

Investigation of Protein-Protein Interactions in the Metal-Reducing Bacterium Desulfovibrio vulgaris Hildenborough Swapnil Chhabra¹, Sara Gaucher², Grant Zane³, Eric Alm⁴, Adam Arkin¹, Terry Hazen¹, Judy Wall³ and Anup Singh²

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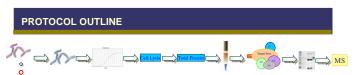
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INTRODUCTION

Desulfovibrio vulgaris is a sulfate reducing bacterium of interest due to its potential use in bioremediation as well as its economic impact in the petroleum industry (bio-corrosion of pumping machinery). This sulfate reducing bacterium has been shown to reduce toxic metals (such as chromium and uranium) to insoluble species making them a good model system for understanding molecular machines involved in bioremediation of contaminated soils and ground water.

We have implemented an approach for the isolation of protein complexes from *D. vulgaris* by generating *D. vulgaris* cell lines that produce a "bait" protein of interest fused to an affinity tag (strep tag). Lysate from these cells is passed over an avidin column, and the bait protein with its associated proteins is captured and can be selectively eluted from contaminating proteins. One advantage of this approach is that complexes are formed and captured under native conditions. Nanoscale liquid chromatography-tandem mass spectrometry (nLC/MS/MS) is used identify the captured proteins.

To determine bait proteins of interest, we computationally identified open reading frames that are involved in stress response (including oxygen, heat, pH and salt) based on homology to known stress related genes from other prokaryotic species. We have also selected proteins that are unique to this sulfate reducer ("signature" genes), expected to yield novel complexes related to sulfate/metal reduction. To validate our methods and address the challenge of non-specific binding, we have included some bait proteins whose interacting partners are well characterized in prokaryotic systems and have validated our methods by isolating the binding partners of these targets.



A. Tagged-Protein Generation

- 1. The gene encoding the target protein is amplified with a modified stop codon.
- 2. The amplified product is introduced in a plasmid containing the tag sequence such that the tag is in the same reading frame as the gene of interest.
- 3. Wild type D. vulgaris is electroporated with the construct from Step 2.
- 4. Positive strains are isolated followed by extraction of the chromosomal DNA.
- 5. Chromosomal presence of the tagged gene is confirmed by PCR/Southern analysis.

B. Tagged Protein Isolation and Identification

1. Mutant strains are grown until mid-late log phase.

- Cells are harvested and lysed using sonication. Protease inhibitors are added to the lysis buffer.
- Total soluble protein is estimated using the BCA assay.
- Extracted protein mixture is fed to a Strep-Tactin resin affinity column followed by gravity flow separation.
- 5. Each column is washed with wash buffer to remove non-specific proteins.
- 6. The tagged protein along with its interacting partners are eluted with a buffer containing a biotin
- analog (desthiobiotin).
- 7. Eluted proteins are either denatured and run on an SDS-PAGE Gel or are digested insolution via trypsin.
- 8. Bands observed after Coomassie-blue staining are excised and tryptically digested.
- 9. Digested peptides (in-gel or in-solution) are identified using nLC/MS/MS analysis.

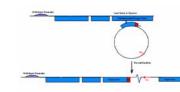
QUALITY CONTROL

- ✓ Sequence verification of PCR-tag constructs
- ✓ Single colony isolation of transformants
- ✓ PCR testing of transformants
- ✓ Southern analysis of transformants

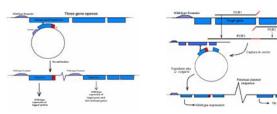
CONSTRUCTION OF TAGGED MUTANTS

Homologous recombination of gene with tagged sequence in circular plasmid with chromosomal copy of the target gene.

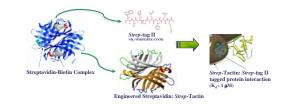
Scheme for last gene in an operon or monocistronic gene



Scheme for first gene in an operon Scheme for penultimate gene in an operor



THE STREP TAG II SYSTEM



· Based on the well known binding of biotin to streptavidin

Strep-tag II ®: consists of 8 amino acids (WSHPQFEK) and binds to the biotin pocket of streptavidin.
Engineered Streptavidin: Strep-Tactin ® has 100 times higher binding affinity to Strep-tag II than Streptavidin

® IBA GmbH, Germany.

STRAINS GENERATED

Strain Name	Target Protein	Gene ID	Description
JW907-0811	DnaK	DVU0811	Molecular chaperone
JW907-2928	RpoB	DVU2928	DNA-directed RNA polymerase, beta subunit
JW907-2929	RpoC	DVU2929	DNA-directed RNA polymerase, beta prime subunit
JW907-1577	HsIV	DVU1577	ATP-dependent protease
JW907-0503	Pnp	DVU0503	Polyribonucleotide nucleotidyltransferase
JW905-3184	Rub	DVU3184	Rubredoxin
JW905-0846	ApsB	DVU0846	Adenylylsulphate reductase, beta subunit
JW907-1568	Ftn	DVU1568	Ferritin
JW913-1397	Bfr 🚽 🚽	DVU1397	Bacterioferritin

Some interacting partners obtained

MODEL SYSTEM: DNA-directed RNA polymerase complex

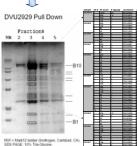
We generated the strain JW907-2929 to validate our protocol for protein complex identification in *D. vulgaris*. DVU2929 encodes for the DNA-directed RNA polymerase, RpoC, and occurs in a two gene



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operon preceded by one of its interacting partners, RpoB (DVU2928). The construct used for transforming the wild type strain was verified yo confirming the sequence of the tagged gene and the recombination was confirmed by southern analysis (left). The pull down experiment generated at least 10 distinct bands on an SDS-PAGE gel. The tagged protein (RpoC) as well as the expected interacting partners, RpoA and RpoB were all detected by MS-MS analysis of the tryptically digested bands.





PROTEIN-PROTEIN INTERACTIONS OBSERVED

core	# peptides	DVU#	Description	MW (Da)
farget: D	VU2928- DNA-	directed RI	NA polymerase, beta subunit (rpoB)	
43	8	01/12/22	transistion elongation factor Tu (tuf)	4385
265	2 6	DVU1834	pyruvate carboxylase, putative	13752
26		DV113025	pyruvate-ferredoxin coldoreductase (poR)	13289
22			DNA-directed RNA polymerase, beta subunit (rppB)	15387
222			sulfate adenvivitransferase (sat)	4806
171	1 9	DV01222	hypothetical protein	2447
17			DNA-directed RNA polymerase, alpha subunit (rppA)	3901
15		DUDING	chosphate acetyltransferase (pta)	7729
14			DNA-directed RNA polymeriase, beta prime subunit (rpoC)	14582
14			type I secretion outer membrane protein. ToIC family	5010
14		0001013	ron-sulfur cluster-binding protein	8091
14			conserved hypothetical protein	5049
11			conserved hypothetical protein chaperonin, 60 kDa (groEL)	5852
110				4254
	r 2	DVU3027	glycolate oxidase, subunit GlcD (glcD) MTH1175-like domain family protein	
10				5062
	VU0811-dnaK			
178			dnaK protein (dnaK)	6878
268			chaperonin, 60 kDa (groEL)	5852
24			MTH1175-like domain family protein	5062
24	5	DVU0812	heat shock protein GrpE (grpE)	2105
225	5	DVU1834	pyruvate carboxytese, putative 1	13752
17-			translation elongation factor Tu (tuf)	4365
17	4	DVU0847	adenylyl-sulphate reductase, alpha subunit (aprA)	7542
2	5 2	DVU2108	MTH1175-like domain family protein	1256
2	2	DVI10846	adenylylaulphate reductase, beta subunit (aprB)	1914
2	2	DVU0775	ATP synthase, F1 beta subunit (atpD)	5082
8			ATP synthase, F1 alpha subunit (atpA)	5471
facate D	UNITETT ATE	anandant.	protease (hslV)	
461	101311-011-0	Los a root to	carbohydrate phosphorylase family protein	2762
40	9 9	DV02349	carbonydrata prosphorytasa tamay protein pvruvate carbonylase, putative	13752
			de nucleotidytransferase (pro)	107.04
14rger: D			polyriborucleotide nucleotid/transferase (pro)	8243
81			ribosomal protein S1 (rpsA)	6489
54			pyruvate carboxytisse, putative	13752
42			translation elongation factor Tu (tuf)	4365
27			polyribonucleotide nucleotidythansferase (prip)	8243
243			ribosomal protein S4 (rpsD)	2417
23			ribosomal protein L2 (rplB)	3030
21			sulfate adanylyltransferase (sat)	4805
21	5	DVU1329	DNA-directed RNA polymerase, alpha subunit (rpoA)	3901
19	2	DVU3150	ribosomal protein S1 (rpsA)	6469
183	2 3	DVU2929	DNA-directed RNA polymerase, beta prime subunit (rpoC)	15562
168	3 3	DVU0607	adenosylhomocysteinase (ahcY)	5343
12		DVU2109	MTH1175-like domain family protein	5082
12	2 2	DVU0503	polyribonucleotide nucleotidytransferase (prip)	8243
100	2	01/10874	ribosomal protein S2 (rosB)	2982
8			incsine-5 -monophosphate dehydrogenase (guaB)	5258

We repeated the approach described above for four other targets - DVU2928, DVU0811. DVU1577 and DVU0503. Typically, 7-10 bands were observed for each of the pull down experiments except for DVU1577. The Table on the left lists significant protein hits that were identified from at least two peptides with MOWSE scores of 48 or higher. In all cases except for DVU1577 we were able to identify the tagged protein. The Table below lists proteins that were identified irrespective of the target protein chosen for the pull down experiment. These proteins might be nonspecifically interacting with the affinity column. Whether these are observed during pull down experiments with other targets remains to be seen

	Description	Average # peptides	Average Score	
DVU1044	nosine-5 -monophosphale dehydrogenase (guaB)	2	92	5255
	suffate adenylytransferase (sat)	4	173	4805
	pyruvate carboxylase, putative	6	283	13752
	MTH1175-like domain family protein	2	127	5082
	translation elongation factor Tu (tuf)	3	180	
DVU3025	pyruyate-ferredoxin coldoreductase (poR)	3	129	13269

We are currently in the process of tagging the various interacting partners observed for the aforementioned targets. Such cross-tagging experiments help confirm weak interactions as well as identify false positives. Expanding this analysis to the entire proteome of *D. vulgaris* will help us map the 'protein interactome' of this sulfate reducing bacterium.

ACKNOWLEDGEMENT

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