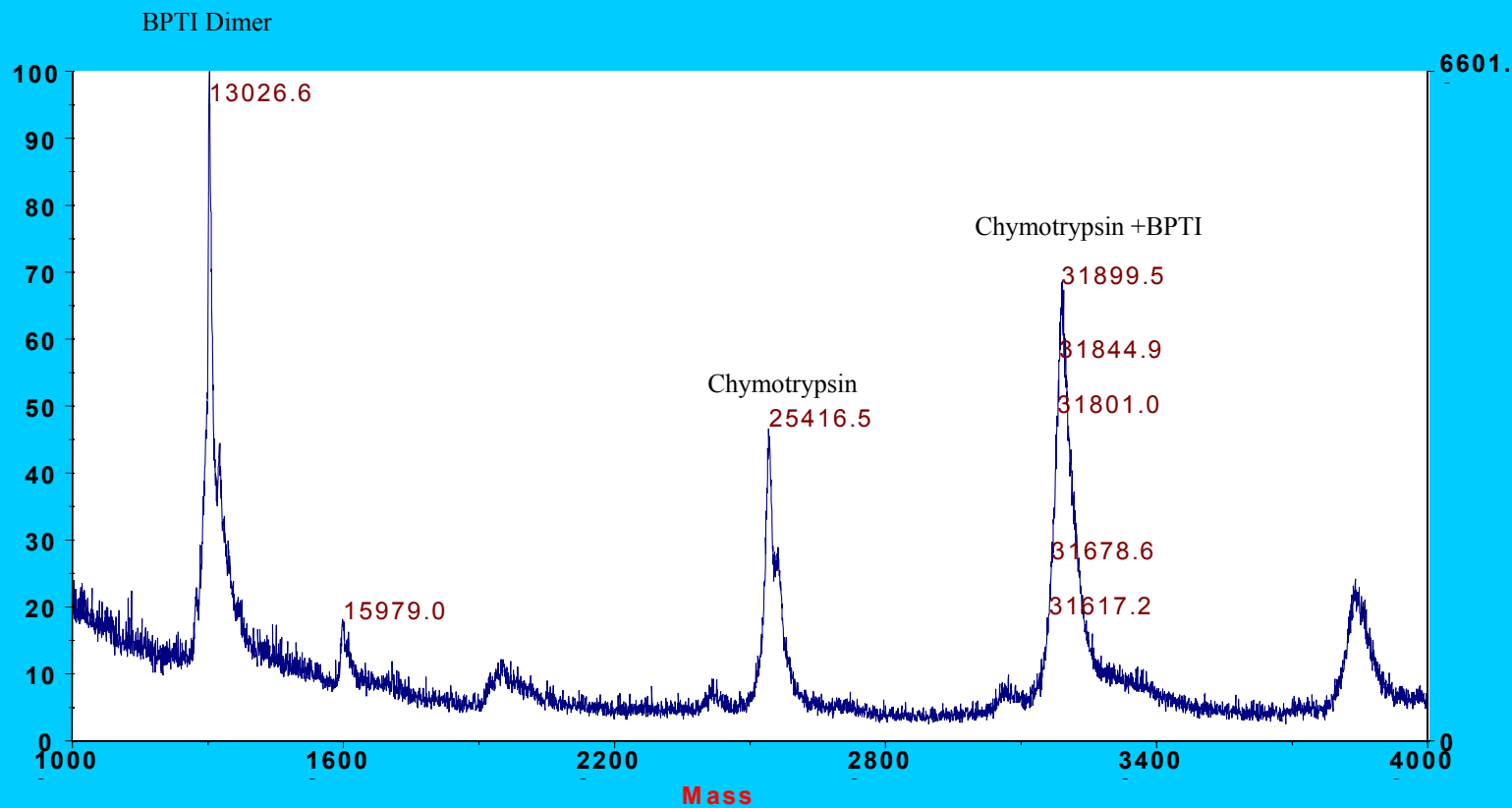
MALDI Spectra of Trypsin (2.4  $\mu\text{M}$ ) with Bovine Pancreatic Trypsin Inhibitor (BPTI) (10  $\mu\text{M}$ )

## Further Dilution Study of Trypsin with Bovine Pancreatic Trypsin Inhibitor (BPTI) Complex



MALDI Spectrum of Trypsin (0.63  $\mu\text{M}$ ) with Bovine Pancreatic Trypsin Inhibitor (BPTI) (2.5  $\mu\text{M}$ )

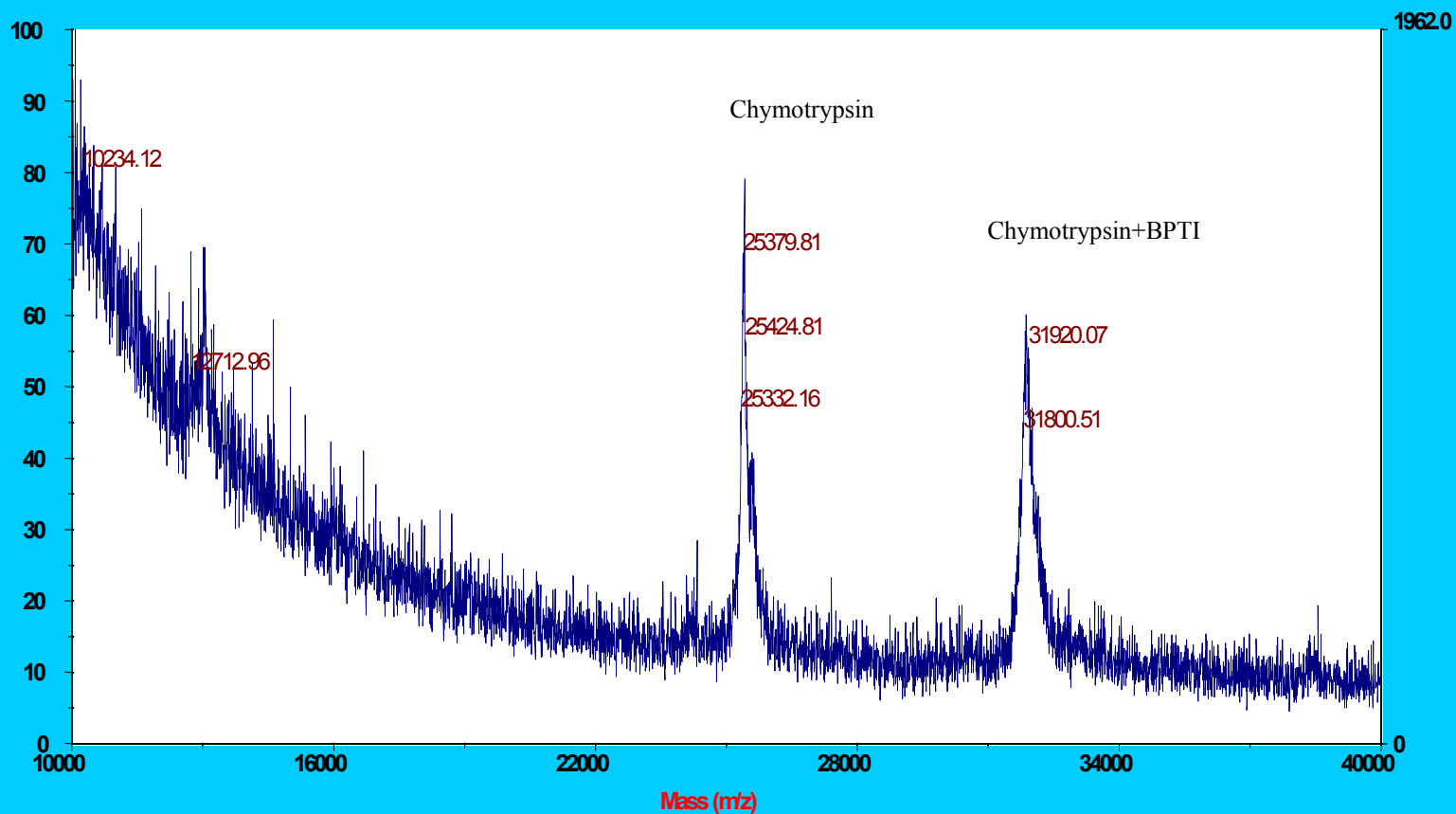
# Dilution Study of Complex of Chymotrypsin with BPTI



MALDI Spectrum of Chymotrypsin (2.5  $\mu$ M) With BPTI (10  $\mu$ M)

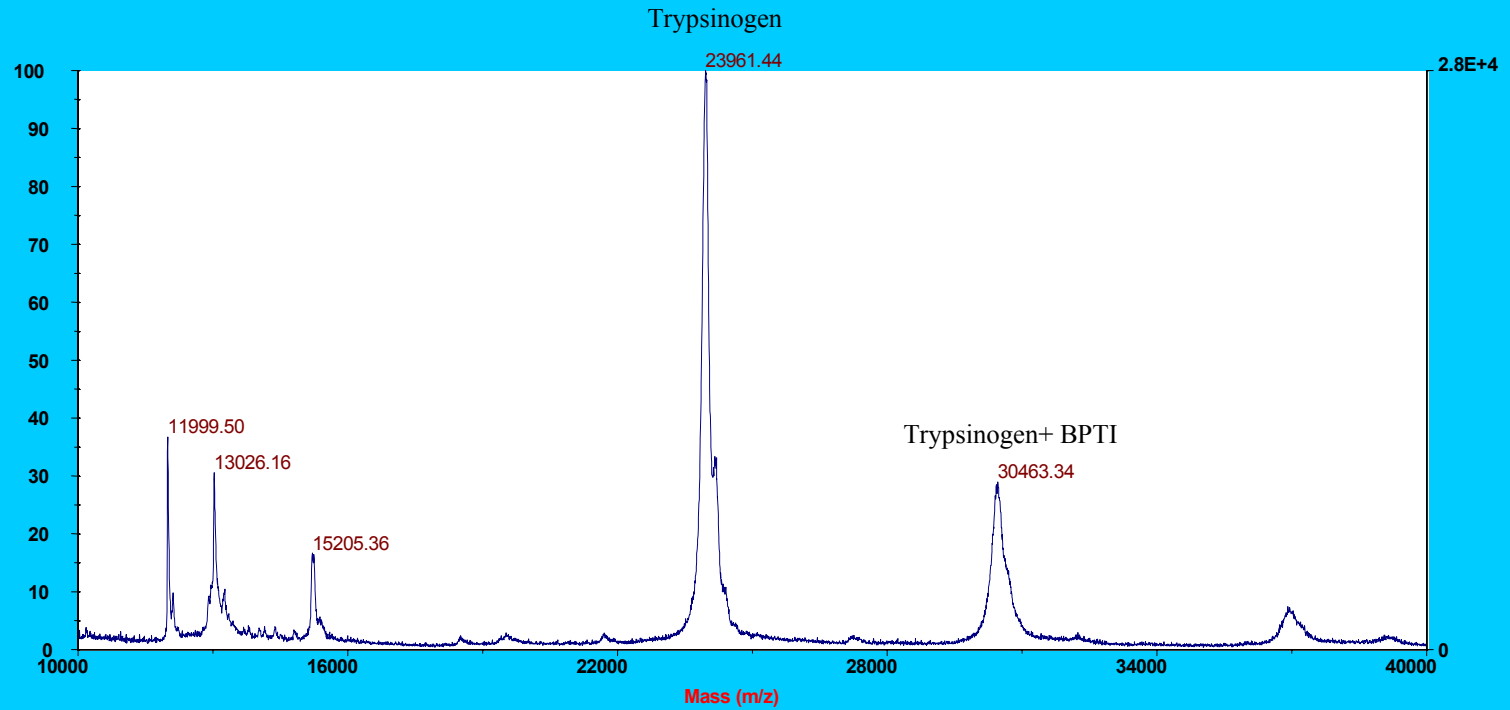


## Further Dilution Study of Complex of Chymotrypsin with BPTI



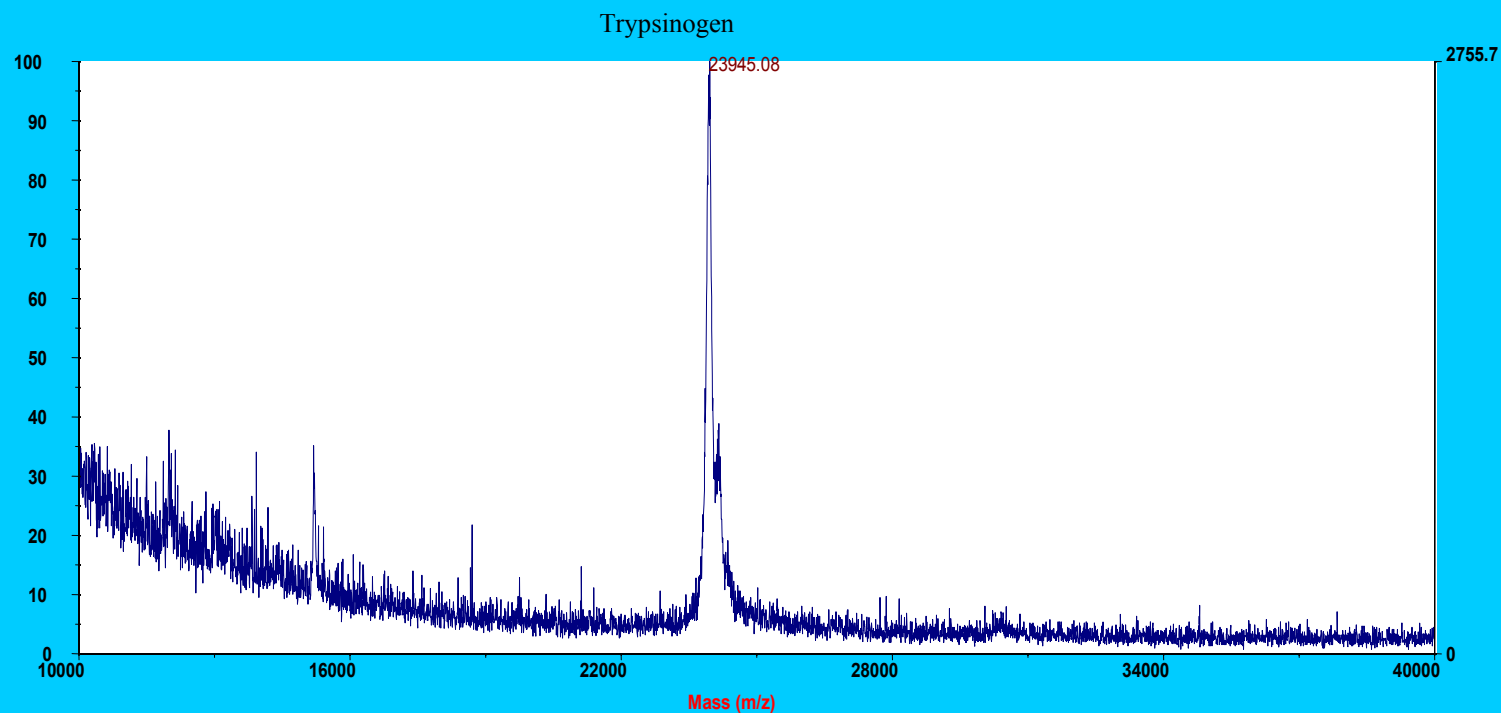
MALDI Spectra of Chymotrypsin (0.63 $\mu$ M) With BPTI (2.5  $\mu$ M)

# Dilution Study of Complex of Trypsinogen with BPTI



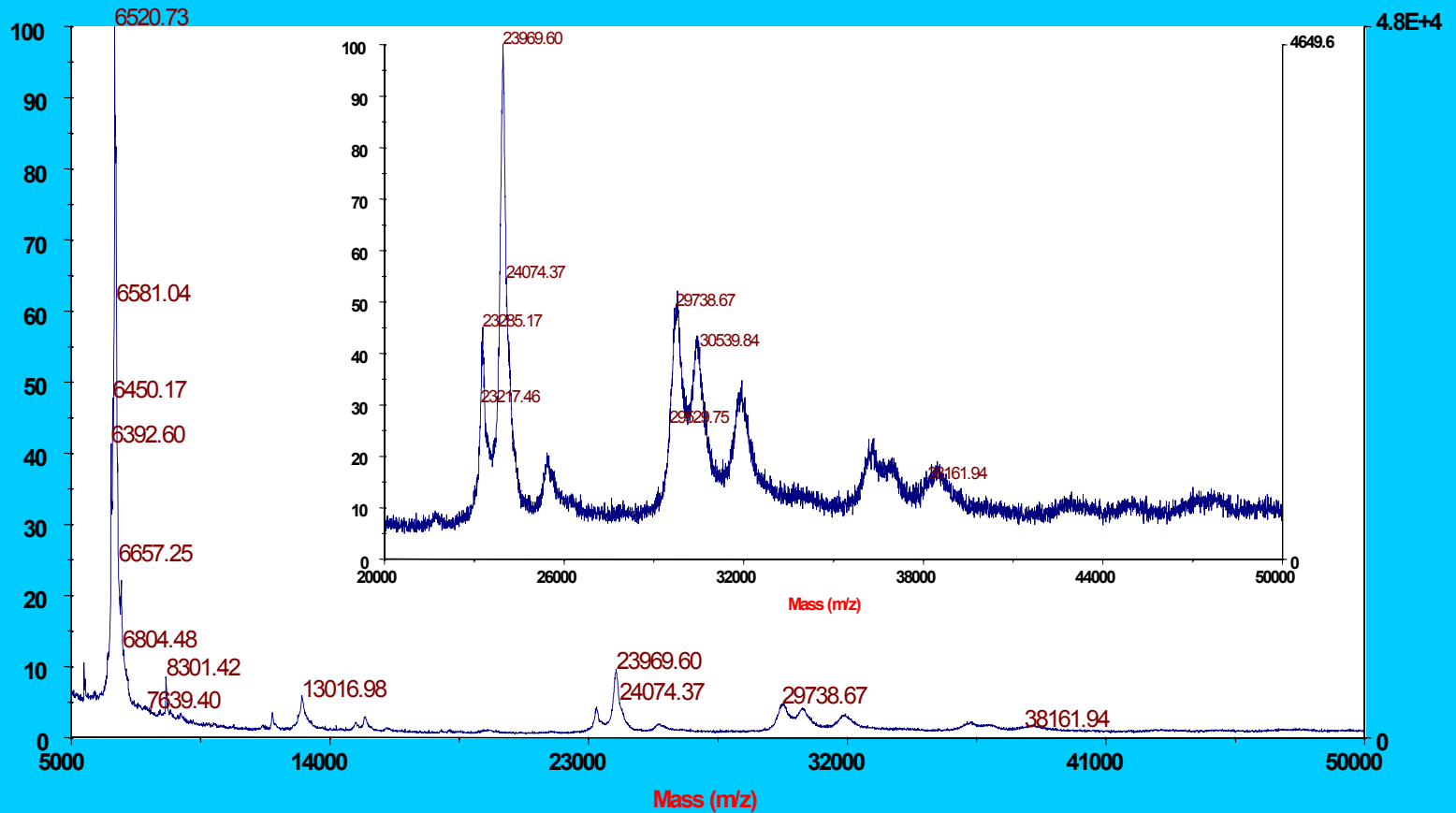
MALDI Spectrum of Trypsinogen (2.5uM) With BPTI (10 uM)

## Further Dilution Study of Complex of Trypsinogen with BPTI



MALDI Spectra of Trypsinogen (0.63  $\mu\text{M}$ ) With BPTI (2.5  $\mu\text{M}$ )

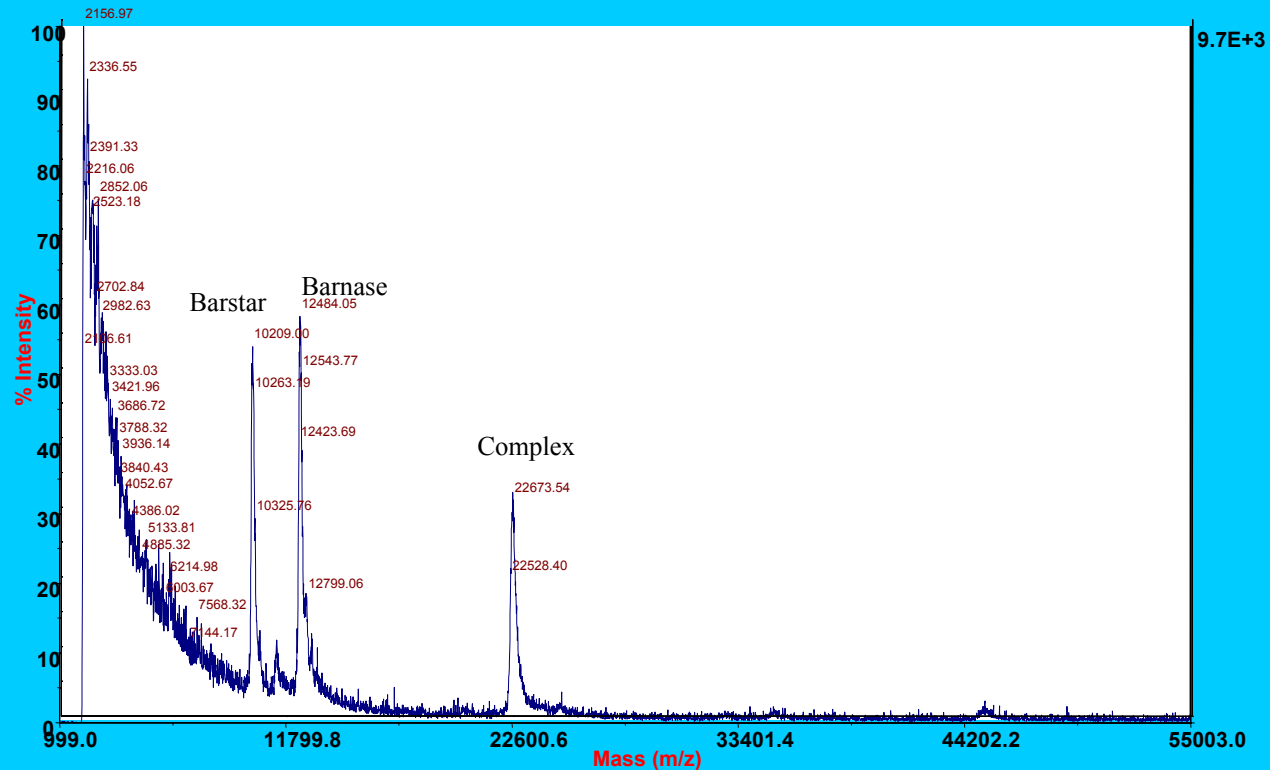
# Competitive Binding Study



MALDI Spectra of Complexes of Trypsin, Chromotrypsin and Trypsinogen in about Equal Molar Ratio with BPTI.

# Complex of Barnase and Barstar

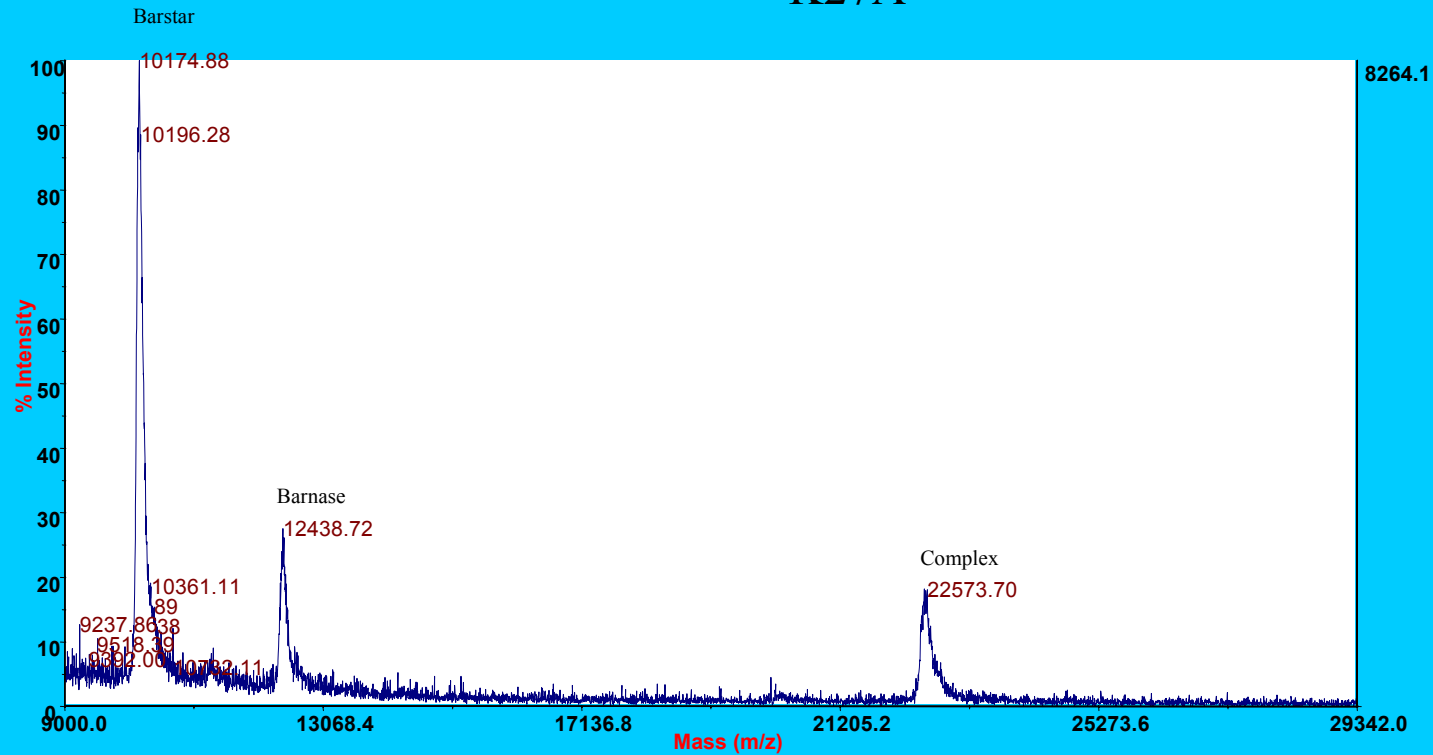
Barnase is a 110-residue extracellular protein found in *Bacillus amyloliquefaciens*. It is a ribonuclease whose potentially lethal functions within the cell are inhibited by barstar, a 90-residue polypeptide.



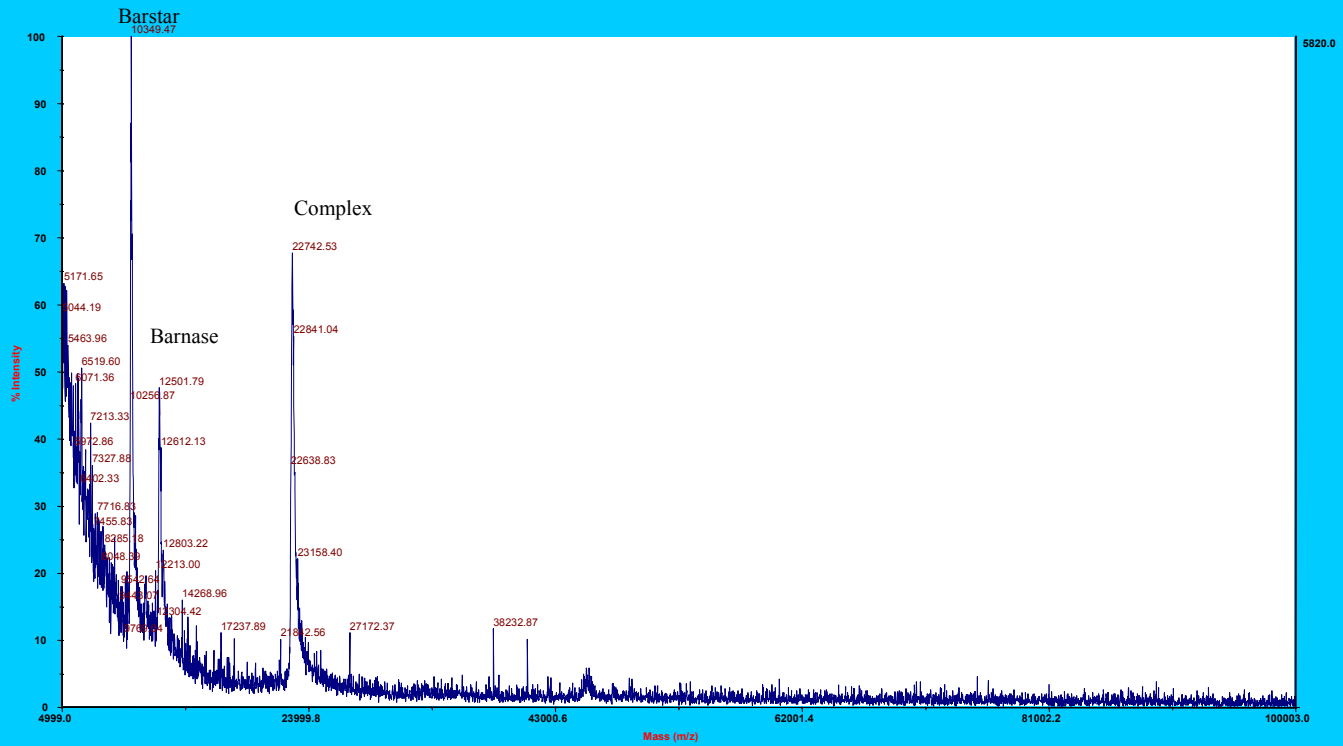
# Complex of Barnase and Barstar Mutants A

Barstar  
Y29A  
Y29F  
D39A

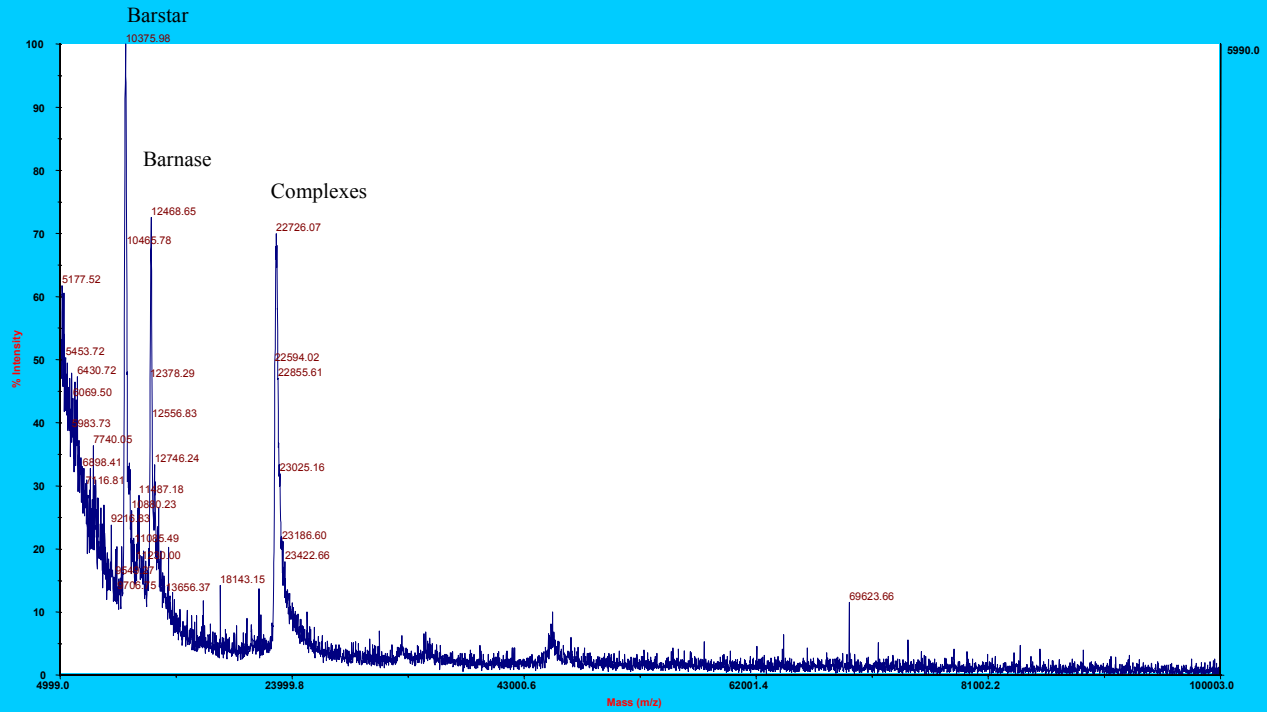
Barnase  
H102A  
R59A  
K27A



# Complex of Barnase and Barstar Mutants B

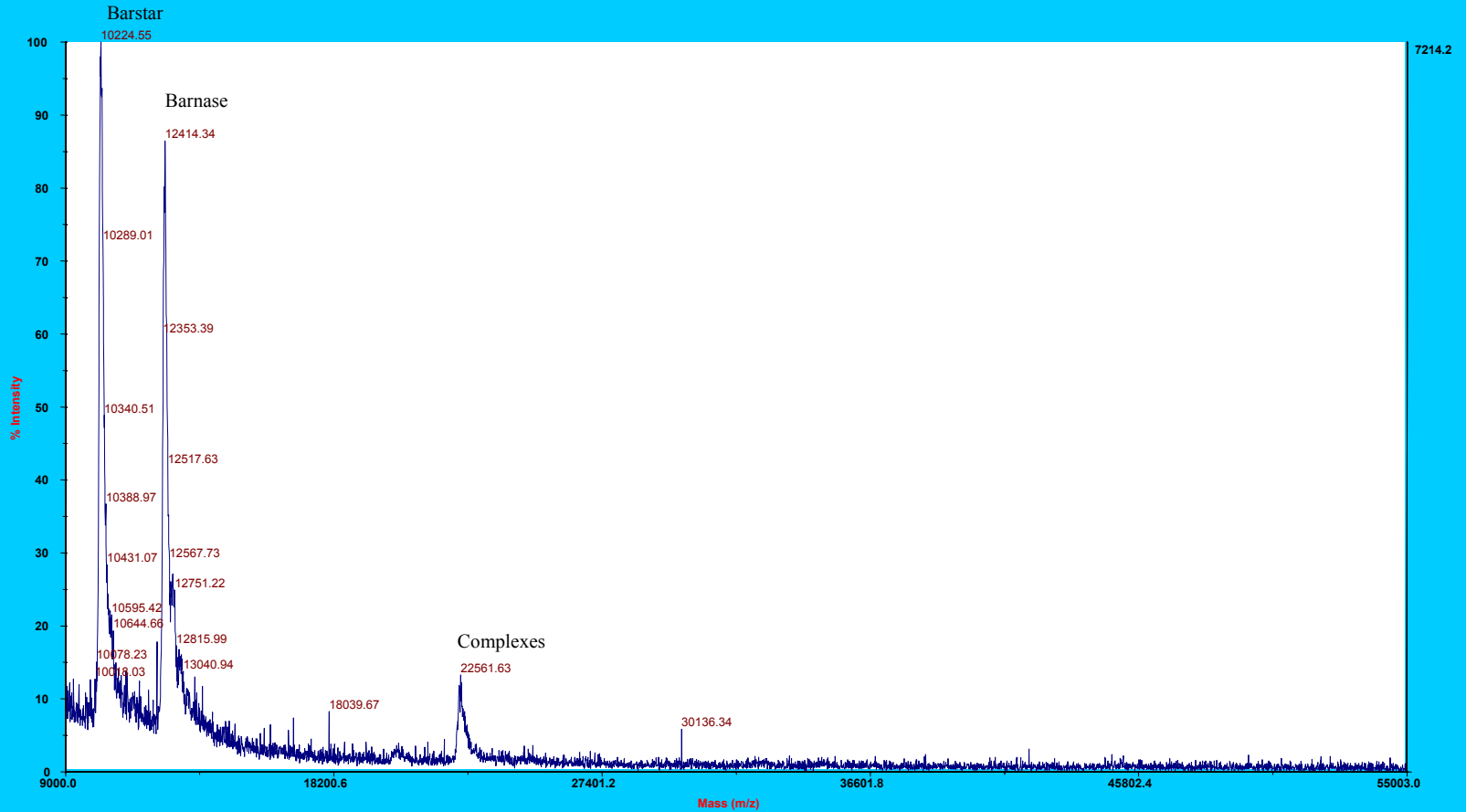


# Complex of Barnase and Barstar Mutants C

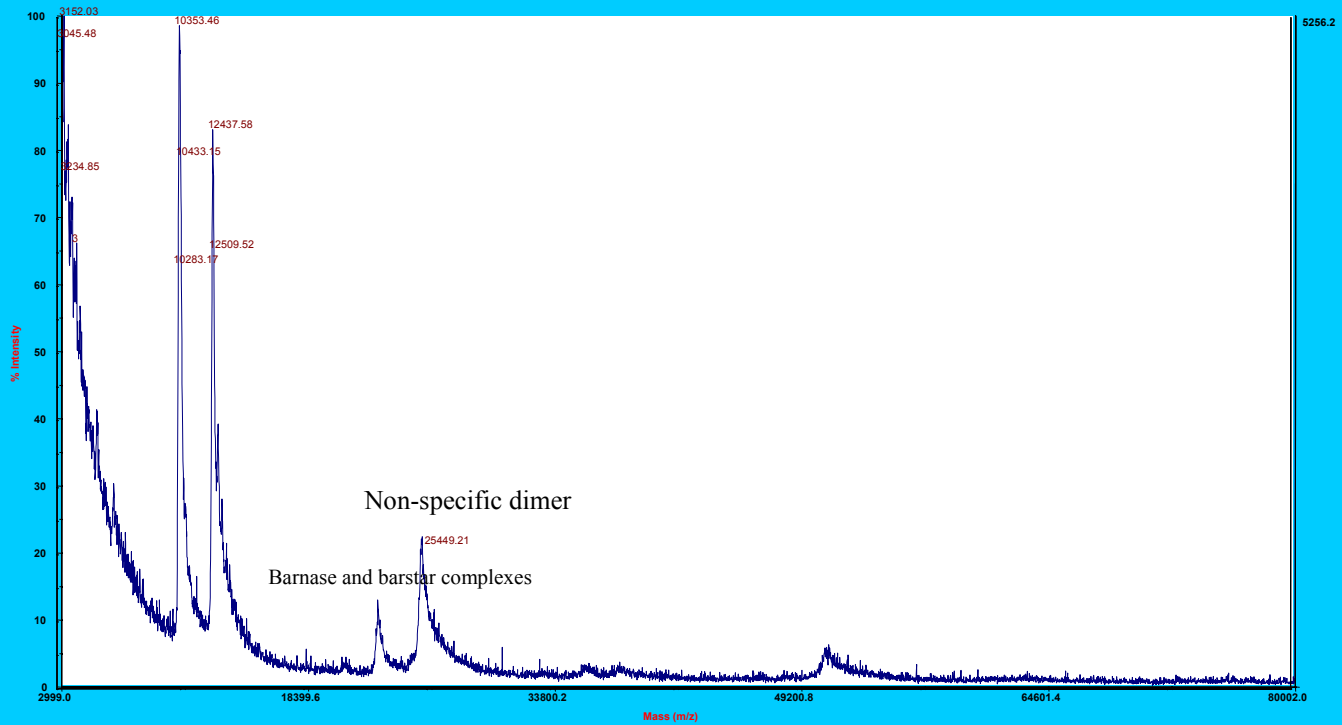




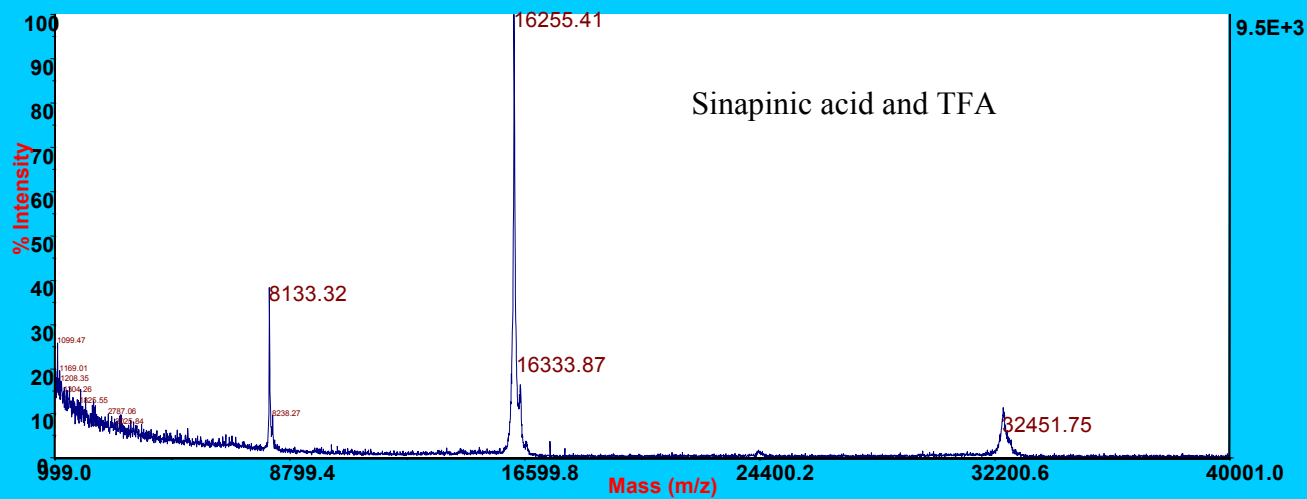
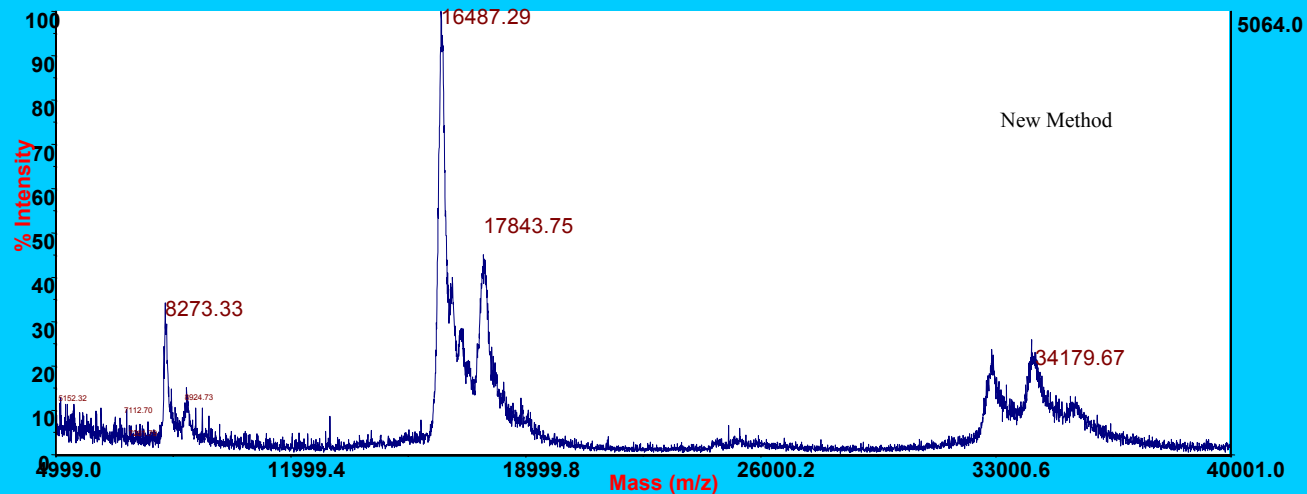
# Complex of Barnase and Barstar Mutants D



# Complex of Barnase and Barstar Mutants E



# A Complexes of PAL Protein and Peptidoglycan



# Conclusions

- Protein-protein complexes were detected by MALDI using a new sample preparation method.
- The new method uses aqueous solution at physiological pH, which can be broadly used to study protein-protein complexes.
- Good correlation was observed between the gas phase complexes detected by MALDI and their known associations in solution by other methods.
- The results suggest that MALDI can be used to study protein quaternary structures.

## Acknowledgements:

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