

Are oiled rock column (ORC) experiments confounded by particulate oil?

Carls et al. NOAA/NMFS Auke Bay Laboratories

Brannon et al. (2006) published a paper in Environmental Toxicology and Chemistry that repeated an embryo toxicity study (Heintz et al. 1999) where we concluded that dissolved polynuclear aromatic hydrocarbons (PAHs) from oil are toxic at concentrations between 1 and 18 $\mu\text{g/L}$ (parts per billion). In contrast, they argue that toxicity is an artifact caused by contact with PAH-laden oil microdroplets instead of dissolved PAHs and that toxic levels are many orders of magnitude higher. We conclude that flawed analysis logic and the lack of any direct evidence of oil droplets in water and eggs led Brannon et al. (2006) to erroneous conclusions (Heintz et al. 2008; Carls et al. 2008a). In their rebuttal, Brannon et al. (2008) reanalyzed a small portion of our data for the presence of oil droplets using phytane as a surrogate indicator, again reaching the scientifically indefensible conclusion that microdroplets explain embryo toxicity. Brannon et al. have presented no evidence of microdroplets as an agent of embryo mortality, whereas we have done so for PAHs (Marty et al. 1997; Heintz et al. 1999, 2000; Carls et al. 1999, 2005) and we have definitively eliminated microdroplets as directly toxic (Carls et al. 2008b). This web document provides the data in question so readers can evaluate the validities of conflicting conclusions based on identical observations

Yes: ORC experiments are confounded by particulate oil

(Brannon et al. 2006)

- The presence of phytane in water is evidence of oil microdroplets.
- Of the 31 exposure water samples, 29 contained phytane above its method detection limit.
- Hence oil droplets were present in exposure water.
- Embryos were exposed to both dissolved and droplet-associated PAH in effluent water
- Consequently, the conclusion that dissolved PAH concentrations as low as 1 ppb were toxic is incorrect
- Because oil droplets were present in exposure water, embryos received a higher dose of high molecular weight PAH than predicted by dissolved PAH.

No: ORC experiments are not confounded by particulate oil

(Carls et al. 2008a)

- The presence of phytane in water is evidence of oil microdroplets.
- Phytane was not detected in 75% of 32 water samples and analytic methods provide no certainty that the remaining 25% contained phytane, despite the non-zero concentrations estimated for them. In the 8 samples where the presence of phytane could not be discounted, estimated concentrations were small fractions of the concentration of the smallest standard (1.6 to 8.2%, mean 3.6%), precluding reliable quantification. Analytic methods for phytane can produce false positives because standard curves are not forced through zero and false positives cannot be eliminated procedurally because phytane is so insoluble that we have been unable to establish method detection limits for it in water.
- Aqueous phytane concentrations were unrelated to oil treatment, hence particulate oil was either absent or negligible in exposure water.
- Embryo exposure to particulate oil was negligible (*see data*). Phytane was above method detection limits in eggs in 1 intermediate dose at 1 intermediate time.
- The absence of phytane in water and eggs and changes in PAH composition from whole oil to water and eggs in multiple experiments (Marty et al. 1997; Heintz et al. 1999, 2000; Carls et al. 1999, 2005) consistently identify dissolved PAHs as the source of embryo toxicity, not particulate oil.
- Higher molecular weight PAH were never observed in eggs, eliminating whole oil as the source of contamination. **All hydrocarbons present in water samples were measured, regardless of phase (solid or dissolved)** in their experiment and ours. Acceptance or rejection of the particulate oil hypothesis has no influence on measured concentration.
 - Damage to embryos in effluent water (without oil contact) was the same as damage to embryos in contact with whole oil (Heintz et al. 1999; Brannon et al. 2006). This indicates direct contact is unimportant, also demonstrated with zebrafish embryos exposed to or isolated from oil droplets (Carls et al. 2008b).

Data

(click to view)

Concluding remarks

Are oiled rock column (ORC) experiments confounded by particulate oil?

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Brannon et al. (2006) published a paper in Environmental Toxicology and Chemistry where they argued that dissolved polynuclear aromatic hydrocarbons (PAHs) from oil are toxic to embryos (Heintz et al. 1999) where we concluded that toxicity was caused by contact with PAH-laden oil microdroplets in concentrations of magnitude higher. We conclude that flawed analysis logic and the lack of any direct evidence of oil droplets in the water column (Heintz et al. 2008; Carls et al. 2008a). In their rebuttal, Brannon et al. (2008) reanalyzed a study that used phytane as a surrogate indicator, again reaching the scientifically indefensible conclusion that microdroplets are toxic as an agent of embryo mortality, whereas we have done so for PAHs (Marty et al. 2008) and used microdroplets as directly toxic (Carls et al. 2008b). This web document presents additional evidence from identical observations

Yes: ORC experiments are confounded by particulate oil

- The presence of phytane in water samples was above the detection limit.
- Of the 31 exposure water samples, 28 were above the detection limit.
- Hence oil droplets were present in the water column.
- Embryos were exposed to oil in effluent water.
- Consequently, the 28 ppb were toxic is in the water column.
- Because oil droplets are present, a higher dose of high PAH.

Data

Phytane in water
(click here to view data)

Phytane in eggs
(click here to view data)

Yes: ORC experiments are confounded by particulate oil

droplets. and analytic methods for phytane, despite the fact that samples where the concentrations were above the detection limit (1.6 to 8.2%, which is above the detection limits for phytane in water forced through a filter because phytane is not removed by filtration limits for phytane in water. Hence, the presence of phytane in water, hence the presence of oil droplets. Phytane was detected in intermediate concentrations in the water column. The composition of the oil was similar to that of the oil used by Heintz et al. 1997; and we have recently identified the oil. The presence of oil droplets in the water column, eliminating the possibility of phytane being present in the water column (or dissolved) in the particulate oil. The presence of oil droplets in the water column (oil contact) was the cause of the mortality of the embryos (Heintz et al. 2008). This indicates direct contact is the cause of the mortality of the embryos exposed to the oil (Heintz et al. 2008b).

Phytane in water

Phytane × QCbatch
(click for illustration)

Phytane × treatment
(click for illustration)

**For comparison,
TPAH × treatment**
(click for illustration)

treatment	level	day	SIN	QCbatch	phytane	QC
control	0	-1	404316	R08034	0.014	a
control	0	0	404436	R08034	0.014	a
control	0	0	404405	R08124	0.018	a
control	0	35	405219	R08224	0.020	a
control	0	35	405217	R08224	0.020	a
control	0	35	405218	R08224	0.020	a
control	0	63	405448	R08264	0.030	a
control	0	63	405437	R08264	0.030	a
control	0	63	405438	R08264	0.030	a
control	0	93	405701	R08124	0.025	b
oil	1	0	404404	R08124	0.018	a
oil	1	35	405237	R08224	0.020	a
oil	1	63	405503	R08264	0.030	a
oil	2	0	404403	R08124	0.018	a
oil	2	35	405244	R08224	0.020	a
oil	2	63	405506	R08264	0.030	a
oil	3	0	404402	R08124	0.032	b
oil	3	36	405313	R08264	0.030	a
oil	3	63	405510	R08264	0.030	a
oil	3	93	405702	R08124	0.018	a
oil	4	0	404401	R08034	0.040	b
oil	4	36	405325	R08264	0.030	a
oil	4	63	405518	R08304	0.020	a
oil	5	0	404350	R08034	0.014	a
oil	5	36	405330	R08264	0.030	a
oil	5	63	405521	R08304	0.028	b
oil	5	93	405703	R08124	0.023	b
oil	6	0	406001	R02285	0.010	b
oil	6	36	405340	R08264	0.041	b
oil	6	63	405522	R02285	0.000	b
VWO	7	0	404349	R02285	0.000	a
VWO	7	63	405513	R08264	0.030	a

SIN: sample identification number QCbatch: quality control batch

QC: a = below level of detection, b = below level of quantification

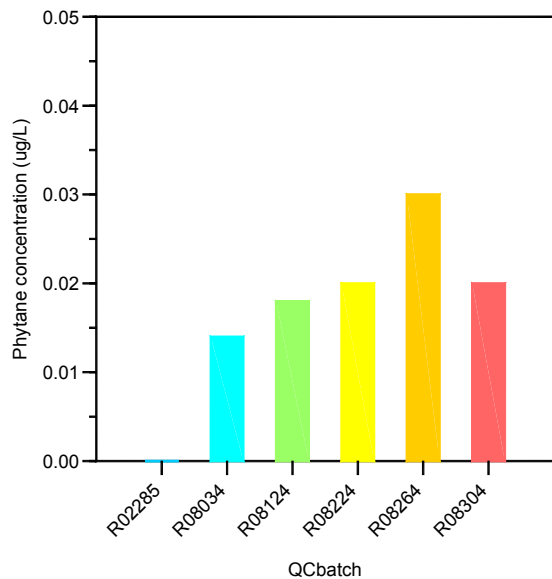
VWO: very weathered oil. Phytane concentration is µg/L.

Level: 1 is low oil, ... 6 is high oil

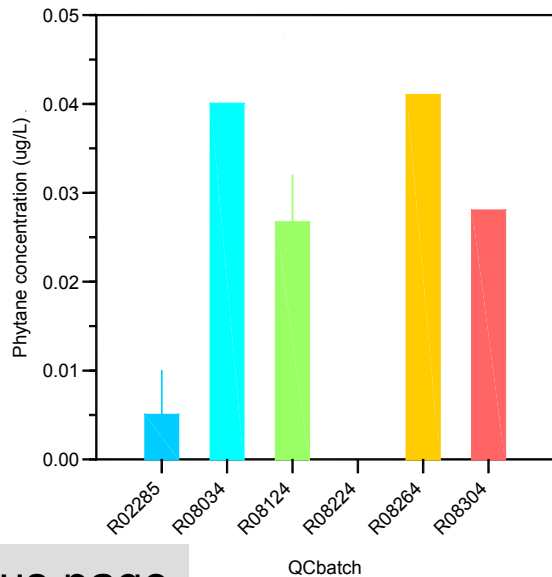
Phytane in water: phytane × QCbatch

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A) Samples where phytane was not detected (n = 24). There was no variance within QCbatch, only among QCbatch.



B) Samples where phytane was below quantification limits (n = 8). Error bars are minimum to maximum range.

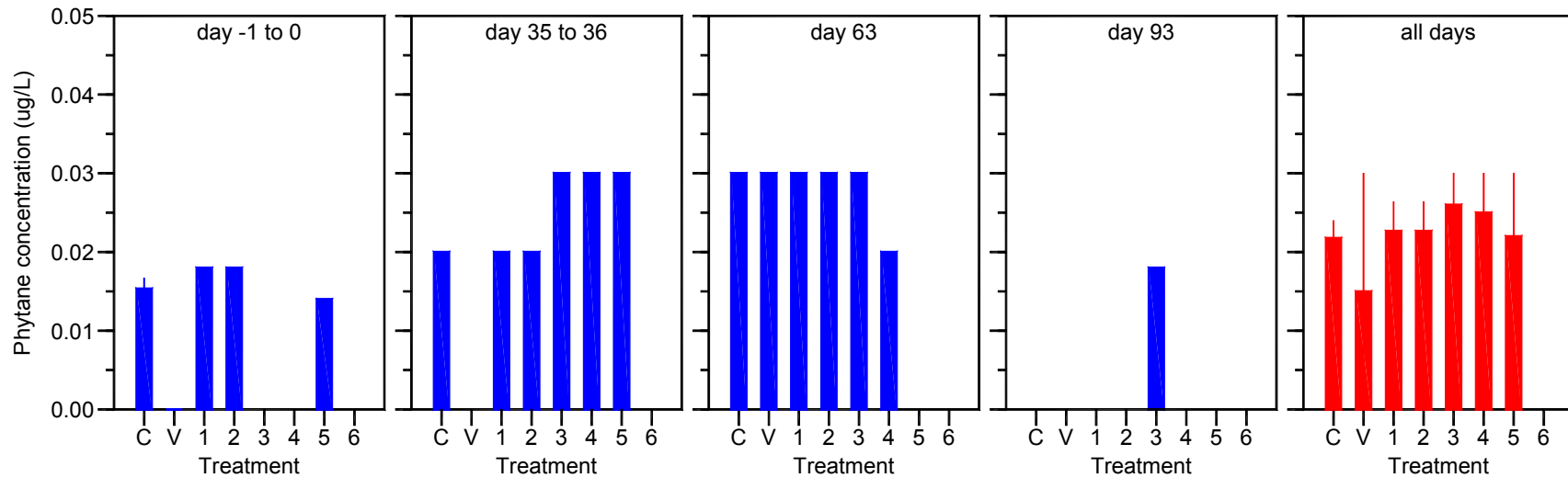


treatment	level	day	SIN	QCbatch	ug/L	
					phytane	QC
VWO	7	0	404349	R02285	0.000	a
oil	5	0	404350	R08034	0.014	a
control	0	0	404436	R08034	0.014	a
control	0	-1	404316	R08034	0.014	a
control	0	0	404405	R08124	0.018	a
oil	1	0	404404	R08124	0.018	a
oil	2	0	404403	R08124	0.018	a
oil	3	93	405702	R08124	0.018	a
control	0	35	405219	R08224	0.020	a
oil	1	35	405237	R08224	0.020	a
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control	0	35	405217	R08224	0.020	a
control	0	35	405218	R08224	0.020	a
control	0	63	405448	R08264	0.030	a
oil	1	63	405503	R08264	0.030	a
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oil	5	36	405330	R08264	0.030	a
VWO	7	63	405513	R08264	0.030	a
control	0	63	405437	R08264	0.030	a
control	0	63	405438	R08264	0.030	a
oil	4	63	405518	R08304	0.020	a
control	0	93	405701	R08124	0.025	b
oil	3	0	404402	R08124	0.032	b
oil	4	0	404401	R08034	0.040	b
oil	5	63	405521	R08304	0.028	b
oil	5	93	405703	R08124	0.023	b
oil	6	0	406001	R02285	0.010	b
oil	6	36	405340	R08264	0.041	b
oil	6	63	405522	R02285	0.000	b

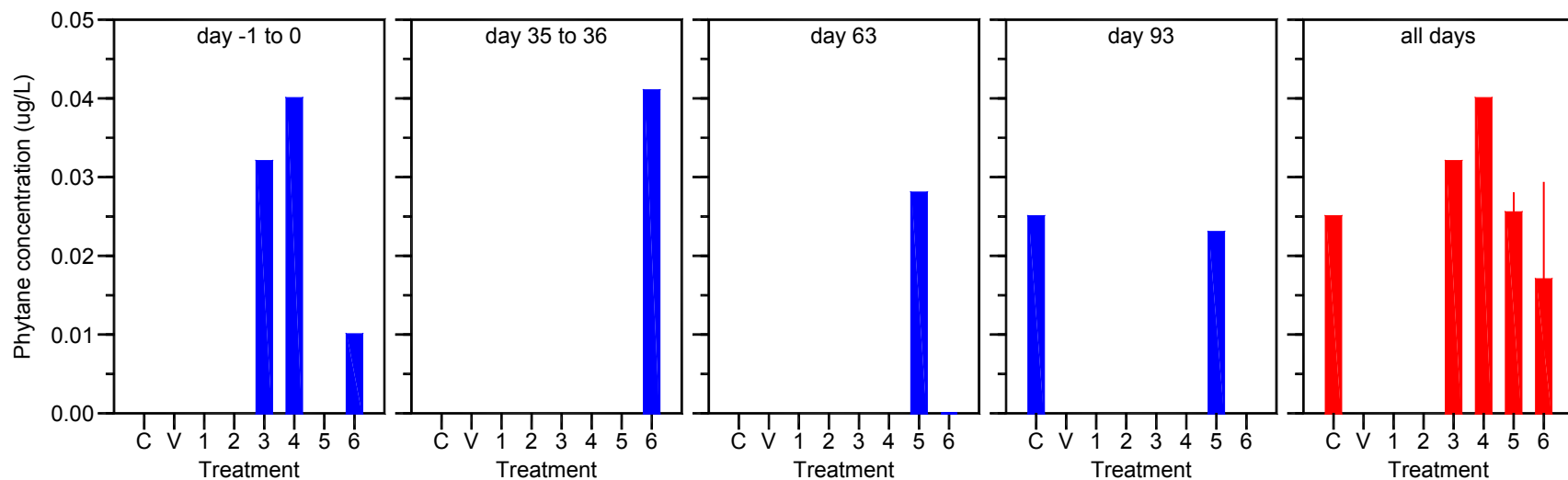
Phytane in water: phytane × treatment

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A) Samples where phytane was not detected (n = 24). Error bars are standard error. Treatment C is control, V is very weathered oil, 1 is low oil, 6 is high oil.



B) Samples where phytane was detected (n = 8). Error bars are standard error.



Phytane in eggs

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treat	sday	ng/g dry weight phytane	treat	sday	ng/g dry weight phytane
0	36	0	4	36	0
0	65	0	4	65	0
0 *	65	0	4	183	0
0	93	0	5	36	0
0	126	0	5 *	42	0
0 *	178	0	5	42	0
0	183	0	5	65	0
1	36	0	5 *	65	0
1	65	0	5	93	0
1	183	0	5	126	0
2	36	0	5 *	178	0
2	65	0	5	183	0
2	183	0	6	36	0
3	42	0	6	65	0
3 *	42	0	6	183	0
3	65	0	7	36	0
3	65	0	7	65	0
3	93	230	7	183	0
3	126	0			
3 *	178	0			
3	183	0			

Treatment 0 = control, 1=lowest oil dose, ... 6=highest oil dose; 7 = very weathered oil.

*Eggs were suspended in effluent and not in contact with whole oil. All other eggs were in contact or potential contact with oiled gravel.

Phytane in water: phytane × QCbatch, our conclusion

All reported aqueous phytane concentrations are likely random noise. Phytane was not detected in 75% of 32 water samples. Aqueous phytane concentrations in the remaining samples were far below the level of quantification (1.6 to 8.2% of the concentration of the smallest standard) and were not significantly different from those where phytane was not detected.

$$P_{\text{ANOVA}} = 0.387$$

The reason undetected phytane concentration estimates are sometimes greater than zero is because standard curves are not forced through zero; positive intercepts result in small, nonzero positive estimates where no phytane was detected.

Phytane in water: phytane × treatment, our conclusion

Aqueous phytane concentrations were unrelated to oil treatment, hence particulate oil was either absent or negligible in exposure water. Aqueous phytane concentrations were not dose-dependent and were unrelated to time.

Phytane in eggs: our conclusion

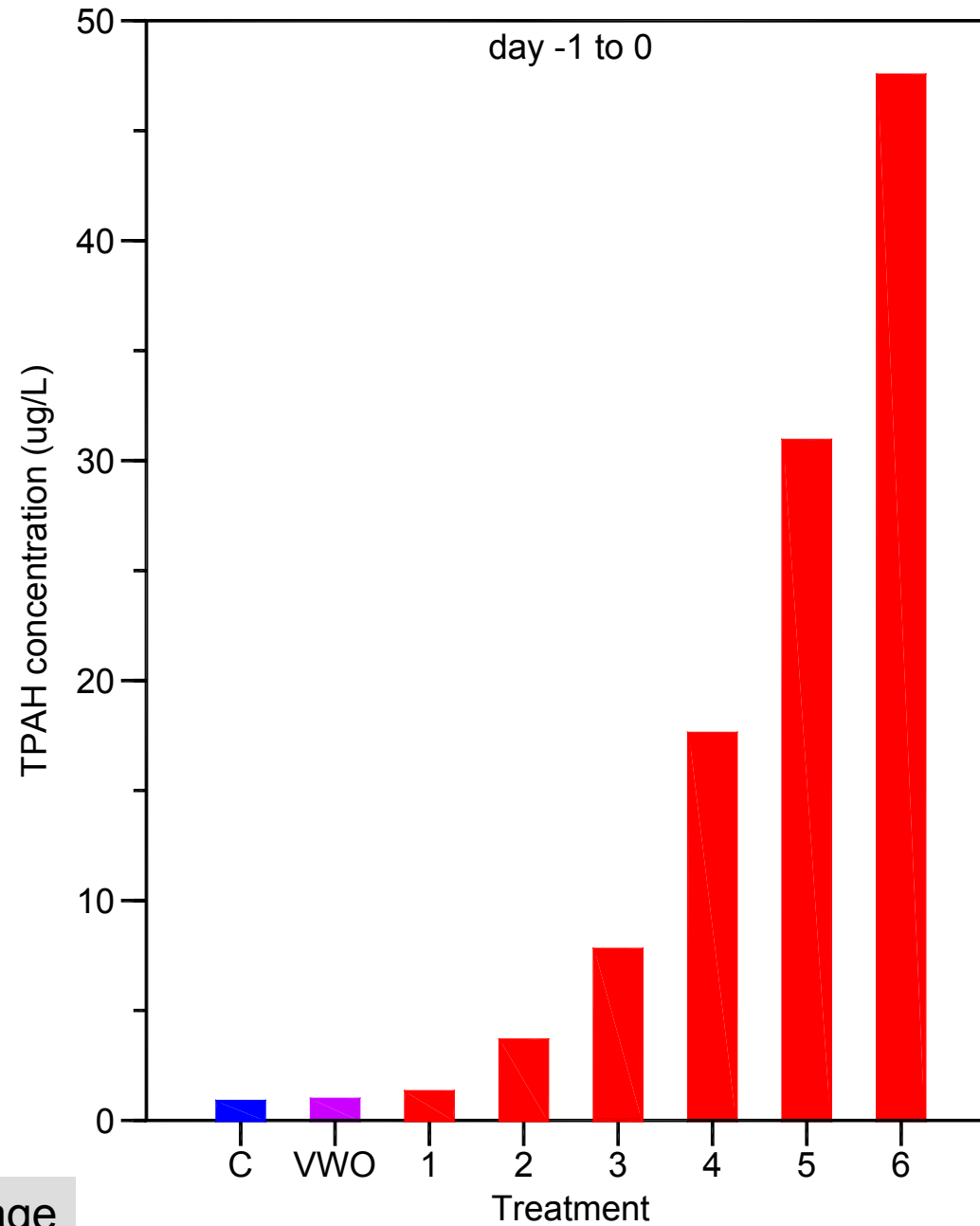
Embryos did not accumulate phytane, indicating exposure to whole oil was either negligible or inconsequential. Phytane concentrations in tissue were neither dose nor time-dependent. Lipophylic eggs are capable of accumulating hydrocarbons to concentrations approximately 1000 times greater than in water, thus they conveniently amplify hydrocarbon levels, improving detection. Despite this amplification (bioaccumulation), phytane was below method detection limits in pink salmon eggs with one exception at an intermediate dose and time ($n_{\text{total}} = 39$). We suspect the exception was caused by sample contamination: this was the only significantly elevated phytane concentration in 147 egg samples collected across five experiments and was more than 5 times greater than any other non-zero phytane concentration in eggs. In contrast, TPAH concentrations in eggs were dose and time dependent.

[Click here to compare to TPAH in eggs](#)

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Total polynuclear aromatic hydrocarbons (TPAH) in water

[Click here for our conclusion](#)



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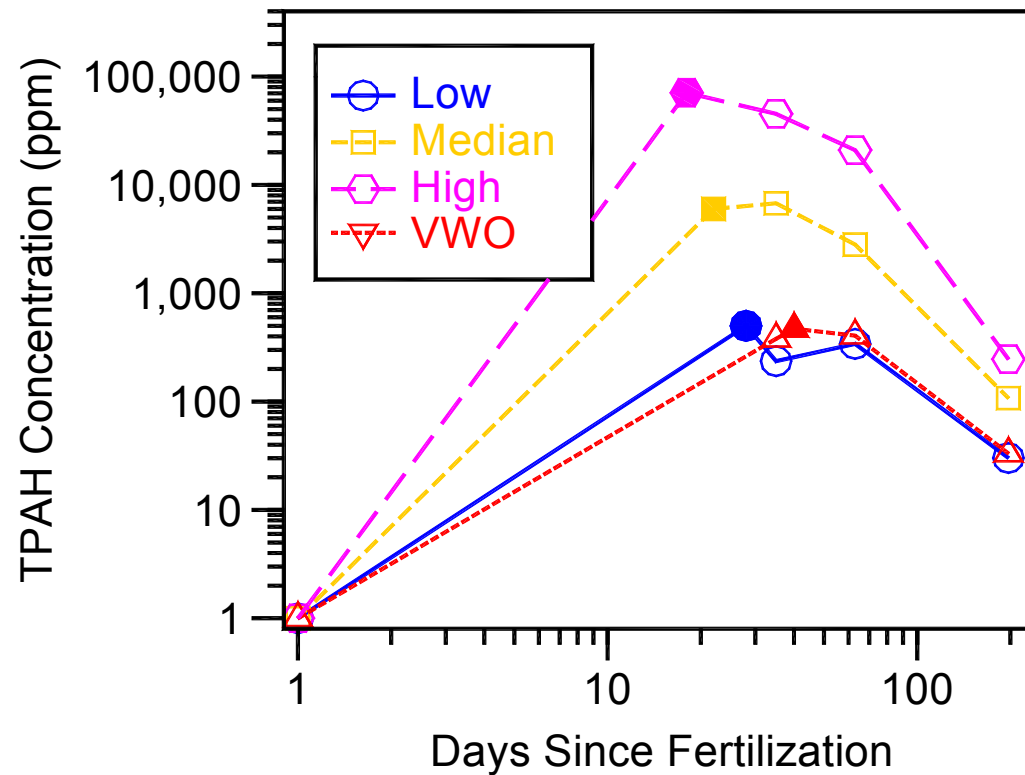


Fig. 3 from Heintz et al. 1999 [Environ Toxicol Chem 18(3):494-503]. Changes in tissue concentrations of total polynuclear aromatic hydrocarbon (TPAH) with time for pink salmon incubated in oil. Lines depict uptake for highest, median, and lowest doses used in the direct exposure experiment and very weathered oil (VWO). Open symbols depict values obtained by gas chromatography and mass spectrometry; filled symbols depict estimates of maximum TPAH concentrations as described in the original paper.

Total polynuclear aromatic hydrocarbons (TPAH) in water: our conclusion

In contrast to aqueous phytane concentrations, total polynuclear aromatic hydrocarbon (TPAH) concentrations in water were dose (and time) dependent.

Total polynuclear aromatic hydrocarbons (TPAH) in eggs: our conclusion

In contrast to phytane concentrations in eggs, PAH concentrations were both dose- and time-dependent.

Concluding remarks

The scientifically indefensible conclusion Brannon et al. (2008) reach, that our phytane data demonstrates our experiments were contaminated by particulate oil, illustrates the interpretive problems we identified in their original paper (Brannon et al. 2006). Their argument that “the multiple ABL studies ... have incorrectly determined the actual PAH dose” because “the dosing columns produce an aqueous effluent containing oil droplets” is illogical and invalid. All hydrocarbons present in water samples were measured, regardless of phase (solid or dissolved) in their experiment and ours. Acceptance or rejection of the particulate oil hypothesis has no influence on these measured concentrations. Measured aqueous TPAH concentrations were < 100 ppb in all treatments across all our experiments (n = 167) and biologically damaging aqueous TPAH concentrations were < 20 ppb in all experiments.

Whether researchers agree or disagree about the toxicological importance of particulate oil, comparison of nominal oil concentrations and aqueous TPAH concentrations as though they were equal measures is invalid and that is how Brannon et al. (2006) began their paper. This invalid comparison is doubly pernicious because measured aqueous TPAH doses in their experiment corroborated our measures. Apparently Brannon et al. believe that hypothetical oil droplets in water explain the orders of magnitude difference between nominal oil application and measured dose in opposition to consistent chemical evidence to the contrary. Apparently they believe that visual observation of rare oil slicks provides a more rigorous basis for interpretation than the wide variety of chemical evidence that repeatedly documents negligible quantities of particulate oil in water (including their own results) and in embryo tissue.

Evidence across multiple independent studies indicates dissolved PAHs are toxic to fish embryos at low concentrations without the presence of particulate oil (Kiparissis et al. 2003; Rhodes et al. 2005; Farwell et al. 2006; Carls et al. 2008b). The reliance on nominal dose for interpretation by Brannon et al. (2006) has resulted in multiple errors as detailed in our [original review](#), including toxicity estimates about five orders of magnitude too low and inconsistent with all other literature (Birtwell and McAllister 2002; Rhodes et al. 2005; Farwell et al. 2006). If reliance on nominal dose were acceptable, then no one would need bother with difficult and costly hydrocarbon measurements; crude measures of applied oil would be sufficient. Most toxicologists rejected this approach decades ago.

References

- Birtwell, I. K. and C. D. McAllister 2002. Hydrocarbons and their effects on aquatic organisms in relation to offshore oil and gas exploration and oil well blowout scenarios in British Columbia, 1985. Canadian Technical Report of Fisheries and Aquatic Sciences 2391: 1-51.
- Brannon, E. L. and A. W. Maki 1996. The *Exxon Valdez* oil spill: analysis of impacts on the Prince William Sound pink salmon. *Reviews in Fisheries Science* 4: 289-337.
- Brannon, E. L., K. M. Collins, J. S. Brown, J. M. Neff, K. R. Parker and W. A. Stubblefield 2006. Toxicity of weathered *Exxon Valdez* crude oil to pink salmon embryos. *Environmental Toxicology and Chemistry* 25(4): 962-972.
- Brannon, E. L., J. S. Brown, J. M. Neff, K. R. Parker and W. A. Stubblefield 2008. Authors' Reply, Letter to the Editor. *Environmental Toxicology and Chemistry*.
- Carls, M. G., S. D. Rice and J. E. Hose 1999. Sensitivity of fish embryos to weathered crude oil: Part I. Low-level exposure during incubation causes malformations, genetic damage, and mortality in larval Pacific herring (*Clupea pallasii*). *Environmental Toxicology and Chemistry* 18(3): 481-493.
- Carls, M. G., R. A. Heintz, G. D. Marty and S. D. Rice 2005. Cytochrome P4501A induction in oil-exposed pink salmon *Oncorhynchus gorbuscha* embryos predicts reduced survival potential. *Marine Ecology-Progress Series* 301: 253-265.
- Carls, M.G., Short, J.W., Rice, S.D., Heintz, R.A. 2008a. Response to Brannon et al. 2006. Web publication: <http://www.afsc.noaa.gov/ABL/Habitat/pdfs/review-3.pdf>.
- Carls, M. G., L. Holland, M. L. Larsen, T. K. Collier, N. L. Scholz and J. P. Incardona 2008b. Fish embryos are damaged by dissolved PAHs, not oil particles. *Aquatic Toxicology* 88: 121-127.
- Farwell, A., V. Nero, M. Croft, P. Bal and D. G. Dixon 2006. Modified Japanese medaka embryo-larval bioassay for rapid determination of developmental abnormalities. *Archives of Environmental Contamination and Toxicology* 51(4): 600-607.
- Heintz, R. A., J. W. Short and S. D. Rice 1999. Sensitivity of fish embryos to weathered crude oil: Part II. Increased mortality of pink salmon (*Oncorhynchus gorbuscha*) embryos incubating downstream from weathered *Exxon Valdez* crude oil. *Environmental Toxicology and Chemistry* 18(3): 494-503.
- Heintz, R. A., S. D. Rice, A. C. Wertheimer, R. F. Bradshaw, F. P. Thrower, J. E. Joyce and J. W. Short 2000. Delayed effects on growth and marine survival of pink salmon *Oncorhynchus gorbuscha* after exposure to crude oil during embryonic development. *Marine Ecology-Progress Series* 208: 205-216.

Heintz, R. A., J. W. Short, S. D. Rice and M. G. Carls 2008. Comment on "Toxicity of weathered *Exxon Valdez* crude oil to pink salmon embryos. *Environmental Toxicology and Chemistry* 27: 1475-1476.

Kiparissis, Y., P. Akhtar, P. V. Hodson and R. S. Brown 2003. Partition-controlled delivery of toxicants: A novel in vivo approach for embryo toxicity testing. *Environmental Science & Technology* 37(10): 2262-2266.

Marty, G. D., J. W. Short, D. M. Dambach, N. H. Willits, R. A. Heintz, S. D. Rice, J. J. Stegeman and D. E. Hinton 1997. Ascites, premature emergence, increased gonadal cell apoptosis, and cytochrome P4501A induction in pink salmon larvae continuously exposed to oil-contaminated gravel during development. *Canadian Journal of Zoology* 75(6): 989-1007.

Neff, J. M. and W. A. Burns 1996. Estimation of polycyclic aromatic hydrocarbon concentrations in the water column based on tissue residues in mussels and salmon: An equilibrium partitioning approach. *Environmental Toxicology and Chemistry* 15(12): 2240-2253.

Rhodes, S., A. Farwell, L. M. Hewitt, M. MacKinnon and D. G. Dixon 2005. The effects of dimethylated and alkylated polycyclic aromatic hydrocarbons on the embryonic development of the Japanese medaka. *Ecotoxicology and Environmental Safety* 60(3): 247-258.