

*In vitro and in vivo*  
molecular imaging

Gulliver multi-scale Imaging Workshop  
18 May, 2007

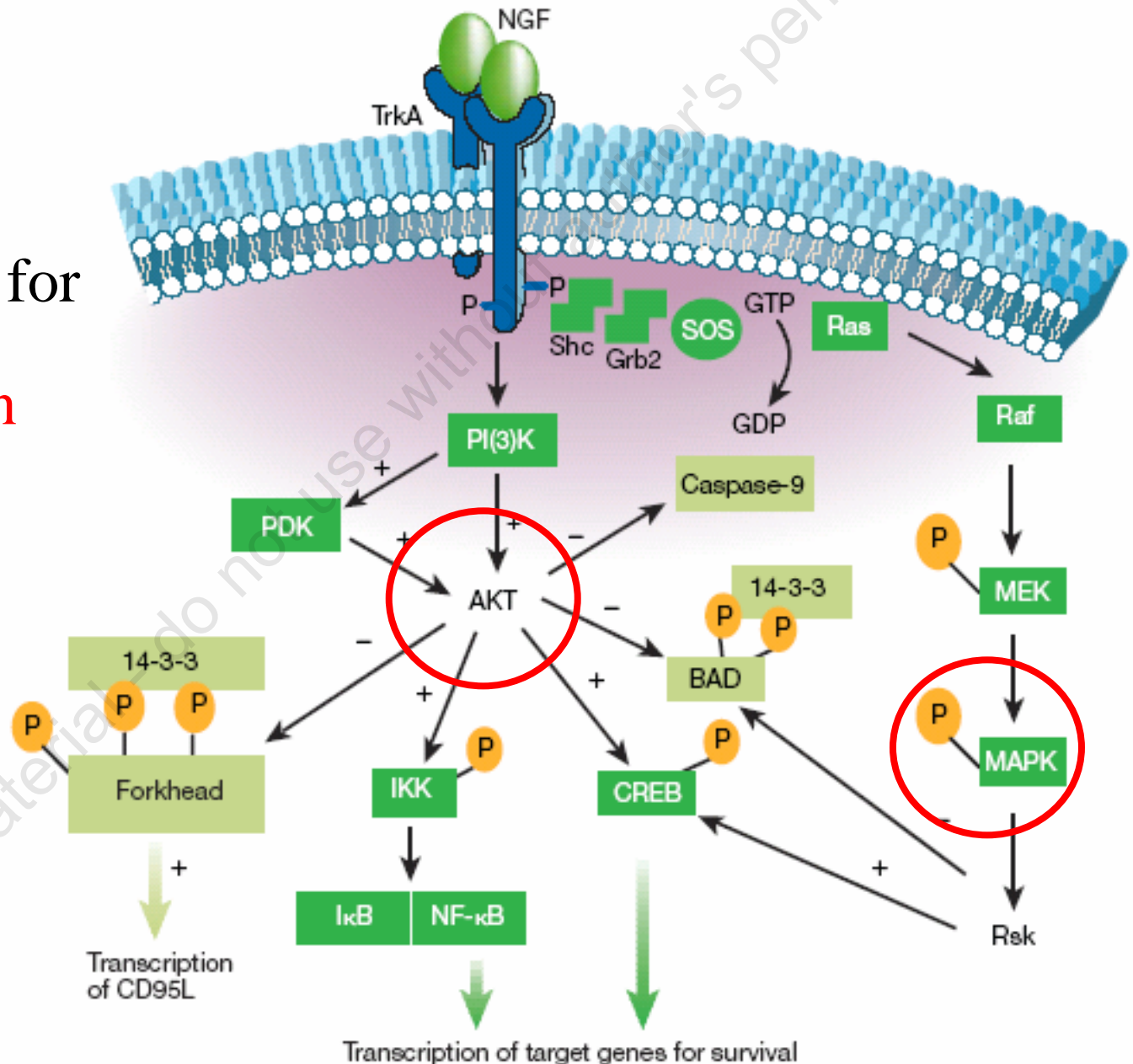
*in vivo* imaging of NGF  
retrograde transport in neurons

**Bianxiao Cui**, Chengbiao Wu, Liang Chen,  
Alfredo Ramirez, Bill Mobley, Steve Chu

# Neural Growth Factor signaling pathways

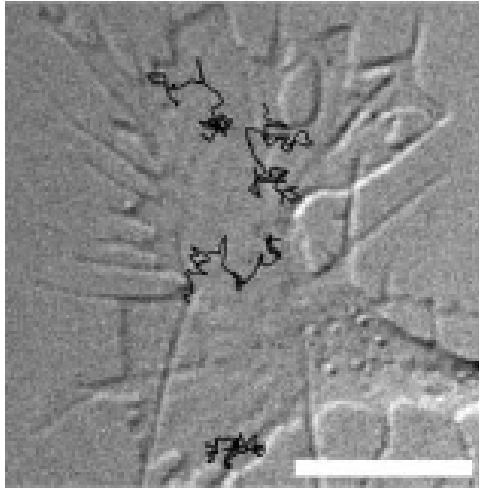
NGF is required for

- Differentiation
- Survival
- Maintenance
- Repair

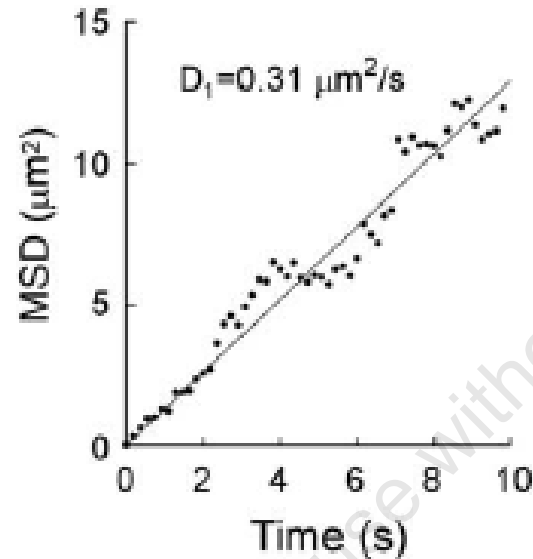


T. Tani, et al. J. Neuroscience **25**, 2181–2191 (2005)

A Lateral diffusion

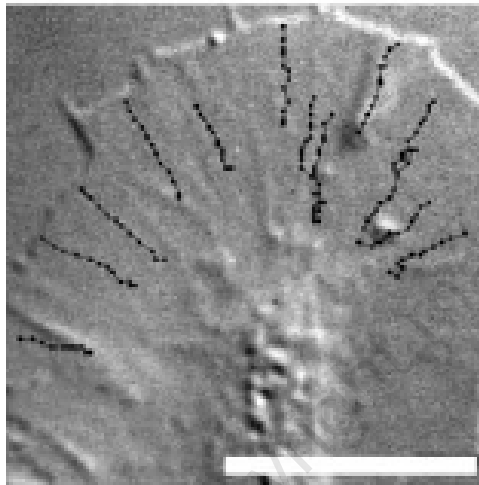


C

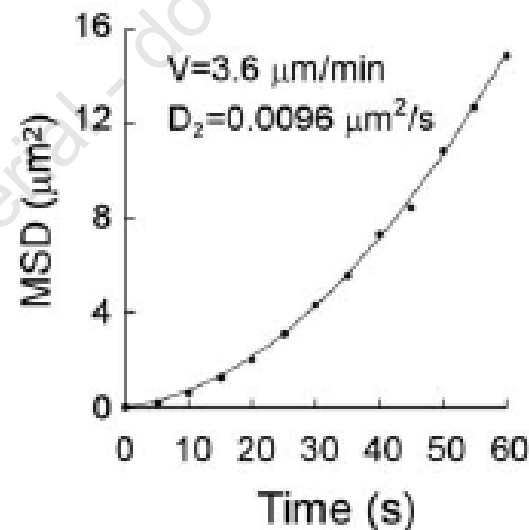


After binding to the receptor, Cy3-NGF diffuses on the membrane of the growth cones

B Rearward movement

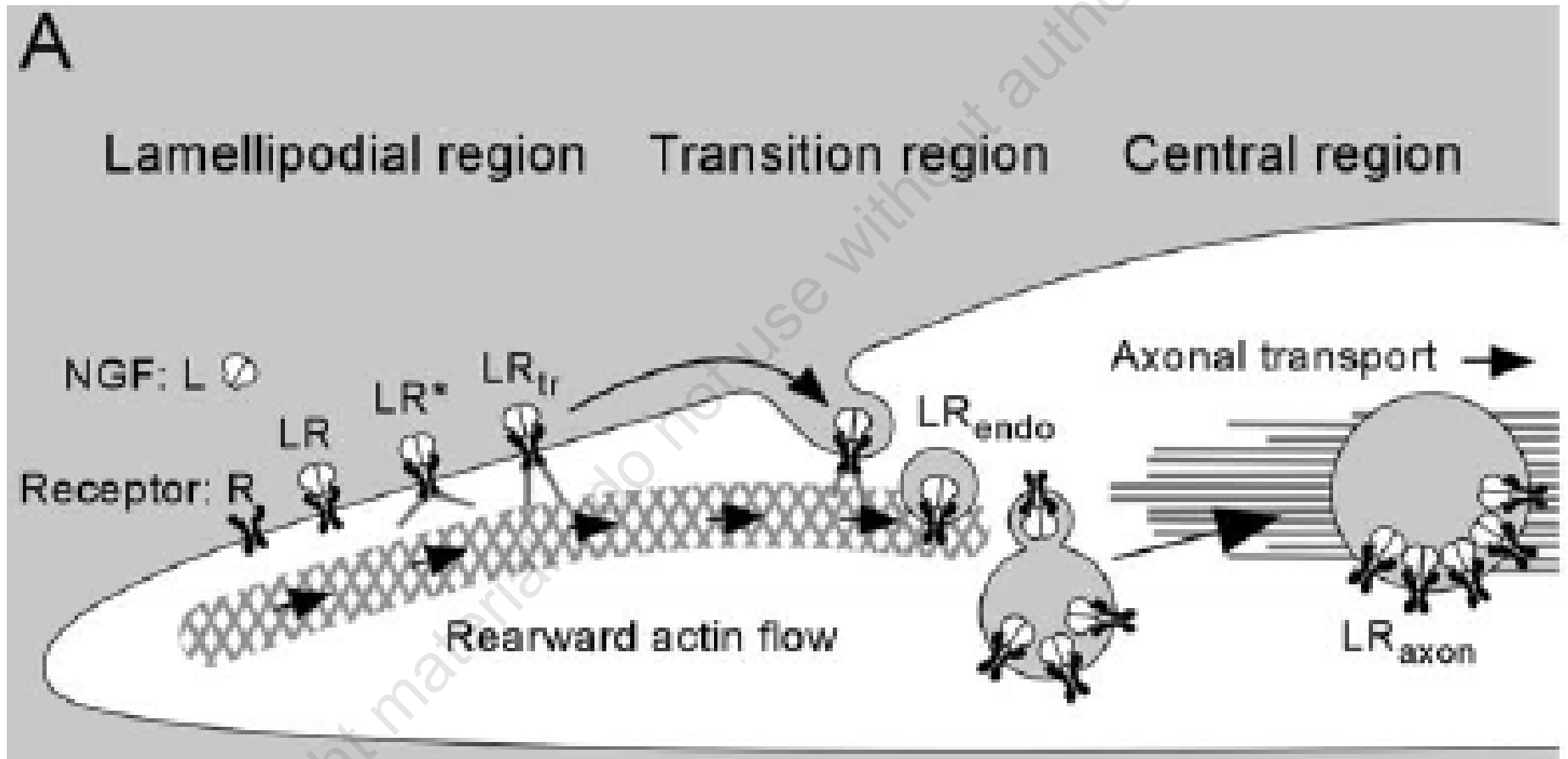


D

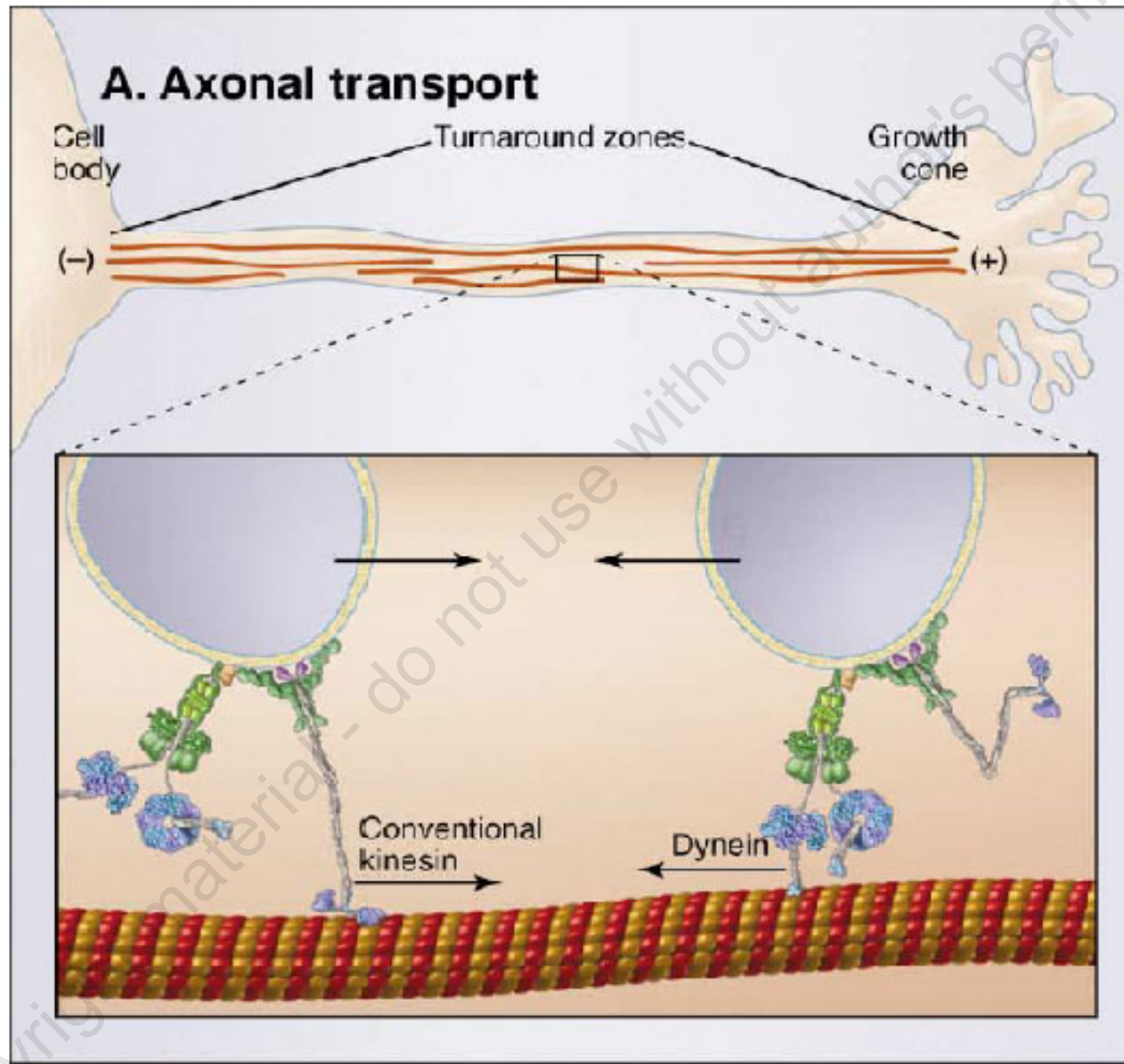


Motion then becomes one-directional toward center of growth cone. Movement could be stopped by adding latrunculin B, a potent inhibitor of actin polymerization.

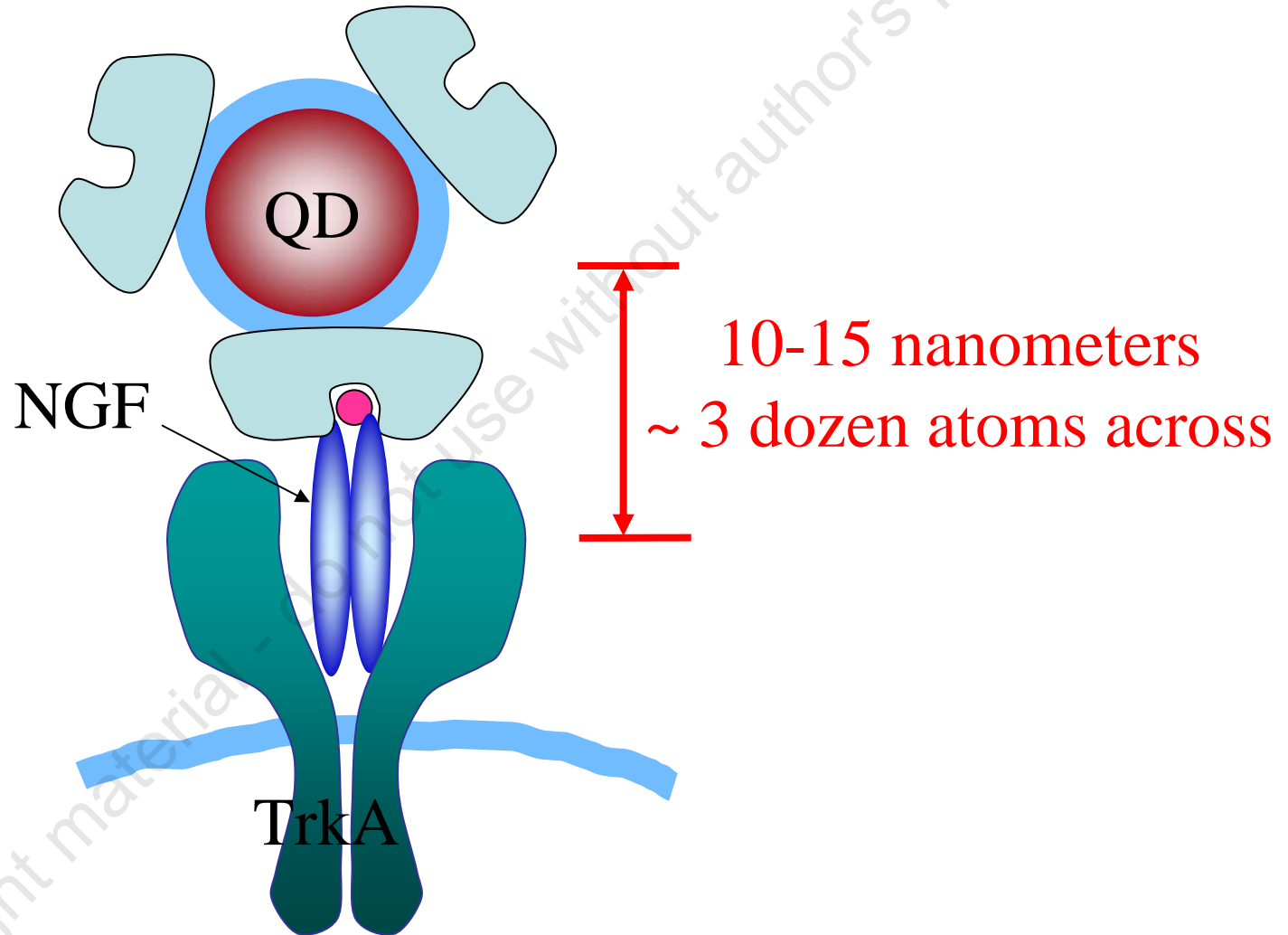
# Model of NGF binding, trafficking, and uptake at the dorsal root ganglion growth cone.



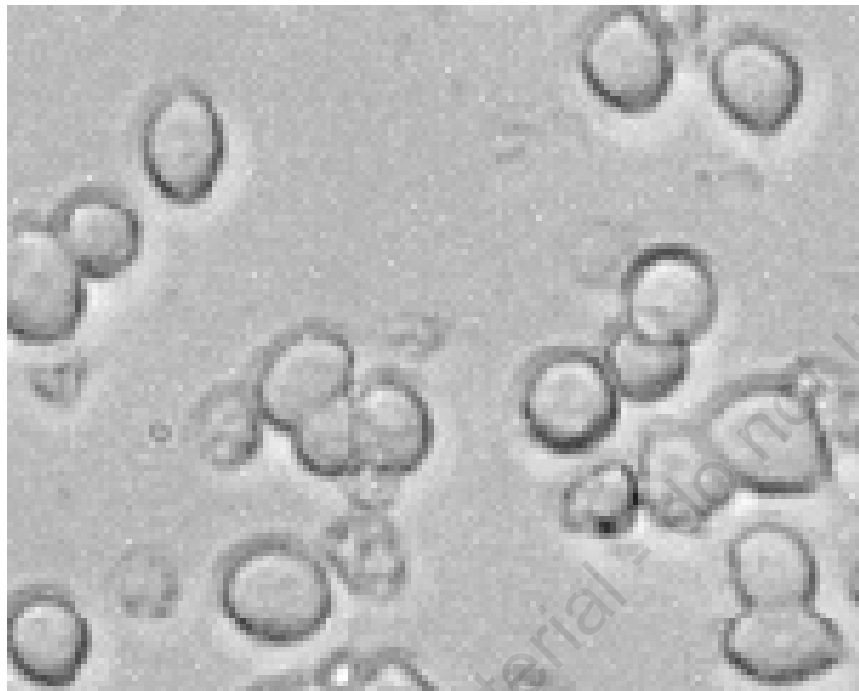
# Axonal transport



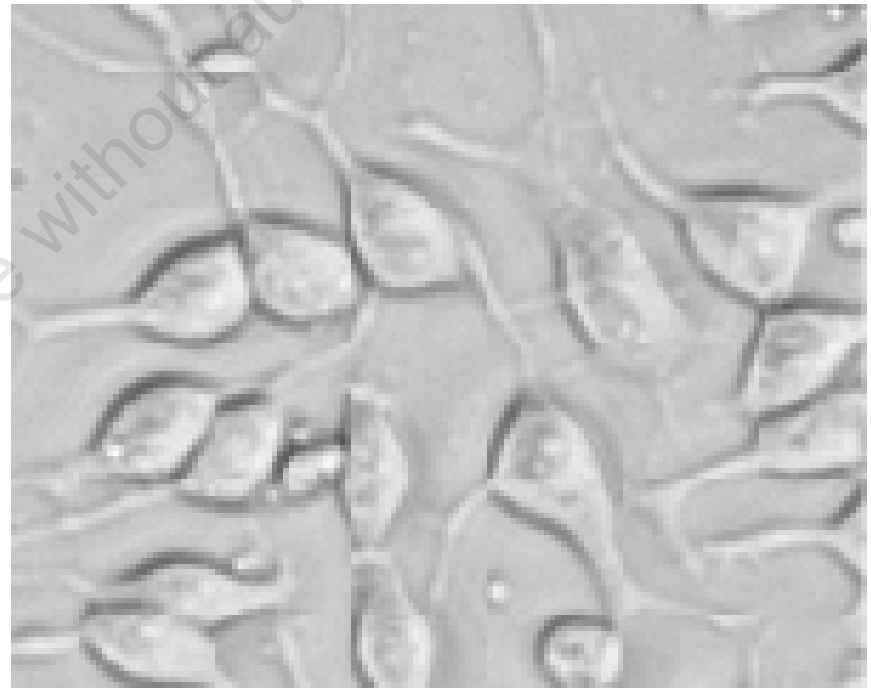
# Quantum dot - Neuro-growth Factor



# Qdot-NGF stimulates neurite outgrowth in PC12 cells



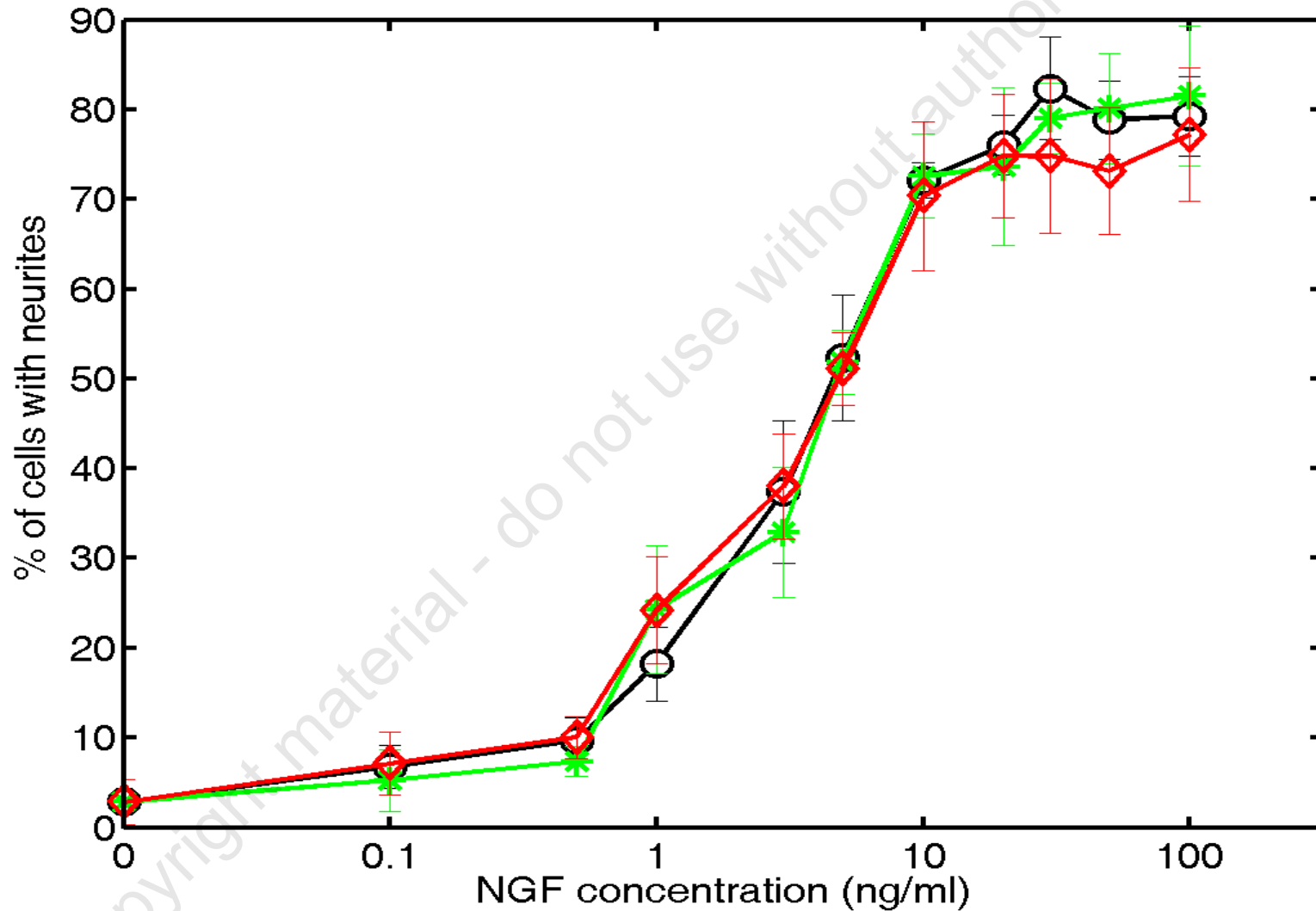
No NGF treatment



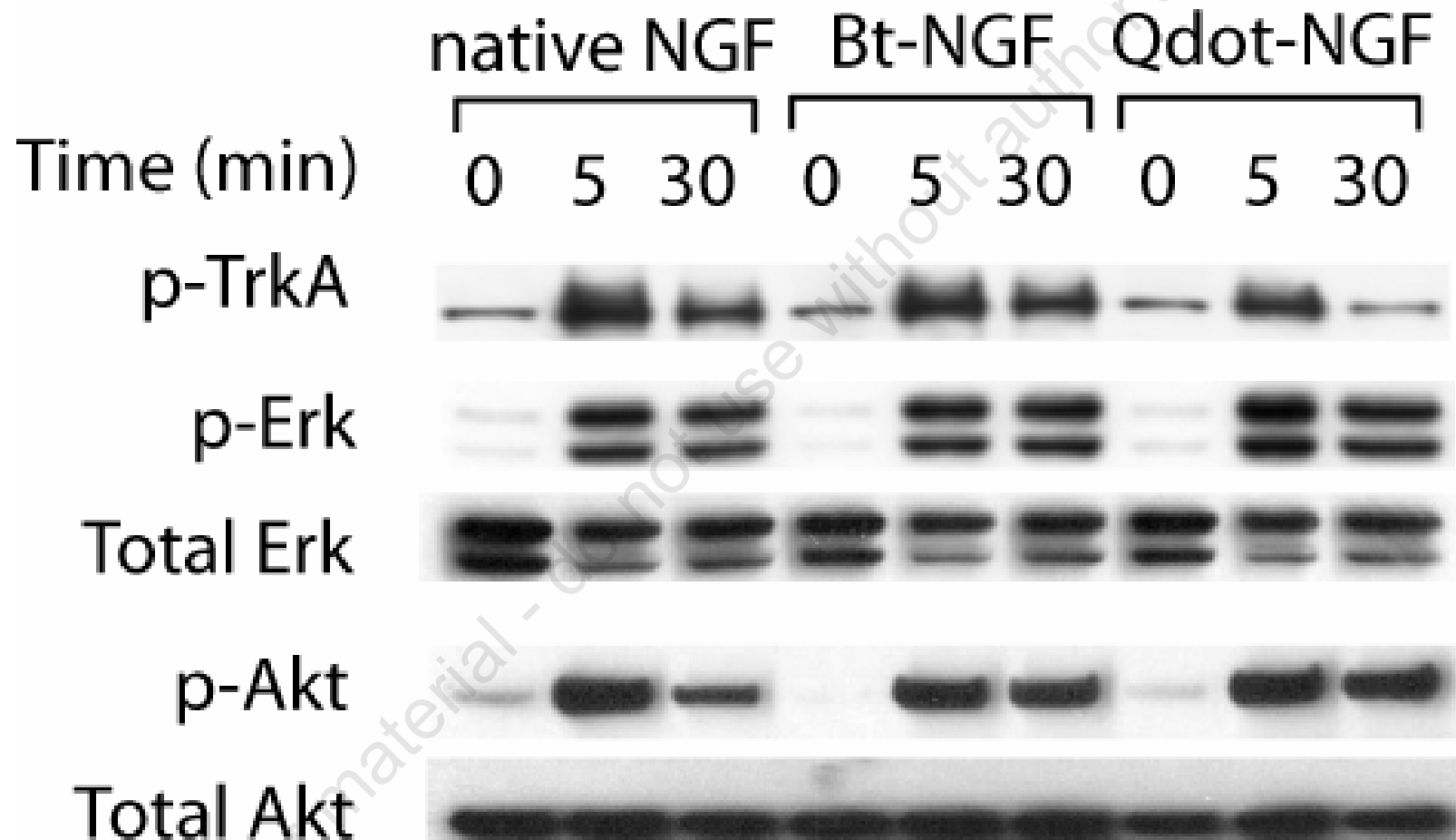
Treated with 20ng/ml Qdot-NGF  
for 48hrs



# Dose response of Qdot-NGF, Bt-NGF and native NGF

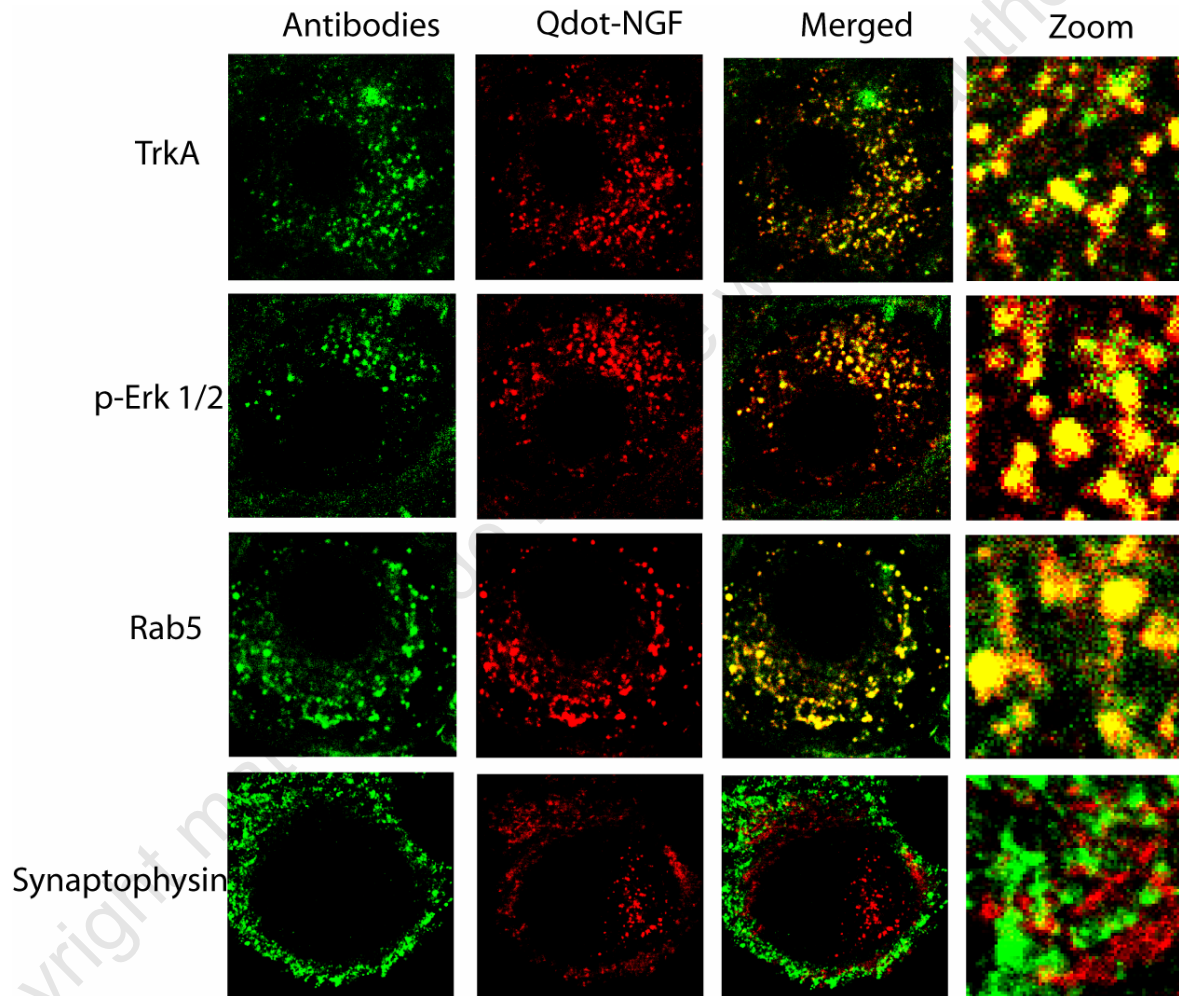


# Qdot-NGF activates signaling proteins

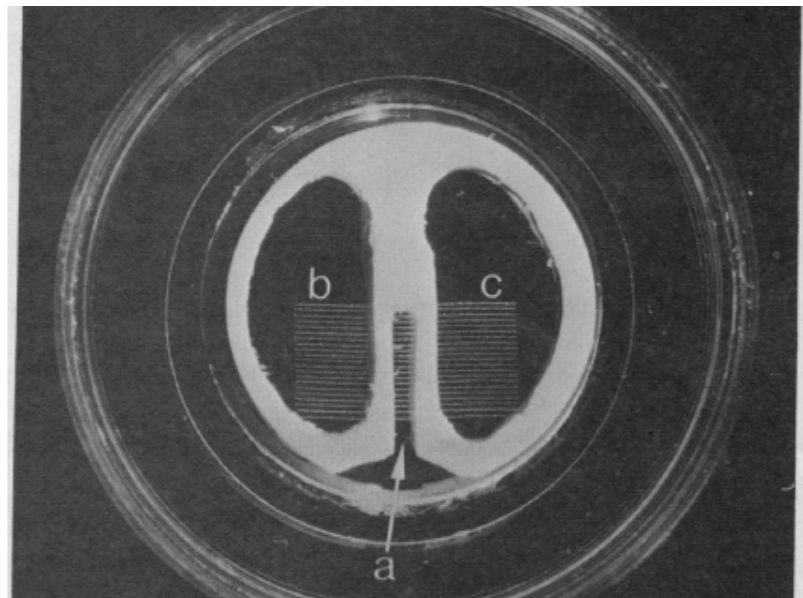
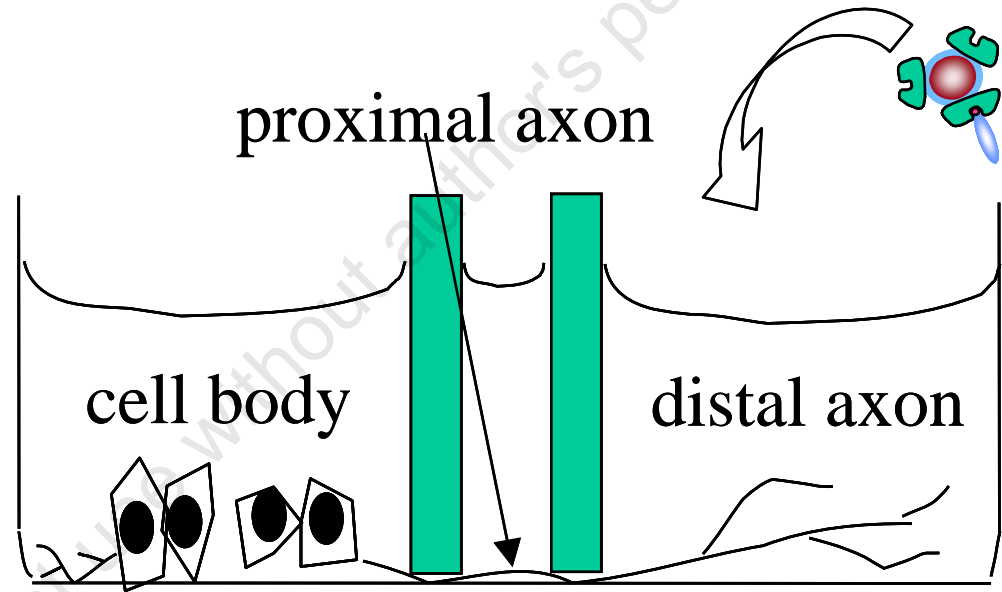
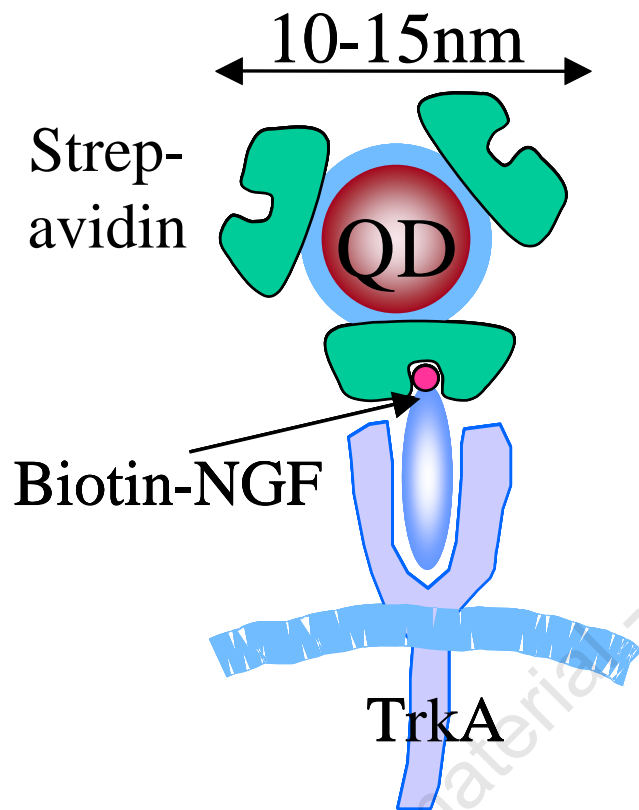


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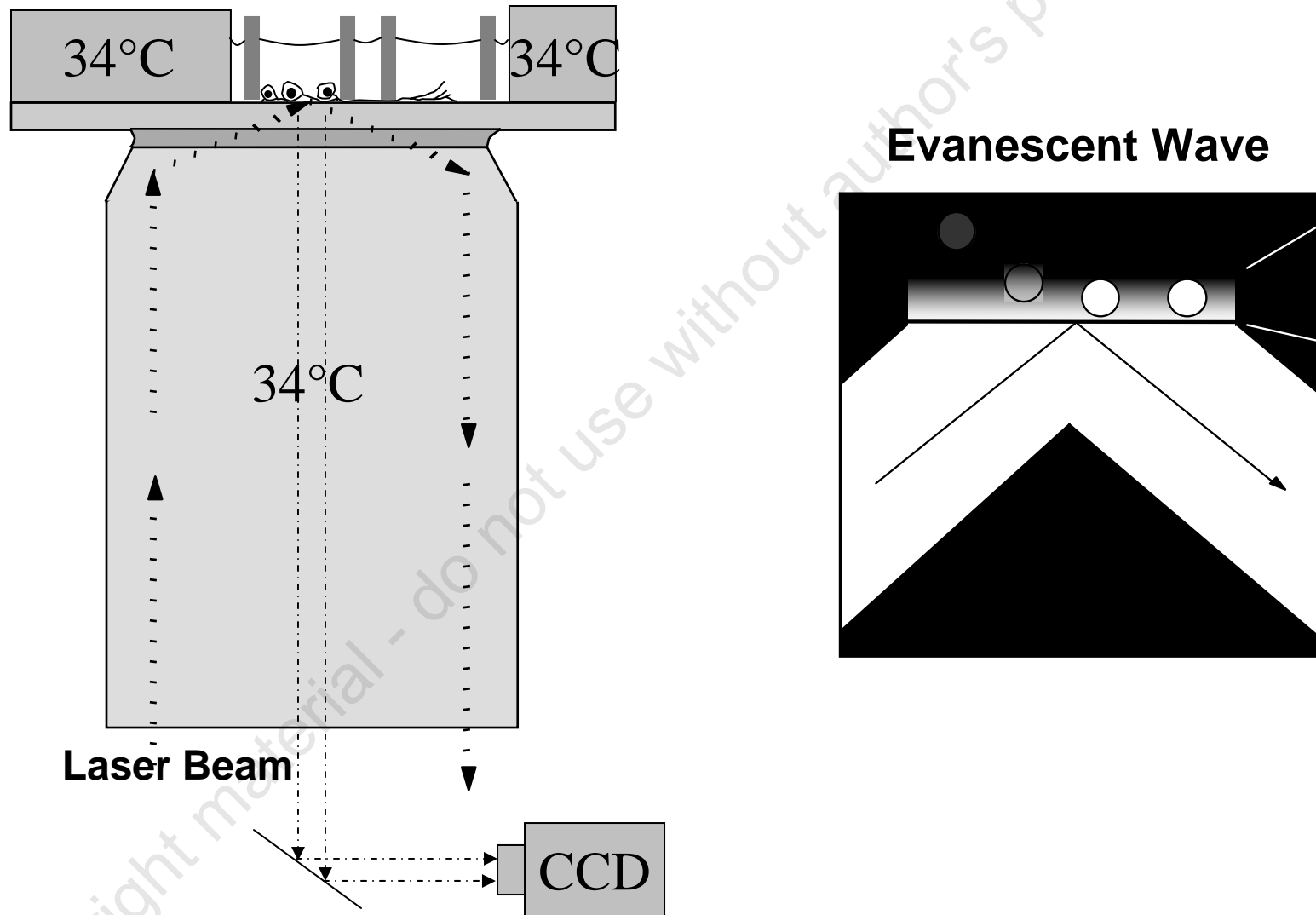
# Co-localization of Qdot-NGF with signaling proteins



# Q-dot tracking of NGF motion

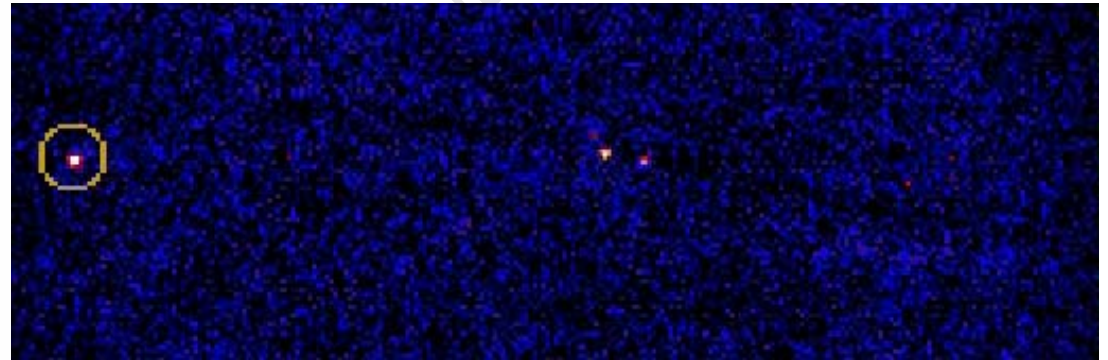
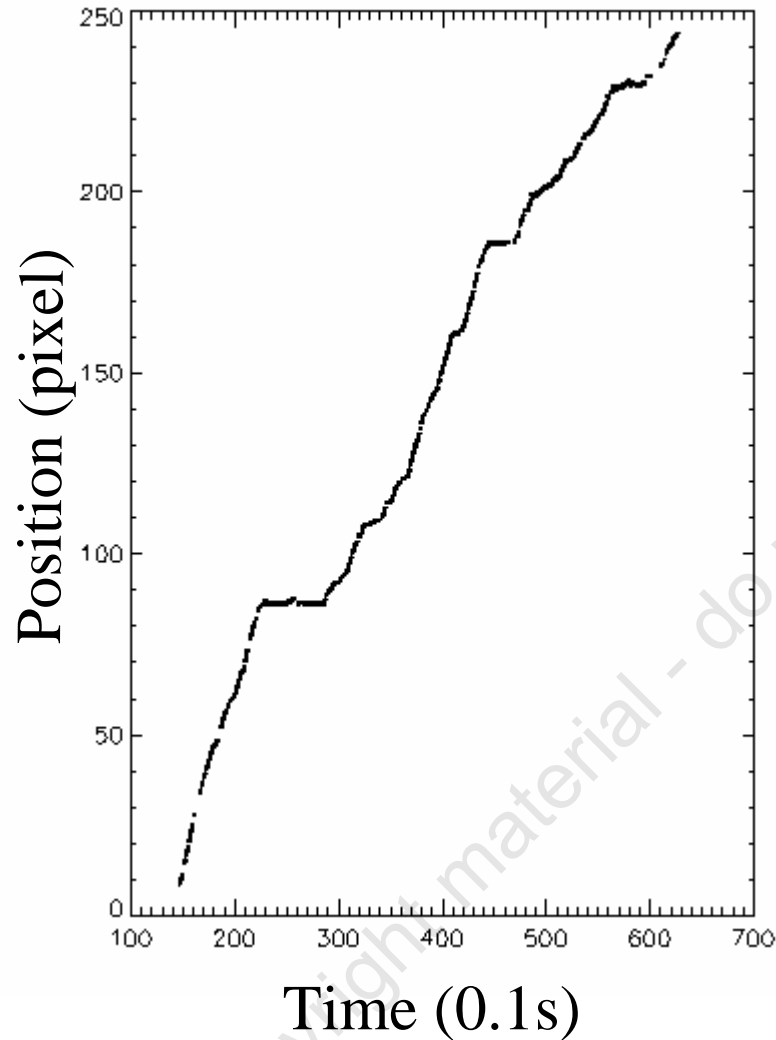


# Live imaging experiment setup



Total Internal Reflection Microscope

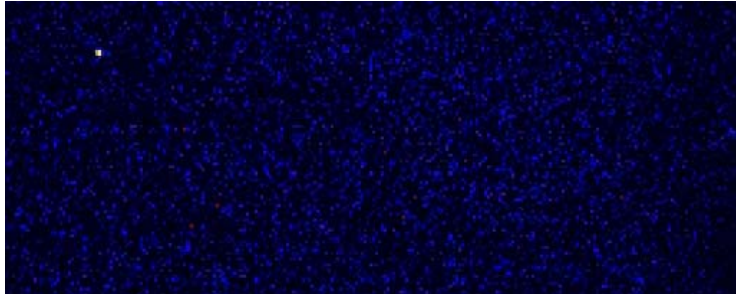
# Tracking an individual endosome



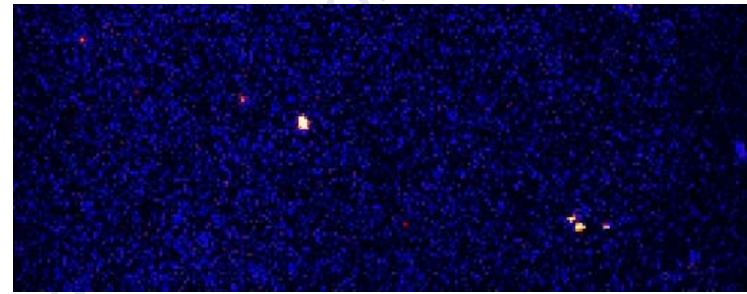
Time corresponds to 30  
seconds of real time  
(~5x faster)

# Live imaging of NGF transport

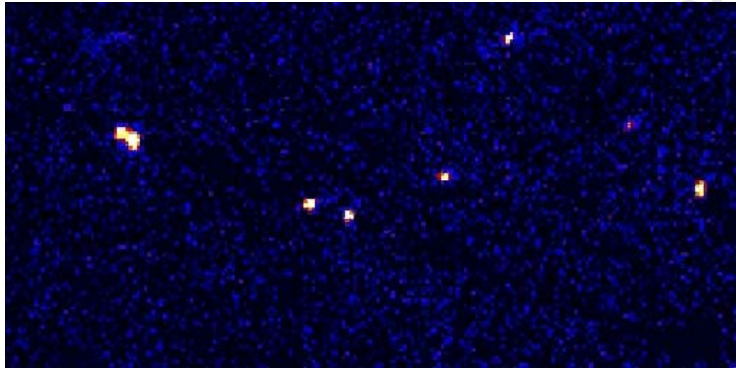
Cui, Wu, Chen, Ramirez, Mobley, Chu



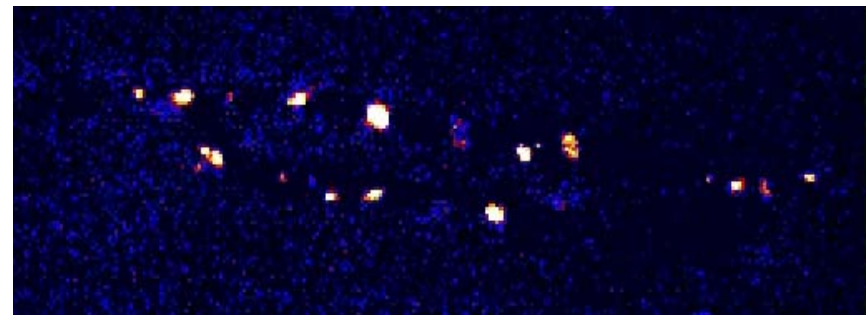
5 ng/ml NGF



25 ng/ml NGF



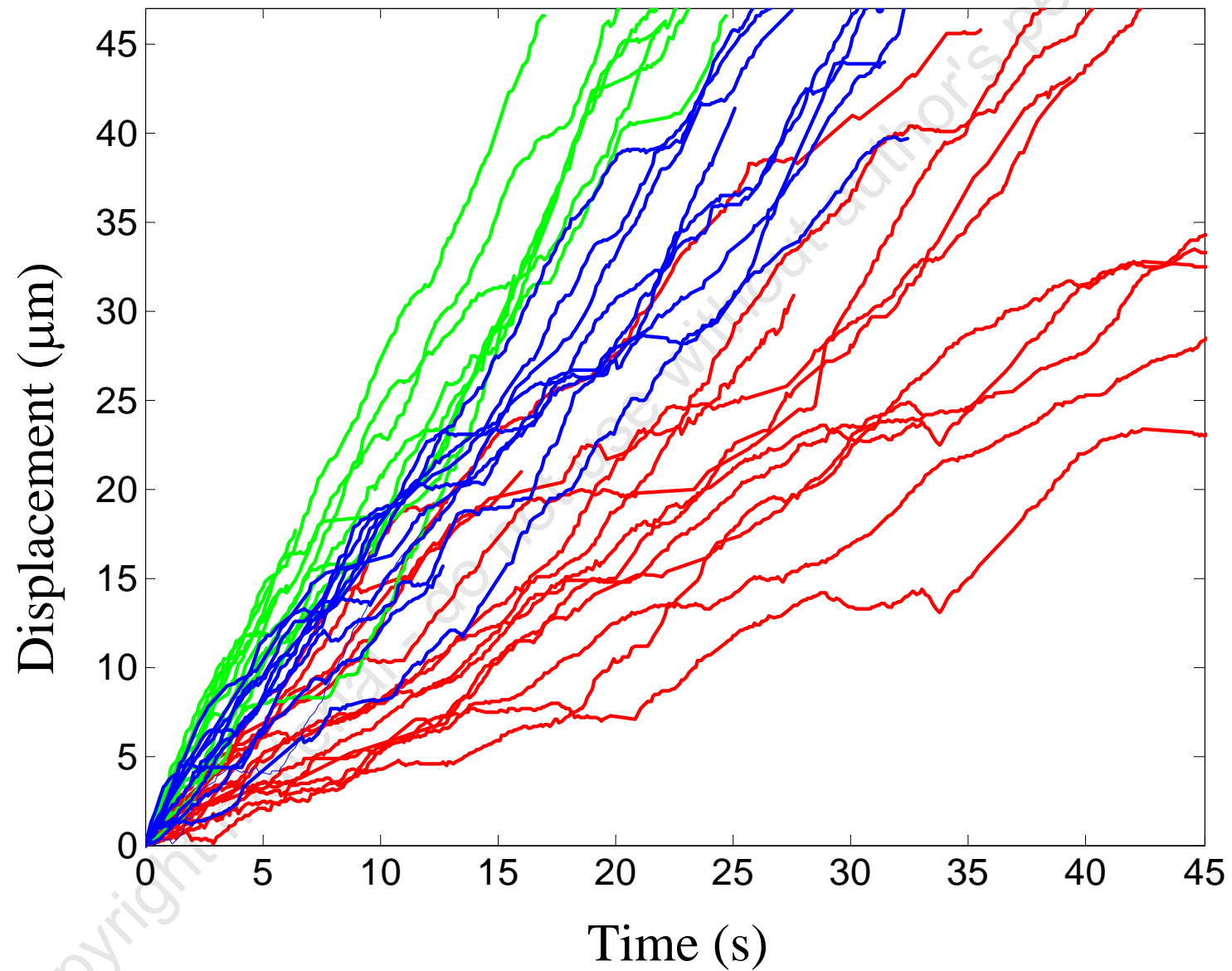
50 ng/ml NGF



500 ng/ml NGF

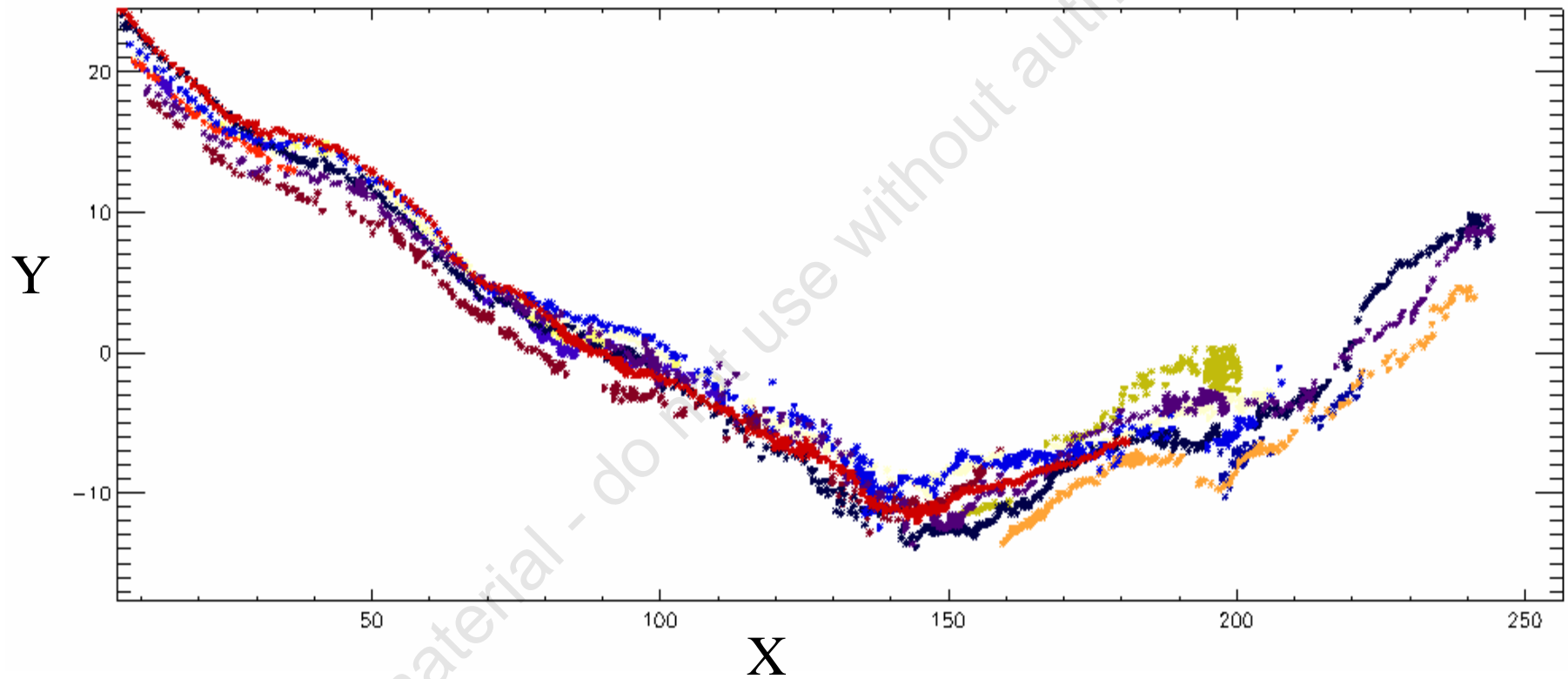
Movies correspond to 30s real time

# Endosomes moving in different axons

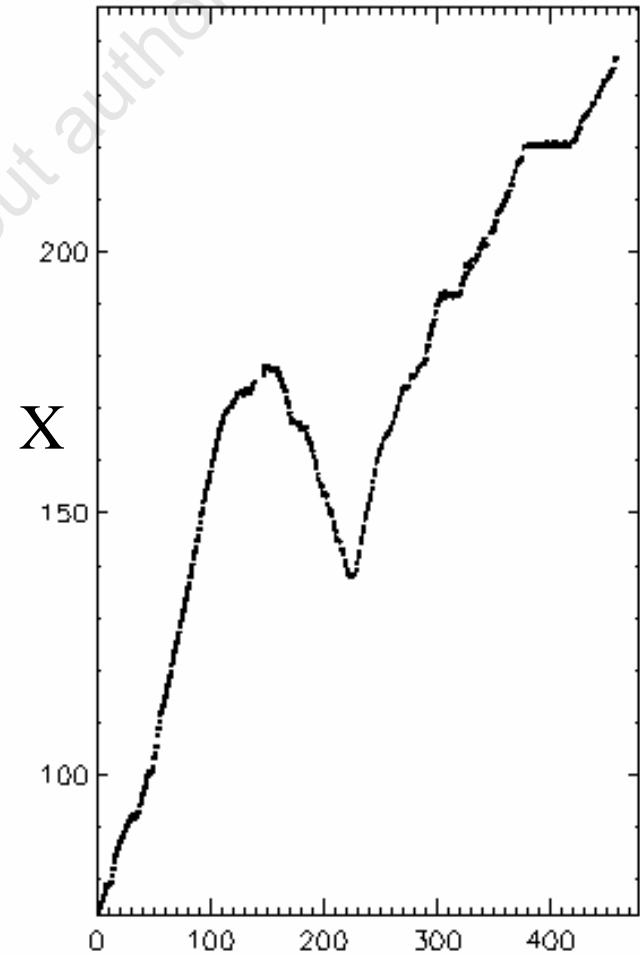
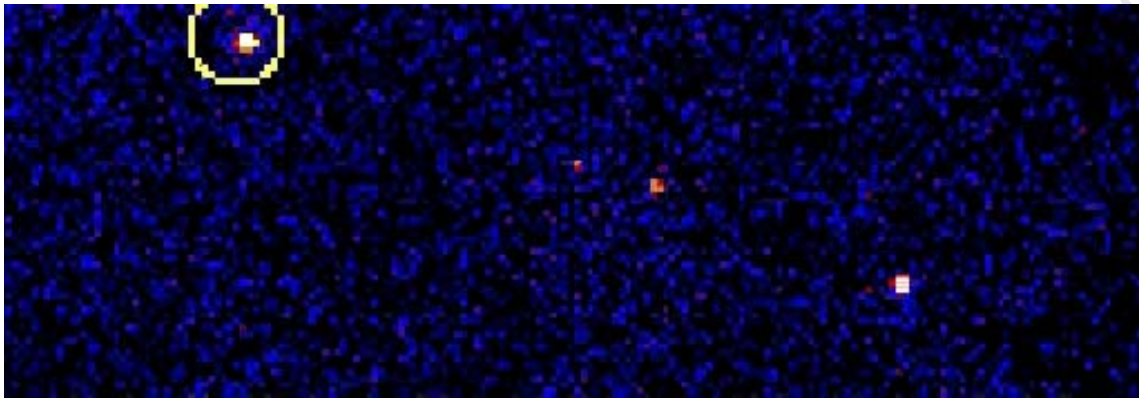




# Each axon is a multi-lane highway

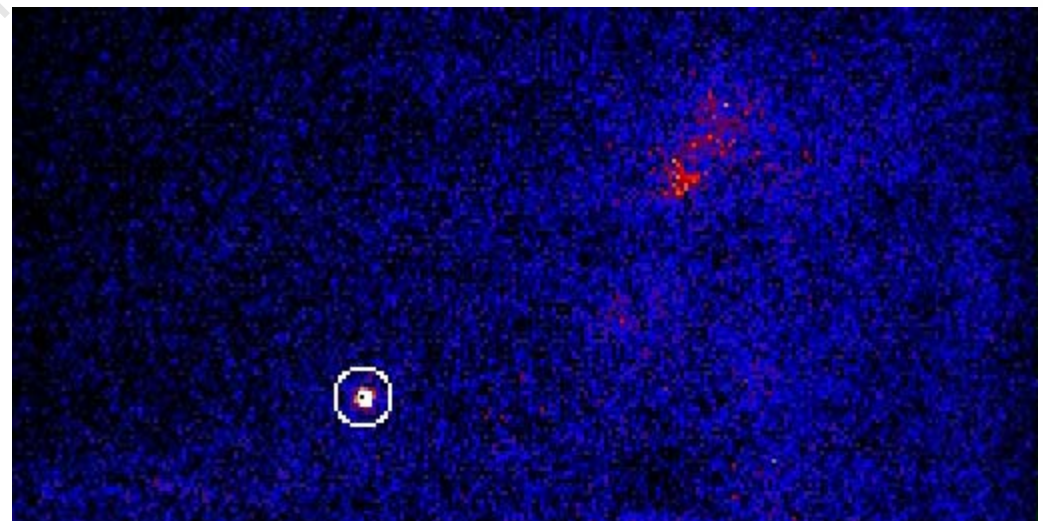
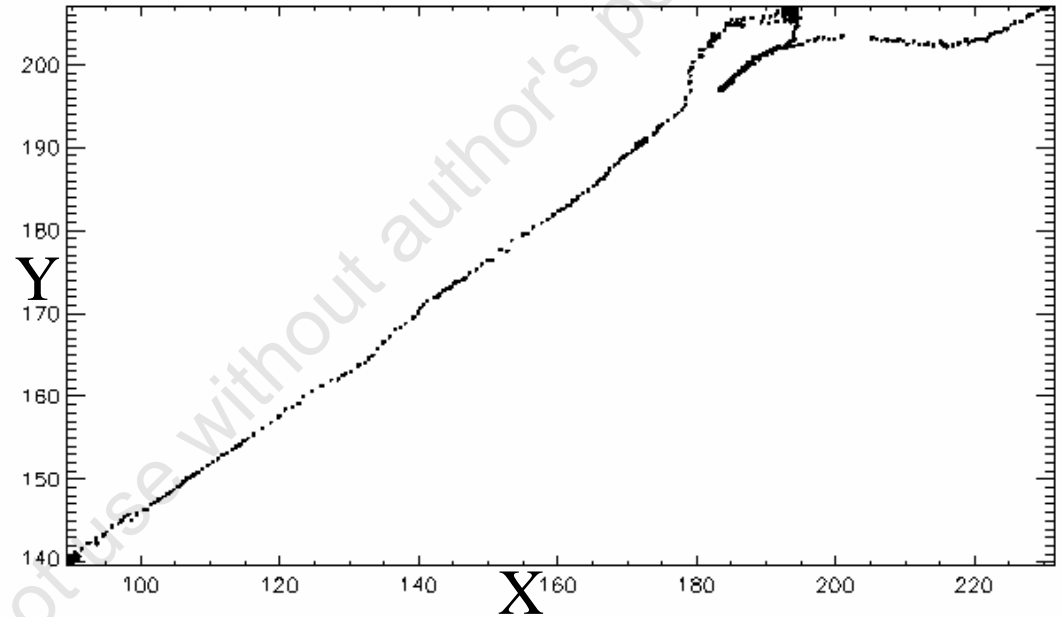
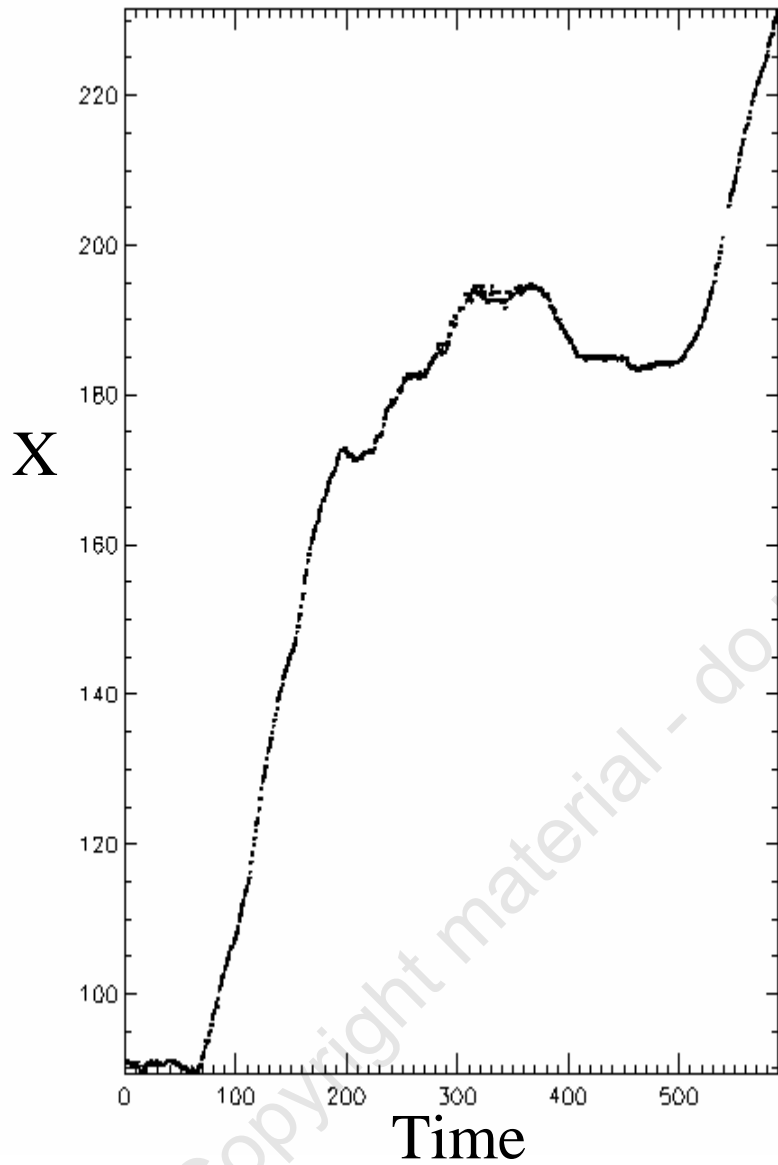


# Anterograde motion

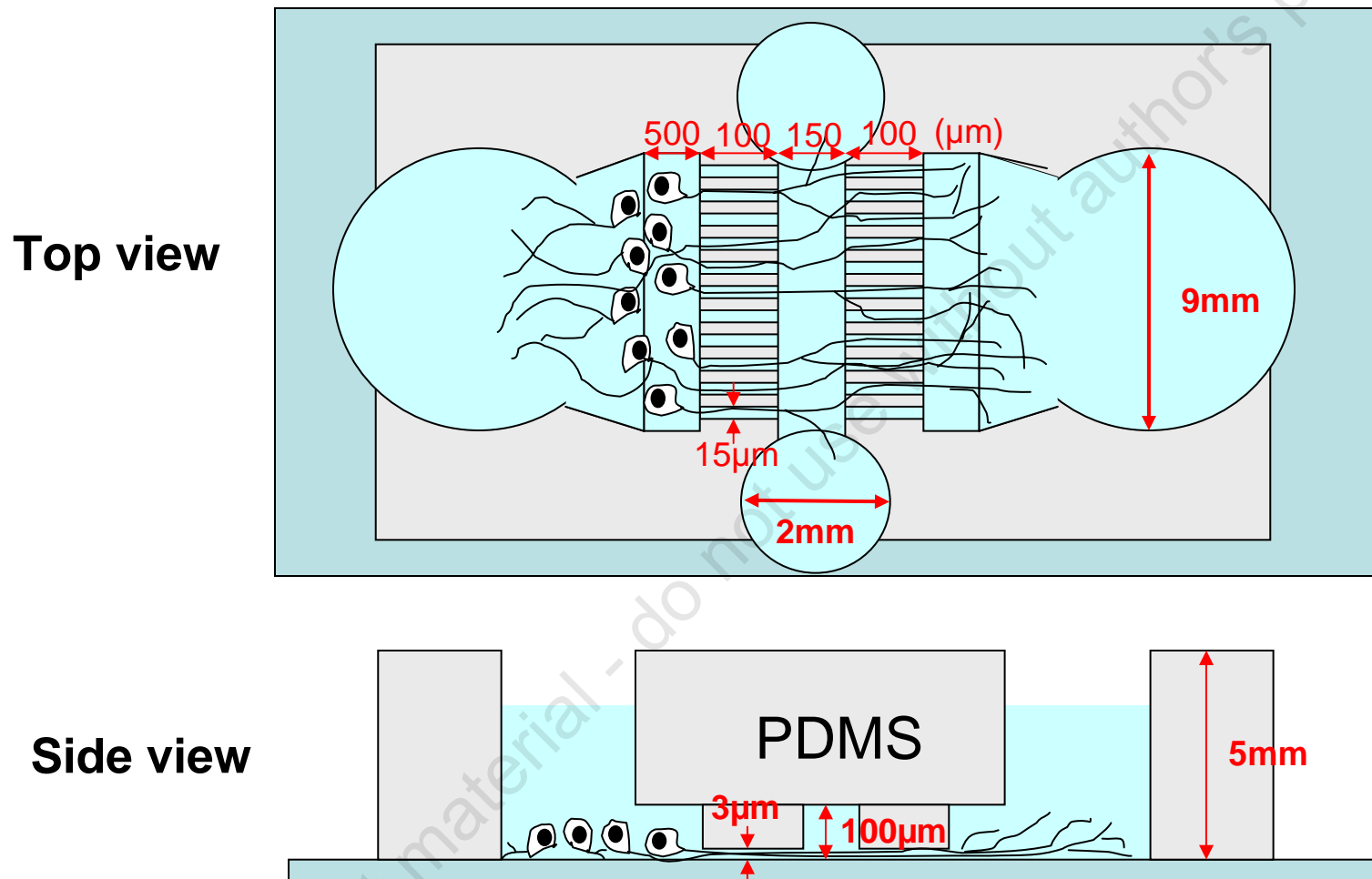


Time

# Endosome changing microtubules?

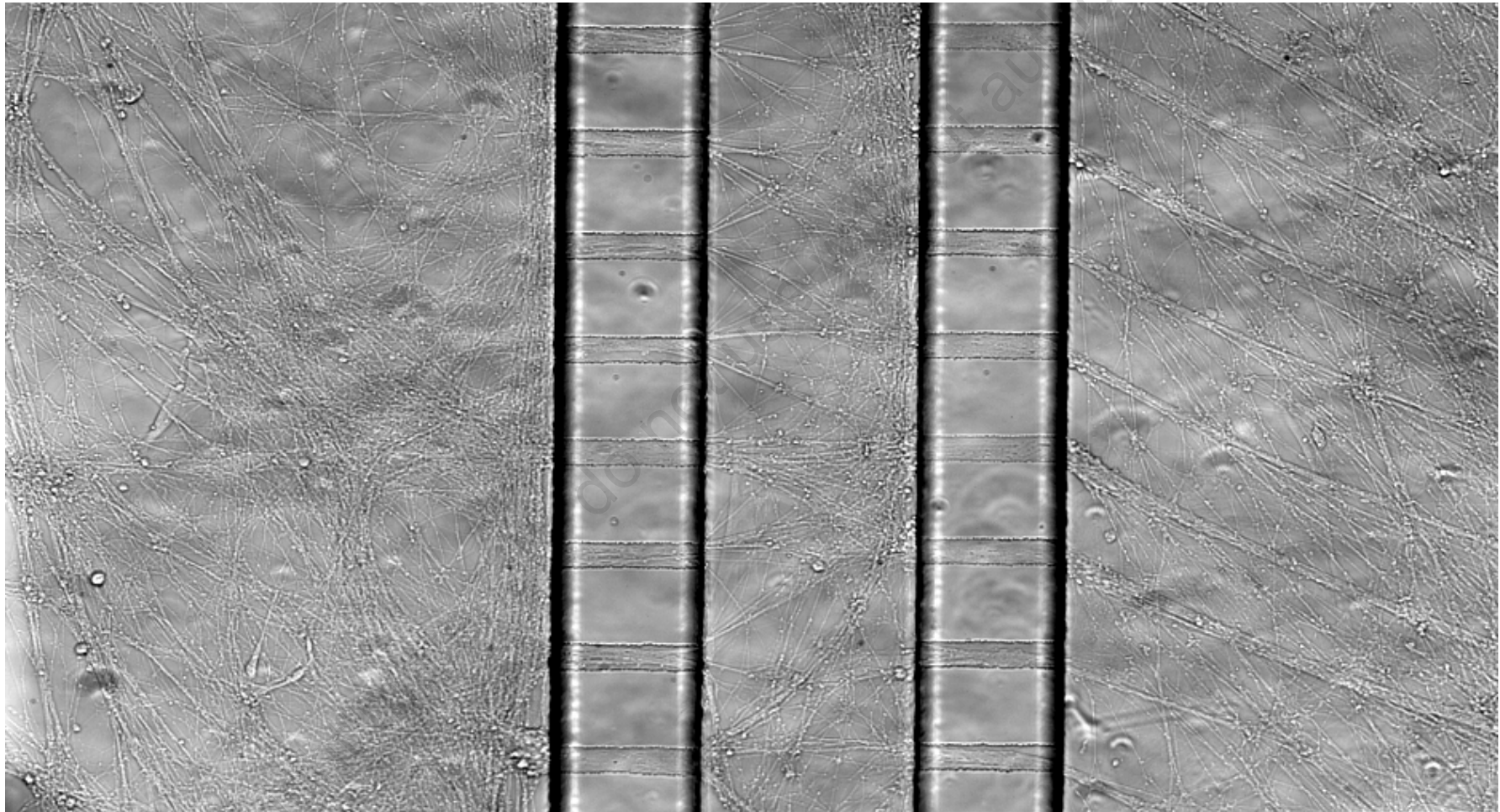


# Microfluidic device diagram



Taylor, AM, et. al. *Langmuir*, **19**, 1551-1556, 2003; *Nature Methods*, **2**, 599, 2005.

# Three-chamber – day 10



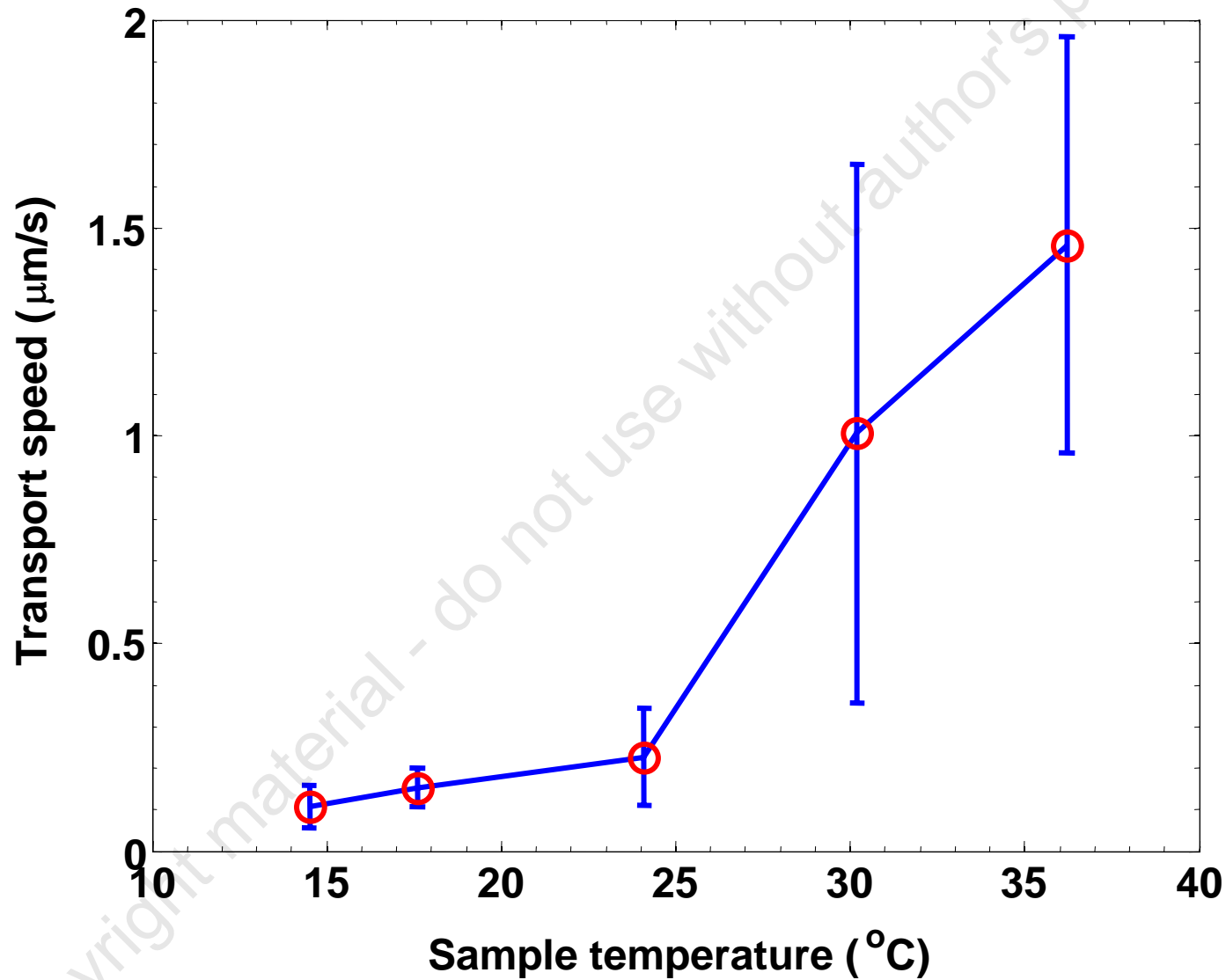
Distal axon chamber

Cell body chamber

# Micro-fluidic advantages

- Axons tend to grow along the flow direction
- No leaking of fluorescent Qdots was detected, even after overnight incubation of 100ng/ml Qdot-NGF in the distal axon well.
- Rat embryonic DRG, cortical and hippocampus neurons, and mouse P0 DRG neurons, have been cultured in microfluidic devices and extend axons to the distal axon well.

# Transport Speed vs. Temperature



# Summary

1. NGF-containing endosomes move retrogradely along axons to cell bodies.
2. Most endosomes exhibit "stop-and-go", unidirectional motion at an average **moving speed** of up to 2.1  $\mu\text{m/s}$ .
3. Motion reversal is seen.
4. A single NGF dimer is sufficient to induce retrograde transport of signaling event.
5. The speed of transport varies by more than an order of magnitude between  $14^\circ$  and  $37^\circ$  C.



*in vivo* imaging and retrograde  
transport of Trk\_A in neurons

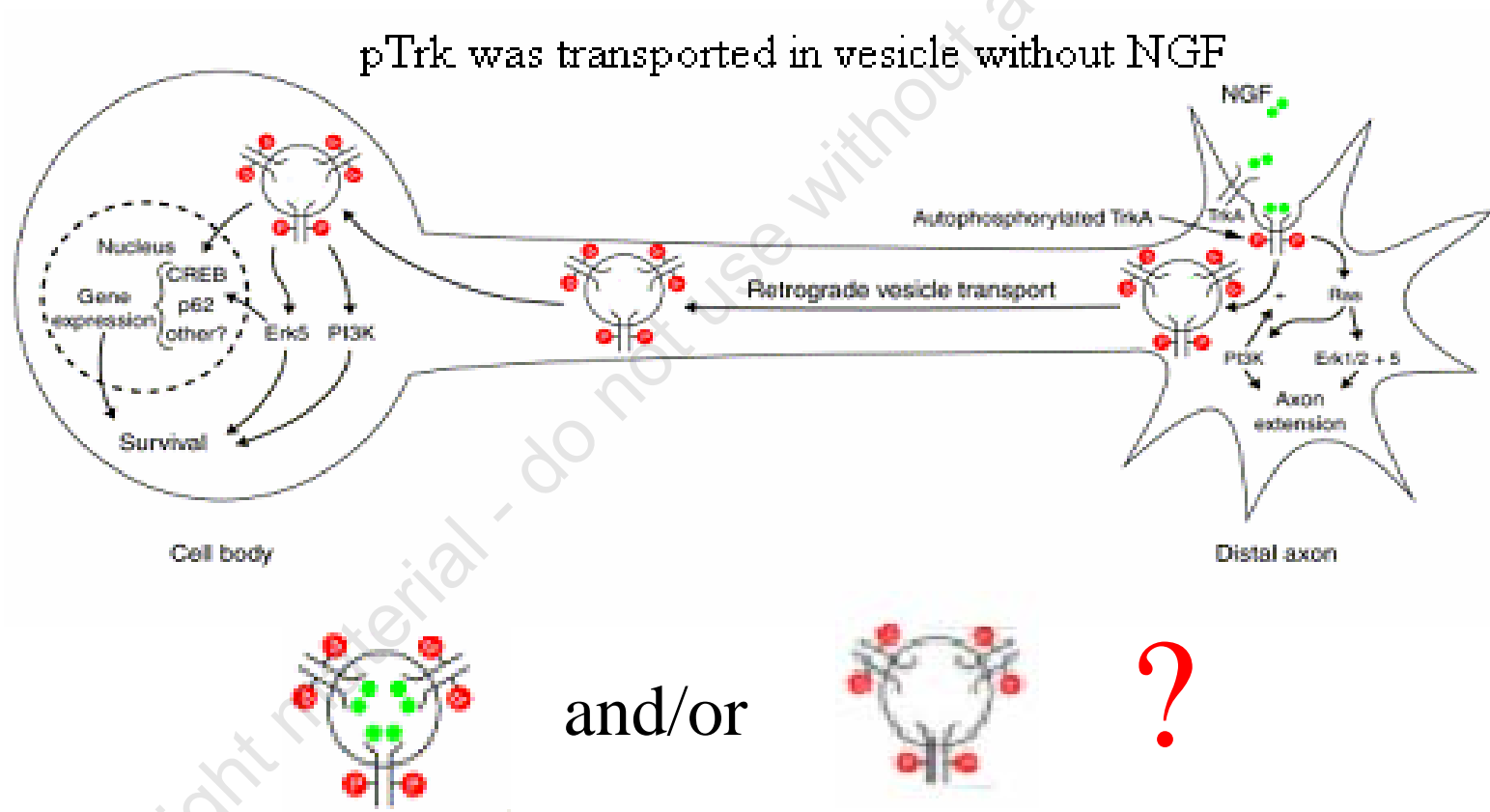
**Liang Chen**, Bianxiao Cui, Steve Chu

Chengbiao Wu, Alfredo Ramirez, William C.  
Mobley

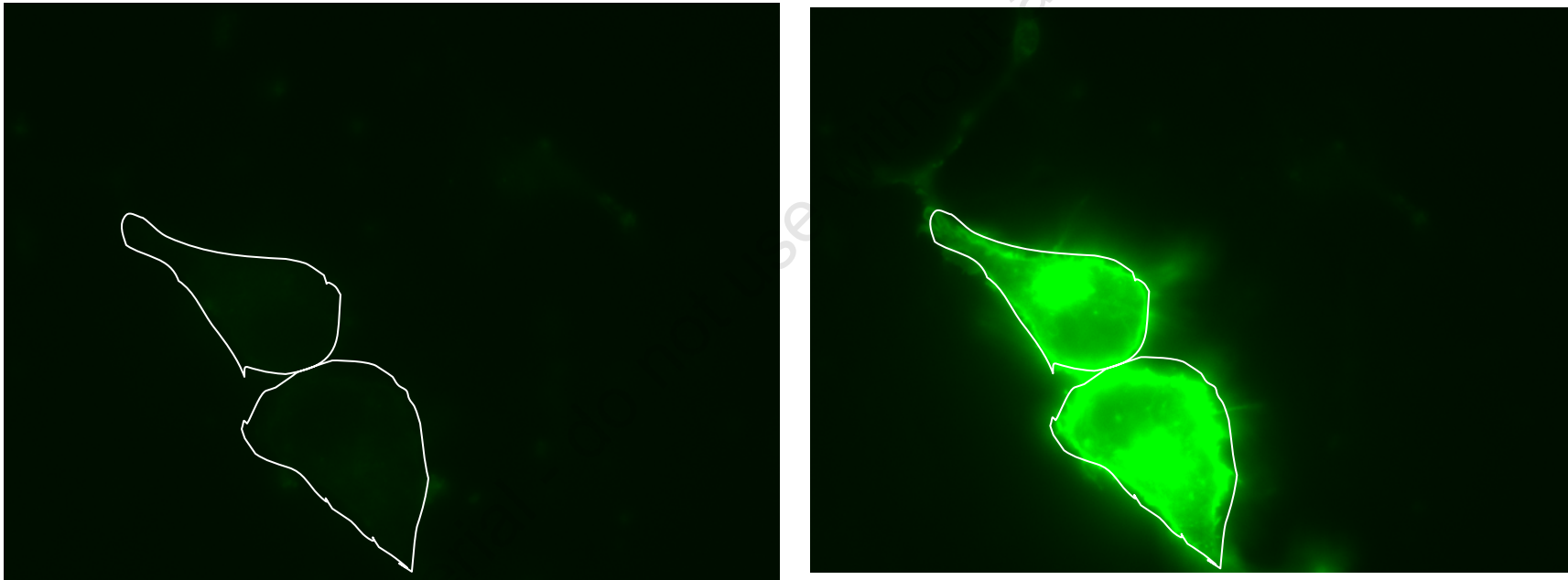
# Is pTrk-A, activated by NGF transported to the cell body?

Senger and Campenot, *JCB* 1997

MacInnis and Campenot, *Science*. 2002



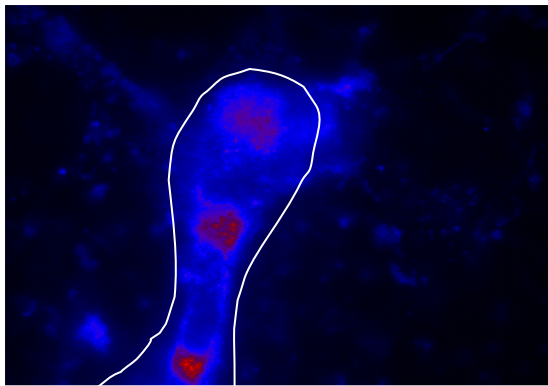
# TrkA\_PAGFP



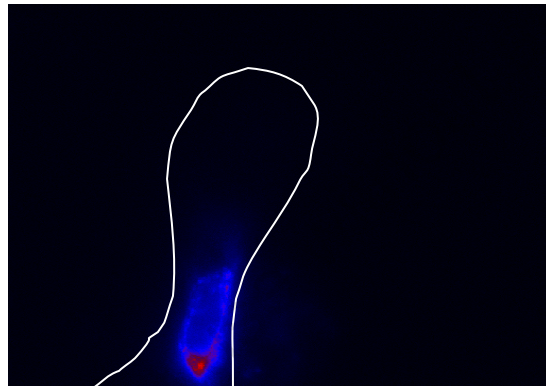
1s after UV excitation

Copyright material

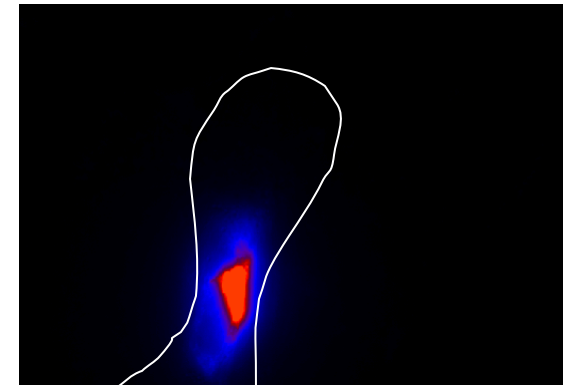
# PAGFP can be partially excited



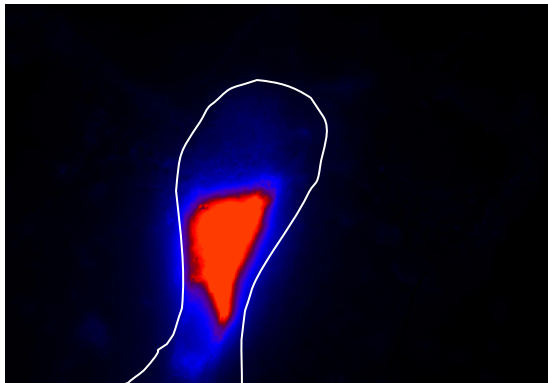
1



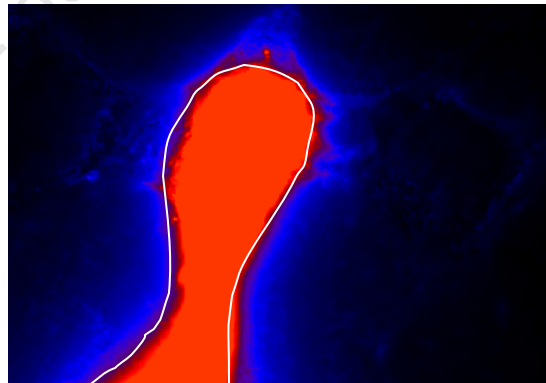
2



3

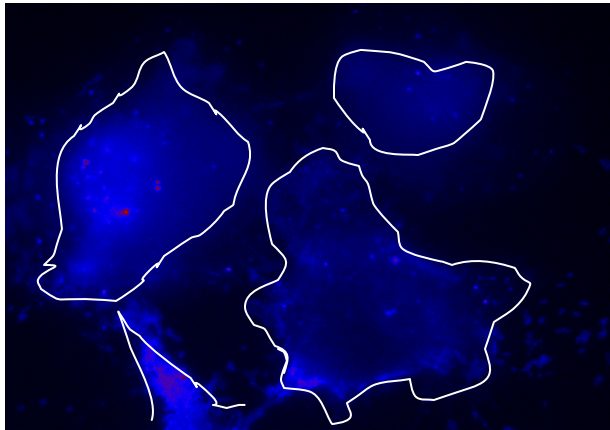


4

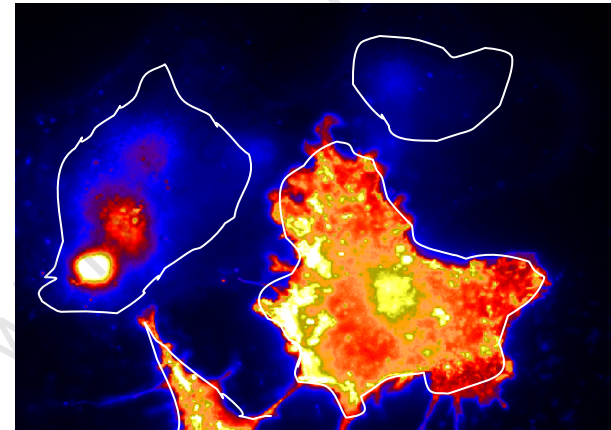
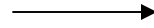


5

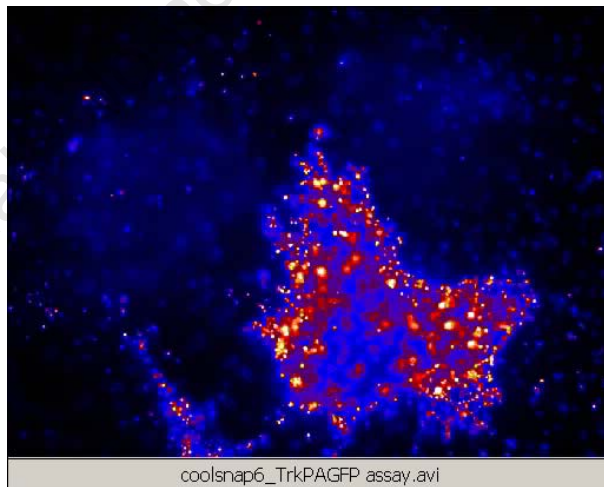
# NGF binds to TrkA\_PAGFP specifically on surface of Cos7 cells



a) PAGFP before UV excitation



b) PAGFP after UV excitation



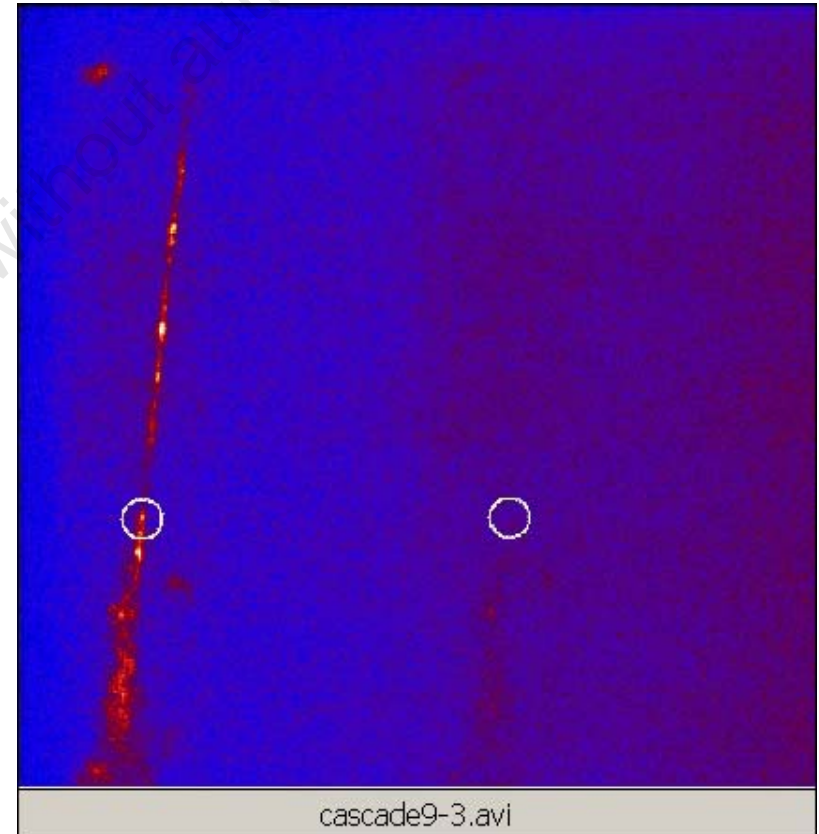
c) QD 605 filter

# Bi-directional transport of TrkAEGFP

W/O NGF

Cell body

↑  
Axon



TrkA\_EGFP

QD605

TrkA\_EGFP

QD605

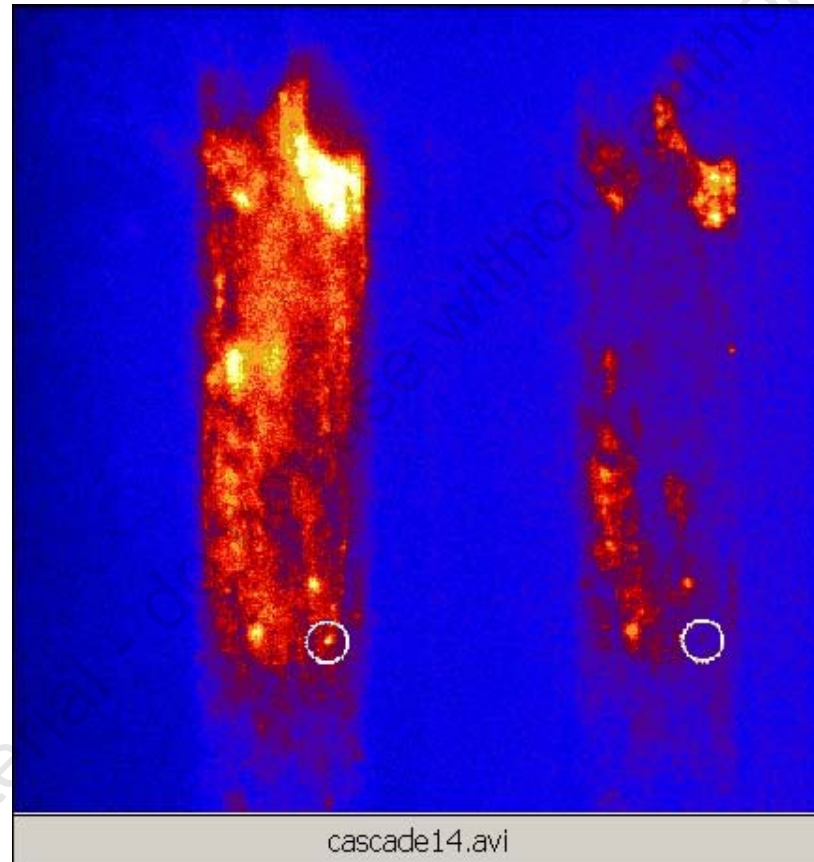
Condition: DIV 8 Over night starvation with GAM, No NGF or Qdot NGF

# Colocalization of TrkAEGFP and Qdot

Cell body



Axon

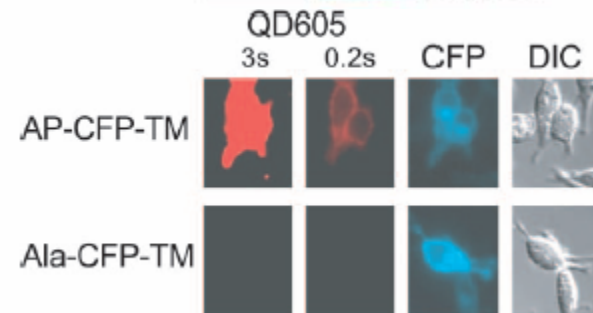
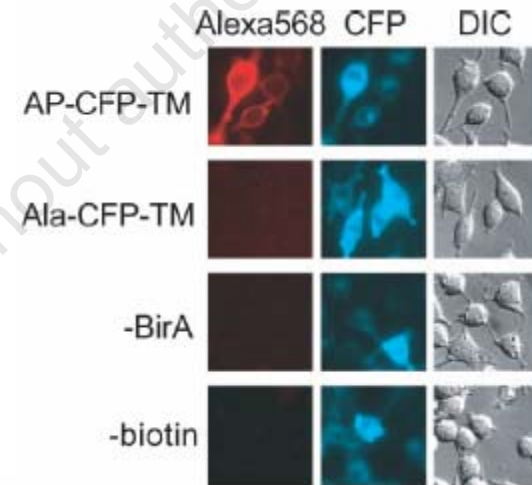
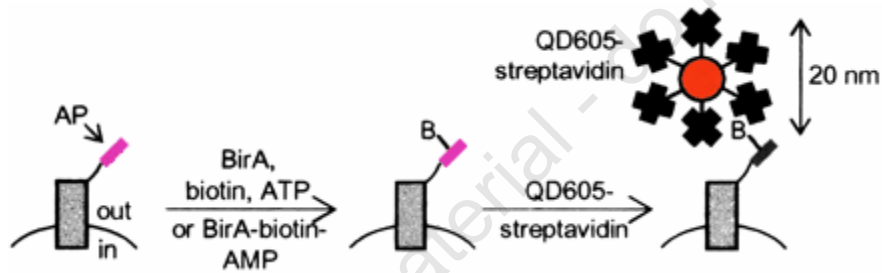
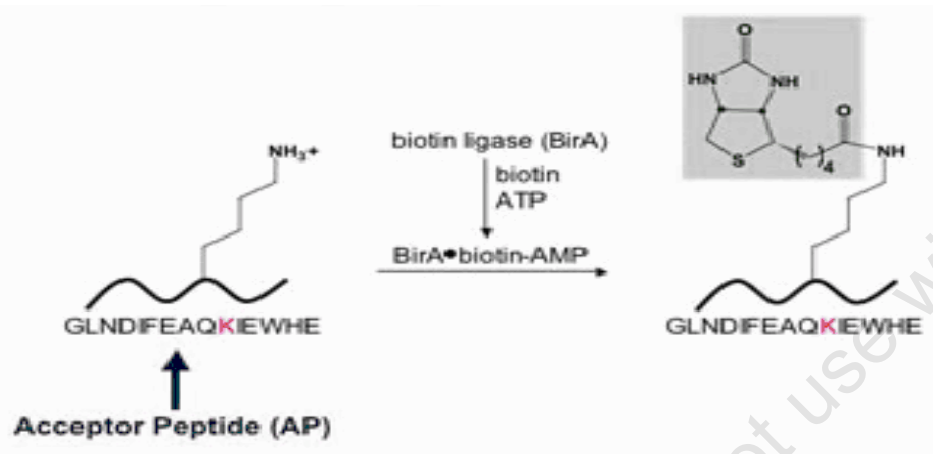


TrkA\_EGFP

QD605

Condition: DIV 8 Over night starvation, add 120ng/ml QdotNGF

# Targeting quantum dots to surface proteins in living cells with biotin ligase

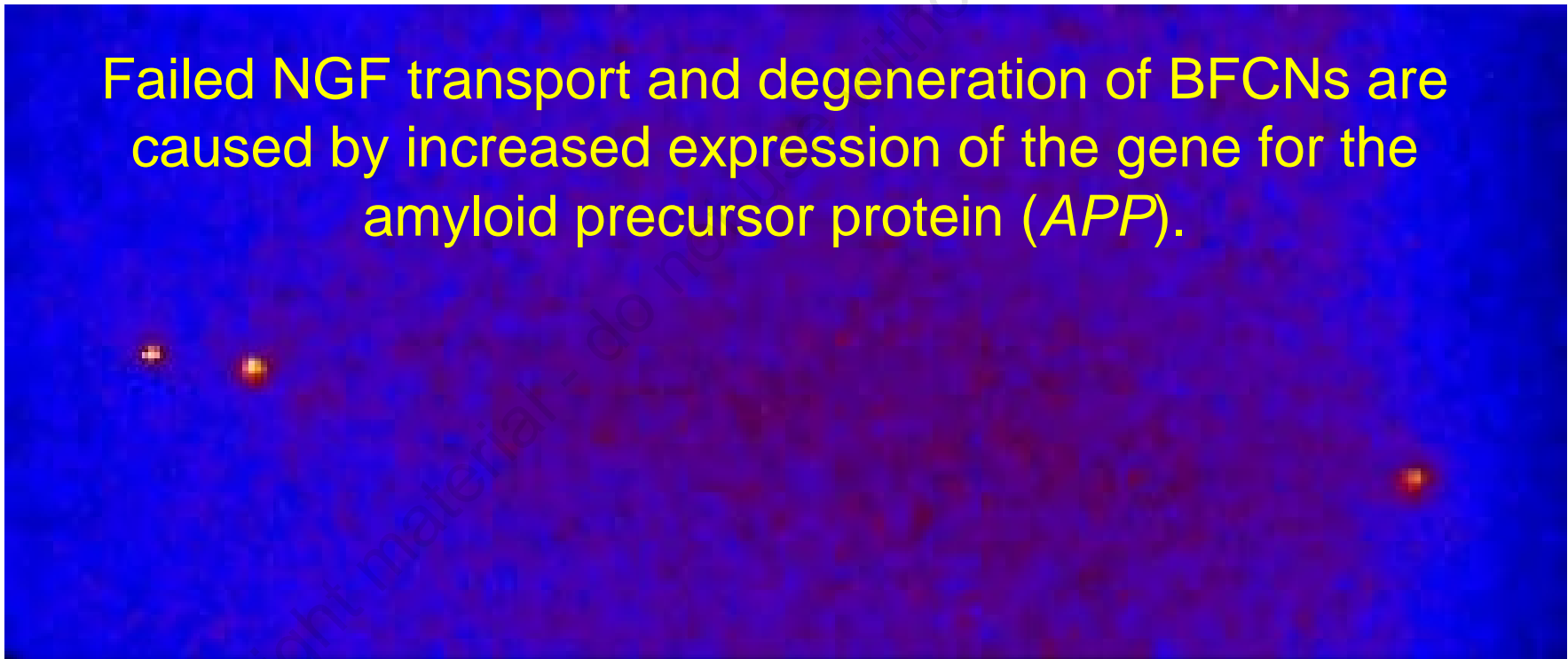


Mark Howarth, Alice Y. Ting etc. *PNAS* May 24, 2005

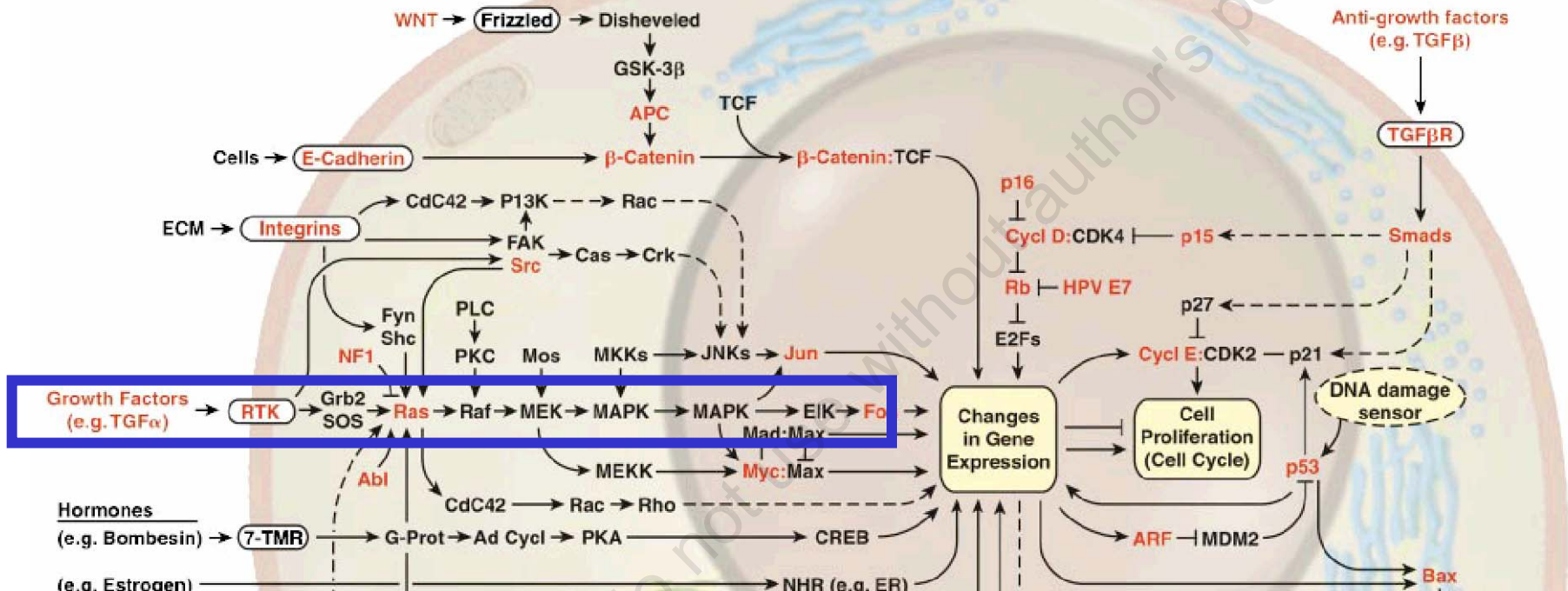


**Alzheimer's disease:** Degeneration of neurons, including basal forebrain cholinergic neurons (BFCNs). BFCN atrophy contributes to loss of attention and memory. Loss of BFCNs is linked to failed retrograde axonal transport of nerve growth factor (NGF).

Failed NGF transport and degeneration of BFCNs are caused by increased expression of the gene for the amyloid precursor protein (*APP*).



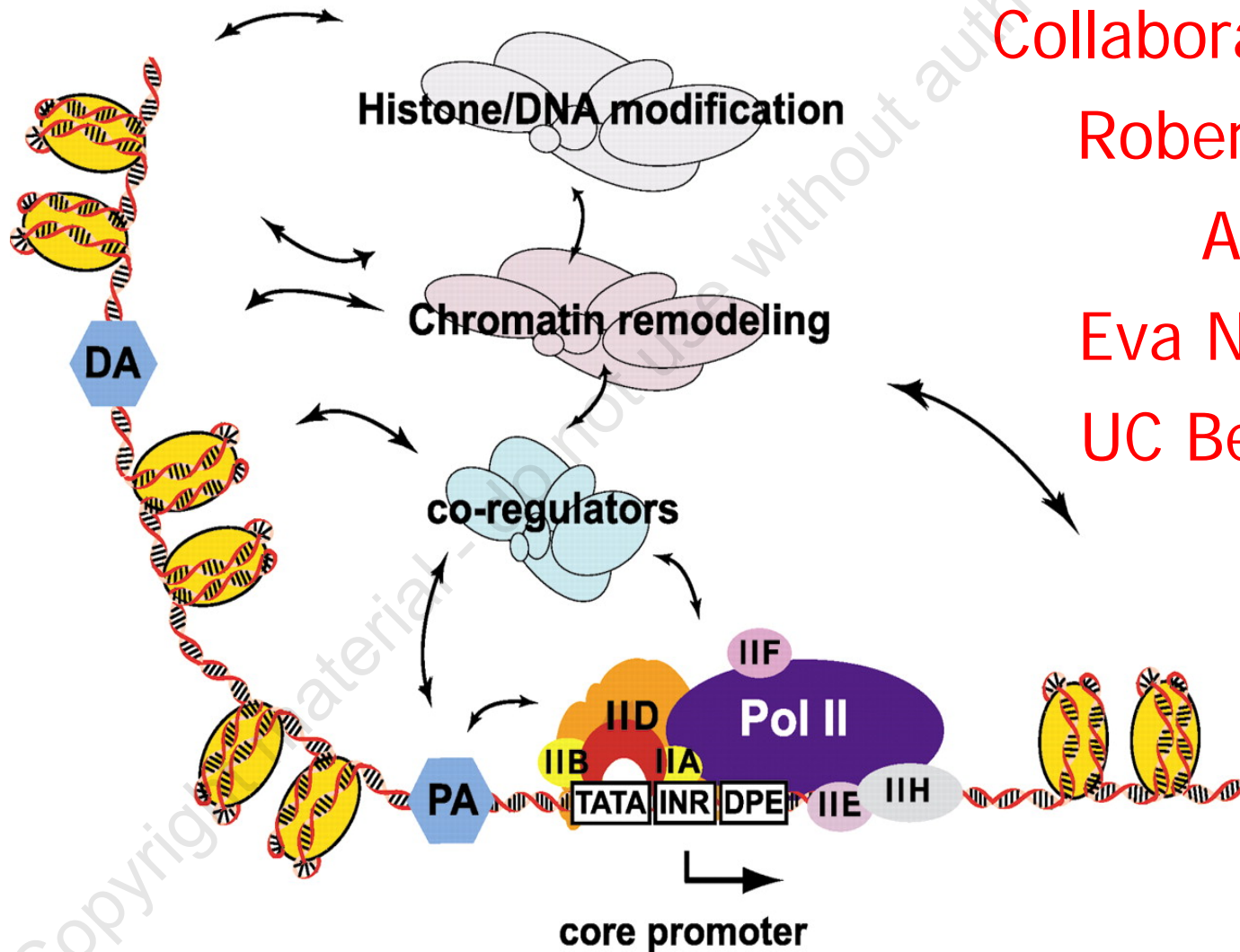
# Cellular signaling networks



“Deregulation of the EGFR-MEK-ERK signaling networks contributes to the initiation or progression of many, if not most, human carcinomas.”

Joe Gray, Assoc. Berkeley Lab Director, Life Sciences,  
Professor, UCSF

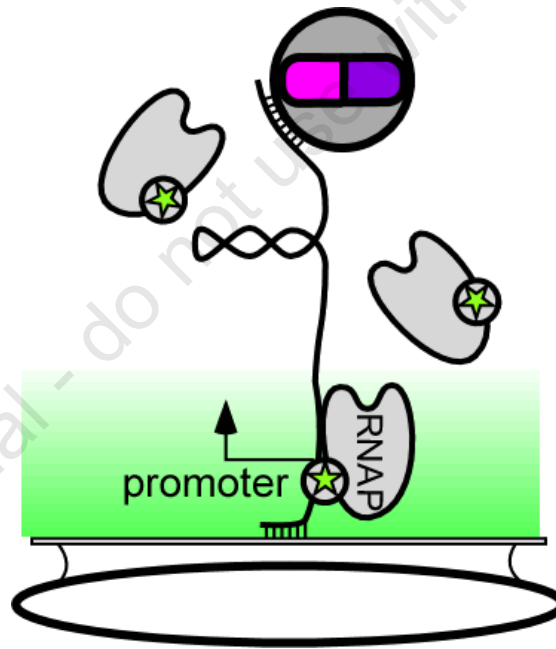
Control of transcription is a complex process:  
proximal and distal promoters, co-regulators,  
chromatin and histone modifications



Collaboration with  
Robert Tjian  
And  
Eva Nogales,  
UC Berkeley

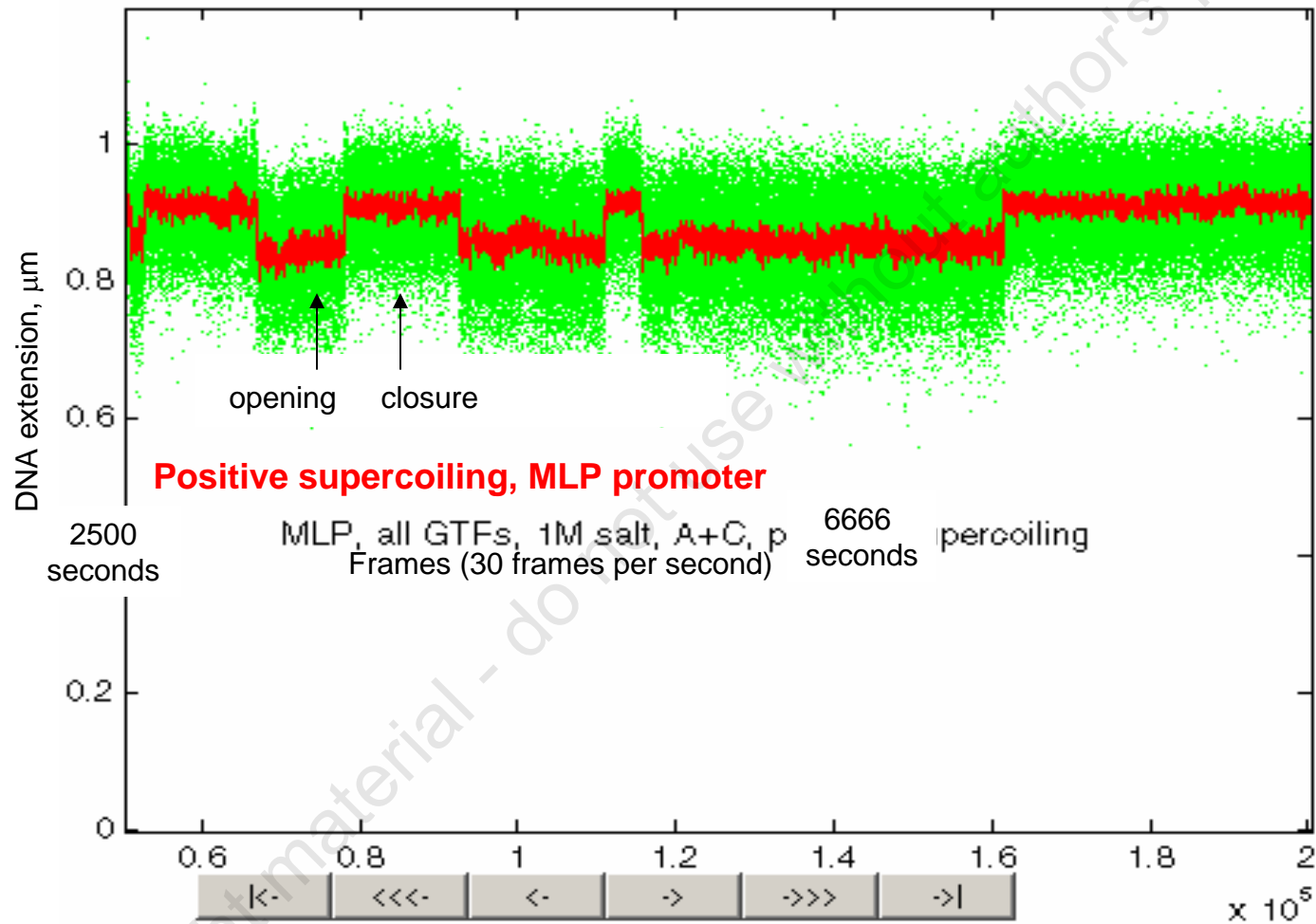
# Promoter melting and promoter binding by fluorescent *E coli* RNAP

Infrared illumination

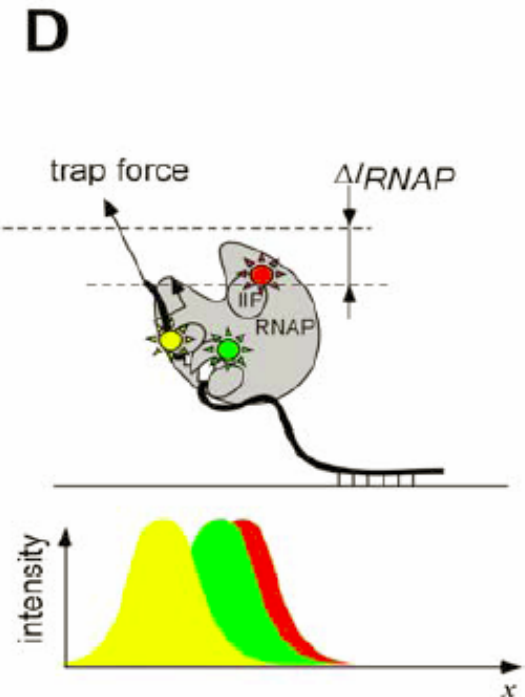
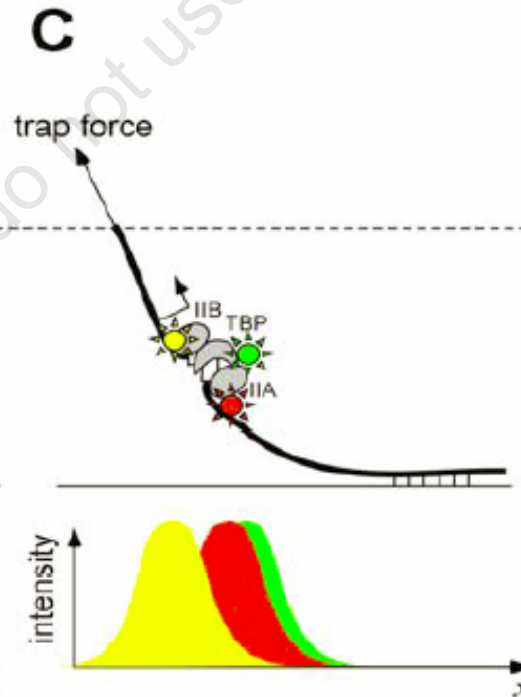
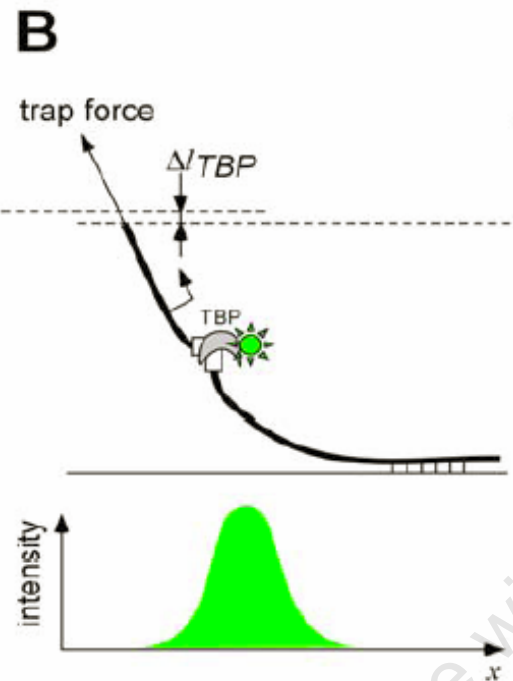
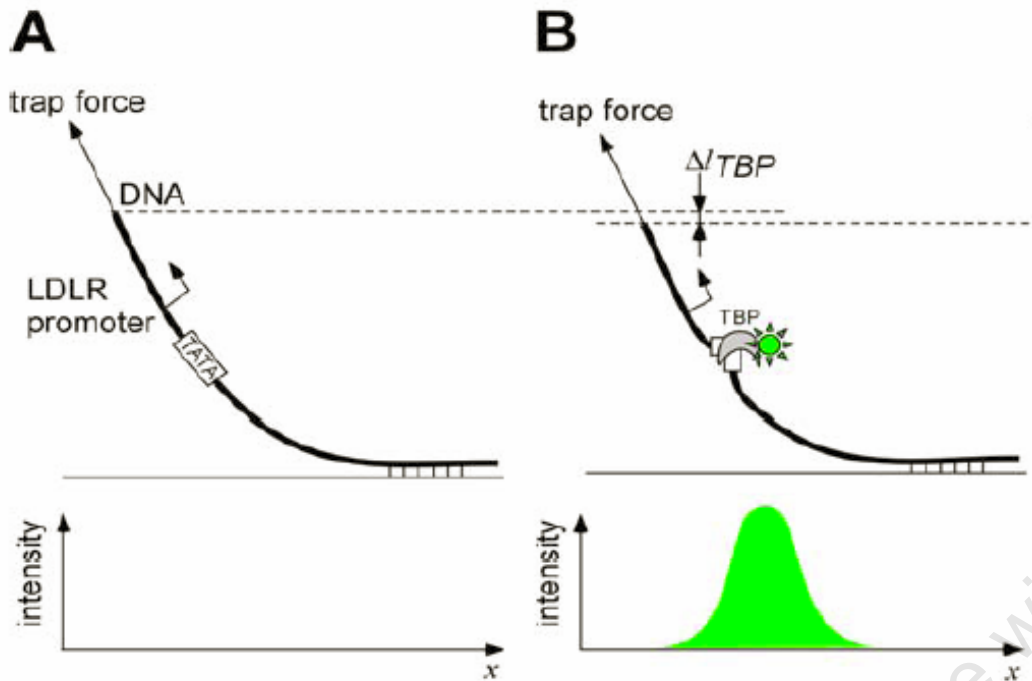


Laser excitation

# Single-molecule detection of pol II activity

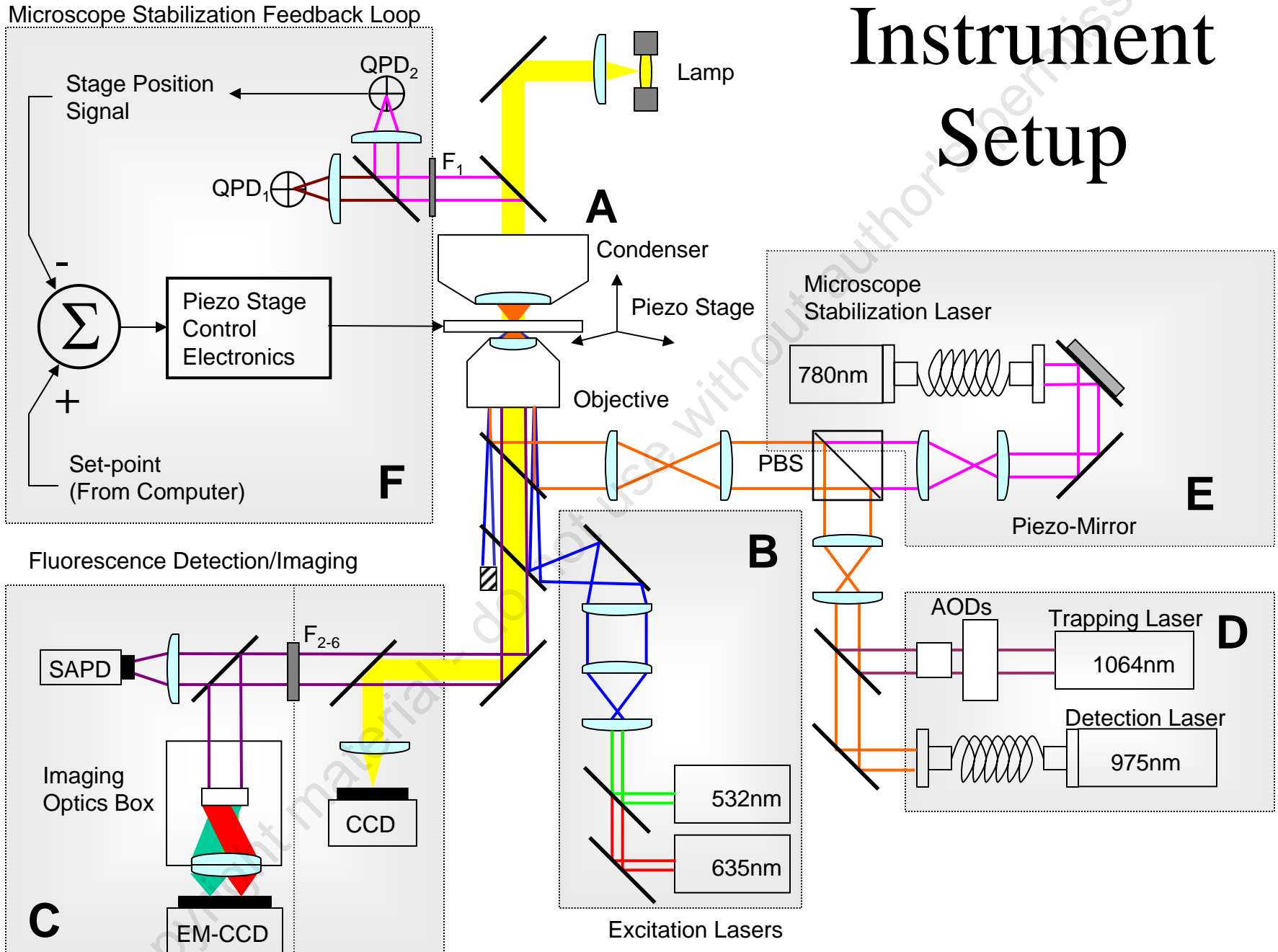


# Nanometer structural information from fluorescence probes



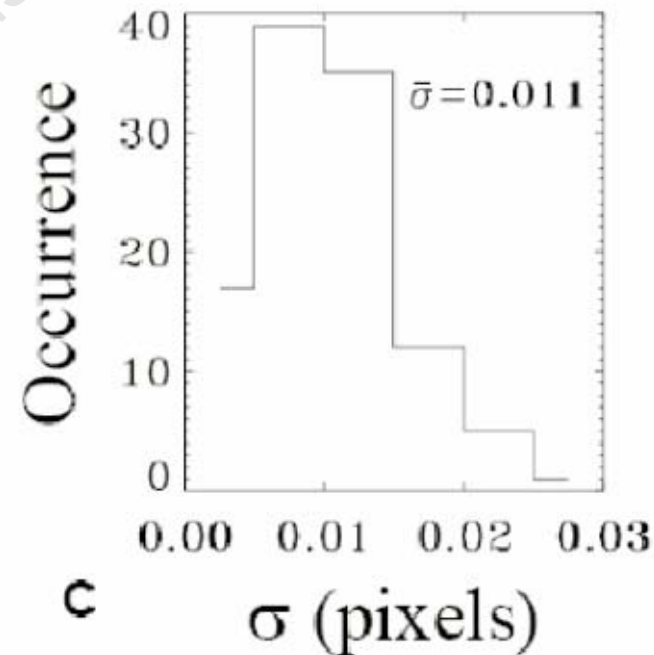
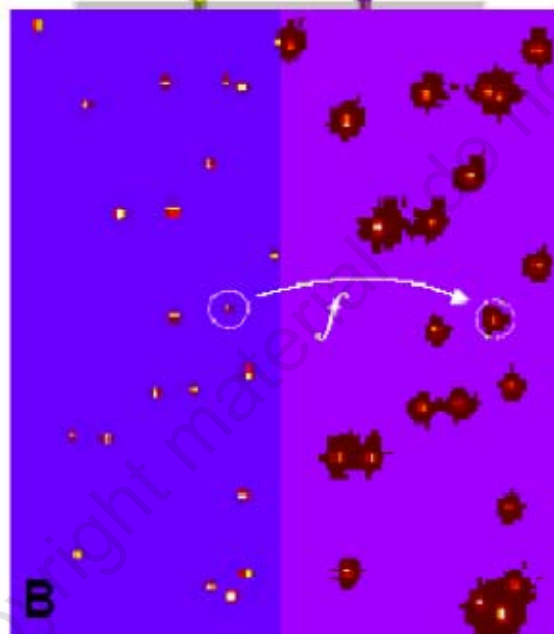
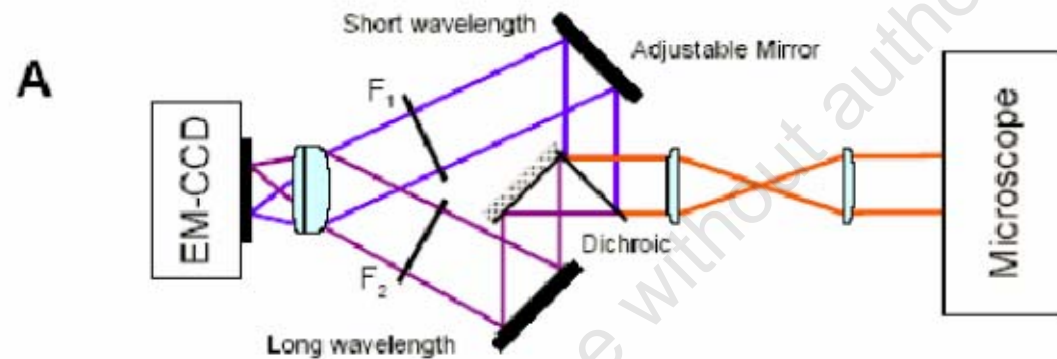
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# Instrument Setup



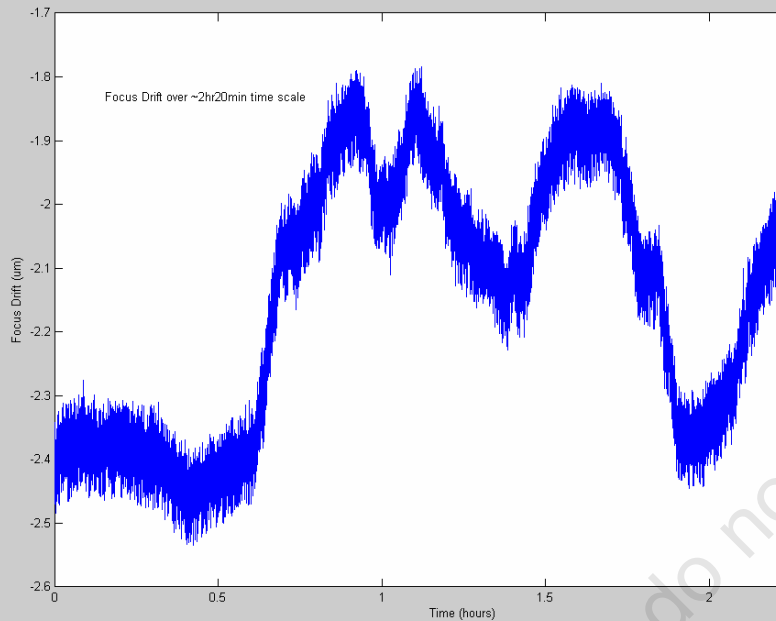
# Mapping of bead positions between green and red images

Current progress:  $\sim 0.8$  nm resolution

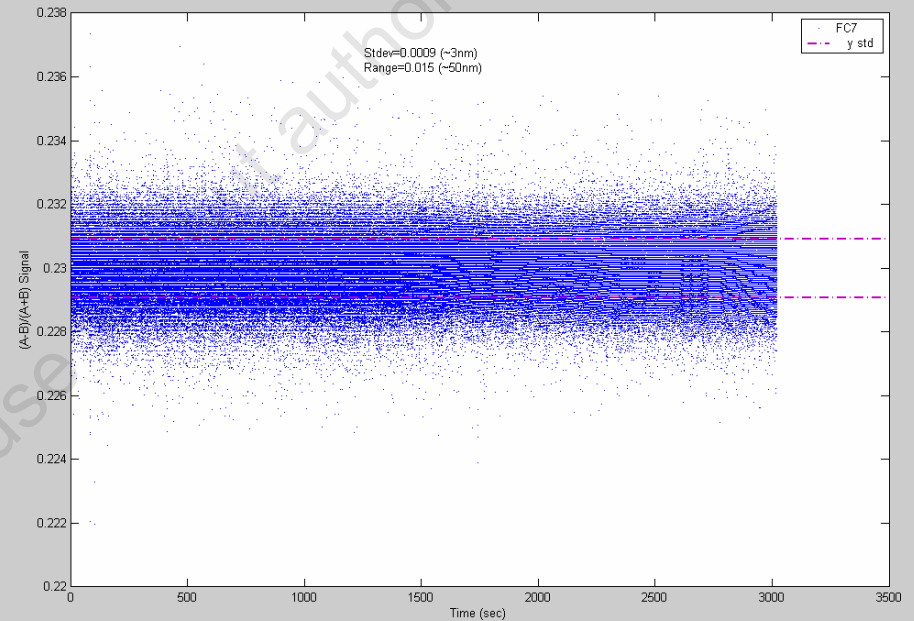




# Microscope Stage Stability



Correction signal



Error signal

~1nm stability in X/Y, 3nm in Z.

# The ribosome is an RNA enzyme !

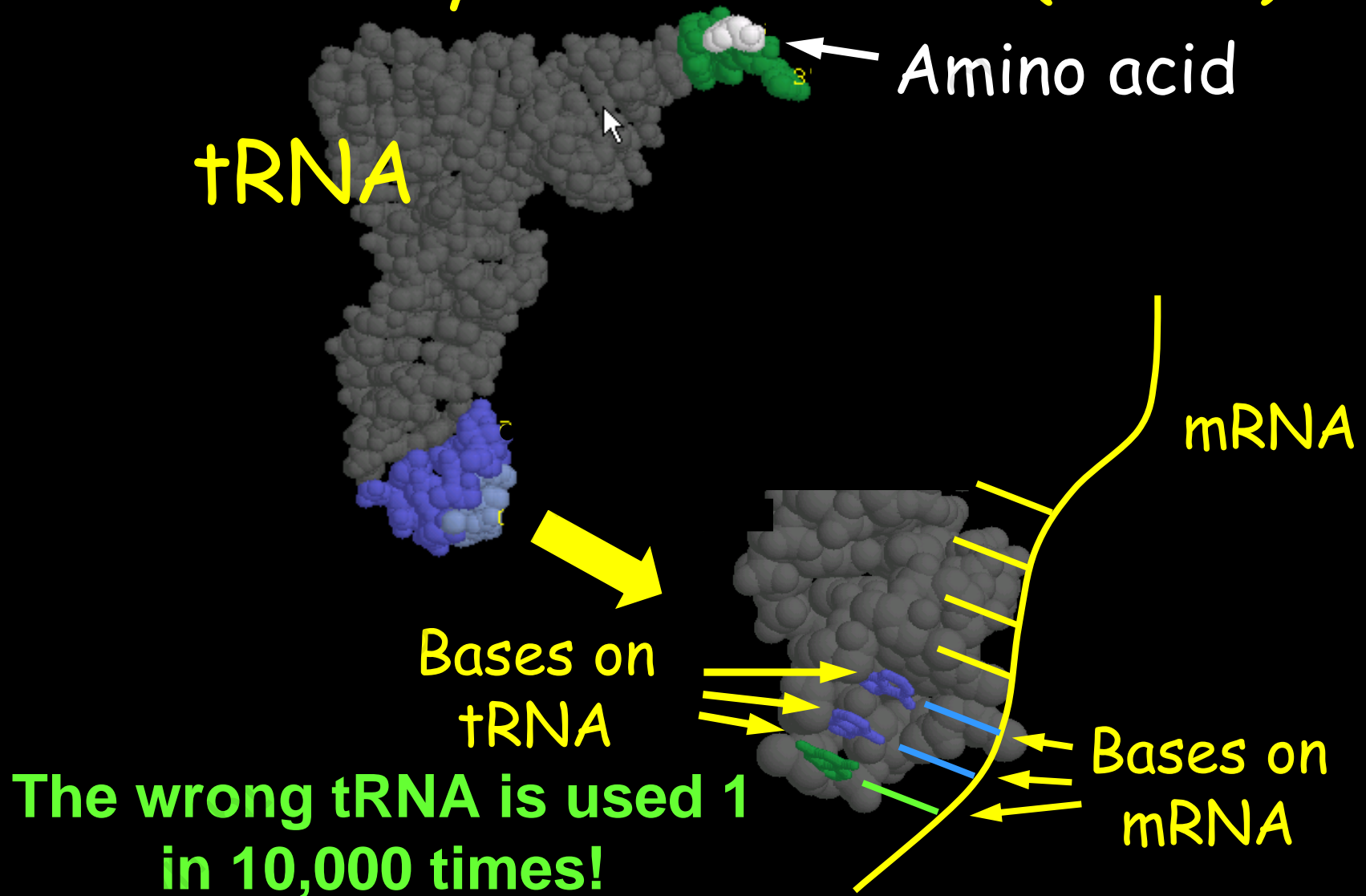
From Harry F. Noller, Marat M. Yusupov, Gulnara Z. Yusupova,  
Albion Baucom and J.H.D. Cate; *FEBS Letters* (2002)

Ribosomal Proteins are located on the Periphery of the Ribosome



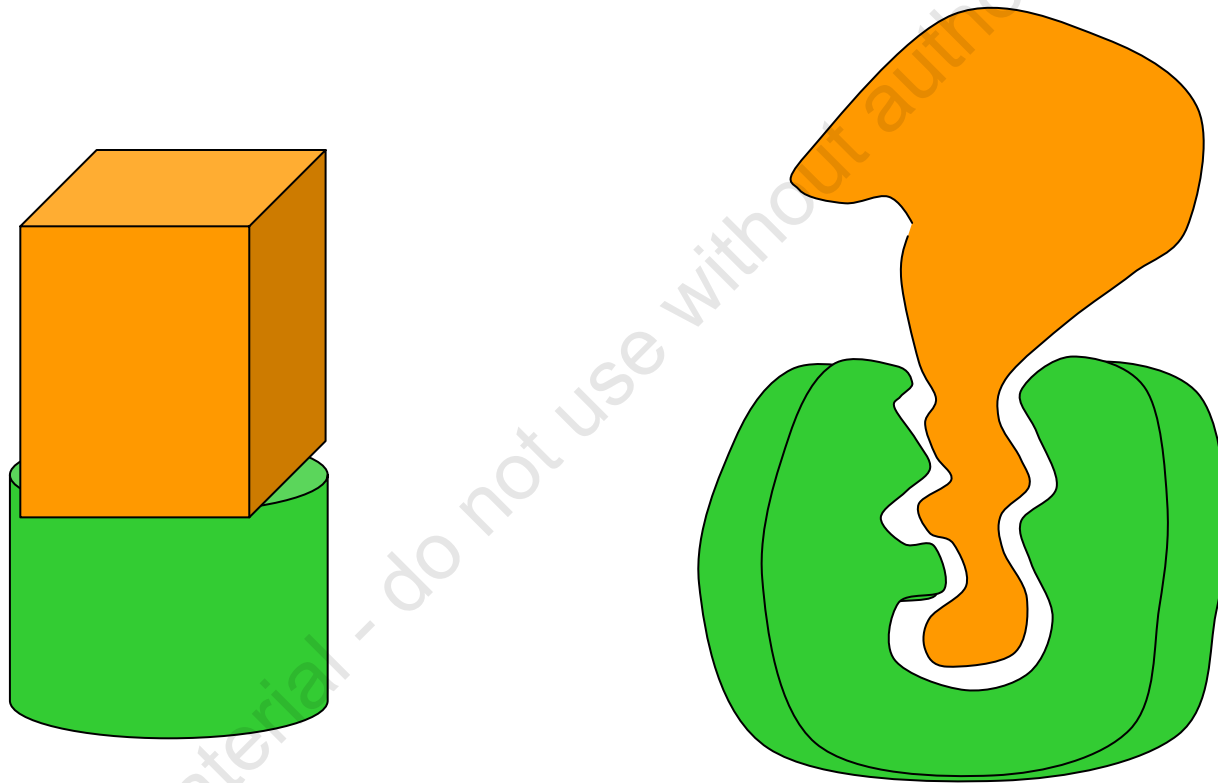
20 nm

Each amino acid is brought into the ribosome by transfer RNA (tRNA)



# Selection based on shape discrimination

J. Davies, W. Gilbert, L. Gorini (1964)



Shape selection can also be due to an  
"induced fit".

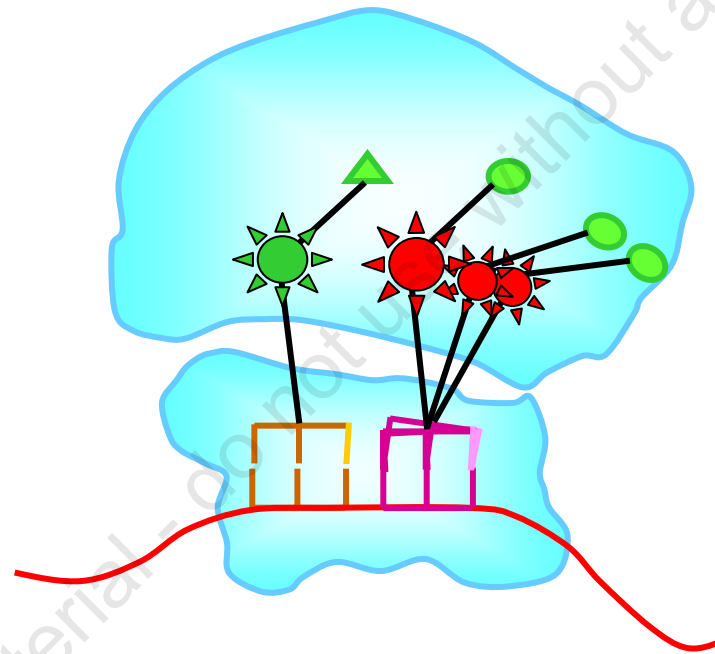


## Previously published work

- tRNA dynamics on the ribosome  
S. Blanchard, H. Kim, R. Gonzalez, J. Puglisi, S. Chu  
*PNAS* 101, 12893–12898 (2004).
- tRNA selection and kinetic proofreading in translation  
S. Blanchard, R. Gonzalez, H. Kim, S. Chu, J. Puglisi  
*Nature Structural Biology* 11, 1008 - 1014 (2004).

- The role of fluctuations in initial selection by the ribosome: *submitted 2007*, T-H. Lee, S. Blanchard, H. Kim, J. Puglisi, S. Chu
- Direct observation of allostery during aminoacyl-tRNA selection by the ribosome: *submitted 2007* R. Gonzalez, S. Chu, J. Puglisi
- tRNA fluctuations between classical and hybrid states *submitted, 2007*, H. Kim, J. Puglisi, S. Chu
- Force measurements on the ribosome and a new allostery: *Nature, 2007*, U. Sotaro, M. Dorywalska, T-H. Lee, J. Puglisi, S. Chu

# Watching t-RNA enter the ribosome using **single molecule** Fluorescence Resonant Energy Transfer (FRET)



S. Blanchard, H. Kim, R. Gonzales, T-H. Lee, J. Puglisi, S. Chu  
*PNAS*, 2005; *Nat. Struct. Bio*, 2004

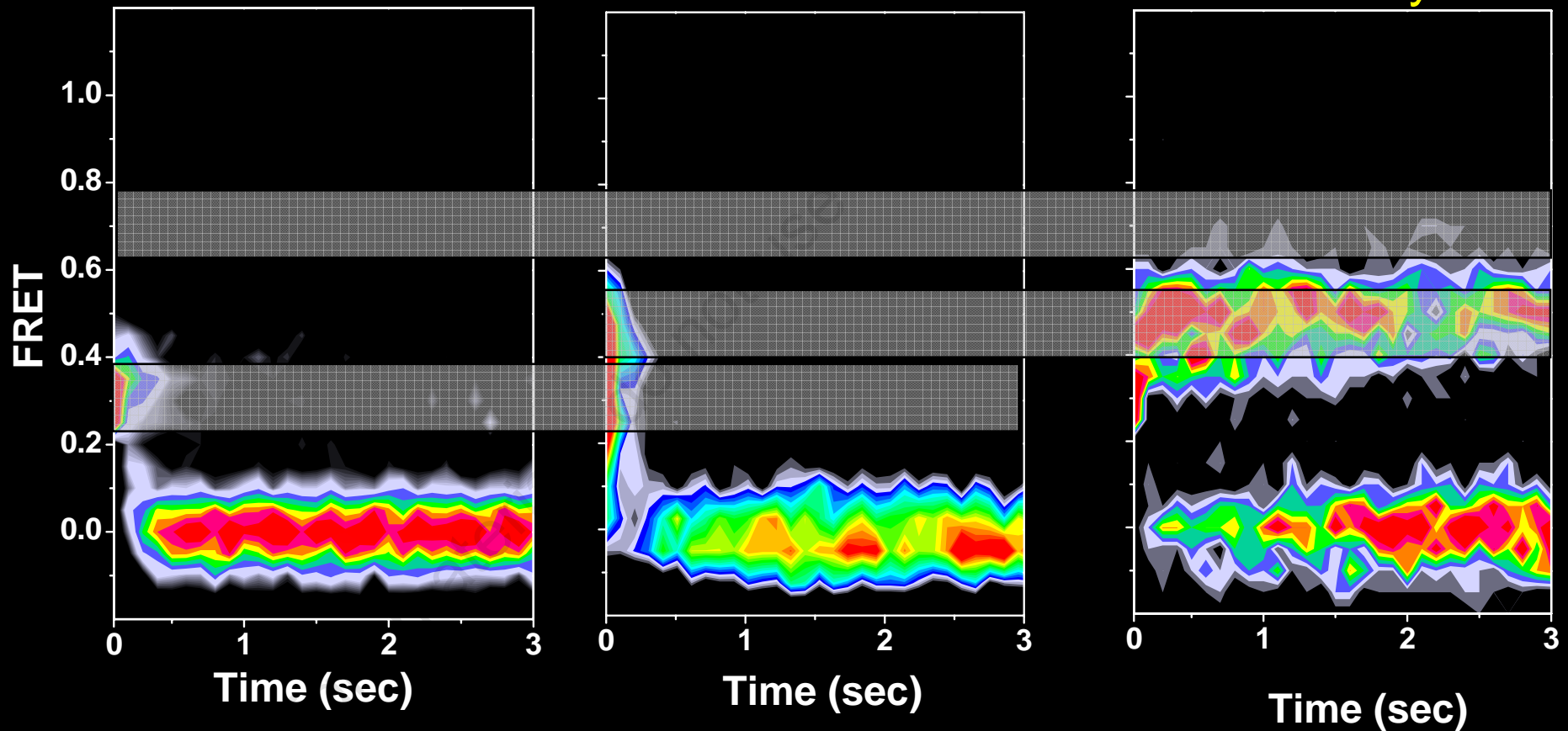


# Separate steps in accommodation are seen by stalling the ribosome

Tetracycline

Thiostrepton

Stalled by GDP-NP  
or Kirromycin



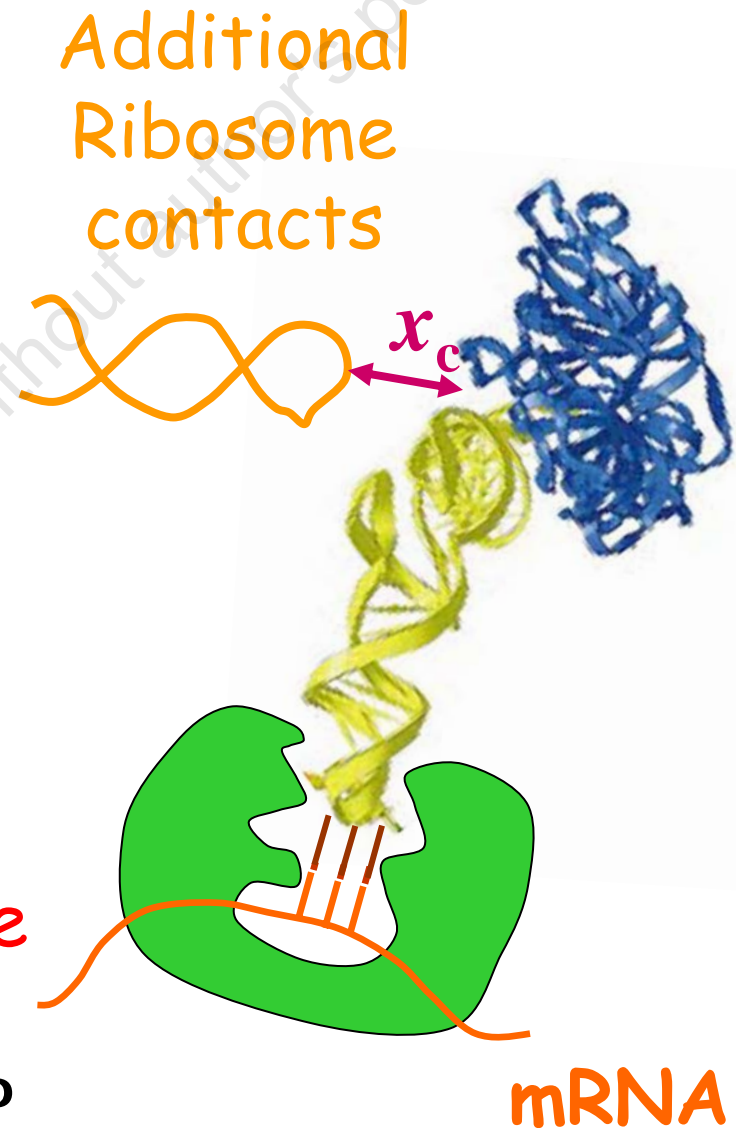
# Our proposed selection mechanism

1) Proper base-pairing causes the ribosome to wrap around the base of the tRNA

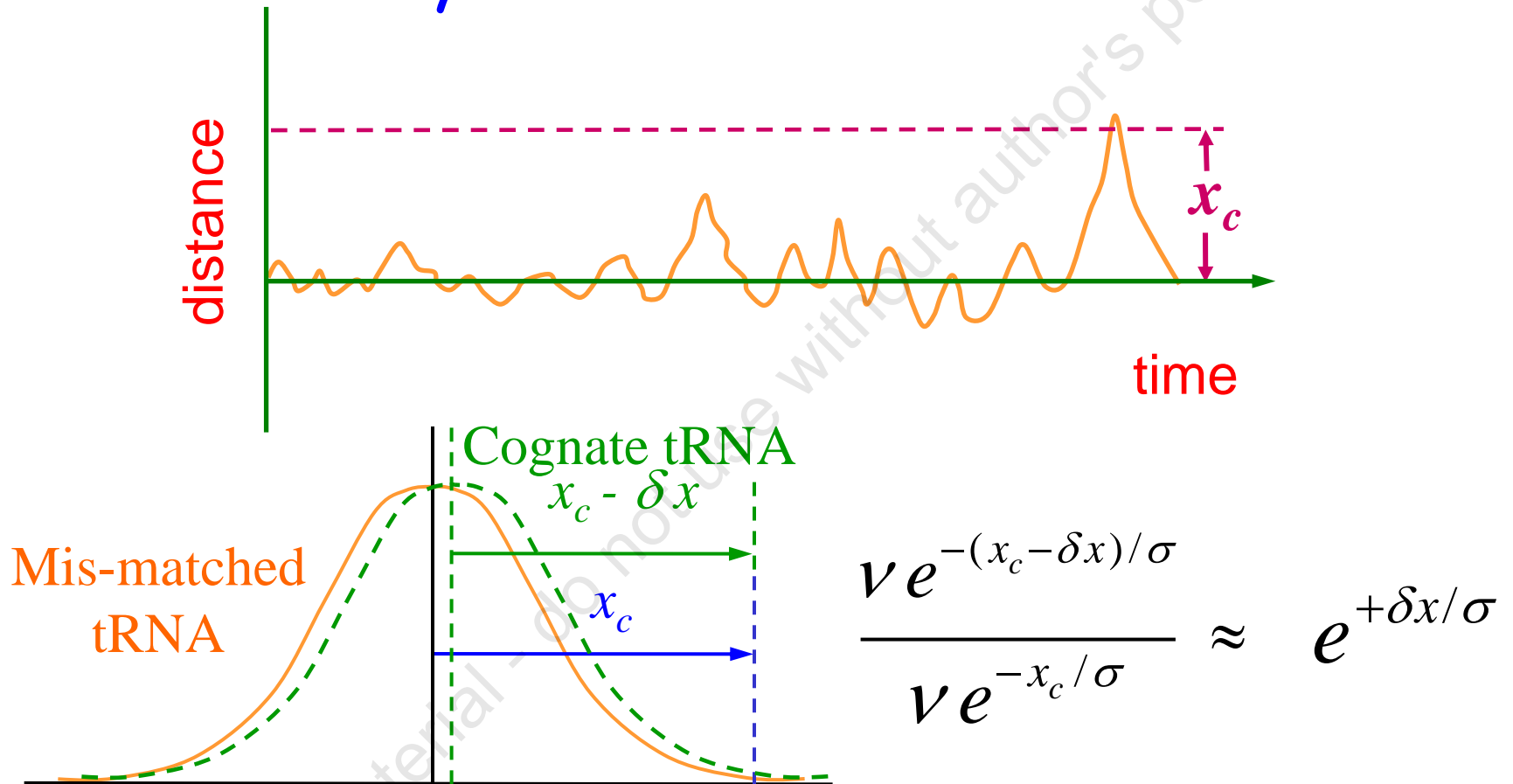
V. Ramakrishnan, *et al.* 2001

2) The wrapping of the ribosome causes the tRNA to move into a position so it is more likely to make stabilizing contacts with the Ribosome.

Tae-Hee Lee, *et. al.* submitted to PNAS, 2007



If you were an **Intelligent Designer**, how would you make the ribosome?

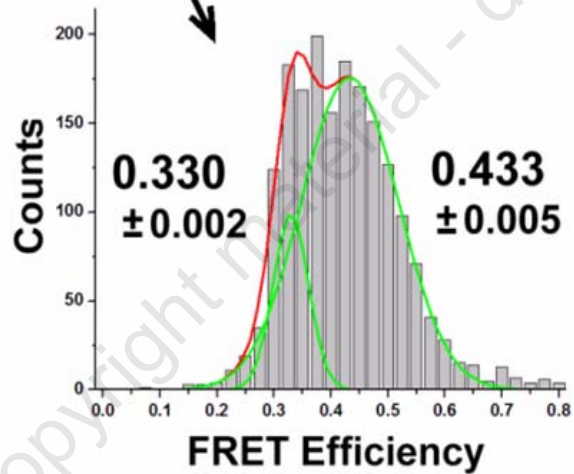
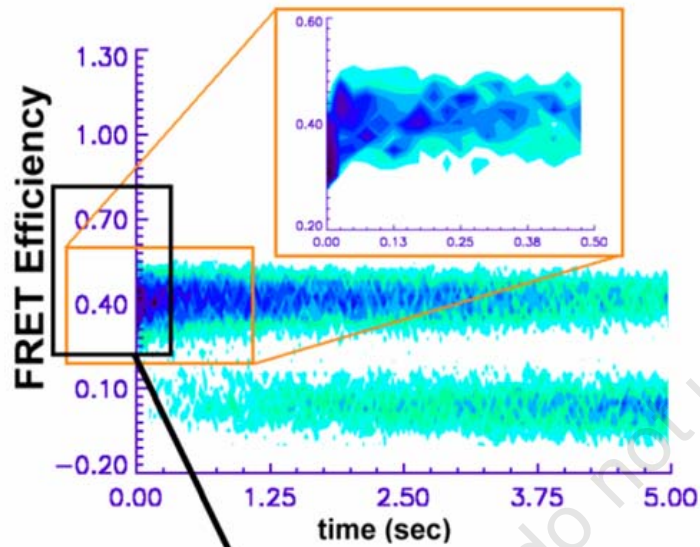


Prediction: the initial 0.3 FRET states of the **correct tRNA** and **incorrect tRNA** are different.

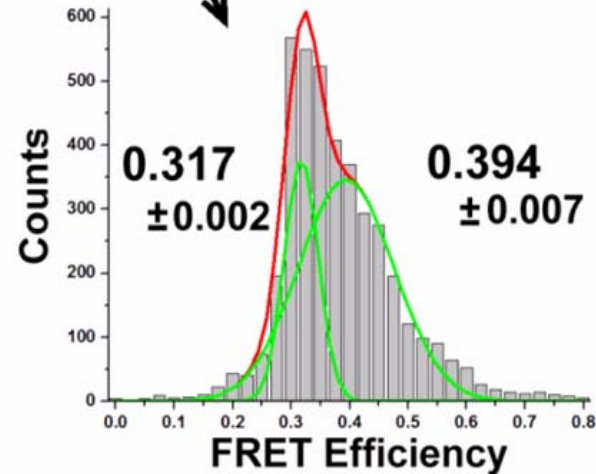
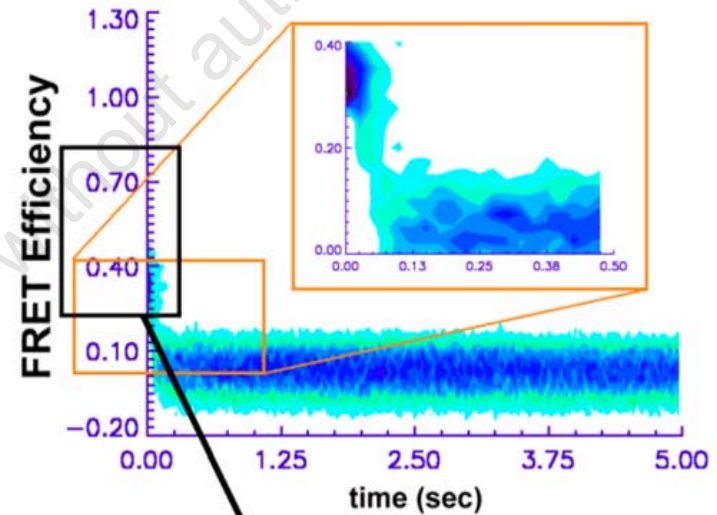
# Post-synchronized FRET > 0.27

Tae-Hee Lee, et al. unpublished (2006)

cognate



near-cognate

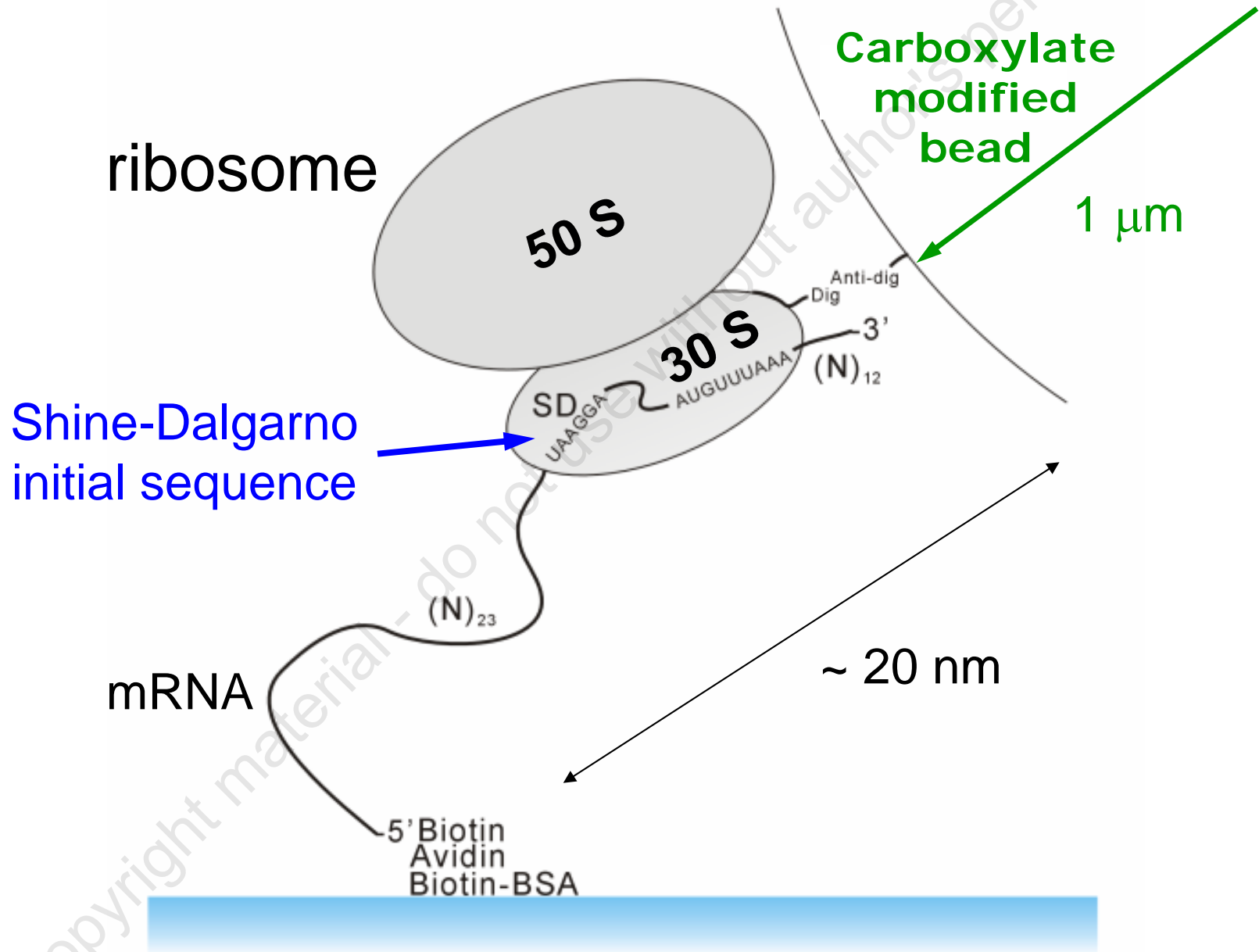


Direct force measurement between  
30S in ribosome and mRNA with  
optical tweezers

*Nature*, 2007

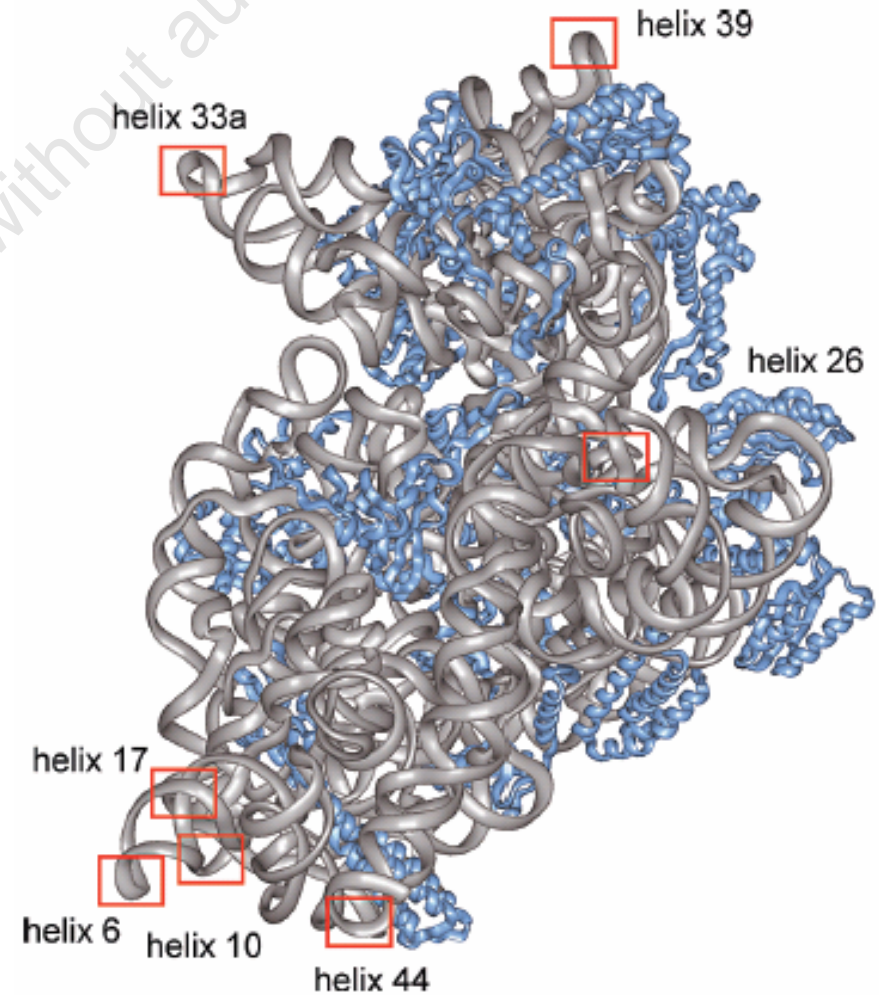
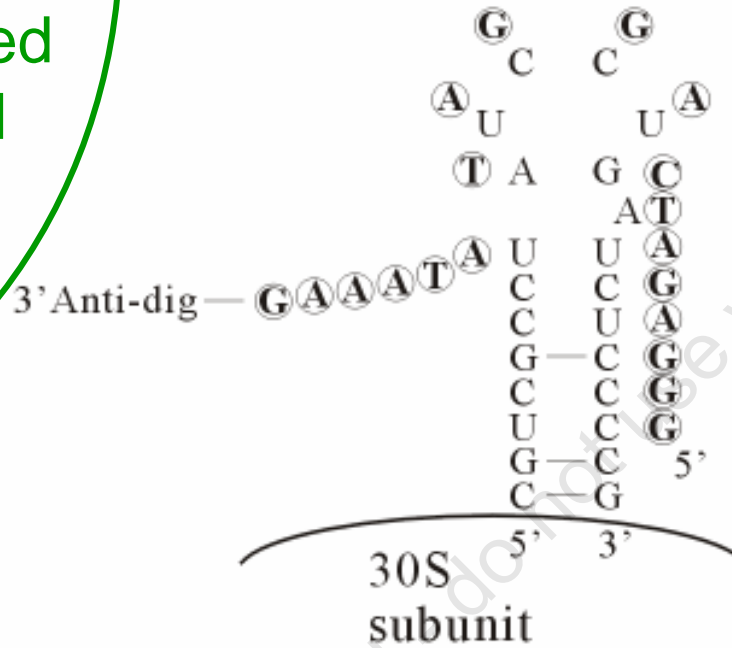
Sotaro Uemura, Magdalena Dorywalska, Tae-  
Hee Lee, Harold Kim, Jody Puglisi and Steve  
Chu

# Pulling on the 30 S subunit of the Ribosome

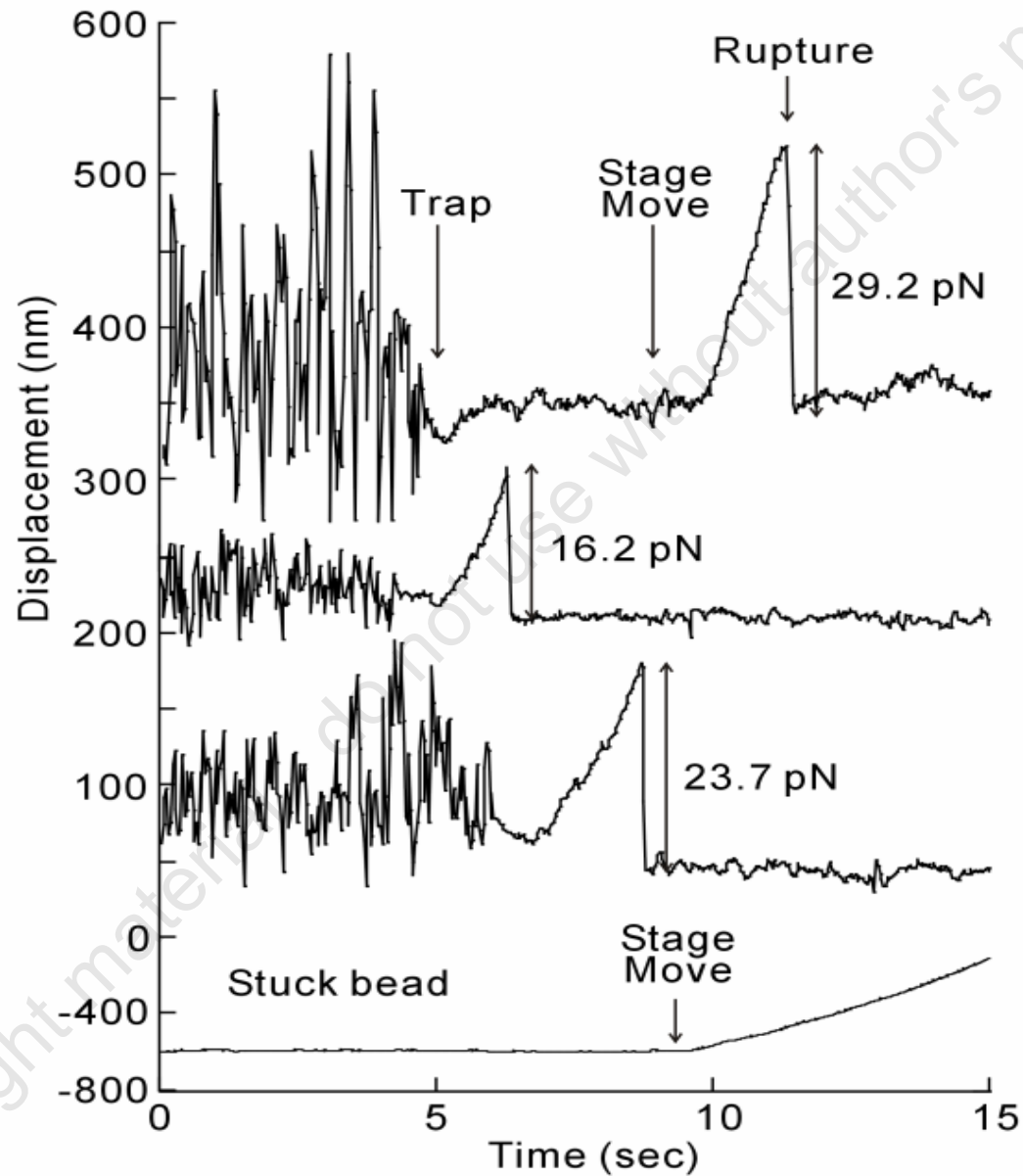


# Bead - ribosome attachment (near beak region of 30S)

Carboxylate  
modified  
Bead

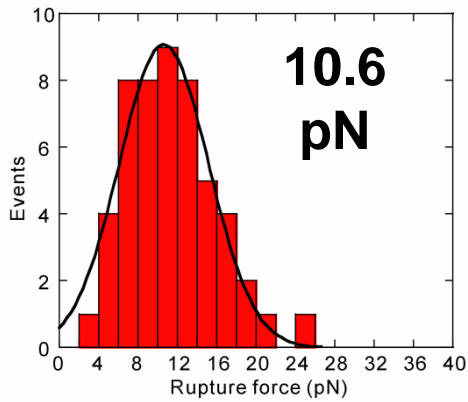
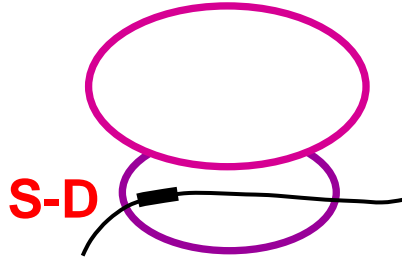


# Sample force measurements

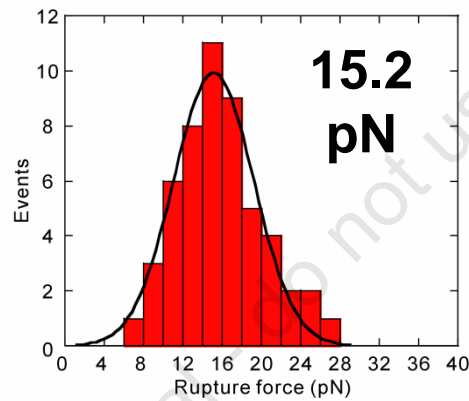
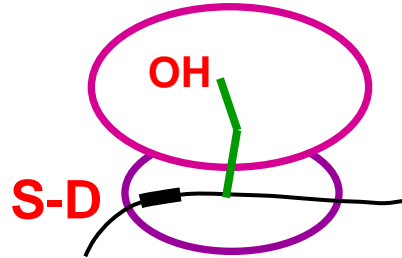




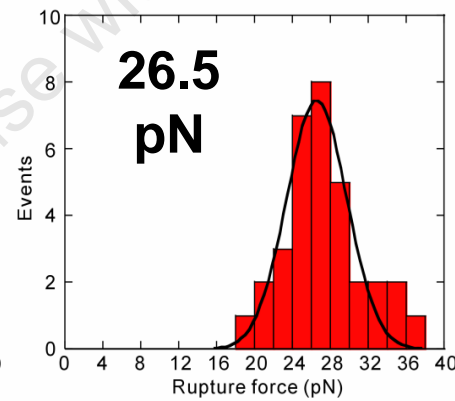
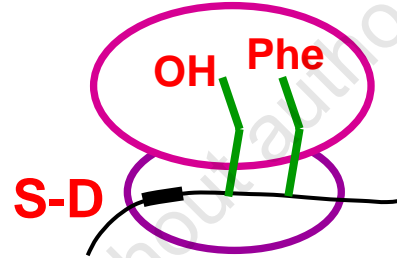
Ribosome + mRNA



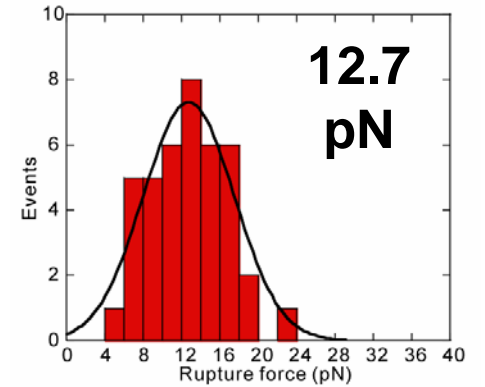
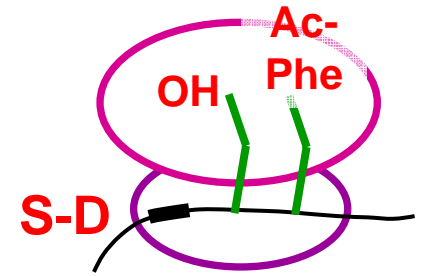
Ribosome + mRNA + tRNA<sup>fMet</sup>



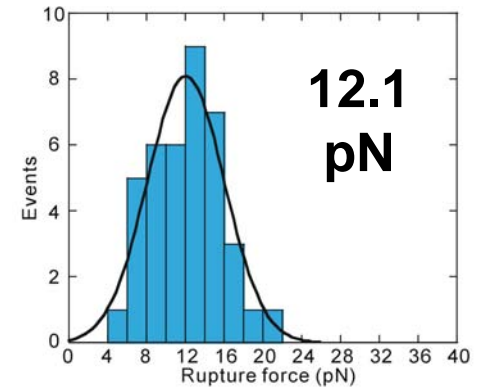
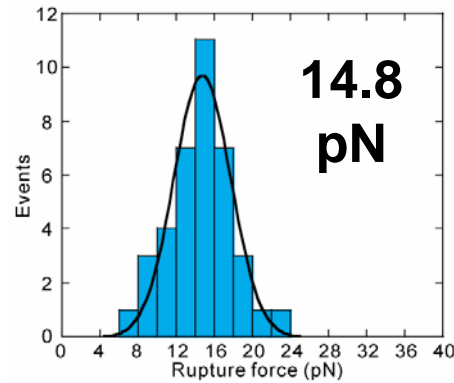
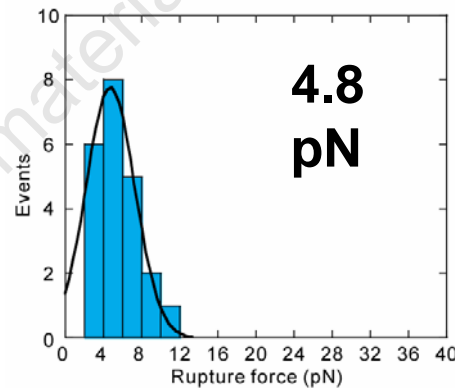
Ribosome + mRNA + tRNA<sup>fMet</sup> + Phe-tRNA<sup>Phe</sup>



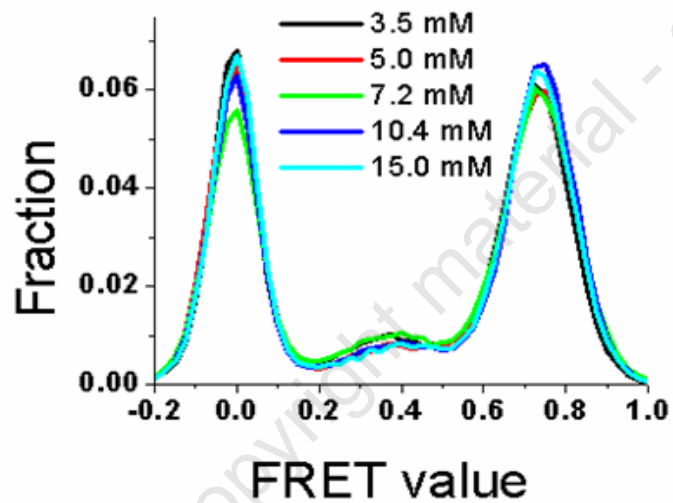
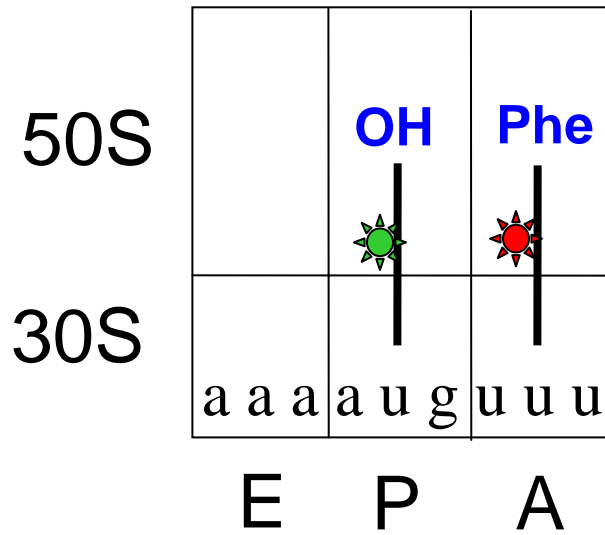
Ribosome + mRNA + tRNA<sup>fMet</sup> + AcPhe-tRNA<sup>Phe</sup>



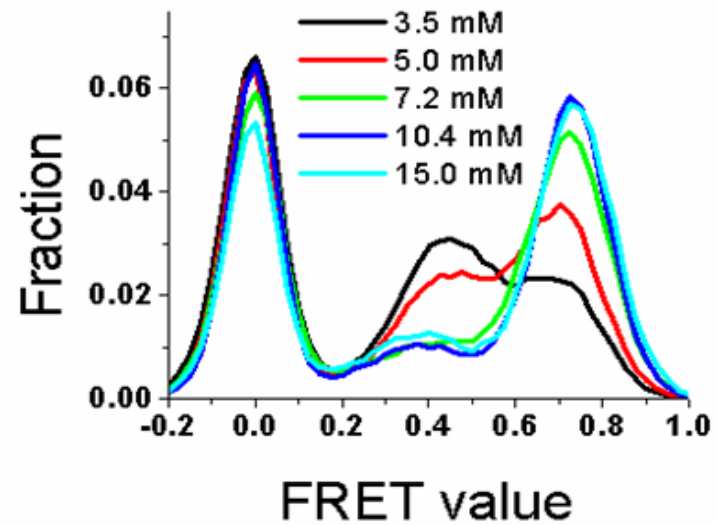
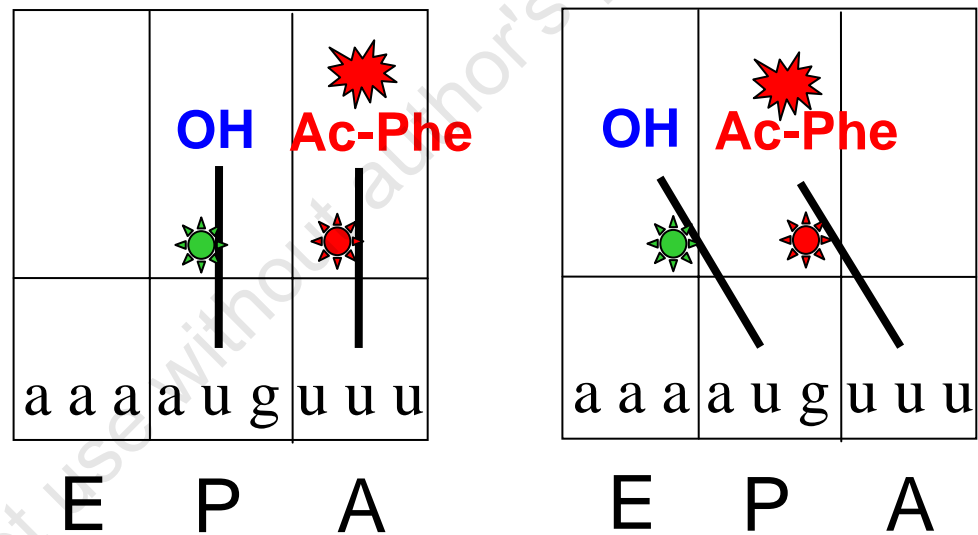
No tethered ribosomes



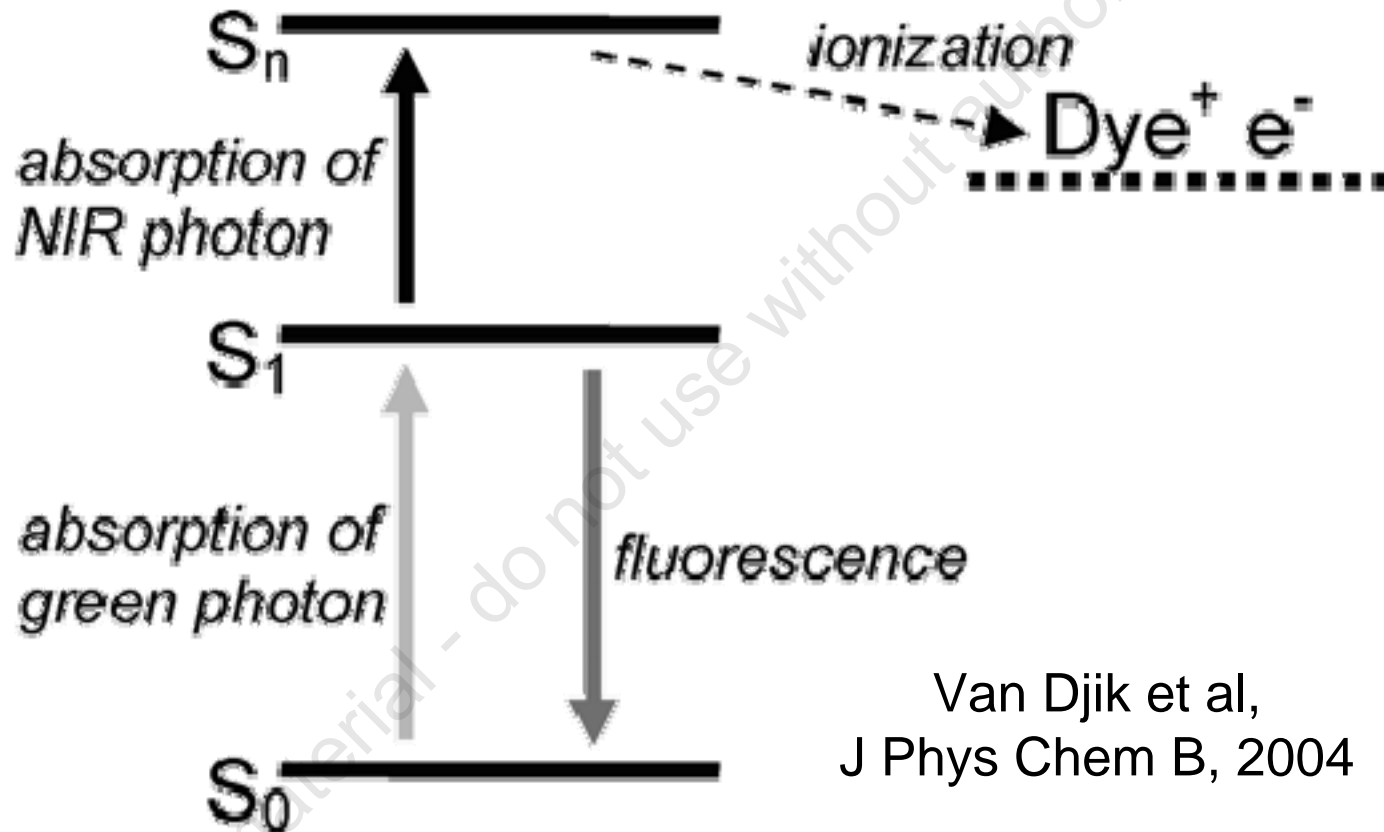
## Phe on A-site tRNA



## aminoacylated Phe on tRNA



# Photo-destruction Pathway



Enhanced Photo-bleaching from Excited State

# Time-Scale Separation

- Excited-state lifetime:  $\sim 10\text{nsec}$
- Viscous Relaxation Time:  $100\mu\text{sec}-1\text{msec}$
- Single molecule biology:  $\text{msec}-\text{sec}$

Take advantage of window at  $\sim 1\text{msec}$

# RF Electronics Layout

