Title: Charcoal effects on soil solution chemistry and growth of *Koeleria macrantha* in the ponderosa pine/Douglas-fir ecosystem

List of Authors:

Michael J. Gundale[1], Thomas H. DeLuca[2]

Department of Ecosystem and Conservation Sciences, University of Montana, Missoula, MT 59812

Corresponding Authors:
[1]E-mail address: mjgundal@yahoo.com
[2]Tel.: 1-406-243-4425; E-mail address: tom.deluca@cfc.umt.edu

Corresponding address during review process:

Michael J. Gundale
Department of Ecosystem and Conservation Sciences
University of Montana
32 Campus Drive
Missoula, MT 59812
Phone: 1-406-243-5326

Abstract

We conducted laboratory and greenhouse experiments to determine whether charcoal derived from the ponderosa pine/Douglas-fir ecosystem may influence soil solution chemistry and growth of *Koeleria macrantha,* a perennial grass that thrives following fire. In our first experiment, we incubated forest soils with a factorial combination of Douglas-fir wood charcoal generated at 350 ºC and extracts of *Arctostapholos uva-ursi,* with and without the addition of glycine as a labile N source. These results showed that charcoal increased N mineralization and nitrification when glycine was added, but reduced N mineralization and nitrification without the addition of glycine. Charcoal significantly reduced the solution concentration of soluble phenols from litter extracts, but may have contributed bio-available C to the soil that resulted in N immobilization in the no-glycine trial. In our second experiment, we grew *K. macrantha* in soil amended with charcoal made at 350 ºC from ponderosa pine and Douglas-fir bark. Growth of *K. macrantha* was significantly diminished by both of these charcoals relative to the control. In our third experiment, we grew *K. macrantha* in soil amended with six concentrations (0, 0.5, 1, 2, 5, and 10%) of charcoal collected from a wildfire. The data showed increasing growth of *K. macrantha* with charcoal addition, suggesting some fundamental differences between laboratory generated charcoal and wildfire produced charcoal. Further, they suggest a need for a better understanding of how temperature and substrate influence the chemical properties of charcoal.

Introduction

It is well established that fire alters N cycling in the ponderosa pine/Douglas-fir (*Pinus ponderosa/Psuedotsuga menziesii*) ecosystem (Neary et al. 1999; Hart et al. 2005). Nitrogen availability has been shown to increase immediately following fire, (Covington and Sackett 1990, 1992; DeLuca and Zouhar 2000), and may remain elevated on the scale of months to years as a result of enhanced mineralization (Covington and Sackett 1990, 1992; Monleon et al. 1997; Kaye and Hart 1998; Gundale et al. 2005). Numerous processes have been identified that increase N mineralization following fire, including: improved substrate quality (White 1991, 1994; Fernandez et al. 1997; Pietikainen et al. 2000a), death of roots and soil organisms resulting in a large labile organic N pool (DeBano et al. 1979; Dunn et al. 1979; Diaz-Ravina et al. 1996; Neary et al. 1999), and a reduction in C:N ratios due to preferential loss of C during combustion (Gundale et al. 2005). A potentially overlooked factor that may also enhance N cycling following fire is the addition of charcoal to soils.

Several recent studies have shown that charcoal has the potential to greatly enhance soil fertility. Amazonian forest soils amended with charcoal and manure centuries ago still maintain some of the highest biodiversity and productivity of any soils within the Amazon basin (Glaser et al. 2001; 2002; Mann 2002). In boreal forest soils, charcoal has been shown to enhance N cycling by ameliorating the inhibitory effects of litter extracts from late-successional species, which in turn promotes growth of early-successional species (Zackrisson et al. 1996; Wardle et al. 1998; DeLuca et al. 2002; Berglund et al. 2004). Recently, DeLuca et al. (2005) found that the addition of wildfire formed charcoal to ponderosa pine forest soils increased nitrification rates.

Charcoal may enhance soil fertility through a variety of mechanisms. Increased N turnover may occur by charcoal sorption of high C:N organic molecules from the soil solution (Zackrisson et al. 1996; Wardle et al. 1998; Glaser et al. 2002), resulting in reduced microbial N immobilization and higher net mineralization and nitrification rates. Additionally, charcoal may remove specific groups of organic molecules, including polyphenol or monoterpene compounds that are thought to inhibit nitrification (Rice and Pancholy 1972; Zackrisson et al. 1996; DeLuca et al. 2002; Berglund et al. 2004). Sorption of organic molecules, along with the gradual breakdown of charcoal, may initiate humus formation and thus enhance long term-soil fertility (Glaser et al. 2002). Charcoal may also enhance soil fertility by creating habitat for microbes within its porous structure (Pietikainen et al. 2000b).

Despite these potential roles that charcoal may have in increasing soil fertility, its ecological role in forest ecosystems, such as ponderosa pine/Douglas-fir, has received little attention. We conducted three separate experiments, using low-temperature charcoal to investigate whether charcoal influences soil solution chemistry, and growth of an early successional species. In our first experiment, our objective was to determine whether charcoal had an influence on soil solution chemistry following addition of the extracts of a late successional species, *Arctostapholos uvi-ursi,* via surface adsorption of phenolic compounds. We hypothesized that charcoal added to a ponderosa pine soil will effectively sorb the phenol fraction in litter extracts, which would correspond with enhanced N cycling

In our second experiment, our objective was to compare the influence of charcoal made from the bark of two species, ponderosa pine and Douglas-fir, on growth of

*Koeleria macrantha***,** a perennial grass species that thrives following fire disturbance in western Montana ponderosa pine/Douglas-fir forests. Bark charring during low intensity wildfire is a potentially significant source of charcoal in this system. Charred bark may gradually slough from trees following fire and become incorporated in the soils surrounding trees. It is recognized that ponderosa pine is a more fire adapted species than Douglas-fir, thus an intriguing hypothesis is that charred bark of the more fire adapted species will have a stronger positive effect on N cycling processes and plant growth.

In our third experiment, our objective was to determine whether charcoal generated during a wildfire would have any effect on *K. macrantha* growth, and to determine whether this relationship is dependent on soil charcoal concentration. We hypothesized that wildfire charcoal will positively influence *K. macrantha*, and that this effect will increase as a function of soil charcoal concentration. Collectively, these three experiments address our central hypothesis, that charcoal will alter solution chemistry by sorbing phenols and enhancing N cycling, which in turn will improve the growth of early successional species.

Methods

All three experiments utilized field collected soil, which was collected from the subsurface horizon (20 – 30 cm, B_w Horizon) of a forest soil associated with low elevation (1,100 m) ponderosa pine/Douglas-fir vegetation in western Montana, USA. The soil is a sandy-skeletal, mixed, frigid Typic Dystrustepts. This ecosystem is characterized by low annual rainfall (<350 mm annually) with approximately 50% falling as snow during the winter months. Soil was collected during the month of September,

returned to the lab, upon which they were sieved (4mm) and homogenized. We then added one part sand to three parts field moist soil (by mass) to decrease fertility and increase porosity and gas exchange, such that nitrification would not be limited by low O_2 availability. The sand fraction was purchased as filter grade silica sand (for pool filters), and was washed with 1 M HCl, followed by distilled water, before being homogenized with field collected soil. This sand amended soil had a pH of 6.8, electrical conductance of 91.2 μS m^{-1}, and had a textural distribution of 71 % sand, 21 % silt and 8 % clay.

All experiments also included the addition of either laboratory-generated charcoal from Douglas-fir and ponderosa pine, or charcoal collected in the field following a wildfire. Laboratory charcoal was generated by burying wood or bark from these species in silica sand, and heating at 350 °C for 2 hours. Charcoal was then ground and sieved as specified for each experiment. Various physical and chemical properties of these charcoals were measured (Table 1). Charcoal pH was measured from a 4:1 slurry of deionized water to charcoal. Electrical conductance (EC) was measured from charcoal paste (2:1 distilled water and charcoal). Cation exchange capacity was estimated on charcoal samples via NH_4^+ replacement, where 1 gram of charcoal was rinsed twice with 25 ml of 1 M ammonium acetate (pH 7) to saturate exchange sites. Excess saturating solution was removed from charcoal samples with three consecutive washes with 25 ml of 95% ethyl alcohol. Sorbed NH_4^+ was then extracted with 25 ml of 2 M KCl and analyzed on a segmented flow analyzer (Auto Analyzer III, Bran Luebbe, Chicago, IL) using the berthelot reaction (Willis et al. 1993). Charcoal density was measured by measuring dry mass of intact charcoal pieces and measuring volume displacement in

deionized water. Total C was measured via dry combustion on a Fissions Elemental Analyzer (Milano, Italy). Soluble and total phenols were measured by extracting 1 g of ground charcoal in 25 ml of deionized water and 50% methanol, respectively, and were analyzed using the Prussian Blue Method (Stern et al. 1996). Extractable NH_4^+ and NO_3^- (Mulvaney 1996) were extracted by shaking 1 g of charcoal for 30 minutes in 25 ml of 2 M KCl, and then filtering through Whatman #2 filters. The extracts were analyzed for NH_4^+-N using the berthelot reaction (Willis et al. 1993) and NO_3^- -N by the cadmium reduction method (Willis and Gentry 1987) on a segmented flow analyzer (Auto Analyzer III, Bran Luebbe, Chicago, IL). Soluble PO_4^- was extracted by placing 1 g of charcoal in 25 ml of 0.01 M $CaCl_2$ for 30 min. Extracts were filtered through Whatman #42 filter paper and then analyzed on a segmented flow analyzer using the molybdate method as described by Kuo (1996).

Experiment 1: Charcoal Sorption Potential

We conducted a laboratory incubation study using the soil described above, where Douglas-fir charcoal and extract of *Arctostapholos uva-ursi* were added in a factorial combination yielding four treatments (Charcoal/Extract, Charcoal/No extract, No Charcoal/Extract, and No Charcoal/No extract). Each treatment was replicated five times and consisted of 300 g of soil and placed into mason jars. The treatments receiving charcoal addition received a two percent charcoal amendment (20 g/kg). Charcoal was generated in a muffle furnace by submerging Douglas-fir wood in sand, and heating it at 350 ºC for 2 hours. Charcoal was ground and sieved through a 4.75 mm sieve. *A. uva-ursi* extract was made by extracting 100 g of *A. uva-ursi* leaves in 1 L of deionized water

for 24 hours and filtering this extract through Whatman #42 filters. The total phenol concentration of this extract was 267.5 mg/l. Extract treatments received 25 ml of this extract. No-extract treatments received an equivalent volume of de-ionized water. Soils were homogenized following this addition. This addition brought the soil in each mason jar to a water content of approximately 60% WHC. Mason jars were dark incubated for 14 days upon which a portion of the soil was extracted and analyzed.

This entire experiment was repeated exactly as described above but with glycine added to all mason jars as a source of highly labile organic N to stimulate a more marked N response. Glycine, a simple amino acid that is readily mineralized to NH_4^+, was added to each mason jar at a rate of 75 mg/jar (250 mg/kg of soil). These two experiments will hereafter be referred to as the glycine and no-glycine trials.

Experiment 2: Effects of Bark Charcoal on Plant Growth

This greenhouse experiment consisted of three treatments (Douglas-fir charcoal, ponderosa pine charcoal, and a control) using the sand amended soil described above to evaluate the influence of charcoal source on *K. macrantha*. Each treatment consisted of 20 replicate pots, where each pot received 1.5 kg of soil, and charcoal treatments received a 2% (by mass) charcoal amendment. One percent of this charcoal was homogenized into the soil while the other one percent was evenly distributed on the soil surface. We made charcoal from Douglas-fir and ponderosa pine in the laboratory by burying bark of each species in silica sand and heating to 350 ºC in a muffle furnace for 2 hours. Charcoal was sieved (< 1 mm) using a Whiley mill. Organic horizons (O_i, O_e, and O_a) were added to the surface of each pot to add an additional and substantial mineralizable

pool of plant essential nutrients, and to provide a source of bio-available organic C that may influence soil nutrient transformations. This organic material was randomly collected (as described in Gundale et al. 2005) from a ponderosa pine/Douglas-fir forest that had not been exposed to fire for approximately 80 years, and originated from numerous species, including understory and overstory species, but appeared to be primarily composed of undecomposed ponderosa pine and Douglas-fir litter. The organic material was homogenized and 100 g were added to the surface of each pot. A mixed bed ionic resin capsule (Unibest Inc., Bozeman, MT) was placed in the center of each pot to sorb nutrients throughout the duration of the experiment.

K. macrantha was grown in these pots between October 2004 and March 2005 under ambient light conditions. An average greenhouse temperature of 21 ºC was maintained. *Koeleria macrantha* seeds (Western Native Seeds, Coaldale, CO) were germinated in a separate soil medium and a single seedling was transplanted into each pot. Pots were watered three days a week throughout the duration of the experiment. At the end of the experiment, resin capsules were recovered and soil was rinsed from roots. Plants were oven dried at 65 ºC and above and belowground mass was measured.

Experiment 3: Effect of Wildfire Charcoal on Plant Growth

Charcoal collected from a wildfire site was added to the soil described above at a rate of 0, 0.5, 1, 2, 5, and 10 %, and placed in greenhouse pots seeded with *K. macrantha* to determine whether an increase in soil charcoal content has any influence on the growth of *K. macrantha*. Each treatment (n=10) was established by adding 1.0 kg of charcoal amended soil per pot. The charcoal used in this experiment differed from both previous

experiments because it was collected following a wildfire rather than generated in the laboratory. Large particles (> 5 cm diameter) of charcoal were collected in the spring of 2004 from the Black Mountain Fire (August 2003), Missoula, MT, (DeLuca et al., 2005). It was impossible to decipher the species origin of this charcoal, but it was likely primarily Douglas-fir and ponderosa pine wood and bark char. The charcoal particles were crushed, using a mallet, producing fragments ranging from a diameter of 2 cm to microscopic. No attempt was made to discriminate against any size class in attempt to simulate the range of charcoal particle sizes likely incorporated into the soil under natural conditions. Organic horizon materials (50 g) were collected from a forest stand not exposed to fire for over 80 years and added to the surface of each pot as described earlier. All other experimental conditions were run identically to experiment number two.

Laboratory Analyses

At the end of experiment one, 30 g of soil were extracted with 2 M KCl and analyzed for NH_4^+ and NO_3^- as described above. Amino N was measured on these same extracts using the ninhydrin method (Moore 1968). Soluble phenols were extracted by shaking 30 g of soil for 1 hour with 50 ml of deionized water followed by filtration. Sorbed phenols were extracted by shaking 30 g of soil with 50 percent methanol for 24 hrs followed by filtration. Phenols in these extracts were measured using the Prussian blue method (Stern et al. 1996). Respiration was measured at the end of the incubation by incubating 50 g dry weight equivalent soil in a sealed container with 20 ml 1 M NaOH traps for three days (Zibilske 1994).

Mixed bed ionic resin capsules (Unibest Inc., Bozeman, MT) were used in experiments two and three to determine solution NH_4^+, NO_3^- and PO_4^- throughout the duration of the experiments. Capsules were placed in the center of each pot, directly beneath each plant, and were removed and extracted in 10 ml 2 M KCl three consecutive times. We analyzed NH_4^+, NO_3^-, and PO_4^- from these extracts as described previously.

Statistical Analyses

Data in experiment one meeting assumptions of normality and homoscedasticity were analyzed using two-factor analysis of variance (ANOVA), where extract and charcoal were entered as fixed factors under the general linear model. Variables not meeting these assumptions were analyzed using a Kruskal-Wallis test (K-W test). This analysis tests for differences among treatments but does not evaluate the significance of individual factors or interactions between factors.

Data in experiment two and three were analyzed using one-factor ANOVA's followed by the Student-Newman-Keuls post-hoc procedure. Different letters are used to display post hoc difference. Data not meeting assumption of normality and homoscedasticity were compared using K-W tests, which were not followed by post-hoc procedures. All analyses were conducted using SPSS 12.0 software.

Results and Discussion

Experiment 1: Low Temperature Charcoal Sorption Potential

Both charcoal and litter extract significantly influenced numerous soil chemical variables (Figure 1). In both glycine and no-glycine trials, litter extract negatively influenced

extractable NO_3^- concentrations. The negative influence of *A. uva-ursi* on extractable NO_3^- reported here is consistent with our previous studies in ponderosa pine forest soils (DeLuca et al. 2005) and with studies that have shown that litter from late-successional boreal species, such as the Ericaceous shrub *Empetrum hermaphroditum*, diminishes net nitrification (DeLuca et al. 2002; Berglund et al. 2004). Charcoal had an unexpected negative effect on NO_3^- in the no-glycine trial. In contrast, the addition of charcoal increased NO_3^- concentrations in the glycine trial. These results may be a function of the charcoal we used in this study, which was generated at a low temperature (350 ºC). Charcoal contains a significant concentration of bio-available C and specifically soluble phenols (Table 1) that may have caused net NO_3^- immobilization (Schimel et al. 1996) in the no-glycine trial, where low NH_4^+ concentrations existed (Rice and Tiedje 1989). The NO_3^- immobilization effect did not occur in the glycine trial because NH_4^+ limitations were drastically reduced with glycine addition. Additionally, higher rates of nitrification in the glycine trial likely occurred because this process was not limited by a lack of substrate availability (glycine additions resulted in high NH_4^+ concentrations).

The higher rate of nitrification associated with charcoal in the glycine trial is consistent with the finding reported by DeLuca et al. (2006), which suggests that charcoal may sorb compounds from litter extract and the soil solution that are inhibitory to nitrifying bacteria, or sorb carbon rich molecules that would otherwise stimulate microbial immobilization of N.

A. uva-ursi extract had a strong positive effect on NH_4^+ in both no-glycine and glycine trials because it likely contained some NH_4^+ as well as substrates that are rapidly mineralized to NH_4^+. Charcoal had a strong negative effect on NH_4^+ in both no-glycine

and glycine trials. The mechanisms for this pattern may differ between the two trials. In the no-glycine trial, the most likely explanation for reduced NH_4^+ is that immobilization occurred as a function of N limitations in these soils. In the glycine trial, higher rates of nitrification associated with charcoal likely contributed to lower NH_4^+ concentrations.

Both charcoal and extract significantly influenced concentrations of amino N, that represent a highly labile fraction of organic N that can be rapidly mineralized. Glycine, which is a simple amino N molecule, stimulated rapid rates of N mineralization and resulted in increased amino N concentrations, which suggests that the added glycine was not completely utilized, and that substrate limitations were eliminated during this trial. In both glycine and no-glycine trials, the litter extract resulted in higher concentrations of amino N to soils. The effect of charcoal on amino N, however, differed in glycine and no-glycine trials. In the no-glycine trial, charcoal significantly increased amino N concentrations. This response may have occurred because charcoal sorbed phenolic molecules that otherwise would form insoluble complexes with amino N groups. In contrast, charcoal had a negative effect on amino N in the glycine trial, which is likely the result of charcoal enhancing microbial utilization of glycine.

As expected, *A. uva-ursi* extract significantly increased phenols (soluble and sorbed) in both trials. The addition of charcoal to soil significantly diminished the soluble phenol concentration, while increasing the pool of sorbed phenol. This result is consistent with several studies in the boreal forest that have demonstrated a high capacity of charcoal to adsorb phenolic compounds (Zackrisson et al. 1996; Wardle et al. 1998; DeLuca et al. 2002; Berglund et al. 2004). Solubility of these fractions likely influences the degree to which they are bio-available, and therefore their ability to interfere with N

transformations (Harborne 1997). Interestingly, total phenols (sorbed and soluble) was higher in the charcoal-only treatment of both trials than the control, demonstrating that charcoal itself adds a substantial amount of total phenol to the soil (Table 1). These phenols are likely derived from the components of wood, such as lignin, that are degraded during charcoal formation. It is unclear what effect these phenols have on soil processes, but it is likely they could be utilized as a food source by microbes, stimulating N immobilization.

Soil respiration showed little response to charcoal in glycine or no-glycine trials. In the no-glycine trial, the extract significantly increased soil respiration. Extract and charcoal had no individual effect on soil respiration in the glycine trial; however, the interaction between charcoal and extract showed a significant effect. We speculate that this response may reflect that amines and degradable carbon substrates were better utilized by microbes when phenolic molecules inhibitory to microbes in the same extract were sorbed by charcoal.

These data demonstrate that low temperature charcoal effectively sorbs soluble phenols from *A. uva-ursi* extracts, which in turn stimulates nitrification, provided nitrification is not substrate limited. Our results are consistent with Bergland et al. (2004) and DeLuca et al. (2002) who showed that the effect of charcoal on nitrification only occurred when a labile N source was also present. These studies are also consistent with the Terra Preta phenomenon reported in the Amazonian basin, where charcoal and manure (high labile N concentration) were historically incorporated into the soil (Glaser et al. 2001; 2002). Today these soils maintain the highest fertility in the region, which may in part be a function of the interactive effect of charcoal and manure.

Experiment 2: Effects of Bark Charcoal on Plant Growth

In this experiment, we unexpectedly found that charcoal from both species diminished growth of *K. macrantha* relative to the control, with reduced mass in both aboveground and belowground growth (Table 2). *K. macrantha* growing in pots with Douglas-fir charcoal had a significantly higher root to shoot ratio than the other treatments which appeared to be primarily driven by low aboveground biomass. This data suggests that there is likely no difference in the effect of ponderosa pine and Douglas-fir charcoal on plant species in this ecosystem.

We found that resin sorbed NH_4^+ and NO_3^- were significantly higher in the Douglas-fir charcoal treatment relative to the ponderosa pine charcoal treatment and the control. Resin sorbed PO_4^- was significantly higher in both Douglas-fir and ponderosa pine charcoal treatments than the control. These results may be interpreted in several ways. First, they may indicate higher mineralization and nitrification rates in the presence of charcoal, as suggested by experiment one. If higher mineralization occurred in the presence of charcoal, it is unclear why a corresponding increase in plant growth did not occur. It is possible that some toxic substance was generated during charcoal formation that inhibited root growth of *K. macrantha,* despite a positive effect on nutrient availability (Fritze et al. 1998; Villar et al. 1998). These toxic substances are likely to be more abundant in low temperature charcoals, such as used in this experiment, and may be prone to volatilization at higher temperatures. An additional explanation is that charcoal may have enhanced soil macroporosity, allowing more soil solution to pass through capsules, resulting in misleading resin-sorbed nutrient concentrations.

Experiment 3: Effect of Wildfire Charcoal on Plant Growth

In support of our hypothesis, natural charcoal collected from a wildfire showed a positive effect on growth of *K. macrantha* (Table 3). Both total mass and aboveground mass were significantly higher in pots amended with 5% and 10% charcoal addition than the control. Pots with lower charcoal content (0.5% - 2%) showed an intermediate growth response. No significant shift in allocation to above or belowground structures was detected across the charcoal gradient. As in experiment two, resin sorbed NO_3^-, and PO_4^- decreased as plant growth increased. These results suggest that these measurements do not reflect any direct effect charcoal may have on nutrient cycling, but rather are indicative of the solution nutrient concentration as influenced by plant uptake. No difference in resin sorbed NH_4^+ occurred across the charcoal gradient.

The different response of *K. macrantha* to charcoal in experiments two and three suggest that charcoal produced in a laboratory may be greatly different from charcoal generated during wildfire. Differences in charring conditions may influence the chemical and structural nature of charcoal, and therefore change its influence on soil solution chemistry. One potentially important difference between laboratory and wildfire collected charcoal was the ratio of soluble phenols to NH_4^+ concentration extracted from the charcoals (Table 1). While all charcoal had relatively similar soluble phenol contents, which may stimulate microbial N immobilization, high NH_4^+ concentrations may have offset this immobilization effect when wildfire charcoal was used. Another potentially important difference is the different pH of laboratory charcoal and wildfire charcoal (Table 1). The low pH associated with the lab charcoals may have indirectly diminished

P availability in these treatments. Another difference between the charcoal used in experiment three and experiment two was the range of charcoal particle size used. Experiment three incorporated charcoal ranging from large (1-2 cm) to microscopic fractions. We noted substantial root penetration into large charcoal particles at the end of this greenhouse experiment, which suggests that some resource such as water is more available inside large charcoal particles. It is also possible that grinding charcoal to a smaller size class in some way eliminates it beneficial effects on soil fertility. For instance, grinding may enhance the availability organic carbon because it is very immobile, whereas N ions are significantly more mobile, and thus nutrient immobilization may be more substantial when charcoal is ground.

Conclusion

It is clear that charcoal has the potential to significantly alter soil solution chemistry and growth of *K. macrantha*. Charcoal did not appear to stimulate N cycling in a low nutrient setting, but when glycine was added to soil, charcoal greatly enhanced N mineralization and nitrification. This result may indicate that low temperature charcoal contributes bio-available carbon that causes N immobilization under low nutrient conditions. As hypothesized, charcoal effectively sorbed soluble phenols from litter extracts. This sorption may effectively reduce the inhibitory effect of litter extracts on soil micro-organisms, plants and biogeochemical processes. Low-temperature laboratory-generated charcoal had a negative effect on growth of *K. macrantha* possibly as a result of a toxicity effect caused by some compound formed during low temperature charring or by N immobilization as suggested by the no-glycine soil incubation. In contrast, charcoal

created during a wildfire had a positive effect on growth of *K. macrantha*, suggesting low temperature laboratory charcoal may not adequately represent field collected charcoal. Field collected charcoal may have been generated in a higher oxygen, higher temperature environment and may have been exposed to leaching by rain water and occlusion by soil organic compounds prior to collection. Further investigation is required to evaluate how charcoal formation conditions alter its affect on soil processes and plant growth and how these processes manifest themselves in natural ecosystems.

Acknowledgments

We thank V. Kurth, D. Mackenzie and T. Burgoyne for their assistance in the laboratory and greenhouse. We also acknowledge funding from the NSF and the USDA Joint Fire Sciences Program for this research.

Literature Cited

Berglund LM, DeLuca TH, Zackrisson O (2004) Activated carbon amendments of soil alters nitrification rates in Scots pine forests. Soil Biol Biochem 36: 2067-2073.

Covington WW, Sackett SS (1990) Fire effects on ponderosa pine soils and their management implications. USDA Forest Service RM-GTR-191., Rocky Mountain Forest and Range Experiment Station, Flagstaff, AZ.**:** 105-111.

Covington WW, Sackett SS (1992) Soil mineral nitrogen changes following prescribed burning in ponderosa pine. Forest Ecol and Manag 54: 175-191.

DeBano LF, Eberlein GE, P.H. D (1979) Effects of burning on chaparral Soils: I. Soil Nitrogen. Soil Sci Soc Am J 43: 504-509.

DeLuca TH, MacKenzie MD, Gundale MJ, Holben WE (2006) Wildfire-produced charcoal directly influences nitrogen cycling in forest ecosystems. S Sci Soc Am J 70: 448-453.

DeLuca TH, Nilsson M-C, Zackrisson O (2002) Nitrogen mineralization and phenol accumulation along a fire chronosequence in northern Sweden. Oecologia 133: 206-214.

DeLuca TH, Zouhar KL (2000) Effects of selection harvest and prescribed fire on the soil nitrogen status of ponderosa pine forests. Forest Ecol Manag 138: 263-271.

Diaz-Ravina M, Prieto A, Baath E (1996) Bacterial activity in a forest soil after soil heating and organic amendments measured bythe thymidine and leucine incorporation techniques. Soil Biol Biochem 28: 419-426.

Dunn PH, DeBano LF, Eberlein GE (1979) Effects of Burning on Chaparral Soils: II. Soil Microbes and Nitrogen Mineralization. Soil Sci Soc Am J 43: 509-514.

Fernandez I, Cabaneiro A, Carballas T (1997) Organic matter changes immediately after a wildfire in an Atlantic forest soil and comparison with laboratory soil heating. Soil Biol and Biochem 29: 1-11.

Fritze H, Pennanen T, Kitunen V (1998) Characterization of dissolved organic carbon from burned humus and its effects on microbial activity and community structure. Soil Biol and Biochem 30: 687-693.

Glaser B, Haumaier L, Guggenberger G, Zech W (2001) The 'Terra Preta' phenomenon: a model for sustainable agriculture in the humid tropics. Naturwissenschaften 88: 37-41.

Glaser B, Lehmann J, Zech W (2002) Amerliorating physical and chemical properties of highly weathered soils in the tropics with charcoal - a review. Biol Fertil Soils 35: 219-230.

Gundale MJ, DeLuca TH, Fiedler CE, Ramsey PW, Harrington MG, Gannon JE (2005) Restoration management in a Montana ponderosa pine forest: effects on soil physical, chemical, and biological properties. Forest Ecol Manag 213: 25-38.

Harborne JB (1997). Role of phenolic secondary metabolites in plants and their degradation in nature. Driven by nature: Plant litter quality and decomposition. Cadisch G, Giller K E. Oxon, UK, CAB International.

Hart SC, DeLuca TH, Newman GS, MacKenzie MD, Boyle SI (2005) Post-fire vegetative dynamics as drivers of microbial community structure and function in forest soils. Forest Ecol Manag 220: 166-184.

Kaye JP, Hart SC (1998) Ecological restoration alters nitrogen transformations in a ponderosa pine-bunchgrass ecosystem. Ecol Appli 8: 1052-1060.

Kuo S (1996). Phosphorus. In Sparks D L. Methods of soil analysis. Part 3 - Chemical methods. SSSA Book Series 5. Soil Science Society of America Madison, WI. pp. 896-919.

Mann CC (2002) The real dirt on rainforest fertility. Science 297: 920-923.

Monleon VJ, Cromack K, Landsberg JD (1997) Short- and long-term effects of prescribed underburning on nitrogen availability in ponderosa pine stands in central Oregon. Can J For Res 27: 369-378.

Moore S (1968) Amino acid analysis: aqueous dimethyl sulfide as a solvent for the ninhydrin reagent. J Biol Chem 243: 6281-6283.

Mulvaney RS (1996). Nitrogen - inorganic forms. In Sparks D L. Methods of soil analysis. Part 3 - Chemical methods. Sparks D L. Soil Science Society of America Madison, WI. pp. 1123-1184.

Neary DG, Klopatek CC, DeBano LF, Ffolliott PF (1999) Fire effects on belowground sustainability: a review and synthesis. Forest Ecol Manag 122: 51-71.

Pietikainen J, Hiukka R, Fritze H (2000a) Does short-term heating of forest humus change its properties as a substrate for microbes? Soil Biol Biochem 32: 277-288.

Pietikainen J, Kiikkila O, Fritze H (2000b) Charcoal as a habitat for microbes and its effect on the microbial community of the underlying humus. Oikos 89: 231-242.

Rice C, Tiedje J (1989) Regulation of nitrate assimilation by ammonium in soils and in isolated soil microorganisms. Soil Biol Biochem 21: 597-602.

Rice EL, Pancholy SK (1972) Inhibition of nitrification by climax ecosystems. Am J Bot 59: 1033-1040.

Schimel JP, VanCleve K, Cates RG, Clausen TP, Reichardt PB (1996) Effects of balsam poplar (Populus balsamifera) tannins and low molecular weight phenolics on microbial activity in taiga floodplain soil: Implications for changes in N cycling during succession. Can J Bot 74: 84-90.

Stern JL, Hagerman AE, Steinberg PD, Winter FC, Estes JA (1996) A new assay for quantifying brown algal phlorotannins and comparisons to previous methods. J Chem Ecol 22: 1273-1293.

Villar MC, González-Prieto SJ, Carballas T (1998) Evaluation of three organic wastes for reclaiming burnt soils: Improvement in the recovery of vegetation cover and soil fertility in pot experiments. Biol Fertil Soils 26: 122-129.

Wardle DA, Zackrisson O, Nilsson M-C (1998) The charcoal effect in boreal forests: mechanisms and ecological consequences. Oecologia 115: 419-426.

White CS (1991) The role of monoterpenes in soil in soil nitrogen cycling processes in ponderosa pine: results from laboratory bioassays and field studies. Biogeochem 12: 43-68.

White CS (1994) Monoterpenes: Their effects on ecosystem nutrient cycling. Journal of Chem Ecol 20: 1381-1406.

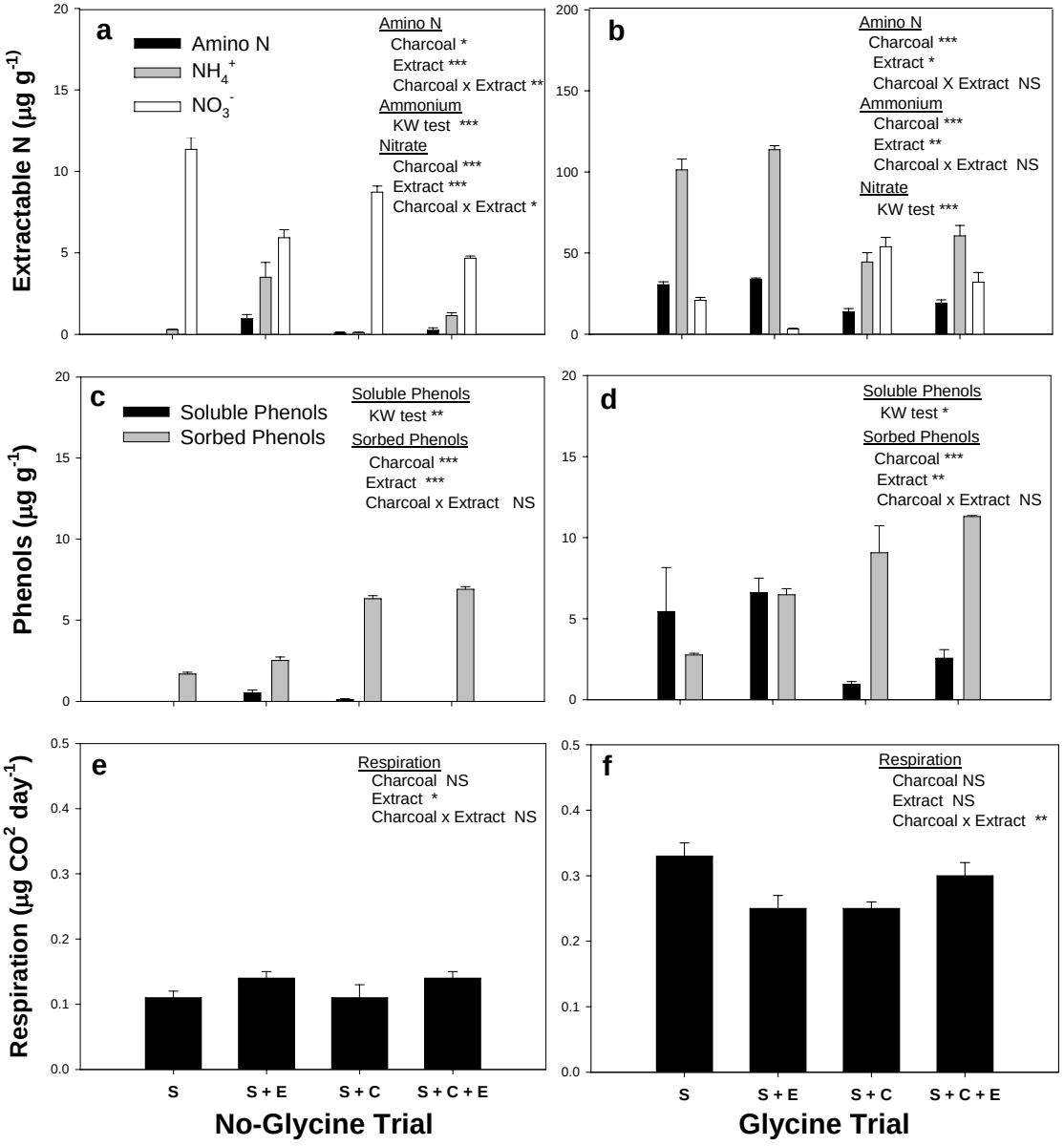
Willis RB, Gentry CE (1987) Automated method for determining nitrate and nitrite in water and soil extracts. Comm Soil Sci Plant Anal 18: 625-636.

Willis RB, Schwab GJ, Gentry CE (1993) Elimination of interferences in the colormetric analysis of ammonium in water and soil extracts. Comm Soil Sci Plant Anal 24: 1009-1019.

Zackrisson O, Nilsson M-C, Wardle DA (1996) Key ecological function of charcoal from wildfire in the Boreal forest. Oikos 77: 10-19.

Zibilske LM (1994). Carbon Mineralization. In Weaver R W, Angle S and Bottomly P. Methods of soil analysis. Part 2: Microbiological and biochemical properties. Soil Science Society America, Madison, WI. pp 835-863.

Figure 1: Extractable amino N, NH_4^+, and NO_3^- (mean (SE)) without (a) and with (b) glycine addition; soluble (water extracted) and sorbed phenols (methanol extracted) (mean (SE)) without (c) and with (d) glycine addition; and basal soil respiration (mean (SE)) without (e) and with (f) glycine addition, from a 14-d soil incubation experiment where soils were amended with a factorial combination of charcoal and extracts from *Arctostaphylos uva-ursi* leaves (S = soil-only; S + E = soil plus extract; S + C = soil plus charcoal; S + C + E = soil plus charcoal plus extract). Data were analyzed with a two-factor ANOVA, where significance was tested for Charcoal, Extract and Charcoal*Extract interaction. Data that did not meet parametric assumptions of normality or homoscedasticity, were analyzed using a Kruskal-Wallis (KW) test. Star notation in each figure represents statistical significance (*p*-value: ns > 0.1, *< 0.05, ** < 0.01, *** < 0.001).


a
Amino N
NH4+
NO3-
Amino N
Charcoal *
Extract ***
Charcoal x Extract **
Ammonium
KW test ***
Nitrate
Charcoal ***
Extract ***
Charcoal x Extract *
b
Amino N
Charcoal ***
Extract *
Charcoal X Extract NS
Ammonium
Charcoal ***
Extract **
Charcoal x Extract NS
Nitrate
KW test ***
Extractable N (µg g-1)
c
Soluble Phenols
Sorbed Phenols
Soluble Phenols
KW test **
Sorbed Phenols
Charcoal ***
Extract ***
Charcoal x Extract NS
d
Soluble Phenols
KW test *
Sorbed Phenols
Charcoal ***
Extract **
Charcoal x Extract NS
Phenols (µg g-1)
e
Respiration
Charcoal NS
Extract *
Charcoal x Extract NS
f
Respiration
Charcoal NS
Extract NS
Charcoal x Extract **
Respiration (µg CO2 day-1)
S
S + E
S + C
S + C + E
No-Glycine Trial
Glycine Trial

1

Table 1: Chemical properties of four charcoal types (df = Douglas-fir; pp = ponderosa pine; wildfire = wildfire collected) used in the laboratory and greenhouse experiments.

	df wood	**df bark**	**pp bark**	**wildfire**
pH	4.15	4.18	4.81	5.74
EC ($\mu S\ g^{-1}$)	20.5	24.2	111.6	141.5
CEC ($cmol_c\ kg^{-1}$)	20.66	19.42	34.48	29.35
Density ($g\ cm^{-3}$)	0.30	0.08	0.21	0.29
Total C (%)	72.9	72.4	71.5	82.3
Soluble Phenols ($\mu g\ g^{-1}$)	34.9	30.7	43.4	48.2
Total Phenols ($\mu g\ g^{-1}$)	441.7	148.1	810.4	393.8
PO_4^- ($\mu g\ g^{-1}$)	0.94	0.84	2.46	0.95
NH_4^+ ($\mu g\ g^{-1}$)	7.12	9.0	5.6	38.3
NO_3^- ($\mu g\ g^{-1}$)	0.12	0.3	0.0	4.4

Table 2: Plant mass and resin sorbed nutrients (Mean ±SE, n=20) from a greenhouse experiment where soil was amended with 2% charcoal made from Douglas-fir (df) and ponderosa pine (pp) bark at 350 ºC. Letters indicate differences using the Student-Newman-Keuls post-hoc procedure.

	df charcoal		**pp charcoal**		**no charcoal**		***p*-value[1]**
Total Mass (g)	**a**1.6	(0.2)	**a**1.9	(0.1)	**b**2.5	(1.0)	***
Root Mass (g)	**a**0.8	(0.1)	**a**0.9	(0.1)	**b**1.2	(0.1)	*
Aboveground Mass (g)	**a**0.7	(0.1)	**b**1.0	(0.1)	**c**1.3	(0.1)	***
Root:Shoot	**a**1.1	(0.1)	**b**0.9	(0.1)	**b**0.9	(0.1)	**
NH_4^+ µg resin capsule^{-1}	**a**4.6	(0.9)	**b**1.4	(0.6)	**b**2.2	(0.8)	*
NO_3^- µg resin capsule^{-1}	**a**1770.8	(286.2)	**b**935.8	(241.9)	**b**581.5	(211.8)	**
PO_4^- µg resin capsule^{-1}	**a**5.1	(1.4)	**a**5.6	(0.9)	**b**0.5	(1.2)	*

[1]*p*-value: *, $p<0.05$; **, $p<0.01$, ***, $p<0.001$

Table 3: Plant mass and resin sorbed nutrients (Mean ±SE, n=10) from a greenhouse experiment where soil was amended with 0, 0.5, 1, 2, 5, and 10% charcoal collected from a wildfire. Letters indicate differences using the Student-Newman-Keuls post-hoc procedure.

	Percent Charcoal												
	0 %		**0.5%**		**1%**		**2%**		**5%**		**10%**		[1]*p*
Total mass (g)	**a**0.5	(0.2)	**ab**1.0	(0.3)	**ab**1.1	(0.2)	**ab**1.1	(0.2)	**b**1.3	(0.1)	**b**1.4	(0.1)	*
Root mass (g)	0.3	(0.2)	0.6	(0.2)	0.7	(0.1)	0.7	(0.1)	0.8	(0.1)	0.8	(0.1)	ns
Aboveground mass (g)	**a**0.2	(0.1)	**ab**0.4	(0.1)	**ab**0.4	(0.1)	**ab**0.4	(0.1)	**b**0.5	(0.1)	**b**0.6	(0.1)	**
Root:Shoot	1.5	(0.2)	1.5	(0.3)	1.8	(0.2)	1.7	(0.3)	1.6	(0.2)	1.3	(0.1)	ns
[3] **NH_4^+ µg resin capsule^{-1}**	55.6	(4.0)	49.8	(4.6)	36.9	(6.0)	42.7	(2.2)	43.0	(1.3)	44.4	(2.9)	[2]*
[3] **NO_3^- µg resin capsule^{-1}**	**a**1539.8	(463.4)	**b**947.9	(128.4)	**bc**552.3	(116.3)	**bc**556.1	(93.4)	**bc**561.8	(278.7)	**c**248.6	(29.8)	***
[3] **PO_4^- µg resin capsule^{-1}**	**a**10.1	(1.4)	**a**8.8	(2.1)	**ab**5.8	(1.2)	**ab**6.5	(1.7)	**bc**1.7	(1.0)	**c**0.0	(1.2)	***

[1]All *p*-values are for one-way ANOVA, unless otherwise noted. *p*-value: ns, $p>0.05$; *, $p<0.05$; **, $p<0.01$, ***, $p<0.001$

[2] Kruskal-Wallis test *p*-value

[3]µg resin