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Supporting Information

Files in this Data Supplement:

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| 1 | MLITVKNSQMVRPAAPTPQRDLWNSNVDLVVPRIHTASVYFYRPTGSPDFFSMNILRDAL | PrHCT |
|-----|---|-------|
| 1 | MKIEVKESTMVKPAAETPQQRLWNSNVDLVVPNFHTPSVYFYRPTGSPNFFDGKVLKEAL | NtHCT |
| 61 | SKLLVPFYPMAGRLKRDPDGRIEINCNGEGVLLVEAITDSVIDDFGDFAPTMELKQLIPK | PrHCT |
| 61 | SKALVPFYPMAGRLCRDEDGRIEIDCKGQGVLFVEAESDGVVDDFGDFAPTLELRQLIPA | NtHCT |
| 121 | VNYSEDISSYPLLVLQVTFFKCGGVSLGVGMQHHVADGYAGIHFINTWSDVARGLDITLP | PrHCT |
| 121 | VDYSQGIQSYALLVLQITHFKCGGVSLGVGMQHHAADGASGLHFINTWSDMARGLDLTIP | NtHCT |
| 181 | PFIDRTLLRARNPPTPKFQHIEYQQPPPLKDTSGIMNGEKTDISVAIFKLTKEQLEIL | PrHCT |
| 181 | PFIDRTLLRARDPPQPQFPHVEYQPPPTLKVTPENTPISEAVPETSVSIFKLTRDQINTL | NtHCT |
| 239 | KGKARENGNNIAYSSYEMLSGHIWRCACKARNLAEDQETKLYIATDGRNRLRPSIPPGYF | PrHCT |
| 241 | KAKSKEDGNTVNYSSYEMLAGHVWRSTCMARGLAHDQETKLYIATDGRSRLRPSLPPGYF | NtHCT |
| 299 | GNVIFTTTPMAVTGDIISKPTYYAASVIHEALGRMDDEYLRSALDYLELQPDLTALVRGA | PrHCT |
| 301 | GNVIFTTTPIAVAGDIQSKPIWYAASKIHDALARMDNDYLRSALDYLELQPDLKALVRGA | NtHCT |
| 359 | HTFRCPNIGITSWSRLPIHDADFGWGRPIFMGPGGIAYEGLAFVLPS <mark>SV</mark> NDGSLSVALGL | PrHCT |
| 361 | HTFKCPNLGITSWSRLPIHDADFGWGRPIFMGPGGIAYEGL <mark>SFI</mark> LPSPTNDGSQSVAISL | NtHCT |
| 419 | QPDHMVRFAKMLYEI. | PrHCT |
| 421 | Qaehmklfekflydf. | NtHCT |

Fig. 5. Alignment of the deduced amino acid sequence of the putative *P. radiata HCT* clone *PrHCT* (top strand; accession no. EF121452) and the amino acid sequence of *N. tabacum HCT* (bottom strand; accession no. AJ507825). Conserved amino acids have gray and identical amino acids have black background.



Fig. 6. Schematic diagrams of the constructs used in *HCT* gene-silencing experiments in *P. radiata* callus cultures. *pAW16* contains the *NPT II* resistance gene controlled by the *Z. mays UB11* promoter and the *GUS* reporter gene driven by the double 35S CaMV promoter; *pHF5* contains an inverted repeat of the *P. radiata HCT* coding region separated by the *Z. mays UB11* intron and controlled by the *Z. mays UB11* promoter. The position of genomic PCR fragments generated to investigate the integration of construct is indicated.



Fig. 7. Pyrogram (total ion chromatogram) of transgenic line pHF5-18 (*A*) compared with a WT control (*B*). Numbers 1-31 refer to the following major thermal breakdown products: 1, 2-furfuryl alcohol; 2, (3*H*) furan-2-one; 3, dihydro-methyl-furanone; 4, 2-hydroxy-1-methyl-1-cyclopentene-3-one; 5, guaiacol; 6, 4-hydroxy-5,6-dihydro-(2*H*)-pyran-2-one; 7, 4-methyl-guaiacol; 8, phenol; 9, 4-ethyl-guaiacol; 10, dimethyl-phenol; 11, 4-methyl-phenol; 12, eugenol; 13, 4-ethyl-phenol; 14, 4-vinyl-guaiacol; 15, *cis*-isoeugenol; 16, 4-allyl-phenol; 17, *trans*-isoeugenol; 18, 4-vinyl-phenol; 19, *cis*-4-propenyl-phenol; 20, 5-hydroxymethyl-2-furaldehyde; 21, *trans*-4-propenyl-phenol; 22, vanillin; 23, 1,5-anhydro-arabinofuranose; 24, acetoguaiacone; 25, guaiacyl acetone; 26, 1,5-anhydro-xylofuranose; 27, *p*-coumaryl alcohol; 28, 4-hydroxy-benzaldehyde; 29, resorcinol; 30, coniferaldehyde; 31, *trans*-coniferyl alcohol. Signals, which largely represent thermal breakdown products of H lignin, such as **8**, **16**, **18**, **19**, and **21** are significantly enriched in pHF5-18 compared with the WT control.



Fig. 8. Partial short-range ${}^{13}C{}^{-1}H$ (HSQC) spectra (side-chain regions) of acetylated enzyme lignins isolated from (*A*) the wild-type control and (*B*) *HCT*-deficient line pHF5-18.