



HUMAN GENOME EPIDEMIOLOGY (HuGE) REVIEW

Cytochrome P-450 1A1 Gene Polymorphisms and Risk of Breast Cancer: A HuGE Review

L. F. Masson, L. Sharp, S. C. Cotton, and J. Little

From the Epidemiology Group, Department of Public Health, University of Aberdeen, Aberdeen, Scotland.

Received for publication July 21, 2004; accepted for publication January 4, 2005.

Cytochrome P-450 (CYP) 1A1 plays a key role in phase I metabolism of polycyclic aromatic hydrocarbons and in estrogen metabolism. It is expressed predominantly in extrahepatic tissues, including the breast. Four *CYP1A1* gene polymorphisms (3801T → C, Ile462Val, 3205T → C, and Thr461Asp) have been studied in relation to breast cancer. The 3801C variant is more common than the Val variant. Both variants occur more frequently in Asians than in White populations. The 3205T → C polymorphism has been observed in African Americans only. Little data are available on the geographic/ethnic distribution of the Thr461Asp polymorphism. The functional significance of the polymorphisms is unclear. In 17 studies, no consistent association between breast cancer and *CYP1A1* genotype was found. Meta-analysis found no significant risk for the genotypes 1) 3801C/C (relative risk (RR) = 0.97, 95% confidence interval (CI): 0.52, 1.80) or 3801T/C (RR = 0.91, 95% CI: 0.70, 1.19) versus 3801T/T, 2) Val/Val (RR = 1.04, 95% CI: 0.63, 1.74) or Ile/Val (RR = 0.92, 95% CI: 0.76, 1.10) versus Ile/Ile, or 3) Asp/Asp (RR = 0.95, 95% CI: 0.20, 4.49) or Thr/Asp (RR = 1.12, 95% CI: 0.87, 1.43) versus Thr/Thr. Future studies should explore possible interactions between *CYP1A1* and sources of polycyclic aromatic hydrocarbons, markers of estrogen exposure, other lifestyle factors influencing hormonal levels, and other genes involved in polycyclic aromatic hydrocarbon metabolism or hormonal biosynthesis.

breast neoplasms; cytochrome P-450 *CYP1A1*; epidemiology; polymorphism, genetic

Abbreviations: CI, confidence interval; COMT, catechol-O-methyltransferase; CYP, cytochrome P-450; GST, glutathione S-transferase; PAH, polycyclic aromatic hydrocarbon; PCB, polychlorinated biphenyl; RR, relative risk.

Editor's note: This paper is also available on the website of the Human Genome Epidemiology Network (<http://www.cdc.gov/genomics/hugenet/>).

GENE

Cytochrome P-450 (CYP) 1A1 is a key enzyme in phase I bioactivation of xenobiotics (1). It contributes to

aryl hydrocarbon hydroxylase activity, catalyzing the first step in the metabolism of a number of polycyclic aromatic hydrocarbons (PAHs), such as the tobacco carcinogen benzo[a]pyrene, to their ultimate DNA-binding forms (2). It is also involved in estrogen metabolism, catalyzing the hydroxylation of 17β-estradiol at the C-2 position (3, 4).

The *CYP1A1* gene, located at 15q22-q24, comprises seven exons and six introns and spans 5,810 base pairs (5). In humans, *CYP1A1* is under the regulatory control of the

aryl hydrocarbon receptor, a transcription factor that regulates gene expression (6).

CYP1A1 expression occurs predominantly in extrahepatic tissue (7). CYP1A1 messenger RNA has been detected in normal and cancerous breast tissue (8, 9) and can be induced in human-breast-derived cell lines (6).

GENE VARIANTS

Several mutations in *CYP1A1* have been described (for *CYP1A1* allele nomenclature, refer to the following website: <http://www.imm.ki.se/CYPalleles>), and four polymorphisms have been studied in relation to breast cancer. Table 1 describes these four polymorphisms and the allele nomenclature system (10–13). The 3801T→C (14, 15) and 3205T→C (16) polymorphisms are located in the 3' noncoding region. The 2455A→G (17) and 2453C→A (13) polymorphisms arise close together in exon 7 and result in the amino acid changes Ile462Val and Thr461Asp, respectively. Because the studies did not always include information on all polymorphisms, it was often not possible to identify which of the *2A, *2B, or *2C alleles were present. Therefore, the “3801T→C,” “Ile462Val,” “3205T→C,” and “Thr461Asp” nomenclature is used throughout this review.

Genotype frequencies

In 2001, Garte et al. (18) estimated *CYP1A1**2A, *CYP1A1**2B, *CYP1A1**2C, and *CYP1A1**3 genotype frequencies in Whites, Asians, and Africans by using data from 33 studies of Whites, nine studies of Asians, and five studies of Africans. In comparison, the present review includes data from 69 articles, including 20 studies published between

2002 and 2004, and also summarizes data for the Thr461Asp polymorphism.

Relevant papers were identified by searching MEDLINE and EMBASE from 1980 to week 4 of 2004 by using the MeSH heading “Cytochrome P-450 CYP1A1” or the text words “CYP1A1” or “P4501A1” combined with the MeSH headings “Polymorphism (Genetics),” “Mutation,” “Point mutation,” “Genotype,” or the text words “polymorph\$,” “mutation\$,” “gene,” “genes,” “genetic\$,” “genotyp\$,” or “allel\$.” Additional articles were identified from the Centers for Disease Control and Prevention Genomics and Disease Prevention Information System and by hand searching reference lists in published papers. Eligible studies presented frequencies for each genotype separately in nondiseased persons. Studies that did not include controls for breast cancer patients were excluded if there were fewer than 200 subjects in each ethnic group, which would limit precision of the estimates of the genotype frequencies. If there appeared to be an overlap in subjects between studies, only the largest study was reported. Hardy-Weinberg equilibrium was assessed by using the Pearson χ^2 test.

Web tables 1, 2, 3, and 4 show homozygous variant and heterozygous genotype frequencies for the 3801T→C, Ile462Val, 3205T→C, and Thr461Asp polymorphisms (13, 17, 19–85). (This information is described in the first four of eight supplementary tables; each is referred to as “Web table” in the text and is posted on the website of the Human Genome Epidemiology Network (<http://www.cdc.gov/genomics/hugenet/reviews.htm>) as well as on the *Journal's* website (<http://aje.oupjournals.org/>.) The subjects in most studies are volunteers (with the sampling frame unspecified) or hospital or clinic patients. It is unclear whether genotype frequencies in such series will reflect those in the general population. Considerable data are available from Japanese, western European, and White American populations. Data are limited, or not available, for other populations.

TABLE 1. CYP1A1 3801T→C, Ile462Val, 3205T→C, and Thr461Asp polymorphisms

Allele nomenclature (10, 11)	Nucleotide change	Amino acid change	Location	Proposed allele nomenclature		Reference to first report: study, year (reference no.)
				Garte (12)	Cascorbi et al. (13)	
<i>CYP1A1</i> *1A	None			<i>CYP1A1</i> *1	<i>CYP1A1</i> *1	
<i>CYP1A1</i> *2A	3801T→C	None	3' noncoding region (downstream of polyadenylation site)	<i>CYP1A1</i> *2	<i>CYP1A1</i> *2A	Bale et al., 1987 (14); Spurr et al., 1987 (15)
<i>CYP1A1</i> *2B	3801T→C	None	3' noncoding region		<i>CYP1A1</i> *2B	Hayashi et al., 1991 (17)
<i>CYP1A1</i> *2C	2455A→G	Isoleucine→valine	Exon 7, codon 462			
	2455A→G	Isoleucine→valine	Exon 7, codon 462 (heme binding region)	<i>CYP1A1</i> *3		Hayashi et al., 1991 (17)
<i>CYP1A1</i> *3	3205T→C	None	3' noncoding region (upstream of polyadenylation site)	<i>CYP1A1</i> *4	<i>CYP1A1</i> *3	Crofts et al., 1993 (16)
<i>CYP1A1</i> *4	2453C→A	Threonine→asparagine	Exon 7, codon 461 (heme binding region)	<i>CYP1A1</i> *5	<i>CYP1A1</i> *4	Cascorbi et al., 1996 (13)

Information is also lacking on genotype frequencies in different age groups. Most studies consider only the 3801T→C and/or Ile462Val polymorphisms, which has the potential to result in misclassification. When individual polymorphisms are assessed, those persons who do not carry the specific variant may not be true wild-type homozygotes; a proportion may carry another variant. Moreover, the presence of the Thr461Asp polymorphism may interfere with detection of the Ile462Val polymorphism, resulting in overestimation of the *Val* allele if the polymerase chain reaction product has not been digested with *Bsr*D1 (13). Genotype frequencies were in Hardy-Weinberg equilibrium, except in two studies of the 3801T→C polymorphism (63, 65) and nine studies of the Ile462Val polymorphism (19, 41, 53, 64, 67, 70, 76, 78, 82).

3801T→C (CYP1A1*2A, CYP1A1*2B). The 3801C variant is most prevalent in Asian populations, where the frequency of the *C/C* genotype is 2–18 percent and that of the *T/C* genotype is 32–55 percent. In European and White American series, 0–5 percent are *C/C* and 9–28 percent are *T/C*. Frequencies in African Americans are intermediate between White and Asian populations (4–6 percent *C/C*, 35–39 percent *T/C*).

In our pooled analysis, the *C/C* genotype frequency was 13 percent (95 percent confidence interval (CI): 12.0, 14.0) in Asians, 1 percent (95 percent CI: 0.9, 1.4) in Whites, and 6 percent (95 percent CI: 3.7, 8.1) in African Americans. The heterozygote frequency was 44 percent (95 percent CI: 42.6, 45.6) in Asians, 17 percent (95 percent CI: 16.5, 18.0) in Whites, and 36 percent (95 percent CI: 31.7, 40.6) in African Americans.

Ile462Val (CYP1A1*2B, CYP1A1*2C). In all ethnic groups, the *Val* variant occurs less frequently than the 3801C variant. Similar to the 3801C variant, it is most common among Asians, where 1–8 percent are *Val/Val* and 15–46 percent are *Ile/Val*. In Europeans and US Whites, at most 3 percent are *Val/Val* and as many as 15 percent are *Ile/Val*. The *Val* variant is less common among African Americans than Whites. In studies including African Americans, no subjects had the *Val/Val* genotype, but up to 6 percent were *Ile/Val*.

Our pooled estimate of *Val/Val* genotype frequency was 5 percent (95 percent CI: 4.0, 5.0) in Asians, 0.7 percent (95 percent CI: 0.5, 0.8) in Whites, and 0 percent in African Americans. The pooled estimate of *Ile/Val* genotype frequency was 31 percent (95 percent CI: 29.5, 31.7), 8 percent (95 percent CI: 7.8, 8.9), and 5 percent (95 percent CI: 3.2, 7.3), respectively.

3205T→C (CYP1A1*3). The 3205C variant was originally thought to occur in African Americans only. This view is supported by studies of Turkish (34), French (84), German (13), Polish (50), Russian (51), and US White subjects (63, 65) in whom the 3205C variant was not found. In four African-American series, less than 1 percent had the *C/C* genotype, while 14–24 percent were heterozygotes. In our pooled analysis, the *C/C* and *T/C* genotype frequencies were 0.1 percent (95 percent CI: 0.0, 0.8) and 15 percent (95 percent CI: 12.8, 18.3), respectively, in African Americans.

Thr461Asp (CYP1A1*4). *Asp/Asp* homozygotes are very rare (≤ 1 percent). The *Thr/Asp* genotype frequency is 4–12

percent in Turkish, European, and White North American populations. The pooled estimates of the *Asp/Asp* and *Thr/Asp* genotype frequencies in Whites were 0.2 percent (95 percent CI: 0.1, 0.4) and 8 percent (95 percent CI: 7.1, 8.8), respectively.

Associations between the CYP1A1 polymorphisms. Studies of linkage between the polymorphisms are limited by the relative rarity of the variants. From the comparatively few studies carried out, the 3801T→C and Ile462Val polymorphisms appear to be closely linked in Asians (17, 21, 25, 34), less closely linked in Europeans (41, 52), and not linked in African Americans (60). In 81 Africans and African Americans carrying the 3205C variant, 23 percent also carried the 3801C variant, and no subjects carried the *Val* variant (18). The Thr461Asp and 3801T→C polymorphisms were not linked in Turkish (34), German (13), or Polish (50) populations. No evidence for linkage between the Thr461Asp and Ile462Val polymorphisms was found in White American (64) or German series (13).

Functional effects

Because the 3801T→C polymorphism is located in the noncoding region, it was originally thought that any apparent functional consequences of the variant were due to linkage with another polymorphism in, for example, the coding region or the aryl hydrocarbon receptor. However, polymorphisms in noncoding sequences may influence gene function by altering the level, location, or timing of gene expression or messenger RNA stability (86).

Studies of the 3801T→C polymorphism and basal and/or induced CYP1A1 messenger RNA expression in lymphocytes and placenta have been inconsistent (87–91). For the Ile462Val polymorphism, one study found that mean messenger RNA (induced/basal) levels increased with number of *Val* variants (92). In another study, heterozygotes for both 3801C and *Val* variants had twofold increased basal CYP1A1 expression compared with homozygotes for the 3801T and *Ile* alleles. (63). In one study, the 3205T→C and Thr461Asp polymorphisms were not associated with steady-state CYP1A1 messenger RNA levels (87).

The *Val* variant caused a twofold increase in complementary DNA-expressed activity in transformed yeast cells (93), but the kinetic properties of the two variants do not differ (94). In purified *Escherichia coli*, there was no difference between the allelic variants in benzo[a]pyrene bioactivation (95).

Studies of genotype and CYP1A1-dependent enzymatic activity in lymphocytes are inconsistent. Studies either suggested high activity associated with the 3801C and *Val* variants (89, 92, 96–98) or produced null findings (99–102). No significant effect of the Thr461Asp polymorphism has been found (101, 102).

The 3801C variant has been associated with higher levels of DNA adducts in breast tissue in some studies (103, 104), but not others (105–107). Findings from one study of Ile462Val and Thr461Asp polymorphisms and breast tissue adducts were null (106). Results of studies of 3801T→C, Ile462Val, and Thr461Asp polymorphisms in other tissues have been inconsistent (91, 108–112), as have those in white

blood cells (111–122). Findings of studies of 3801T→C and/or Ile462Val and levels of DNA damage, as assessed by 8-hydroxydeoxyguanosine in breast tissue (123), urine (124, 125) or leukocytes (126), or DNA-protein cross-links (33) have been null.

The 3801T→C polymorphism does not appear to be associated with serum estrone or estradiol levels (127) or the ratio of baseline urinary estrogen metabolites (2-hydroxyestrone/16-hydroxyestrone) (63). However, after indole-3-carbinol was ingested, the 2-hydroxyestrone/16-hydroxyestrone ratio increased significantly for persons with the 3801TT genotype; heterozygotes showed no significant increase (63).

In some (74, 128–132), but not all (75, 111, 115, 116, 133–135), studies, the 3801C and Val variants are associated with higher urinary levels of 1-hydroxypyrene, a biomarker for PAH exposure. Results of studies of Ile462Val and urinary levels of 2-naphthol, another PAH biomarker, are inconsistent (74, 135). Findings of studies of the *CYP1A1* genotype and urinary levels of cotinine (136), malondialdehyde (124), and biomarkers for organic solvent exposure (137) have been null.

Mammographic breast density is positively related to breast cancer risk (138, 139). In one study, neither the 3801T→C nor the Ile462Val polymorphisms were associated with breast density (140).

DISEASE

In 2002, over 1 million new cases of breast cancer were diagnosed worldwide (141). In both developed and developing countries, it is the most common cancer in women (142). In developed countries, incidence increases rapidly with age to about age 50 years; thereafter, rates rise less rapidly with age (143). There is a 16-fold variation in incidence between the population with the highest rate (Montevideo, Uruguay, world age-standardized incidence 114.9 per 100,000 in 1993–1995) and that with the lowest (The Gambia, 7.0 per 100,000 in 1997–1998) (144). In many populations, there has been a consistent long-term rise in incidence, which cannot be entirely attributed to the introduction of mammographic screening (145).

The autosomal dominant susceptibility genes, *BRCA1* and *BRCA2*, account for about 5 percent of breast cancers (146, 147). Familial aggregation, which confers increased risk for first- and second-degree relatives (148, 149), does not appear to be entirely due to *BRCA1* and *BRCA2* (150), suggesting that other aspects of genetic susceptibility are important.

The products of *CYP1A1* are involved in estrogen and PAH metabolism. The most firmly established risk factors for breast cancer relate to cumulative exposure of the breast to endogenous hormones, particularly estrogen (143). Risk is increased for women with longer cumulative exposure, that is, for those experiencing early menarche, late menopause, late first full-term pregnancy, or no pregnancies (151).

Exogenous hormones have also been associated with increased risk of breast cancer. Risk is increased among current users of hormone replacement therapy (152, 153) and current users of oral contraceptives (154, 155). Other

lifestyle risk factors, such as postmenopausal obesity, lack of physical activity (156), and alcohol intake (157), may influence risk via effects on estrogen levels.

PAHs may be involved in breast cancer etiology. These substances are lipophilic and are stored in adipose tissue, including that of the breast (158), and they are activated and metabolized by breast epithelial cells (159). Adduct levels are higher in normal breast tissue of breast cancer cases than in that of healthy controls (160), although it is unclear whether this is a cause or effect of disease. PAHs also affect estrogen production and metabolism, thereby acting as xenoestrogens; many xenoestrogenic compounds induce mammary carcinogenesis in experimental animals (161). PAHs themselves are powerful mammary carcinogens in mice (162).

Tobacco smoke is a major environmental source of PAH exposure (163). Most studies of breast cancer and smoking show a weak positive or null association (164–170), although the association may be stronger for premenopausal women or for those who started smoking at an early age (171, 172) or smoked before their first full-term pregnancy (173). Although positive associations with passive smoking have been reported (168, 174, 175), a recent review concluded that this factor was unlikely to increase risk (176).

PAHs (and heterocyclic amines) are formed when meats are exposed to temperatures that cause pyrolysis (177). An expert review of observational evidence and a recent meta-analysis suggested that high-meat diets increase breast cancer risk (178, 179), and, whereas most investigators have not considered cooking methods, some studies found raised risk with increased consumption of fried, broiled, and/or well-done meat (180–183).

ASSOCIATIONS

Web tables 5, 6, 7, and 8 summarize 17 studies of *CYP1A1* and breast cancer risk (25, 32, 33, 38, 45, 56, 63, 65, 66, 73, 79, 81, 83, 107, 127, 184, 185) identified by using the search strategy described earlier, with the addition of the MeSH heading “Breast neoplasms” or the text word “breast.” The subjects included in the studies of Huang et al. (32, 186), Taioli et al. (63, 187), Li et al. (105) and Zhu et al. (107), and Ritchie et al. (188) and Bailey et al. (65) may overlap. Therefore, only the largest of each set was included in Web tables 5, 6, 7, and 8 and in our meta-analyses.

Meta-analyses of studies of 3801T→C, Ile462Val, and Thr461Asp were carried out. From the papers, we abstracted the odds ratios or relative risks for homozygous variants (3801C/C, Val/Val, or Asp/Asp) and heterozygotes (3801T/C, Ile/Val, or Thr/Asp) versus homozygous wild types (3801T/T, Ile/Ile, or Thr/Thr). When reported, the adjusted effect estimate was included in the analysis in preference to the unadjusted one. If odds ratios were not reported, we computed unadjusted odds ratios from the data presented. Analyses were conducted by using Stata statistical software, release 7.0 (189). Heterogeneity was assessed by the *Q* test, with a fixed-effects model used if $p \geq 0.1$ and a random-effects model used if $p < 0.1$. The I^2 statistic was also calculated as a measure of consistency between studies

(190). Except for the association between breast cancer and the 3801T → C polymorphism, the estimates of effect in the first published study were similar to those for the cumulative meta-analyses.

Study characteristics

Four studies took place in Japan, two in Taiwan, six in the United States, and one each in Canada, Brazil, France, Greece, and the United Kingdom. Thirteen studies analyzed the 3801T → C polymorphism (2,484 cases), 10 analyzed the Ile462Val polymorphism (3,535 cases), two analyzed the 3205T → C polymorphism (280 cases), and three analyzed the Thr461Asp polymorphism (2,245 cases).

In one study, case DNA was derived from tumor specimens (38); in the remainder, and for all control series, DNA came from blood samples. Of the US studies, two involved subjects of whom the majority (or all) were White, three included more than one ethnic group (analyzed separately in two studies), and, in one, ethnicity was not reported. One study included postmenopausal women only; all others either consisted of both pre- and postmenopausal women ($n = 6$) or did not describe the subjects' menopausal status ($n = 10$). Eleven studies included fewer than 200 breast cancer cases.

In 15 studies, cases were recruited from clinics or hospital series; in one study, cases were identified from a cancer registry; and one study was nested within the Nurses' Health Study. Without information on all potentially eligible cases in the population, it is difficult to assess the generalizability of the results. At least four control series included "volunteers" from either an unspecified source or a convenient population such as medical workers—a potential source of bias. Seven studies presented estimates adjusted for potential confounding factors.

In general, the studies considered the polymorphisms separately. Therefore, the effect of one polymorphism may have been overshadowed by the effects of others, whereas construction of haplotypes may have revealed effects that were not apparent by analyzing single polymorphisms. Studies of the Ile462Val polymorphism, with the exception of those by Bailey et al. (65), Krajcinovic et al. (56), and Basham et al. (79), may have suffered from some minor misclassification due to the undetected presence of the Thr461Asp polymorphism.

3801T → C (CYP1A1*2A, CYP1A1*2B)

Most studies found no evidence of an association between the 3801T → C polymorphism and breast cancer risk (33, 38, 45, 56, 65, 107, 127, 184, 185) (Web table 5). In Taiwan, women with the C/C genotype had a raised risk compared with other genotypes combined (32). African-American women with the C/C genotype also had an increased risk compared with those with the T/T genotype (63), but this study included only 25 cases. The 3801C variant was associated with reduced risk for Japanese and non-White Brazilian women (25, 66). However, in both studies, the cases were surgical series, and controls were not population based.

Our meta-analysis included eight studies for which data were available for all three genotypes separately (25, 32, 38, 45, 56, 63, 65, 66). Breast cancer risk did not differ from unity for C/C versus T/T (random-effects relative risk (RR) = 0.97, 95 percent CI: 0.52, 1.80; $Q = 15.26$, $p = 0.08$) or for T/C versus T/T (random-effects RR = 0.91, 95 percent CI: 0.70, 1.19; $Q = 17.34$, $p = 0.07$). The I^2 statistics for these analyses were 41 percent and 42 percent, respectively, indicating moderate heterogeneity across studies.

Ile462Val (CYP1A1*2B, CYP1A1*2C)

A Japanese study found a significantly reduced risk for women with the Ile/Val genotype compared with the Ile/Ile genotype (RR = 0.66, 95 percent CI: 0.44, 0.99) (25). However, meta-analysis found no association between breast cancer risk and the Val/Val (fixed-effects RR = 1.04, 95 percent CI: 0.63, 1.74; $Q = 4.59$, $p = 0.33$, $I^2 = 13$ percent) (25, 32, 73, 79, 81) or Ile/Val (fixed-effects RR = 0.92, 95 percent CI: 0.76, 1.10; $Q = 11.57$, $p = 0.17$, $I^2 = 31$ percent) (25, 32, 56, 63, 65, 73, 79, 81) genotypes versus the Ile/Ile genotype.

3205T → C (CYP1A1*3)

There was no association between the 3205C variant and breast cancer in the two available studies (Web table 7). However, these studies each included small series ($n = 27$ and $n = 59$) of African-American breast cancer cases (63, 65).

Thr461Asp (CYP1A1*4)

In a Canadian study, carriers of the Asp variant had an increased breast cancer risk (adjusted RR = 3.3, 95 percent CI: 1.1, 9.7) (56) (Web table 8). Results of the other studies, in White American women and African-American women, and in White women in England, were null. Meta-analysis found no association between disease risk and the Asp/Asp (fixed-effects RR = 0.95, 95 percent CI: 0.20, 4.49; $Q = 0.52$, $p = 0.77$, $I^2 = 0$ percent) or Thr/Asp (fixed-effects RR = 1.12, 95 percent CI: 0.87, 1.43; $Q = 0.89$, $p = 0.64$, $I^2 = 0$ percent) genotypes versus the Thr/Thr genotype (56, 65, 79).

Combinations of genotypes

Taioli et al. (187) assessed the impact of combinations of 3801T → C, Ile462Val, and 3205T → C genotypes on breast cancer risk. Compared with homozygotes for the 3801T, Ile, and 3205T alleles, only the 3801C/C genotype was associated with increased risk for African-American women (RR = 5.8, 95 percent CI: 1.0, 36.0), but the effect estimate was imprecise.

One study combined Ile462Val and Thr461Asp genotypes and found no significant effect in any of the three combined genotype groups relative to the group with the Ile/Ile and Thr/Thr genotypes (79). Another study assessed disease risk for subjects with either the Val or Asp variant; no significant association was found in White women or African-American women (65).

Subgroup analyses

Menopausal status, age at menarche, and estrogen and progesterone receptor status. In a Taiwanese study, the association of the 3801C/C genotype with raised disease risk was evident in postmenopausal, but not premenopausal, women, and further analysis suggested that the relation might be more pronounced for women experiencing early menarche (32). Other studies found no association between the 3801C variant and breast cancer when subjects were stratified by menopausal status (66, 185) or age at menarche (66). There was no evidence for an association with the Ile462Val polymorphism for either pre- or postmenopausal women (32, 65, 79, 185). In a Canadian study, the increased risk associated with the Thr461Asp polymorphism was evident for postmenopausal women only (56); however, a study in the United Kingdom found no difference in Thr461Asp genotypic risks by menopausal status (79). There were no significant associations between *CYP1A1* polymorphisms and estrogen or progesterone receptor status (25, 65, 127, 185).

Age at diagnosis and clinical characteristics. Studies investigating *CYP1A1* genotype and age at diagnosis of breast cancer have produced inconsistent results (25, 32, 38, 65, 81). The 3801C variant has been significantly associated with a higher frequency of lymph-node metastasis and the *Val* variant with a higher frequency of small tumors (<2 cm), while neither variant was associated with histology or histologic grade (25). Other studies found no association between the four polymorphisms and tumor size, stage, type, grade, or nodal status (65) or between 3801T→C or Ile462Val and tumor type or stage of disease (185).

Survival

In a British study of 1,793 incident or prevalent breast cancer cases, the Ile462Val polymorphism was not related to survival (191). The hazard ratio was reduced for *Thr/Asp* heterozygotes compared with *Thr/Thr* homozygotes, but not significantly (hazard ratio = 0.67, 95 percent CI: 0.33, 1.37) (191).

Other diseases

CYP1A1 has been explored in relation to several cancers, particularly those in which smoking is implicated. In pooled and meta-analyses, the 3801C and *Val* variants were associated with increased lung cancer risk in Whites, but not Asians (192–195). Neither the 3205C (196) nor the *Asp* (13, 197) variants were associated with lung cancer risk. Also investigated, with mainly either inconsistent or unconfirmed results, have been tumors of the head and neck (29, 31, 40, 43, 48, 82, 198–215), large bowel (24, 53, 58, 216–221), prostate (72, 222–224), female gynecologic sites (62, 76, 225–230), skin (231, 232), and kidney (84) and liver (28), as well as leukemias and lymphomas (36, 57, 78, 233–238). Results of studies of bladder (44), brain (239), and pancreatic (64, 108, 240–242) cancer have been null.

Associations have been found between *CYP1A1* and other diseases, including male infertility (243), systemic lupus

erythematosus (244), type II porphyria cutanea tarda (245), psoriasis (39), ankylosing spondylitis (246), and rheumatoid arthritis (247). Findings from studies of endometriosis (46, 248, 249) and Parkinson's disease (54, 68, 250, 251) have been inconsistent, while those for asthma (252), atherosclerosis (253), cirrhosis (37), Crohn's disease (254), age-related macular degeneration (255), leukoplakia (20), early pregnancy loss (256), acne (41), and oral clefting (257) have been null.

INTERACTIONS

If *CYP1A1* is involved in breast cancer, it may influence disease risk by interacting with exposure (or indicators of exposure) to PAHs or estrogen, for example, or with other genes involved in the metabolism of carcinogens, estrogens, or other hormones. Sample size is particularly important in this context. For instance, to detect a multiplicative interaction, very large sample sizes are required for adequate power (258). Although the sample size needed to detect other types of interactions may be smaller (259), a priori it is not usually clear what model of interaction would be predicted.

Gene-environment interactions

Smoking. In five studies investigating genotype-smoking interactions (63, 65, 79, 81, 185), two found evidence of an interaction (81, 185). In Ambrosone et al.'s study (81), adjusted relative risks for *Val* carriers versus *Ile/Ile* homozygotes among nonsmokers, light smokers (<29 pack-years of exposure), and heavy smokers (≥29 pack-years) were 1.3 (95 percent CI: 0.62, 2.70), 5.2 (95 percent CI: 1.16, 23.56), and 0.9 (95 percent CI: 0.24, 3.09), respectively, but no formal test of interaction was conducted. Ishibe et al. (185) found no interaction between pack-years of smoking and either 3801T→C or Ile462Val polymorphisms, but they observed effect modification for smoking status at diagnosis and age at which smoking started. Risk was significantly raised for current smokers carrying the 3801C variant versus 3801T/T nonsmokers (*p* for interaction = 0.06) and for women with either variant who started smoking before age 18 years versus 3801T/T nonsmokers (*p* for interaction = 0.04) and *Ile/Ile* nonsmokers (*p* for interaction = 0.08).

The numbers analyzed in the studies of genotype-smoking interactions were small, and interpretation is difficult because of differences in the way in which interactions were assessed (stratifying by smoking status (63, 81) or genotype (65), or using a single reference group of smoking status and genotype combined (185)) and in categorization of smoking status. For example, the interaction patterns observed by Ambrosone et al. (81) or Ishibe et al. (185) would not be detectable by using an ever/never smoking categorization, as has been used in other studies (63, 65).

Polychlorinated biphenyls (PCBs). PCBs have been linked to breast cancer risk because of their estrogenic (260) and tumor-promoting (261) properties. In the Nurses' Health Study, a modest interaction between the Ile462Val polymorphism and plasma PCBs was found for postmenopausal,

but not premenopausal, breast cancer (262). Among postmenopausal subjects, the adjusted relative risk for *Val* carriers in the upper tertile of plasma PCB levels, compared with *Ile/Ile* homozygotes in the lowest PCB tertile, was 2.78 (95 percent CI: 0.99, 7.82, p for interaction = 0.05). There was no interaction between PCBs and 3801T→C (262). In a subset of a study in western New York (81), *Val* carriers with an above-median PCB body burden had an increased risk compared with *Ile/Ile* homozygotes with a below-median PCB burden (adjusted RR = 2.9, 95 percent CI: 1.18, 7.45; p for interaction = 0.13) (263).

Alcohol. Basham et al. (79) reported no interactions between Ile462Val or Thr461Asp polymorphisms and alcohol consumption. However, results were not shown.

Gene-gene interactions

CYP1A1 and glutathione *S*-transferase (*GST*) gene activities may be interrelated. The *GST* genes belong to the *Ah* gene battery, since *GST* is one of six enzymes regulated by the aryl hydrocarbon receptor (1). In human B-cell lines, absence of *GSTM1* was associated with induction of high levels of *CYP1A1* messenger RNA by 2,3,7,8-tetrachlorodibenzo-*p*-dioxin, and presence of *GSTM1* was associated with induction of low levels (264). Four breast cancer studies found no evidence of a *CYP1A1-GSTM1* interaction (56, 65, 66, 81), although *CYP1A1-GSTM1* genotype combinations have been associated with age at presentation (38). In two relatively small studies assessing *CYP1A1-GSTT1* genotype combinations, risk estimates were not significant, and tests for interaction were not reported (65, 66). Another study (45) found no significant differences in combined *CYP1A1* 3801T→C, *GSTM1*, and *GSTT1* genotype frequencies between breast cancer patients and controls.

CYP17, *CYP19*, and catechol-*O*-methyltransferase (*COMT*) are involved in steroid hormone metabolism (265, 266). One study found an increased breast cancer risk associated with the presence of two "high-risk" genotypes, defined as homozygosity for the *CYP1A1* 3801C, *CYP17A*₂, or *COMT* low-activity alleles (RR = 3.5, 95 percent CI: 1.06, 12.04), but no test for interaction was conducted (186). In another study, 3801T homozygotes carrying the *CYP19* (*TTTA*)_{7(-3bp)} allele had increased risk of estrogen-receptor-positive breast cancer (adjusted RR = 3.00, 95 percent CI: 1.56, 5.74) compared with women carrying the 3801C variant but not the *CYP19* (*TTTA*)_{7(-3bp)} allele (127).

LABORATORY TESTS

CYP1A1 3801T→C, Ile462Val, 3205T→C, and Thr461Asp polymorphisms are detected by using polymerase chain reaction followed by digestion with *MspI* for 3801T→C (17, 38, 65); *NcoI* (65, 81, 185), *HincII* (32), or *BsrD1* (65) for Ile462Val; *MspI* for 3205T→C (16); and *BsaI* for Thr461Asp (65). The polymorphisms 3801T→C and 3205T→C can be detected simultaneously from one polymerase chain reaction product by using *MspI* and *SphI* (65). For accurate genotyping of Ile462Val, the presence of *Val* (and absence of *Asp*) can be verified by *BsrD1* digestion (13).

Success rates for DNA extraction and genotype assignment, and reproducibility, are important indicators of analytic validity of genotyping (267), but few breast cancer studies reported this information. Taioli et al. (187) and Ishibe et al. (185) successfully assigned 3801T→C genotype to 99.7 percent and 99.8 percent, respectively, of subjects providing samples, and Ile462Val genotype to 95.9 percent and 96.6 percent, respectively; however, Ambrosone et al. (81) obtained interpretable polymerase chain reaction assays for only 69 percent of subjects consenting to phlebotomy.

POPULATION TESTING

Current evidence does not suggest that there would be value in testing for the *CYP1A1* genotype in isolation to predict breast cancer risk. In addition, the evidence on joint effects of *CYP1A1* variants and variants of other genes is very limited. The possibility of raised risk associated with some genotypes in combination with tobacco exposure should be addressed via standard public health advice on smoking cessation.

CONCLUSIONS AND RECOMMENDATIONS FOR RESEARCH

The *CYP1A1* 3801C variant is more common than the *Val* variant. Both variants occur more frequently in Asian than in White populations. The 3205C variant has been observed in African Americans only, and little data are available on the geographic or ethnic distribution of the Thr461Asp polymorphism. The 3205T→C and Thr461Asp polymorphisms should be investigated in African, Asian, and Hispanic populations. The functional significance of all four polymorphisms is unclear, which could be due to the small sample sizes of most studies. Further investigation is warranted.

No consistent associations between breast cancer and *CYP1A1* polymorphisms were found. While meta-analyses have greater power and precision for detecting gene-disease associations, our meta-analyses were limited by different genotype categorizations between studies. For the purposes of future meta-analyses, authors should provide results for all genotypes separately.

The 3801T→C and Ile462Val polymorphisms may modify the smoking-disease association, although the evidence is limited and inconsistent. A similar "inverse dose effect" has been observed in studies of *CYP1A1*, smoking, and lung cancer (60, 184, 268), and it has been suggested that the genetic variant might confer increased sensitivity to lower levels of exposure (269). Additional investigation is needed. The *Val* variant may interact with PCB levels to affect breast cancer risk, but confirmation is necessary. There was no evidence that *GSTM1* or *GSTT1* and *CYP1A1* genotypes have a joint effect on disease risk. Studies suggesting interactions with *CYP17*, *CYP19*, and *COMT* should be replicated. Interpreting the studies of interaction was difficult because of the different approaches used; adopting a more unified approach (e.g., Botto and Khoury (270)) in future studies would aid interpretation and synthesis of evidence.

Studies are needed to explore joint effects on breast cancer risk of the *CYP1A1* genotype and 1) sources of PAH exposure other than tobacco, 2) markers of exposure to endogenous estrogens, 3) exposure to exogenous estrogens, 4) other lifestyle factors that influence hormone levels, 5) other genes encoding enzymes involved in PAH metabolism, and 6) other genes involved in hormonal biosynthesis. To detect gene-environment or gene-gene interactions, future studies must be large, and pooled analyses should be considered.

REFERENCES

- Nebert DW. Role of genetics and drug metabolism in human cancer risk. *Mutat Res* 1991;247:267–81.
- McManus ME, Burgess WM, Veronese ME, et al. Metabolism of 2-acetylaminofluorene and benzo(a)pyrene and activation of food-derived heterocyclic amine mutagens by human cytochrome P-450. *Cancer Res* 1990;50:3367–76.
- Dannan GA, Porubek DJ, Nelson SD, et al. 17 beta-estradiol 2- and 4-hydroxylation catalyzed by rat hepatic cytochrome P-450: roles of individual forms, inductive effects, developmental patterns, and alterations by gonadectomy and hormone replacement. *Endocrinology* 1986;118:1952–60.
- Spink DC, Eugster HP, Lincoln DW II, et al. 17 beta-estradiol hydroxylation catalyzed by human cytochrome P450 1A1: a comparison of the activities induced by 2,3,7,8-tetrachlorodibenzo-p-dioxin in MCF-7 cells with those from heterologous expression of the cDNA. *Arch Biochem Biophys* 1992;293:342–8.
- Kawajiri K, Watanabe J, Gotoh O, et al. Structure and drug inducibility of the human cytochrome P-450c gene. *Eur J Biochem* 1986;159:219–25.
- Spink DC, Spink BC, Cao JQ, et al. Differential expression of *CYP1A1* and *CYP1B1* in human breast epithelial cells and breast tumor cells. *Carcinogenesis* 1998;19:291–8.
- Smith CAD, Smith G, Wolf CR. Genetic polymorphisms in xenobiotic metabolism. *Eur J Cancer* 1994;30A:1921–35.
- Huang Z, Fasco MJ, Figge HL, et al. Expression of cytochromes P450 in human breast tissue and tumors. *Drug Metab Dispos* 1996;24:899–905.
- McKay JA, Murray GI, Ah-See AK, et al. Differential expression of *CYP1A1* and *CYP1B1* in human breast cancer. *Biochem Soc Trans* 1996;24:327S.
- Nebert DW, Ingelman-Sundberg M, Daly AK. Genetic epidemiology of environmental toxicity and cancer susceptibility: human allelic polymorphisms in drug-metabolizing enzyme genes, their functional importance, and nomenclature issues. *Drug Metab Rev* 1999;31:467–87.
- Human Cytochrome P 450 Allele Nomenclature Committee. *CYP1A1* allele nomenclature. 2004. (<http://www.imm.ki.se/CYPalleles/cyp1a1.htm>).
- Garte S. The role of ethnicity in cancer susceptibility gene polymorphisms: the example of *CYP1A1*. *Carcinogenesis* 1998;19:1329–32.
- Cascorbi I, Brockmoller J, Roots I. A C 4887A polymorphism in exon 7 of human *CYP1A1*: population frequency, mutation linkages, and impact on lung cancer susceptibility. *Cancer Res* 1996;56:4965–9.
- Bale AE, Nebert DW, McBride OW. Subchromosomal localization of the dioxin-inducible P1450 locus (*CYP1*) and description of two RFLPs detected within a 3' P1450 cDNA probe. *Cytogenet Cell Genet* 1987;46:574–5.
- Spurr NK, Gough AC, Stevenson K, et al. *Msp*-I polymorphism detected with a cDNA probe for the P-450 I family on chromosome 15. *Nucleic Acids Res* 1987;15:5901.
- Crofts F, Cosma GN, Currie D, et al. A novel *CYP1A1* gene polymorphism in African-Americans. *Carcinogenesis* 1993;14:1729–31.
- Hayashi S, Watanabe J, Nakachi K, et al. Genetic linkage of lung cancer-associated *Msp*I polymorphisms with amino acid replacement in the heme binding region of the human cytochrome P4501A1 gene. *J Biochem* 1991;110:407–11.
- Garte S, Gaspari L, Alexandrie AK, et al. Metabolic gene polymorphism frequencies in control populations. *Cancer Epidemiol Biomarkers Prev* 2001;10:1239–48.
- Song N, Tan W, Xing D, et al. *CYP 1A1* polymorphism and risk of lung cancer in relation to tobacco smoking: a case-control study in China. *Carcinogenesis* 2001;22:11–16.
- Sikdar N, Mahmud SA, Paul RR, et al. Polymorphism in *CYP1A1* and *CYP2E1* genes and susceptibility to leukoplakia in Indian tobacco users. *Cancer Lett* 2003;195:33–42.
- Kotekar A, Bhisey R. *CYP1A1**2 and *CYP1A1**3 polymorphisms cosegregate in the Indian population. *Anthropol Anz* 2002;60:255–60.
- Kiyohara C, Wakai K, Mikami H, et al. Risk modification by *CYP1A1* and *GSTM1* polymorphisms in the association of environmental tobacco smoke and lung cancer: a case-control study in Japanese nonsmoking women. *Int J Cancer* 2003;107:139–44.
- Gorai I, Tanaka K, Inada M, et al. Estrogen-metabolizing gene polymorphisms, but not estrogen receptor-alpha gene polymorphisms, are associated with the onset of menarche in healthy postmenopausal Japanese women. *J Clin Endocrinol Metab* 2003;88:799–803.
- Inoue H, Kiyohara C, Marugame T, et al. Cigarette smoking, *CYP1A1* *Msp*I and *GSTM1* genotypes, and colorectal adenomas. *Cancer Res* 2000;60:3749–52.
- Miyoshi Y, Takahashi Y, Egawa C, et al. Breast cancer risk associated with *CYP1A1* genetic polymorphisms in Japanese women. *Breast J* 2002;8:209–15.
- Nakachi K, Imai K, Hayashi S, et al. Genetic susceptibility to squamous cell carcinoma of the lung in relation to cigarette smoking dose. *Cancer Res* 1991;51:5177–80.
- Kihara M, Kihara M, Noda K. Risk of smoking for squamous and small cell carcinomas of the lung modulated by combinations of *CYP1A1* and *GSTM1* gene polymorphisms in a Japanese population. *Carcinogenesis* 1995;16:2331–6.
- Yu MW, Chiu YH, Yang SY, et al. Cytochrome P450 1A1 genetic polymorphisms and risk of hepatocellular carcinoma among chronic hepatitis B carriers. *Br J Cancer* 1999;80:598–603.
- Wu M, Lee J, Wu D, et al. Genetic polymorphisms of cytochrome P4501A1 and oesophageal squamous-cell carcinoma in Taiwan. *Br J Cancer* 2002;87:529–32.
- Lin P, Wang SL, Wang HJ, et al. Association of *CYP1A1* and microsomal epoxide hydrolase polymorphisms with lung squamous cell carcinoma. *Br J Cancer* 2000;82:852–7.
- Cheng YJ, Chien YC, Hildesheim A, et al. No association between genetic polymorphisms of *CYP1A1*, *GSTM1*, *GSTT1*, *GSTP1*, *NAT2*, and nasopharyngeal carcinoma in Taiwan. *Cancer Epidemiol Biomarkers Prev* 2003;12:179–80.
- Huang CS, Shen CY, Chang KJ, et al. Cytochrome P4501A1 polymorphism as a susceptibility factor for breast cancer in postmenopausal Chinese women in Taiwan. *Br J Cancer* 1999;80:1838–43.

33. Wu FY, Lee YJ, Chen DR, et al. Association of DNA-protein crosslinks and breast cancer. *Mutat Res* 2002;501:69–78.
34. Aynacioglu AS, Cascorbi I, Mrozikiewicz PM, et al. High frequency of CYP1A1 mutations in a Turkish population. *Arch Toxicol* 1998;72:215–18.
35. Sarmanova J, Tynkova L, Susova S, et al. Genetic polymorphisms of biotransformation enzymes: allele frequencies in the population of the Czech Republic. *Pharmacogenetics* 2000;10:781–8.
36. Sarmanova J, Benesov K, Gut I, et al. Genetic polymorphisms of biotransformation enzymes in patients with Hodgkin's and non-Hodgkin's lymphomas. *Hum Mol Genet* 2001;10:1265–73.
37. Lucas D, Menez C, Floch F, et al. Cytochromes P450E1 and P4501A1 genotypes and susceptibility to cirrhosis or upper aerodigestive tract cancer in alcoholic Caucasians. *Alcohol Clin Exp Res* 1996;20:1033–7.
38. Fontana X, Peyrottes I, Rossi C, et al. Study of the frequencies of CYP1A1 gene polymorphisms and glutathione S-transferase mu1 gene in primary breast cancers: an update with an additional 114 cases. *Mutat Res* 1998;403:45–53.
39. Richter-Hintz D, Their R, Steinwachs S, et al. Allelic variants of drug metabolizing enzymes as risk factors in psoriasis. *J Invest Dermatol* 2003;120:765–70.
40. Ko Y, Abel J, Harth V, et al. Association of CYP1B1 codon 432 mutant allele in head and neck squamous cell cancer is reflected by somatic mutations of p53 in tumor tissue. *Cancer Res* 2001;61:4398–404.
41. Paraskevaidis A, Drakoulis N, Roots I, et al. Polymorphisms in the human cytochrome P-450 1A1 gene (CYP1A1) as a factor for developing acne. *Dermatology* 1998;196:171–5.
42. Harth V, Bruning T, Abel J, et al. Real-time genotyping of cytochrome P4501A1 A4889G and T6235C polymorphisms. *Mol Cell Probes* 2001;15:93–7.
43. Matthias C, Bockmuhl U, Jahnke V, et al. Polymorphism in cytochrome P450 CYP2D6, CYP1A1, CYP2E1 and glutathione S-transferase, GSTM1, GSTM3, GSTT1 and susceptibility to tobacco-related cancers: studies in upper aerodigestive tract cancers. *Pharmacogenetics* 1998;8:91–100.
44. Brockmoller J, Cascorbi I, Kerb R, et al. Combined analysis of inherited polymorphisms in arylamine N-acetyltransferase 2, glutathione S-transferases M1 and T1, microsomal epoxide hydrolase and cytochrome P450 enzymes as modulators of bladder cancer risk. *Cancer Res* 1996;56:3915–25.
45. Dialyna IA, Arvanitis DA, Spandidos DA. Genetic polymorphisms and transcriptional pattern analysis of CYP1A1, AhR, GSTM1, GSTP1 and GSTT1 genes in breast cancer. *Int J Mol Med* 2001;8:79–87.
46. Arvanitis DA, Koumantakis GE, Goumenou AG, et al. CYP1A1, CYP19, and GSTM1 polymorphisms increase the risk of endometriosis. *Fertil Steril* 2003;79:702–9.
47. Taioli E, Mari D, Franceschi C, et al. Polymorphisms of drug-metabolizing enzymes in healthy nonagenarians and centenarians: difference at GSTT1 locus. *Biochem Biophys Res Commun* 2001;280:1389–92.
48. van Lieshout EM, Roelofs HM, Dekker S, et al. Polymorphic expression of the glutathione S-transferase P1 gene and its susceptibility to Barretts' esophagus and esophageal carcinoma. *Cancer Res* 1999;59:586–9.
49. Tefre T, Ryberg D, Haugen A, et al. Human CYP1A1 (cytochrome P1450) gene: lack of association between the MspI restriction fragment length polymorphism and incidence of lung cancer in a Norwegian population. *Pharmacogenetics* 1991;1:20–5.
50. Mrozikiewicz PM, Cascorbi I, Brockmoller J, et al. CYP1A1 mutations 4887A, 4889G, 5639C and 6235C in the Polish population and their allelic linkage, determined by peptide nucleic acid-mediated PCR clamping. *Pharmacogenetics* 1997;7:303–7.
51. Gaikovitch EA, Cascorbi I, Mrozikiewicz PM, et al. Polymorphisms of drug-metabolizing enzymes CYP2C9, CYP2C19, CYP2D6, CYP1A1, NAT2 and of P-glycoprotein in a Russian population. *Eur J Clin Pharmacol* 2003;59:303–12.
52. Alexandrie AK, Sundberg MI, Seidegard J, et al. Genetic susceptibility to lung cancer with special emphasis on CYP1A1 and GSTM1: a study on host factors in relation to age at onset, gender and histological cancer types. *Carcinogenesis* 1994;15:1785–90.
53. Sachse C, Smith G, Wilkie MJV, et al. A pharmacogenetic study to investigate the role of dietary carcinogens in the etiology of colorectal cancer. *Carcinogenesis* 2002;23:1839–49.
54. Nicholl DJ, Bennett P, Hiller L, et al. A study of five candidate genes in Parkinson's disease and related neurodegenerative disorders. *Neurology* 1999;53:1415–21.
55. Brockton NT. Genetic polymorphisms in folate and xenobiotic metabolism and susceptibility to colorectal cancer. Aberdeen, United Kingdom: University of Aberdeen, 2002.
56. Krajcinovic M, Ghardirian P, Richer C, et al. Genetic susceptibility to breast cancer in French-Canadians: role of carcinogen-metabolizing enzymes and gene-environment interactions. *Int J Cancer* 2001;92:220–5.
57. Krajcinovic M, Labuda D, Richer C, et al. Susceptibility to childhood acute lymphoblastic leukemia: influence of CYP1A1, CYP2D6, GSTM1, and GSTT1 genetic polymorphisms. *Blood* 1999;93:1496–501.
58. Ishibe N, Stampfer M, Hunter DJ, et al. A prospective study of cytochrome P450 1A1 polymorphisms and colorectal cancer risk in men. *Cancer Epidemiol Biomarkers Prev* 2000;9:855–6.
59. Garcia-Closas M, Kelsey KT, Wiencke JK, et al. A case-control study of cytochrome P450 1A1, glutathione S-transferase M1, cigarette smoking and lung cancer susceptibility (Massachusetts, United States). *Cancer Causes Control* 1997;8:544–53.
60. Taioli E, Ford J, Trachman J, et al. Lung cancer risk and CYP1A1 genotype in African Americans. *Carcinogenesis* 1998;19:813–17.
61. Xu X, Kelsey KT, Wiencke JK, et al. Cytochrome P450 CYP1A1 MspI polymorphism and lung cancer susceptibility. *Cancer Epidemiol Biomarkers Prev* 1996;5:687–92.
62. Terry KL, Titus-Ernstoff L, Garner EO, et al. Interaction between CYP1A1 polymorphic variants and dietary exposures influencing ovarian cancer risk. *Cancer Epidemiol Biomarkers Prev* 2003;12:187–90.
63. Taioli E, Bradlow HL, Garbers SV, et al. Role of estradiol metabolism and CYP1A1 polymorphisms in breast cancer risk. *Cancer Detect Prevent* 1999;23:232–7.
64. Duell EJ, Holly EA, Bracci PM, et al. A population-based, case-control study of polymorphisms in carcinogen-metabolizing genes, smoking, and pancreatic adenocarcinoma risk. *J Natl Cancer Inst* 2002;94:297–306.
65. Bailey LR, Roodi N, Verrier CS, et al. Breast cancer and CYP1A1, GSTM1, and GSTT1 polymorphisms: evidence of a lack of association in Caucasians and African Americans. *Cancer Res* 1998;58:65–70.
66. da Fonte de Amorim LM, Rossini A, Mendonca GAS, et al. CYP1A1, GSTM1, and GSTT1 polymorphisms and breast

- cancer risk in Brazilian women. *Cancer Lett* 2002;181:179–86.
67. London SJ, Yuan JM, Coetzee GA, et al. CYP1A1 I462V genetic polymorphisms and lung cancer risk in a cohort of men in Shanghai, China. *Cancer Epidemiol Biomarkers Prev* 2000;9:987–91.
 68. Chan DK, Mellick GD, Buchanan DD, et al. Lack of association between CYP1A1 polymorphism and Parkinson's disease in a Chinese population. *J Neural Transm* 2002;109:35–9.
 69. Buch S, Kotekar A, Kawle D, et al. Polymorphisms at CYP and GST gene loci. Prevalence in the Indian population. *Eur J Clin Pharmacol* 2001;57:553–5.
 70. Oyama T, Mitsudomi T, Kawamoto T, et al. Detection of CYP1A1 gene polymorphism using designed RFLP and distributions of CYP1A1 genotypes in Japanese. *Int Arch Occup Environ Health* 1995;67:253–6.
 71. Oyama T, Kawamoto T, Mizoue T, et al. p53 mutations of lung cancer are not significantly affected by CYP1A1 or GSTM1 polymorphisms. *Int J Oncol* 1997;11:305–9.
 72. Murata M, Shiraishi T, Fukutome K, et al. Cytochrome P4501A1 and glutathione S-transferase M1 genotypes as risk factors for prostate cancer in Japan. *Jpn J Clin Oncol* 1998;28:657–60.
 73. Hayashi S, Watanabe J, Kawajiri K. High susceptibility to lung cancer analyzed in terms of combined genotypes of P4501A1 and Mu-class glutathione S-transferase genes. *Jpn J Cancer Res* 1992;83:866–70.
 74. Lee CY, Lee JY, Kang JW, et al. Effects of genetic polymorphisms of CYP1A1, CYP2E1, GSTM1, and GSTT1 on the urinary levels of 1-hydroxypyrene and 2-naphthol in aircraft maintenance workers. *Toxicol Lett* 2001;123:115–24.
 75. Yang M, Jang JY, Kim S, et al. Genetic effects on urinary 1-hydroxypyrene levels in a Korean population. *Carcinogenesis* 2003;24:1085–9.
 76. Aktas D, Guney I, Alikasifoglu M, et al. CYP1A1 gene polymorphism and risk of epithelial ovarian neoplasm. *Gynecol Oncol* 2002;86:124–8.
 77. Ratnasinghe D, Tangrea JA, Stewart C, et al. Influence of antioxidants and the CYP1A1 isoleucine to valine polymorphism on the smoking-lung cancer association. *Anticancer Res* 2001;21:1295–300.
 78. Roddam PL, Rollinson S, Kane E, et al. Poor metabolizers at the cytochrome P450 2D6 and 2C19 loci are at increased risk of developing adult acute leukaemia. *Pharmacogenetics* 2000;10:605–15.
 79. Basham VM, Pharoah PD, Healey CS, et al. Polymorphisms in CYP1A1 and smoking: no association with breast cancer risk. *Carcinogenesis* 2001;22:1797–800.
 80. Cantlay AM, Lamb D, Gillooly M, et al. Association between the CYP1A1 gene polymorphism and susceptibility to emphysema and lung cancer. *J Clin Pathol Mol Pathol* 1995;48:M210–14.
 81. Ambrosone CB, Freudenheim JL, Graham S, et al. Cytochrome P4501A1 and glutathione S-transferase (M1) genetic polymorphisms and postmenopausal breast cancer risk. *Cancer Res* 1995;55:3483–5.
 82. Evans AJ, Henner WD, Eilers KM, et al. Polymorphisms of GSTT1 and related genes in head and neck cancer risk. *Head Neck* 2004;26:63–70.
 83. Rebbeck TR, Rosvold EA, Duggan DJ, et al. Genetics of CYP1A1: coamplification of specific alleles by polymerase chain reaction and association with breast cancer. *Cancer Epidemiol Biomarkers Prev* 1994;3:511–14.
 84. Longuemaux S, Delomenie C, Gallou C, et al. Candidate genetic modifiers of individual susceptibility to renal cell carcinoma: a study of polymorphic human xenobiotic-metabolizing enzymes. *Cancer Res* 1999;59:2903–8.
 85. London SJ, Daly AK, Fairbrother KS, et al. Lung cancer risk in African-Americans in relation to a race-specific CYP1A1 polymorphism. *Cancer Res* 1995;55:6035–7.
 86. Tabor HK, Risch NJ, Myers RM. Opinion: candidate-gene approaches for studying complex genetic traits: practical considerations. *Nat Rev Genet* 2002;3:391–7.
 87. Garte S, Ganguly S, Taioli E. Effect of genotype on steady-state CYP1A1 gene expression in human peripheral lymphocytes. *Biochem Pharmacol* 2003;65:441–5.
 88. Cosma G, Crofts F, Currie D, et al. Racial differences in restriction fragment length polymorphisms and messenger RNA inducibility of the human CYP1A1 gene. *Cancer Epidemiol Biomarkers Prev* 1993;2:53–7.
 89. Landi MT, Bertazzi PA, Shields PG, et al. Association between CYP1A1 genotype, mRNA expression and enzymatic activity in humans. *Pharmacogenetics* 1994;4:242–6.
 90. Rumsby PC, Yardley-Jones A, Anderson D, et al. Detection of CYP1A1 mRNA levels and CYP1A1 MspI polymorphisms as possible biomarkers of exposure and susceptibility in smokers and non-smokers. *Teratog Carcinog Mutagen* 1996;16:65–74.
 91. Whyatt RM, Bell DA, Jedrychowski W, et al. Polycyclic aromatic hydrocarbon-DNA adducts in human placenta and modulation by CYP1A1 induction and genotype. *Carcinogenesis* 1998;19:1389–92.
 92. Crofts F, Taioli E, Trachman J, et al. Functional significance of different human CYP1A1 genotypes. *Carcinogenesis* 1994;15:2961–3.
 93. Kawajiri K, Nakachi K, Imai K, et al. The CYP1A1 gene and cancer susceptibility. *Crit Rev Oncol Hematol* 1993;14:77–87.
 94. Persson I, Johansson I, Ingelman-Sundberg M. In vitro kinetics of two human CYP1A1 variant enzymes suggested to be associated with interindividual differences in cancer susceptibility. *Biochem Biophys Res Commun* 1997;231:227–30.
 95. Zhang ZY, Fasco MJ, Huang L, et al. Characterization of purified human recombinant cytochrome P4501A1-Ile462 and -Val462: assessment of a role for the rare allele in carcinogenesis. *Cancer Res* 1996;56:3926–33.
 96. Cosma G, Crofts F, Taioli E, et al. Relationship between genotype and function of the human CYP1A1 gene. *J Toxicol Environ Health* 1993;40:309–16.
 97. Kiyohara C, Hirohata T, Inutsuka S. The relationship between aryl hydrocarbon hydroxylase and polymorphisms of the CYP1A1 gene. *Jpn J Cancer Res* 1996;87:18–24.
 98. Kiyohara C, Nakanishi Y, Inutsuka S, et al. The relationship between CYP1A1 aryl hydrocarbon hydroxylase activity and lung cancer in a Japanese population. *Pharmacogenetics* 1998;8:315–23.
 99. Jacquet M, Lambert V, Baudoux E, et al. Correlation between P450 CYP1A1 inducibility, MspI genotype and lung cancer incidence. *Eur J Cancer* 1996;32A:1701–6.
 100. Wedlund PJ, Kimura S, Gonzalez FJ, et al. I462V mutation in the human CYP1A1 gene: lack of correlation with either the Msp I 1.9 kb (M2) allele or CYP1A1 inducibility in a three-generation family of East Mediterranean descent. *Pharmacogenetics* 1994;4:21–6.
 101. Daly AK, Fairbrother KS, Smart J. Recent advances in understanding the molecular basis of polymorphisms in genes encoding cytochrome P450 enzymes. *Toxicol Lett* 1998;102–103:143–7.

102. Smart J, Daly AK. Variation in induced CYP1A1 levels: relationship to CYP1A1, Ah receptor and GSTM1 polymorphisms. *Pharmacogenetics* 2000;10:11–24.
103. Firozi PF, Bondy ML, Sahin AA, et al. Aromatic DNA adducts and polymorphisms of CYP1A1, NAT2 and GSTM1 in breast cancer. *Carcinogenesis* 2002;23:301–6.
104. Li D, Wang M, Firozi PF, et al. Characterization of a major aromatic DNA adduct detected in human breast tissues. *Environ Mol Mutagen* 2002;39:193–200.
105. Li D, Walcott FL, Chang P, et al. Genetic and environmental determinants on tissue response to in vitro carcinogen exposure and risk of breast cancer. *Cancer Res* 2002;62:4566–70.
106. Brockstedt U, Krajcinovic M, Richer C, et al. Analyses of bulky DNA adduct levels in human breast tissue and genetic polymorphisms of cytochromes P450 (CYPs), myeloperoxidase (MPO), quinone oxidoreductase (NQO1), and glutathione S-transferases (GSTs). *Mutat Res* 2002;516:41–7.
107. Zhu J, Chang P, Bondy ML, et al. Detection of 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine-DNA adducts in normal breast tissues and risk of breast cancer. *Cancer Epidemiol Biomarkers Prev* 2003;12:830–7.
108. Li D, Firozi PF, Zhang W, et al. DNA adducts, genetic polymorphisms, and K-ras mutation in human pancreatic cancer. *Mutat Res* 2002;513:37–48.
109. Cheng YW, Chen CY, Lin P, et al. DNA adduct level in lung tissue may act as a risk biomarker of lung cancer. *Eur J Cancer* 2000;36:1381–8.
110. Cheng YW, Hsieh LL, Lin PP, et al. Gender difference in DNA adduct levels among nonsmoking lung cancer patients. *Environ Mol Mutagen* 2001;37:304–10.
111. Schoket B, Papp G, Levay K, et al. Impact of metabolic genotypes on levels of biomarkers of genotoxic exposure. *Mutat Res* 2001;482:57–69.
112. Rojas M, Alexandrov K, Cascorbi I, et al. High benzo(a)pyrene diol-epoxide DNA adduct levels in lung and blood cells from subjects with combined CYP1A1 MspI/MspI-GSTM1*0/*0 genotypes. *Pharmacogenetics* 1998;8:109–18.
113. Ichiba M, Hagmar L, Rannug A, et al. Aromatic DNA adducts, micronuclei and genetic polymorphism for CYP1A1 and GST1 in chimney sweeps. *Carcinogenesis* 1994;15:1347–52.
114. Zhang J, Ichiba M, Feng Y, et al. Aromatic DNA adducts in coke-oven workers, in relation to exposure, lifestyle and genetic polymorphism of metabolic enzymes. *Int Arch Occup Environ Health* 2000;73:127–35.
115. Hemminki K, Dickey C, Karlsson S, et al. Aromatic DNA adducts in foundry workers in relation to exposure, life style and CYP1A1 and glutathione transferase M1 genotype. *Carcinogenesis* 1997;18:345–50.
116. Kuljukka-Rabb T, Nylund L, Vaaranrinta R, et al. The effect of relevant genotypes on PAH exposure-related biomarkers. *J Expo Anal Environ Epidemiol* 2002;12:81–91.
117. Grzybowska E, Butkiewicz D, Motykiewicz G, et al. The effect of the genetic polymorphisms of CYP1A1, CYP2D6, GSTM1 and GSTP1 on aromatic DNA adduct levels in the population of healthy women. *Mutat Res* 2000;469:271–7.
118. Palli D, Vineis P, Russo A, et al. Diet, metabolic polymorphisms and DNA adducts: the EPIC-Italy cross-sectional study. *Int J Cancer* 2000;87:444–51.
119. Whyatt RM, Perera FP, Jedrychowski W, et al. Association between polycyclic aromatic hydrocarbon-DNA adduct levels in maternal and newborn white blood cells and glutathione S-transferase P1 and CYP1A1 polymorphisms. *Cancer Epidemiol Biomarkers Prev* 2000;9:207–12.
120. Teixeira JP, Gaspar J, Martinho G, et al. Aromatic DNA adduct levels in coke oven workers: correlation with polymorphisms in genes GSTP1, GSTM1, GSTT1 and CYP1A1. *Mutat Res* 2002;517:147–55.
121. Rojas M, Cascorbi I, Alexandrov K, et al. Modulation of benzo[a]pyrene diol-epoxide-DNA adduct levels in human white blood cells by CYP1A1, GSTM1 and GSTT1 polymorphism. *Carcinogenesis* 2000;21:35–41.
122. Mooney LA, Bell DA, Santella RM, et al. Contribution of genetic and nutritional factors to DNA damage in heavy smokers. *Carcinogenesis* 1997;18:503–9.
123. Matsui A, Ikeda T, Enomoto K, et al. Increased formation of oxidative DNA damage, 8-hydroxy-2'-deoxyguanosine, in human breast cancer tissue and its relationship to GSTP1 and COMT genotypes. *Cancer Lett* 2000;151:87–95.
124. Hong YC, Lee KH, Yi CH, et al. Genetic susceptibility of term women to oxidative damage. *Toxicol Lett* 2002;129:255–62.
125. Hong YC, Park HS, Ha EH. Influence of genetic susceptibility on the urinary excretion of 8-hydroxydeoxyguanosine of firefighters. *Occup Environ Med* 2000;57:370–5.
126. Kim YD, Lee CH, Nan HM, et al. Effects of genetic polymorphisms in metabolic enzymes on the relationships between 8-hydroxydeoxyguanosine levels in human leukocytes and urinary 1-hydroxypyrene and 2-naphthol concentrations. *J Occup Health* 2003;45:160–7.
127. Miyoshi Y, Ando A, Hasegawa S, et al. Association of genetic polymorphisms in CYP19 and CYP1A1 with the oestrogen receptor-positive breast cancer risk. *Eur J Cancer* 2003;39:2531–7.
128. Adonis M, Martinez V, Riquelme R, et al. Susceptibility and exposure biomarkers in people exposed to PAHs from diesel exhaust. *Toxicol Lett* 2003;144:3–15.
129. Alexandrie AK, Warholm M, Carstensen U, et al. CYP1A1 and GSTM1 polymorphisms affect urinary 1-hydroxypyrene levels after PAH exposure. *Carcinogenesis* 2000;21:669–76.
130. Wu MT, Huang SL, Ho CK, et al. Cytochrome P450 1A1 MspI polymorphism and urinary 1-hydroxypyrene concentrations in coke-oven workers. *Cancer Epidemiol Biomarkers Prev* 1998;7:823–9.
131. Gil L, Martinez V, Riquelme R, et al. Occupational and environmental levels of mutagenic PAHs and respirable particulate matter associated with diesel exhaust in Santiago, Chile. *J Occup Environ Med* 2003;45:984–92.
132. Nerurkar PV, Okinaka L, Aoki C, et al. CYP1A1, GSTM1, and GSTP1 genetic polymorphisms and urinary 1-hydroxypyrene excretion in non-occupationally exposed individuals. *Cancer Epidemiol Biomarkers Prev* 2000;9:1119–22.
133. Apostoli P, Neri G, Lucas D, et al. Influence of genetic polymorphisms of CYP1A1 and GSTM1 on the urinary levels of 1-hydroxypyrene. *Toxicol Lett* 2003;144:27–34.
134. Pan G, Hanaoka T, Yamano Y, et al. A study of multiple biomarkers in coke oven workers—a cross-sectional study in China. *Carcinogenesis* 1998;19:1963–8.
135. Nan HM, Kim H, Lim HS, et al. Effects of occupation, lifestyle and genetic polymorphisms of CYP1A1, CYP2E1, GSTM1 and GSTT1 on urinary 1-hydroxypyrene and 2-naphthol concentrations. *Carcinogenesis* 2001;22:787–93.
136. Yang M, Kunugita N, Kitagawa K, et al. Individual differences in urinary cotinine levels in Japanese smokers: relation to genetic polymorphisms of drug-metabolizing enzymes. *Cancer Epidemiol Biomarkers Prev* 2001;10:589–93.
137. Kawamoto T, Koga M, Oyama T, et al. Habitual and genetic factors that affect urinary background levels of biomarkers for organic solvent exposure. *Arch Environ Contam Toxicol* 1996;30:114–20.

138. Ursin G, Ma H, Wu AH, et al. Mammographic density and breast cancer in three ethnic groups. *Cancer Epidemiol Biomarkers Prev* 2003;12:332–8.
139. Boyd NF, Lockwood GA, Byng JW, et al. Mammographic densities and breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 1998;7:1133–44.
140. Haiman CA, Hankinson SE, De Vivo I, et al. Polymorphisms in steroid hormone pathway genes and mammographic density. *Breast Cancer Res Treat* 2003;77:27–36.
141. GLOBOCAN. 2002—Cancer incidence, mortality and prevalence worldwide, 2005. (<http://www-depdb.iarc.fr/globocan/GLOBOframe.htm>).
142. Parkin DM, Pisani P, Ferlay J. Estimates of the worldwide incidence of 25 major cancers in 1990. *Int J Cancer* 1999;80:827–41.
143. Pike MC, Spicer DV, Dahmouch L, et al. Estrogens, progestogens, normal breast cell proliferation, and breast cancer risk. *Epidemiol Rev* 1993;15:17–35.
144. Parkin DM, Whelan SL, Ferlay J, et al, eds. *Cancer incidence in five continents*. Vol VIII. Lyon, France: International Agency for Research on Cancer, 2002. (IARC scientific publication no. 155).
145. Coleman MP, Esteve J, Damiecki P, et al. Trends in cancer incidence and mortality. *IARC Sci Publ* 1993;121:1–806.
146. Easton DF. The inherited component of cancer. *Br Med Bull* 1994;50:527–35.
147. Miki Y, Swensen J, Shattuck-Eidens D, et al. A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1. *Science* 1994;266:66–71.
148. Bishop DT. Family history of breast cancer: how important is it? *J Med Genet* 1992;29:152–3.
149. Colditz GA, Willett WC, Hunter DJ, et al. Family history, age and risk of breast cancer. *JAMA* 1993;270:338–43.
150. Weber BL, Garber JE. Family history and breast cancer: probabilities and possibilities. *JAMA* 1993;270:1602–3.
151. Henderson BE, Pike MC, Bernstein L, et al. Breast cancer. In: Schottenfeld D, Fraumeni JF Jr, eds. *Cancer epidemiology and prevention*. 2nd ed. New York, NY: Oxford University Press, 1996:1022–39.
152. Beral V: Million Women Study Collaborators. Breast cancer and hormone-replacement therapy in the Million Women Study. *Lancet* 2003;362:419–27.
153. Rossouw JE, Anderson GL, Prentice RL, et al. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results from the Women's Health Initiative randomized controlled trial. *JAMA* 2002;288:321–33.
154. Breast cancer and hormonal contraceptives: collaborative reanalysis of individual data on 53 297 women with breast cancer and 100 239 women without breast cancer from 54 epidemiological studies. Collaborative Group on Hormonal Factors in Breast Cancer. *Lancet* 1996;347:1713–27.
155. Newcomer LM, Newcomb PA, Trentham-Dietz A, et al. Oral contraceptive use and risk of breast cancer by histologic type. *Int J Cancer* 2003;106:961–4.
156. Weight control and physical activity. IARC handbooks of cancer prevention. Vol. 6. Lyon, France: International Agency for Research on Cancer, 2002.
157. Smith-Warner SA, Spiegelman D, Yaun SS, et al. Alcohol and breast cancer in women. A pooled analysis of cohort studies. *JAMA* 1998;279:535–40.
158. Morris JJ, Seifter E. The role of aromatic hydrocarbons in the genesis of breast cancer. *Med Hypotheses* 1992;38:177–84.
159. MacNicol AD, Easty GC, Neville AM, et al. Metabolism and activation of carcinogenic polycyclic hydrocarbons by human mammary cells. *Biochem Biophys Res Commun* 1980;95:1599–606.
160. Li DH, Wang MY, Dhingra K, et al. Aromatic DNA adducts in adjacent tissues of breast cancer patients—clues to breast cancer etiology. *Cancer Res* 1996;56:287–93.
161. Davis DL, Bradlow HL, Wolff M, et al. Medical hypothesis: xenoestrogens as preventable causes of breast cancer. *Environ Health Perspect* 1993;101:372–7.
162. Yuspa SH, Poirier MC. Chemical carcinogenesis: from animal models to molecular models in one decade. *Adv Cancer Res* 1988;50:25–70.
163. Hemminki K, Pershagen G. Cancer risk of air pollution: epidemiological evidence. *Environ Health Perspect* 1994;102:187–92.
164. Tobacco smoking. IARC monographs on the evaluation of carcinogenic risks to humans. Vol 38. Lyon, France: International Agency for Research on Cancer, 1986.
165. Bannicke K, Conrad C, Sabroe S, et al. Cigarette smoking and breast cancer. *BMJ* 1995;310:1431–3.
166. Hirose K, Tajima K, Hamajima N, et al. A large scale, hospital based case-control study of risk factors of breast cancer according to menopausal status. *Jpn J Cancer Res* 1995;86:146–54.
167. Ambrosone CB, Freudenheim JL, Graham S, et al. Cigarette smoking, N-acetyltransferase 2 genetic polymorphisms, and breast cancer risk. *JAMA* 1996;276:1494–501.
168. Morabia A, Bernstein M, Héritier S, et al. Relation of breast cancer with passive and active exposure to tobacco smoke. *Am J Epidemiol* 1996;143:918–28.
169. Baron JA, Newcomb PA, Longnecker MP, et al. Cigarette smoking and breast cancer. *Cancer Epidemiol Biomarkers Prev* 1996;5:399–403.
170. Hunter DJ, Hankinson SE, Hough H, et al. A prospective study of NAT2 acetylation genotype, cigarette smoking, and risk of breast cancer. *Carcinogenesis* 1997;18:2127–32.
171. Khuder SA, Mutgi AB, Nugent S. Smoking and breast cancer: a meta-analysis. *Rev Environ Health* 2001;16:253–61.
172. Palmer JR, Rosenberg L, Clarke EA, et al. Breast cancer and cigarette smoking: a hypothesis. *Am J Epidemiol* 1991;134:1–13.
173. Terry PD, Rohan TE. Cigarette smoking and the risk of breast cancer in women: a review of the literature. *Cancer Epidemiol Biomarkers Prev* 2002;11:953–71.
174. Tredaniel J, Boffetta P, Saracci R, et al. Environmental tobacco smoke and the risk of cancer in adults. *Eur J Cancer* 1993;29A:2058–68.
175. Khuder SA, Simon VJ Jr. Is there an association between passive smoking and breast cancer? *Eur J Epidemiol* 2000;16:1117–21.
176. Tobacco smoke and involuntary smoking. IARC monographs on the evaluation of carcinogenic risks to humans. Vol 83. Lyon, France: International Agency for Research on Cancer, 2004.
177. Vineis P, McMichael A. Interplay between heterocyclic amines in cooked meat and metabolic phenotype in the etiology of colon cancer. *Cancer Causes Control* 1996;7:479–86.
178. World Cancer Research Fund/American Institute for Cancer Research. *Food, nutrition and the prevention of cancer: a global perspective*. Menasha, WI: Banta Book Group, 1997.
179. Boyd NF, Stone J, Vogt KN, et al. Dietary fat and breast cancer risk revisited: a meta-analysis of the published literature. *Br J Cancer* 2003;89:1672–85.
180. De Stefani E, Ronco A, Mendilaharsu M, et al. Meat intake, heterocyclic amines, and risk of breast cancer: a case-control

- study in Uruguay. *Cancer Epidemiol Biomarkers Prev* 1997;6:573–81.
181. Jarvinen R, Knekt P, Seppanen R, et al. Diet and breast cancer risk in a cohort of Finnish women. *Cancer Lett* 1997;114:251–3.
 182. Zheng W, Gustafson DR, Sinha R, et al. Well-done meat intake and the risk of breast cancer. *J Natl Cancer Inst* 1998;90:1724–9.
 183. Dai Q, Shu XO, Jin F, et al. Consumption of animal foods, cooking methods, and risk of breast cancer. *Cancer Epidemiol Biomarkers Prev* 2002;11:801–8.
 184. Nakachi K, Imai K, Hayashi S, et al. Polymorphisms of the CYP1A1 and glutathione S-transferase genes associated with susceptibility to lung cancer in relation to cigarette dose in a Japanese population. *Cancer Res* 1993;53:2994–9.
 185. Ishibe N, Hankinson SE, Colditz GA, et al. Cigarette smoking, cytochrome P450 1A1 polymorphisms, and breast cancer risk in the Nurses' Health Study. *Cancer Res* 1998;58:667–71.
 186. Huang CS, Chern HD, Chang KJ, et al. Breast cancer risk associated with genotype polymorphism of the estrogen-metabolizing genes CYP17, CYP1A1, and COMT: a multi-genic study on cancer susceptibility. *Cancer Res* 1999;59:4870–5.
 187. Taioli E, Trachman J, Chen X, et al. A CYP1A1 restriction fragment length polymorphism is associated with breast cancer in African-American women. *Cancer Res* 1995;55:3757–8.
 188. Ritchie MD, Hahn LW, Roodi N, et al. Multifactor-dimensionality reduction reveals high-order interactions among estrogen-metabolism genes in sporadic breast cancer. *Am J Hum Genet* 2001;69:138–47.
 189. StataCorp. Stata statistical software, release 7.0. College Station, TX: Stata Corporation, 2001.
 190. Higgins JP, Thompson SG, Deeks JJ, et al. Measuring inconsistency in meta-analyses. *BMJ* 2003;327:557–60.
 191. Goode EL, Dunning AM, Kuschel B, et al. Effect of germline genetic variation on breast cancer survival in a population-based study. *Cancer Res* 2002;62:3052–7.
 192. Vineis P, Veglia F, Benhamou S, et al. CYP1A1 T3801C polymorphism and lung cancer: a pooled analysis of 2451 cases and 3358 controls. *Int J Cancer* 2003;104:650–7.
 193. Le Marchand L, Guo C, Benhamou S, et al. Pooled analysis of the CYP1A1 exon 7 polymorphism and lung cancer (United States). *Cancer Causes Control* 2003;14:339–46.
 194. Taioli E, Gaspari L, Benhamou S, et al. Polymorphisms in CYP1A1, GSTM1, GSTT1 and lung cancer below the age of 45 years. *Int J Epidemiol* 2003;32:60–3.
 195. Hung RJ, Boffetta P, Brockmoller J, et al. CYP1A1 and GSTM1 genetic polymorphisms and lung cancer risk in Caucasian non-smokers: a pooled analysis. *Carcinogenesis* 2003;24:875–82.
 196. Kelsey KT, Wiencke JK, Spitz MR. A race-specific genetic polymorphism in the CYP1A1 gene is not associated with lung cancer in African Americans. *Carcinogenesis* 1994;15:1121–4.
 197. Gsur A, Haidinger G, Hollaus P, et al. Genetic polymorphisms of CYP1A1 and GSTM1 and lung cancer risk. *Anticancer Res* 2001;21:2237–42.
 198. Sato M, Sato T, Izumo T, et al. Genetic polymorphism of drug-metabolizing enzymes and susceptibility to oral cancer. *Carcinogenesis* 1999;20:1927–31.
 199. Sato M, Sato T, Izumo T, et al. Genetically high susceptibility to oral squamous cell carcinoma in terms of combined genotyping of CYP1A1 and GSTM1 genes. *Oral Oncol* 2000;36:267–71.
 200. Kao SY, Wu CH, Lin SC, et al. Genetic polymorphism of cytochrome P4501A1 and susceptibility to oral squamous cell carcinoma and oral precancer lesions associated with smoking/betel use. *J Oral Pathol Med* 2002;31:505–11.
 201. Tanimoto K, Hayashi S, Yoshiga K, et al. Polymorphisms of the CYP1A1 and GSTM1 gene involved in oral squamous cell carcinoma in association with a cigarette dose. *Oral Oncol* 1999;35:191–6.
 202. Sreelekha TT, Ramadas K, Pandey M, et al. Genetic polymorphism of CYP1A1, GSTM1 and GSTT1 genes in Indian oral cancer. *Oral Oncol* 2001;37:593–8.
 203. Park JY, Muscat JE, Ren Q, et al. CYP1A1 and GSTM1 polymorphisms and oral cancer risk. *Cancer Epidemiol Biomarkers Prev* 1997;6:791–7.
 204. Morita S, Yano M, Tsujinaka T, et al. Genetic polymorphisms of drug-metabolizing enzymes and susceptibility to head-and-neck squamous-cell carcinoma. *Int J Cancer* 1999;80:685–8.
 205. Katoh T, Kaneko S, Kohshi K, et al. Genetic polymorphisms of tobacco- and alcohol-related metabolizing enzymes and oral cavity cancer. *Int J Cancer* 1999;83:606–9.
 206. Hahn M, Hagedorn G, Kuhlisch E, et al. Genetic polymorphisms of drug-metabolizing enzymes and susceptibility to oral cavity cancer. *Oral Oncol* 2002;38:486–90.
 207. Worrall SF, Corrigan M, High A, et al. Susceptibility and outcome in oral cancer: preliminary data showing an association with polymorphism in cytochrome P450 CYP2D6. *Pharmacogenetics* 1998;8:433–9.
 208. Hori H, Kawano T, Endo M, et al. Genetic polymorphisms of tobacco- and alcohol-related metabolizing enzymes and human esophageal squamous cell carcinoma susceptibility. *J Clin Gastroenterol* 1997;25:568–75.
 209. Jahnke V, Matthias C, Fryer A, et al. Glutathione S-transferase and cytochrome P450 polymorphism as risk factors for squamous cell carcinoma of the larynx. *Am J Surg* 1996;172:671–3.
 210. Oude Ophuis MB, van Lieshout EMM, Roelofs HMJ, et al. Glutathione S-transferase M1 and T1 and cytochrome P4501A1 polymorphisms in relation to the risk for benign and malignant head and neck lesions. *Cancer* 1998;82:936–43.
 211. Gronau S, Koenig-Greger D, Jerg M, et al. Gene polymorphisms in detoxification enzymes as susceptibility factor for head and neck cancer? *Otolaryngol Head Neck Surg* 2003;128:674–80.
 212. McWilliams JE, Evans AJ, Beer TM, et al. Genetic polymorphisms in head and neck cancer risk. *Head Neck* 2000;22:609–17.
 213. Olshan AF, Weissler MC, Watson MA, et al. GSTM1, GSTT1, GSTP1, CYP1A1, and NAT1 polymorphisms, tobacco use, and the risk of head and neck cancer. *Cancer Epidemiol Biomarkers Prev* 2000;9:185–91.
 214. Roth MJ, Dawsey SM, Wang GQ, et al. Association between GSTM1*0 and squamous dysplasia of the esophagus in the high risk region of Linxian, China. *Cancer Lett* 2000;156:73–81.
 215. Nimura Y, Yokoyama S, Fujimori M, et al. Genotyping of the CYP1A1 and GSTM1 genes in esophageal carcinoma patients with special reference to smoking. *Cancer* 1997;80:852–7.
 216. Sivaraman L, Leatham MP, Yee J, et al. CYP1A1 genetic polymorphisms and in situ colorectal cancer. *Cancer Res* 1994;54:3692–5.
 217. Fritsch E, Bruning T, Jonkmanns C, et al. Detection of cytochrome P450 1B1 Bfr I polymorphism: genotype

- distribution in healthy German individuals and in patients with colorectal carcinoma. *Pharmacogenetics* 1999;9:405–8.
218. Butler WJ, Ryan P, Roberts-Thomson IC. Metabolic genotypes and risk for colorectal cancer. *J Gastroenterol Hepatol* 2001;16:631–5.
 219. Dolzan V, Ravnik-Glavac M, Breskvar K. Genetic polymorphisms of xenobiotic metabolizing enzymes in human colorectal cancer. *Radiol Oncol* 1998;32:35–9.
 220. Kiss I, Sandor J, Pajkos G, et al. Colorectal cancer risk in relation to genetic polymorphism of cytochrome P450 1A1, 2E1, and glutathione-S-transferase M1 enzymes. *Anticancer Res* 2000;20:519–22.
 221. Ye Z, Parry JM. Genetic polymorphisms in the cytochrome P450 1A1, glutathione S-transferase M1 and T1, and susceptibility to colon cancer. *Teratog Carcinog Mutagen* 2002;22:385–92.
 222. Suzuki K, Matsui H, Nakazato H, et al. Association of the genetic polymorphism in cytochrome P450 (CYP) 1A1 with risk of familial prostate cancer in a Japanese population: a case-control study. *Cancer Lett* 2003;195:177–83.
 223. Acevedo C, Opazo JL, Huidobro C, et al. Positive correlation between single or combined genotypes of CYP1A1 and GSTM1 in relation to prostate cancer in Chilean people. *Prostate* 2003;57:111–17.
 224. Beer TM, Evans AJ, Hough KM, et al. Polymorphisms of GSTP1 and related genes and prostate cancer risk. *Prostate Cancer Prostatic Dis* 2002;5:22–7.
 225. Kinoshita M, Seno T, Shin S, et al. Polymorphic genotypes of enzymes involved in carcinogen metabolism as risk factors for the development of human cervical cancer. *Jpn J Clin Chem* 1994;23:195–202.
 226. Kim JW, Lee CG, Park YG, et al. Combined analysis of germline polymorphisms of p53, GSTM1, GSTT1, CYP1A1, and CYP2E1: relation to the incidence rate of cervical carcinoma. *Cancer* 2000;88:2082–91.
 227. Esteller M, Garcia A, Martinez-Palones JM, et al. Susceptibility to endometrial cancer: influence of allelism at p53, glutathione S-transferase (GSTM1 and GSTT1) and cytochrome P-450 (CYP1A1) loci. *Br J Cancer* 1997;75:1385–8.
 228. Esteller M, Garcia A, Martinez-Palones JM, et al. Germ line polymorphisms in cytochrome-P450 1A1 (C4887 CYP1A1) and methylenetetrahydrofolate reductase (MTHFR) genes and endometrial cancer susceptibility. *Carcinogenesis* 1997;18:2307–11.
 229. Esteller M, Garcia A, Martinez-Palones JM, et al. Clinicopathologic features and genetic alterations in endometrioid carcinoma of the uterus with villoglandular differentiation. *Am J Clin Pathol* 1999;111:336–42.
 230. Goodman MT, McDuffie K, Hernandez B, et al. CYP1A1, GSTM1, and GSTT1 polymorphisms and the risk of cervical squamous intraepithelial lesions in a multiethnic population. *Gynecol Oncol* 2001;81:263–9.
 231. Yengi L, Inskip A, Gilford J, et al. Polymorphism at the glutathione S-transferase locus GSTM3: interactions with cytochrome P450 and glutathione S-transferase genotypes as risk factors for multiple cutaneous basal cell carcinoma. *Cancer Res* 1996;56:1974–7.
 232. Clairmont A, Sies H, Ramachandran S, et al. Association of NAD(P)H:quinone oxidoreductase (NQO1) null with numbers of basal cell carcinomas: use of a multivariate model to rank the relative importance of this polymorphism and those at other relevant loci. *Carcinogenesis* 1999;20:1235–40.
 233. Krajcinovic M, Labuda D, Mathonnet G, et al. Polymorphisms in genes encoding drugs and xenobiotic metabolizing enzymes, DNA repair enzymes, and response to treatment of childhood acute lymphoblastic leukemia. *Clin Cancer Res* 2002;8:802–10.
 234. Balta G, Yuksek N, Ozyurek E, et al. Characterization of MTHFR, GSTM1, GSTT1, GSTP1, and CYP1A1 genotypes in childhood acute leukemia. *Am J Hematol* 2003;73:154–60.
 235. Garte S, Taioli E, Crost F, et al. Deletion of parental GST genes as a possible susceptibility factor in the etiology of infant leukemia. *Leuk Res* 2000;24:971–4.
 236. Tsukasaki K, Miller CW, Kubota T, et al. Tumor necrosis factor alpha polymorphism associated with increased susceptibility to development of adult T-cell leukemia/lymphoma in human T-lymphotropic virus type 1 carriers. *Cancer Res* 2001;61:3770–4.
 237. Kerridge I, Lincz L, Scorgie F, et al. Association between xenobiotic gene polymorphisms and non-Hodgkin's lymphoma risk. *Br J Haematol* 2002;118:477–81.
 238. Soucek P, Sarmanova J, Kristensen VN, et al. Genetic polymorphisms of biotransformation enzymes in patients with Hodgkin's and non-Hodgkin's lymphomas. *Int Arch Occup Environ Health* 2002;75:S86–92.
 239. Trizna Z, de Andrade M, Kyritsis AP, et al. Genetic polymorphisms in glutathione S-transferase mu and theta, N-acetyltransferase, and CYP1A1 and risk of gliomas. *Cancer Epidemiol Biomarkers Prev* 1998;7:553–5.
 240. Lee HC, Yoon YB, Kim CY. Association between genetic polymorphisms of the cytochromes P-450 (1A1, 2D6, and 2E1) and the susceptibility to pancreatic cancer. *Korean J Intern Med* 1997;12:128–36.
 241. Okada T, Kawashima K, Fukushi S, et al. Association between a cytochrome P450 CYP1A1 genotype and incidence of lung cancer. *Pharmacogenetics* 1994;4:333–40.
 242. Liu G, Ghadirian P, Vesprini D, et al. Polymorphisms in GSTM1, GSTT1 and CYP1A1 and risk of pancreatic adenocarcinoma. *Br J Cancer* 2000;82:1646–9.
 243. Fritsche E, Schuppe HC, Dohr O, et al. Increased frequencies of cytochrome P4501A1 polymorphisms in infertile men. *Andrologia* 1998;30:125–8.
 244. von Schmiedeberg S, Fritsche E, Ronnau AC, et al. Polymorphisms of the xenobiotic-metabolizing enzymes CYP1A1 and NAT-2 in systemic sclerosis and lupus erythematosus. *Adv Exp Med Biol* 1999;455:147–52.
 245. Gardlo K, Selimovic D, Bolsen K, et al. Cytochrome P4501A1 polymorphisms in a Caucasian population with porphyria cutanea tarda. *Exp Dermatol* 2003;12:843–8.
 246. Yen J, Tsai W, Chen C, et al. Cytochrome P450 1A1 and manganese superoxide dismutase genes polymorphisms in ankylosing spondylitis. *Immunol Lett* 2003;88:113–16.
 247. Yen JH, Chen CJ, Tsai WC, et al. Manganese superoxide dismutase and cytochrome P450 1A1 genes polymorphisms in rheumatoid arthritis in Taiwan. *Hum Immunol* 2003;64:366–73.
 248. Hadfield RM, Manek S, Weeks DE, et al. Linkage and association studies of the relationship between endometriosis and genes encoding the detoxification enzymes GSTM1, GSTT1 and CYP1A1. *Mol Hum Reprod* 2001;7:1073–8.
 249. Iizuka S, Kosugi Y, Isaka K, et al. Could polymorphisms of N-acetyltransferase 2 (NAT2), glutathione S-transferase M1 (GSTM1), and cytochrome P450 (CYP1A1) be responsible for genetic predisposition to endometriosis among Japanese? *Zasshi/Tokyo Ika Daigaku* 2003;61:59–66.
 250. Takakubo F, Yamamoto M, Ogawa N, et al. Genetic association between cytochrome P450IA1 gene and susceptibility to Parkinson's disease. *J Neural Transm* 1996;103:843–9.
 251. Kurth MC, Kurth JH. Variant cytochrome P450 CYP2D6 allelic frequencies in Parkinson's disease. *Am J Med Genet* 1993;48:166–8.

252. Ivaschenko TE, Sideleva OG, Baranov VS. Glutathione-S-transferase mu and theta gene polymorphisms as new risk factors of atopic bronchial asthma. *J Mol Med* 2002;80:39–43.
253. Binkova B, Smerhovsky Z, Strejc P, et al. DNA-adducts and atherosclerosis: a study of accidental and sudden death males in the Czech Republic. *Mutat Res* 2002;501:115–28.
254. de Jong DJ, van der Logt EM, van Schaik A, et al. Genetic polymorphisms in biotransformation enzymes in Crohn's disease: association with microsomal epoxide hydrolase. *Gut* 2003;52:547–51.
255. Kimura K, Isashiki Y, Sonoda S, et al. Genetic association of manganese superoxide dismutase with exudative age-related macular degeneration. *Am J Ophthalmol* 2000;130:769–73.
256. Zusterzeel PLM, Nelen WLD, Roelofs HMJ, et al. Polymorphisms in biotransformation enzymes and the risk for recurrent early pregnancy loss. *Mol Hum Reprod* 2000;6:474–8.
257. van Rooij IALM, Wegerif MJM, Roelofs HMJ, et al. Smoking, genetic polymorphisms in biotransformation enzymes, and nonsyndromic oral clefting: a gene-environment interaction. *Epidemiology* 2001;12:502–7.
258. Smith PG, Day NE. The design of case-control studies: the influence of confounding and interaction effects. *Int J Epidemiol* 1984;13:356–65.
259. Khoury MJ, Beaty TH, Hwang SJ. Detection of genotype-environment interaction in case-control studies of birth defects: how big a sample size? *Teratology* 1995;51:336–43.
260. McKinney JD, Waller CL. Polychlorinated biphenyls as hormonally active structural analogues. *Environ Health Perspect* 1994;102:290–7.
261. Norback DH, Weltman RH. Polychlorinated biphenyl induction of hepatocellular carcinoma in the Sprague-Dawley rat. *Environ Health Perspect* 1985;60:97–105.
262. Laden F, Ishibe N, Hankinson SE, et al. Polychlorinated biphenyls, cytochrome P450 1A1, and breast cancer risk in the Nurses' Health Study. *Cancer Epidemiol Biomarkers Prev* 2002;11:1560–5.
263. Moysich KB, Shields PG, Freudenheim JL, et al. Polychlorinated biphenyls, cytochrome P4501A1 polymorphism, and postmenopausal breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 1999;8:41–4.
264. Vaury C, Lain R, Noguez P, et al. Human glutathione S-transferase M1 null genotype is associated with a high inducibility of cytochrome P450 1A1 gene transcription. *Cancer Res* 1995;55:5520–3.
265. Mitrunen K, Hirvonen A. Molecular epidemiology of sporadic breast cancer. The role of polymorphic genes involved in oestrogen biosynthesis and metabolism. *Mutat Res* 2003;544:9–41.
266. Sharp L, Cardy AH, Cotton SC, et al. CYP17 gene polymorphisms: prevalence and associations with hormone levels and related factors: a HuGE review. *Am J Epidemiol* 2004;160:729–40.
267. Little J, Bradley L, Bray MS, et al. Reporting, appraising, and integrating data on genotype prevalence and gene-disease associations. *Am J Epidemiol* 2002;156:300–10.
268. Ishibe N, Wiencke JK, Zuo Z, et al. Susceptibility to lung cancer in light smokers associated with CYP1A1 polymorphisms in Mexican- and African-Americans. *Cancer Epidemiol Biomarkers Prev* 1997;6:1075–80.
269. Garte S, Zocchetti C, Taioli E. Gene-environment interactions in the application of biomarkers of cancer susceptibility in epidemiology. In: Toniolo P, Boffetta P, Shuker DEG, et al, eds. *Application of biomarkers in cancer epidemiology*. Lyon, France: International Agency for Research on Cancer, 1997:251–64.
270. Botto LD, Khoury MJ. Commentary: facing the challenge of gene-environment interaction: the two-by-four table and beyond. *Am J Epidemiol* 2001;153:1016–20.

APPENDIX

Internet Sites

Data on cancer incidence, survival, and mortality

International Agency for Research on Cancer—Cancer Mondial: <http://www-dep.iarc.fr/>
 Surveillance, Epidemiology, and End Results Program: <http://www.seer.cancer.gov/publicdata/>
 National Programme of Cancer Registries (NPCR): <http://www.cdc.gov/cancer/npcr>

Information on cancer

Cancer Research UK: <http://www.cancerresearchuk.org>
 American Association of Cancer Research: <http://www.aacr.org/>
 National Cancer Institute: <http://cancer.gov/cancerinfo/>
 International Union against Cancer: <http://www.uicc.ch/>
 American Cancer Society: <http://www.cancer.org/docroot/home/index.asp>

Genetic information

Human Genome Epidemiology Network (HuGENet): <http://www.cdc.gov/genomics/hugenet/default.htm>
 Centers for Disease Control and Prevention Office of Genomics and Disease Prevention—medical literature search: <http://www.cdc.gov/genomics/info/medlit.htm>
 Public Health Genetics Unit: <http://www.phgu.org.uk/index.php>
 Human Gene Mutation Database: <http://archive.uwcm.ac.uk/uwcm/mg/hgmd0.html>
 OMIM (Online Mendelian Inheritance in Man): <http://www.ncbi.nlm.nih.gov/Omim/>
 GenAtlas: <http://www.dsi.univ-paris5.fr/genatlas/>
 GeneCards: <http://www.cgal.icnet.uk/genecards/>
 The National Center for Biotechnology Information: <http://www.ncbi.nlm.nih.gov/>
 Human cytochrome P-450 allele nomenclature: <http://www.imm.ki.se/CYPalleles/>
 MRC Rosalind Franklin Centre for Genomics Research (includes links to other sites via The Genome Web): <http://www.hgmp.mrc.ac.uk/>
 Links to other sites: http://cedar.genetics.soton.ac.uk/public_html/links.html