FINAL REPORT

Microbiological Sampling Report

for

National Oceanic & Atmospheric Administration

Small Abatement Conducted in the Room 15327 of Building SSMC-3 in December 1999 and January 2000

Interagency Agreement #: D8H00CO31200

Task: 9903

January 22, 2000

Prepared by US Public Health Service Division of Federal Occupational Health Bethesda Central Office

Executive Summary

At the request of the National Oceanic & Atmospheric Administration (NOAA), Federal Occupational Health (FOH) conducted a small-scale drywall removal project in room 15327 of Building SSMC-3, located at 1315 East-West Highway, Silver Spring, Maryland. The visibly contaminated drywall was removed according to FOH's protocol (Attachment A) in the evening of December 10, 1999. The room was then thoroughly cleaned by wet-wiping and High Efficiency Particulate Air (HEPA) vacuuming. Air (both Andersen^â and Zefon^â), swab, contact plate, and vacuum dust samples were collected from this room and an indoor reference room 15212. Air samples were also collected from outdoors. Initially, samplings were conducted at three time periods: before (refer as Pre-), immediately after (refer as Post-I), and one-week after (refer as Post-II) removal of the contaminated drywall and thorough cleaning of this room. Based on results from the Post-II sampling, a second thorough cleaning was conducted in room 15327 during the weekend of January 21 – 23, 2000. Another full-scale post-cleaning sampling was conducted on January 24, 2000 and February 1, 2000 (refer as Post-III).

Some findings are as follows:

• *Stachybotrys chartarum* and *Penicillium* were the predominant fungi recovered from the interior side of removed drywall, perforated paper and plastic surfaces on the window frame, exterior paper wrapping of fiberglass insulation materials, and surfaces of concrete pre-cast in the wall cavity.

• Indoor fungal levels were lower than those of outdoors and fungi detected indoors were similar to those detected outdoors.

• *Stachybotrys chartarum* was present at low levels in the air of room 15327 during Post-I sampling and this fungus was not detected from Post-II and Post-III samplings.

· In general, fungal burden on vertical surfaces was lower than that of horizontal surfaces.

• Fungal genera recovered from horizontal and vertical surfaces and carpet and furniture dust were similar to those recovered from outdoor air samples.

• *Cladosporium* was the predominant fungal genus recovered from surfaces in reference room 15212. However, *Penicillium* was the predominant one recovered from room 15327.

• *Stachybotrys chartarum* was detected once, from one contact plate sample collected from the vertical surface of room 15327 on December 21, 1999, one week after drywall removal and cleaning. This fungus was not detected from samples collected after second cleaning.

• Fungal levels on horizontal and vertical surfaces were very low after second cleaning of room 15327.

 \cdot Very low fungal burden was detected from wipe samples collected from surfaces of supply diffusers and return troughers in light fixture.

• Fungal burden on dust samples collected from carpet and furniture were at $10^3 - 10^5$ CFU/g levels.

· Consistent presence of *Stachybotrys chartarum* in carpet dust of room 15327 was detected throughout this project.

• Stachybotrys chartarum was detected from furniture dust in room 15327 after abatement and cleaning, but was not detected after second cleaning.

Overall, fungal burden on various surfaces in room 15327 was low after the second cleaning of this room. Airborne fungal levels and total fungal spores in the air in room 15327 were lower than those of outdoors and were compatible to those of the reference room 15212. Fungal levels in composite furniture dust decreased from 10^4 to 10^3 CFU/g of fine dust after second cleaning of room 15327. However, fungal levels in carpet dust remained at the 10^4 CFU/g of fine dust level and consistent presence of *Stachybotrys chartarum* was detected.

Recommendations were as follows:

• HEPA vacuum and wet-wipe of the open edge of bookshelf in room 15212.

• Revise the small-scale drywall removal protocol to prevent spores release to the carpet during drywall removal (Attachment D).

• Use HEPA vacuum cleaner as a routine housekeeping of the carpeting in the building.

 \cdot Implement an emergency water intrusion protocol for this building to adequately manage the unexpected water intrusion in order to prevent any fungal proliferation.

INTRODUCTION

At the request of the National Oceanic & Atmospheric Administration (NOAA), Federal Occupational Health (FOH) conducted a small-scale drywall removal project in room 15327 of Building SSMC-3, located at 1315 East-West Highway, Silver Spring, Maryland. The visibly contaminated drywall was removed according to FOH's protocol (Attachment A) in

the evening of December 10, 1999. The room was then thoroughly cleaned by wet-wiping and High Efficiency Particulate Air (HEPA) vacuuming. Air (both Andersen^â and Zefon^â), swab, contact plate, and vacuum dust samples were collected from this room and an indoor reference room 15212. Air samples were also collected from outdoors. Initially, samplings were conducted at three time periods: before (refer as Pre-), immediately after (refer as Post-I), and one-week after (refer as Post-II) removal of the contaminated drywall and thorough cleaning of this room.

Based on results from the Post-II sampling, a second thorough cleaning was conducted in room 15327 during the weekend of January 21 - 23, 2000. Another full-scale post-cleaning sampling was conducted on January 24, 2000 and February 1, 2000 (refer as Post-III).

EVALUATION METHODOLOGY

Pre-abatement Sampling on December 9, 1999

Various types of samples were collected from room 15327 and a reference room 15212 which was selected by NOAA based on no history of water damage and no visible fungal growth on the window sill areas. Sampling was conducted on December 9, 1999, before the initiation of drywall removal. Results from this sampling were used as baseline information.

In each room, two air sampling locations were selected, one by the window area and the other in the center of the room. Outdoor air samples were collected near the entrance of the building. Two types of air samples were collected: (1) culturable method using Andersen^â N-6 samplers at a flow rate of 28.3 L/min, and (2) non-culturable method using Zefon^â Air-O-Cell cassettes at a flow rate of 15 L/min. Indoor Andersen^â air samples were collected for 3 minutes and outdoor samples were collected for both one and three minutes. Two percent (2 %) malt extract agar (MEA) and cellulose Czapek agar (CCA) was used to recover general fungi and cellulose-loving fungi, respectively. Non-culturable air samples were collected for both five and ten minutes. Temperature and relative humidity measurements were collected from each air sampling location by a battery operated, direct readout Hygroskop^â meter.

To determine fungal burden on horizontal and vertical surfaces of these rooms, eight contact plate samples were collected from each room. Four samples were collected from randomly selected horizontal surfaces and four from the randomly selected vertical surfaces. Sampling was conducted by pressing the MEA-filled Rodac^â plate against the surface of interest for five seconds.

Swab samples were collected from surfaces of each supply diffusers and return troughers in each room. They were collected by wiping a known area of surface with a sterile cotton swab (Culturette^â) wetted with holding media. Approximately 5 in² area was wiped for return trougher and 4 in² for supply diffusers. The swab was then placed directly into its holder. Each holder was labeled with an identifiable number. Nine wipe samples were collected from room 15327 and four samples from room 15212.

Dust accumulated on carpeting and chairs and fabric system furniture were collected with a High Efficiency Particulate Air (HEPA) vacuum attached with a special "sock" device. For each carpet sample, a 3-ft by 3-ft area was vacuumed for

at least five minutes. Dust on surfaces of system furniture and chairs were vacuumed, with a total area of 9 ft^2 , and composite as one sample. One carpet sample and one composite furniture sample were collected from each room.

All samples collected were sent to FOH's Environmental Microbiology Laboratory (EML) in Philadelphia, Pennsylvania for analysis.

Contaminated Drywall Removal on December 10, 1999

Personnel from Facility Management and FOH observed a representative from the Foulger Pratt performed the small-scale removal of drywall near the windowsill of room 15327 on Friday, December 10, 1999. This small-scale removal of contaminated drywall began at about 7:00 p.m. after shut down of the ventilation system. The plastic sheeting and duct tape, which covered the visibly fungal contaminated area, were peeled off. Representative from the Foulger Pratt used a utility knife to cut a drywall of about 1-ft by 1-ft area which included visibly fungal growth areas (Exhibit 1). The black fungal growth was observed about 6 inches above the base of the window frame (Exhibit 1). Fungal growth also penetrated into the plastic layer on the window frame. Visible fungal growth was also detected from the paper wrapping outside of the insulation materials in the wall cavity (Exhibit 1). With the aids of a flashlight and a hand-held mirror, the interior surfaces of the drywall immediately adjacent to the cut-off areas were visually inspected (Exhibit 2). Due to detection of black fungal growth on the interior surfaces of the drywall (from top) was cut. Visible fungal growth was not detected from any other areas inspected. Total area of the removed drywall was about 14" x 16". Then, the drywall cut-off and surrounding areas were cleaned with a HEPA vacuum cleaner. The representative from Foulger Pratt patched the open drywall area with a green drywall and screwed it in place (Exhibit 3). The room was then thoroughly cleaned according to FOH's cleaning protocol (Attachment A) during that weekend.

Two wipe samples were collected from the concrete pre-cast in the wall cavity. Two wipe samples were collected on plastic surfaces behind removed drywall. Two tape-lift samples were collected for microscopic examination: one from the plastic surface and the other from the exterior paper wrapping of fiberglass insulation materials. The removed drywall with paper and plastic materials on the window frame, and a small piece of exterior paper wrapping, the wipes, and tape-lift samples were expressed mailed to FOH's EML in Philadelphia, PA.

Post-abatement Microbiological Samplings on December 21, 1999

The exact same sampling was conducted on Monday, December 13, 1999, after this small-scale drywall removal and cleaning. Due to delay receipt of wipe samples at the EML, wipe samples on diffuser surfaces were retaken on December 15, 1999. Another full-scale sampling was conducted on December 21, 1999, approximately one week after drywall removal and cleaning.

Samplings Conducted after Second Cleaning

Another full-scale microbiological sampling was conducted on January 24, 2000 after the second thorough cleaning of this room during the weekend of January 21-23, 2000. Wipe samples were re-collected on February 1, 2000 due to delay receipt of these samples at the EML.

Laboratory Procedures

Upon receipt, all Andersen^â air and contact plate samples were incubated in a 25°C incubator. Sub-samples were collected from the removed drywall and the paper-wrapping samples. Three sub-samples of 1-inch by 1-inch area were cut from the

interior surface of the removed drywall sample. Three sub-samples, each of 1 square inch, were cut from the paper wrapping for analysis. Six wipe samples were also collected from the removed drywall and perforated paper and plastic materials on the window, by wiping one square inch of black fungal growth area with Culturettes^â.

Each swab sample was suspended in sterile distilled water, diluted serially, and inoculated onto agar plates. Both MEA and CCA were used for retrieving fungi. At least three dilution series were used for each sample. Each vacuum dust sample was sieved through a 250 mm sieve. The fine dust (< 250 mm) retrieved was then weighed and followed the dilution plating for fungal analysis. All bulk samples were weighed and followed the aforementioned dilution plating.

All plates were incubated in a 25°C incubator. They were examined every other day for up to 10 days to ensure the full recovery of fungi. Fungal identification was based on colony morphology, spores and conidia formation. Total fungal colonies formed on each MEA plate and *Stachybotrys chartarum* on CCA plates were counted and recorded. Fungal levels in samples were presented as colony forming units (CFUs) per measuring unit. For example, CFU/m³ for Andersen^â air samples, CFU/in² for wipe samples, CFU/plate for contact plate samples, CFU/g for bulk, and CFU/g of fine dust for dust samples.

All Zefon^â cassettes and tape-lift samples were analyzed by the Environmental Microbiology Laboratory in Escondido, California for direct microscopic examination. Fungal spores were identified and their airborne levels were presented as spores/m³. Qualitative information was provided for tape-lift samples.

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RESULTS AND DISCUSSION

Temperature and Relative Humidity

Temperature and relative humidity measurements are presented in Table 1. In general, outdoors had lower temperature and higher relative humidity readings. Indoor temperature and relative humidity readings ranged from 71.4°F to 75.1°F, and 16.6% - 25.1%, respectively.

Table 1. Temperature (Temp.) and relative humidity (Rh) readings in outdoors and rooms 15212 and 15327 of SSMC-3, at different dates.

Locations	152	12	153	27	Outd	loors
Parameters	Temp.	Rh	Temp.	Rh	Temp.	Rh
Dates	(°F)	(%)	(°F)	(%)	(°F)	(%)
12/9/1999	74.4	25.1	73.3	24.7	52.0	49.4
12/13/1999	72.4	24.9	73.4	23.2	49.8	53.8
12/21/1999	72.1	31.8	NA	NA	44.5	52.6
1/24/2000	71.4	18.2	75.1	16.6	NA*	NA

* Not available.

Microbiological Results

All laboratory analytical reports from FOH's EML are presented in Attachment B. Results from swab and bulk samples collected from the removed drywall and paper wrapping are presented in laboratory reports #NOAA-00-12R and #NOAA-00-14R. The laboratory report #NOAA-00-11R-A, -B, and -C contain results of air and contact plate, surface wipe, and vacuum dust samples collected on December 9, 1999 (Pre-abatement). Similarly, reports #NOAA-00-13R and #NOAA-00-15R contained results, respectively, from post-abatement sampling conducted immediately (Post-I) and one-week after (Post-II) the abatement and cleaning. Results from air, contact plate, and dust samples collected on January 24, 2000 are presented in a laboratory report #NOAA-00-21R. Results from wipe samples collected on February 1, 2000 are presented in a report #NOAA-00-23R.

Results from microscopic examination of Zefon^â cassettes and tape-lift samples are presented in Attachment C.

The Extent of Contamination

Stachybotrys chartarum and Penicillium were the predominant fungi recovered from the removed drywall and paper wrapping (Table 2). Total fungal levels on these samples were at $10^6 - 10^7$ CFU/g. All wipe and bulk samples showed presence of Stachybotrys chartarum on CCA plates. Wipe samples collected from the interior side of the removed drywall showed a fungal level of 10^6 CFU/in². Fungal levels on visibly fungal contaminated surfaces behind the drywall were $10^5 - 10^6$ CFU/in². Stachybotrys chartarum were observed on tape-lift samples as the predominant fungus, followed by colorless Penicillium/Aspergillus group. These results confirm that Stachybotrys chartarum and Penicillium/Aspergillus dominated these samples.

Samples collected from concrete pre-cast also showed predominance of *Penicillium* and presence of *Stachybotrys chartarum*. Two samples were collected from plastic surfaces on the window frame behind the removed drywall: one on visible growth area and the other on surfaces of no visible growth. *Penicillium* and *Stachybotrys chartarum* were detected from both samples. The sample collected from the visible growth area had a fungal level at least 1,000 times higher than the control one (report #NOAA-00-12R, samples #W03 and W04).

Table 2. Mean fungal levels of bulk and wipe samples collected from room 15327 of SSMC-3, on December 10, 1999.

Sample Types/Descriptions	Penicillium	SC* levels on	Total levels on	Presence of
	levels on MEA	MEA	MEA	SC on CCA
Bulk Samples Removed drywall (CFU/g)	10,720,010	1,235,613	11,955,622	+**
Paper wrapping outside of fiberglass				
insulation materials (CFU/g)	881,766	874,250	1,756,015	+
Wipe Samples				
Removed drywall interior surfaces (CFU/in ²)	2,990,000	270,000	3,260,000	+

Perforated paper surfaces behind the removed drywall (CFU/in ²)	2,000	5,092,000	5,094,000	+
Perforated plastic surfaces behind the removed drywall (CFU/in ²)	6,000	754,000	760,000	+
Concrete pre-cast (CFU/in ²)	5,450	205	5,655	+

* SC: Stachybotrys chartarum was detected.

** SC was detected on CCA plates of these samples.

Air Samples

Andersen Results

Mean outdoor airborne fungal levels were much higher than those of indoors (Table 3). *Cladosporium, Penicillium,* and Basidiomycetes dominated outdoor fungal flora. Fungi detected indoors were *Cladosporium, Aspergillus,* and Basidiomycetes.

Stachybotrys chartarum was detected on CCA plates, from one of eight outdoor samples (sample #OM2-120999 in report #NOAA-00-11R-A), with a level of 71 CFU/m³. This fungus was also detected on CCA plates, from both samples collected in room 15327 immediately after (Post-I) abatement and cleaning (samples #15327M1, M2-121399 in report #NOAA-00-13R-A), both at 12 CFU/m³. But, *Stachybotrys chartarum* was not detected during Post-II and Post-III samplings.

Zefon Results

Mean outdoor fungal levels were higher than those of indoors (Table 3). Fungal spores detected indoors were similar to those of outdoors. Basidiospores and Ascospores dominated outdoor fungal flora followed by *Cladosporium*, and *Penicillium/Aspergillus* types. Indoor fungal flora were dominated by *Cladosporium*, and *Penicillium/Aspergillus* types.

Stachybotrys chartarum was detected from one sample collected in room 15327 immediately after (Post-I) abatement and cleaning (samples #90749, in Attachment B), at 7 spores/m³. *Stachybotrys chartarum* was not detected from Post-II and Post-III samplings.

Results from both Andersen^â and Zefon^â air samplings showed similar trends: (1) indoor fungal levels were lower than those of outdoors, (2) fungi detected indoors were similar to those detected outdoors, and (3) *Stachybotrys chartarum* was present at low levels in the air of room 15327 during Post-I sampling and this fungus was not detected from Post-II and Post-III samplings.

Table 3.Mean airborne fungal levels in indoor and outdoors, collected by Andersen^â N-6 and Zefon^â samplers at

different periods of time.

	Andersen ^â N-6				Zefonâ			
	(CFU/m ³)					(Spc	ores/m ³)	
Room	Pre-*	Post-I*	Post-II*	Post-III*	Pre-	Post-I	Post-II	Post-III
15327	6	24 (+@)	47	< 12#	74	68 (+)	51	7#
15212	42	30	36	< 12#	134	63	37	< 7#
Outdoors	377	335	684	94	2,283	3,237	5,710	27
	(+)							

* Pre-: Pre-abatement sampling on December 9, 1999.

Post-I: Post-abatement sampling on December 13, 1999.

Post-II: Post-abatement sampling on December 21, 1999.

Post-III: Sampling conducted on January 24, 2000, after second cleaning.

[@] Stachybotrys chartarum was detected.

[#] Only one indoor sample was collected from each room for Post-III sampling.

Wipe Samples

Most (49 out of 52) samples collected from surfaces of supply diffusers and return troughers in light fixtures were below the detection limits of 2 CFU/in². Fungal levels of the three samples, where fungi were detected, ranged from 2 CFU/in² to 10 CFU/in². Fungi detected were yeast, *Penicillium*, and *Chaetomium*. *Stachybotrys chartarum* was not detected from any samples.

Contact Plate Samples

In general, higher fungal levels were detected from the horizontal surfaces than vertical surfaces (Table 4). In each room, on each horizontal or vertical surface, mean fungal level differences were not detected from different sampling periods (p > 0.05) (Table 4).

Table 4. Mean fungal levels (CFU/plate) on horizontal and vertical surfaces of rooms 15327 and 15212 by contact plate sampling collected at different period of time.

Room	Surfaces	Pre-*	Post-I*	Post-II*	Post-III*
15327	Vertical	$1.0 \pm 0.7^{\#}$	0.0 ± 0.0	3.0 ± 2.0	0.0 ± 0.0
15212	Vertical	1.5 <u>+</u> 0.9	1.0 ± 0.7	1.0 ± 0.4	2.5 <u>+</u> 1.7
15327	Horizontal	9.3 <u>+</u> 3.8	25.3 <u>+</u> 19.0	7.8 <u>+</u> 5.3	0.5 ± 0.5
15212	Horizontal	6.5 <u>+</u> 2.7	10.3 ± 7.0	12.3 <u>+</u> 8.6	19.8 <u>+</u> 17.1

* Pre-: Pre-abatement sampling on December 9, 1999.

Post-I: Post-abatement sampling on December 13, 1999.

Post-II: Post-abatement sampling on December 21, 1999.

Post-III: Sampling conducted on January 24, 2000, after second cleaning.

[#] Mean \pm standard error (sample number = 4).

In reference room 15212, fungal levels ranged from below the detection limits (BDL) of 1 CFU/plate to 71 CFU/plate. Fungal genera detected in a descending order, were *Cladosporium, Alternaria, Chaetomium,* Basidiomycetes, *Penicillium,* and *Aureobasidium.* The consistently higher fungal levels (13 - 71 CFU/plate) were detected from open edge of bookshelf with *Penicillium* and *Chaetomium* as dominant fungi.

Fungal levels in room 15327 ranged from BDL of 1 CFU/plate to 82 CFU/plate. *Penicillium* followed by *Alternaria* and *Cladosporium*, were the predominant fungal genera detected in this room. The highest fungal level (82 CFU/plate), with predominant *Penicillium* (81 CFU/plate), was detected from a sample collected from horizontal surfaces in room 15327 immediately after cleaning (Post-I) (sample #15327CP8-121399 in report #NOAA-00-13R-A). One of four samples collected after one-week of cleaning (Post-II) also showed predominant *Penicillium* (23 CFU/plate) (sample #15327CP5-122199 in report #NOAA-00-15R-A). Very low fungal levels were detected from samples collected from this room after the second cleaning (Table 4). *Stachybotrys chartarum* (4 CFU/plate) was detected once from one sample collected on vertical surfaces of room 15327 on Post-II sampling (sample #15327CP4-122199 in report #NOAA-00-15R-A). This fungus was not detected during Post-III sampling after second cleaning.

Vacuum Dust Samples

Fungal analysis was not performed on carpet dust sample collected from room 15212 on December 21, 1999 due to insufficient fine dust collected. Fungal levels in the fine dust of carpet and furniture ranged from 10³ CFU/g to 10⁵ CFU/g of fine dust (Table 5). Fungi detected were *Cladosporium, Alternaria, Aureobasidium, Epicoccum, Paecilomyces,* and *Penicillium.*

Fungal levels in carpet dust of the reference room 15212 were at $10^3 - 10^4$ CFU/g of fine dust levels (Table 5). *Cladosporium* was the predominant fungal genus detected from carpet dust in this room. Dust collected from carpet in room 15327 had fungal levels consistently at 10^4 CFU/g of fine dust levels with *Penicillium* as the predominant fungal genus. *Stachybotrys chartarum* was consistently detected from carpet dust samples collected from room 15327 regardless of sampling time. These results showed carpet in room 15327 harbor fungi such as *Penicillium* and *Stachybotrys chartarum*. The consistent presence of these fungi in carpet dust samples in room 15327 may have resulted from (1) continual supply of these fungal spores in this room, and (2) repeated HEPA vacuuming of these carpet surfaces did not effectively remove the residual spores.

Fungal levels in furniture dust ranged from 10^3 CFU/g to 10^5 CFU/g of fine dust (Table 5). *Cladosporium* was the predominant fungal genus recovered from these samples. *Stachybotrys chartarum* was not detected from samples collected from the reference room 15212. This fungus was detected from room 15327 during the Post-I and Post-II samplings.

Table 5.Fungal levels (CFU/g of fine dust) in fine dust collected from carpet and furniture of rooms 15327 and 15212

by vacuum dust sampling, collected at different period of time.

Room	Surfaces	Pre-*	Post-I*	Post-II*	Post-III*
15327	Carpet	11,600 (+**)	19,200 (+)	13,200 (+)	19,294 (+)
15212	Carpet	5,200 (-)	16,000 (-)	NA [#]	3,168 (-)
15327	Furniture	13,056 (-)	29,057 (+)	18,919 (+)	6,536 (-)
15212	Furniture	65,753 (-)	15,000 (-)	112,000 (-)	11,707 (-)

* Pre-: Pre-abatement sampling on December 9, 1999.

Post-I: Post-abatement sampling on December 13, 1999.

Post-II: Post-abatement sampling on December 21, 1999.

Post-III: Sampling on January 24, 2000, after second cleaning.

** +: *Stachybotrys chartarum* was detected on CCA or MEA plates.

-: Stachybotrys chartarum was not detected on CCA and MEA plates.

[#] Data not available.

FINDINGS

• *Stachybotrys chartarum* and *Penicillium* were the predominant fungi recovered from the interior side of removed drywall, perforated paper and plastic surfaces on the window frame, exterior paper wrapping of fiberglass insulation materials, and surfaces of concrete pre-cast in the wall cavity.

 \cdot Indoor fungal levels were lower than those of outdoors and fungi detected indoors were similar to those detected outdoors.

• *Stachybotrys chartarum* was present at low levels in the air of room 15327 during Post-I sampling and this fungus was not detected from Post-II and Post-III samplings.

· In general, fungal burden on vertical surfaces was lower than that of horizontal surfaces.

• Fungal genera recovered from horizontal and vertical surfaces and carpet and furniture dust were similar to those recovered from outdoor air samples.

• *Cladosporium* was the predominant fungal genus recovered from surfaces in reference room 15212. However, *Penicillium* was the predominant one recovered from room 15327.

• *Stachybotrys chartarum* was detected once, from one contact plate sample collected from the vertical surface of room 15327 on December 21, 1999, one week after removal and cleaning. This fungus was not detected from samples collected after second cleaning.

• Fungal levels on horizontal and vertical surfaces were very low after second cleaning of room 15327.

• Very low fungal burden was detected from wipe samples collected from surfaces of supply diffusers and return troughers in light fixture.

Fungal burden on dust samples collected from carpet and furniture were at $10^3 - 10^5$ CFU/g levels.

Consistent presence of *Stachybotrys chartarum* in carpet dust of room 15327 was detected throughout this project.

• Stachybotrys chartarum was detected from furniture dust in room 15327 after abatement and cleaning, but was not detected after second cleaning.

CONCLUSIONS

Overall, fungal burden on various surfaces in room 15327 was low after the second cleaning of this room. Airborne fungal levels and total fungal spores in the air in room 15327 were lower than those of outdoors and were compatible to those of the reference room 15212. Fungal levels in composite furniture dust decreased from 10^4 to 10^3 CFU/g of fine dust after second cleaning of room 15327. However, fungal levels in carpet dust remained at the 10^4 CFU/g of fine dust level and consistent presence of *Stachybotrys chartarum* was detected.

RECOMMENDATIONS

• HEPA vacuum and wet-wipe of the open edge of bookshelf in room 15212.

• Revise the small-scale drywall removal protocol to prevent spores release to the carpet during drywall removal (Attachment D).

• Use HEPA vacuum cleaner as a routine housekeeping of the carpeting in the building.

 \cdot Implement an emergency water intrusion protocol for this building to adequately manage the unexpected water intrusion in order to prevent any fungal proliferation.

ATTACHMENT A

SSMC-3 small-scale drywall removal protocol.

ATTACHMENT B

Microbiological laboratory reports for samples collected

from rooms 15327 and 15212 of SSMC-3,

in December, 1999 and January 2000.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA LABORATORY REPORT #NOAA-00-11R-A

http://www.rdc.noaa.gov/~facmd/phs%20final%20reports/15327.htm (12 of 42) [2/6/2002 3:34:55 PM]

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903 Sampling date: 12/9/99 Dates of inoculation: 12/9/99 General location: SSMC-3, Silver Spring, MD Specific location: 15th floor Sampling techniques: Air (Andersen N-6 sampler) and contact plate samplings Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi Samples submitted by: J. Sobelman Date characterization completed: 12/20/99

(A) Air samples on MEA and CCA plates

Sample ID	Sampling Location	Air	Fungi on MEA	Presence of	
		Volume	@ 25º C	<i>Stachybotrys</i> <i>chartarum</i> *** on	
15327M1, C1	Room 15327, near window casing	(L) 84.9	No fungal growth	CCA @ 25º C No	
15327M2, C2	Room 15327, near center of room	84.9	CFU/m ³ < 12 1. <i>Alternaria</i> (1*)	No	
15212M1, C1	Room 15212, near window	84.9	$CFU/m^{3} = 12$ 1. Cladosporium (4) 2. Alternative (1)	No	
15212M2, C2	Room 15212, table in center	84.9	 Alternaria (1) CFU/m³ = 59 Aureobasidium (1) Scopulariopsis (1) 	No	
			$CFU/m^3 = 24$		
Sample ID	Sampling Location	Air	Fungi on MEA	Presence of Stachybotrys	
		Volume (L)	@ 25º C	chartarum*** or CCA @ 25º C	

OM1, OC1	Outside bldg. 3	84.9	1.	Penicillium (30)	No
			2.	Cladosporium (15)	
			3.	Aspergillus sp. (2)	
			4.	Aureobasidium (1)	
			5.	Paecilomyces (1)	
OM2, OC2	Outside bldg. 3	28.3	CF 1.	$U/m^3 = 577$ Penicillium (2)	Yes (2)
			2.	Aspergillus niger** (1)	$CFU/m^{3} = 71$
			3.	Cladosporium (1)	
			4.	Rhizopus (1)	
			CF	$U/m^3 = 177$	

(B) Contact plate samples on MEA plates

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15327CP1	Room 15327, door to room (entrance/exit)	No fungal growth CFU/plate < 1
15327CP2	Room 15327, wall near window	No fungal growth CFU/plate < 1
15327CP3	Room 15327, front of system furniture, 2 nd station	 Alternaria (1) Aspergillus sp. (1) Basidiomycetes (1) CFU/plate = 3
15327CP4	Room 15327, front of black metal cabinet	1. <i>Alternaria</i> (1) CFU/plate = 1

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15327CP5	Room 15327, top of color laser	1. Alternaria (1)
		2. Epicoccum (1)
		CFU/plate = 2

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15327CP6	Room 15327, top of grey file	1.	Cladosporium (9)
	cabinet	2.	Alternaria (3)
		3.	Aspergillus sp. (2)
		4.	Penicillium (2)
		5.	Aspergillus flavus*** (1)
		6.	yeast (1)
		CFU	f/plate = 18
15327CP7	Room 15327, top of table in front of	1.	Aureobasidium (3)
	window	2.	Cladosporium (3)
		3.	Penicillium (3)
		4.	Aspergillus sp. (2)
		5.	Alternaria (1)
		6.	Basidiomycetes (1)
		CFU	l/plate = 13
15327CP8	Room 15327, top of scanner	1.	Alternaria (2)
		2.	Cladosporium (1)
		3.	Penicillium (1)
		CFU	/plate = 4
15212CP1	Room 15212, wall near entrance	1.	Cladosporium (1)
		CFU	1/plate = 1
15212CP2	Room 15212, wall above coffee pot	1.	Alternaria (1)
		CFU	l/plate = 1
15212CP3	Room 15212, wall near window	1.	Alternaria (2)
		2.	Chaetomium (1)
		3.	Basidiomycetes (1)
		CFU	l/plate = 4

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15212CP4	Room 15212, front of shelves	No fungal growth
	(door) opposite desk	
		CFU/plate < 1
15212CP5	Room 15212, top of desk	1. Chaetomium (6)
		2. Cladosporium (1)
		CFU/plate = 7

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		,	,	
	15212CP6	Room 15212, top of computer	1.	Alternaria (3)
			2.	Aureobasidium (1)
			3.	Cladosporium (1)
			4.	Epicoccum (1)
			CFU	I/plate = 6
	15212CP7	Room 15212, file cabinet of coffee	No f	ungal growth
		pot		
		r	CFU	1/plate < 1
ĺ	15212CP8	Room 15212, book shelf (open).	1.	Alternaria (2)
	10212010	front of books	··	/
			2.	Chaetomium (2)
			3.	Aspergillus niger**(1)
			4.	Cladosporium (1)
			5.	Basidiomycetes (7)
			CFU	1/plate = 13

* Colony counts.

** Opportunistic fungi.

*** Toxigenic fungi.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-11R-B</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/9/99

Dates of inoculation: 12/10/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling technique: Wipe samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 12/20/99

Wipe samples on MEA and CCA plates

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA
<u></u>					@ 25°C
15327S1	Room 15327, supply	4	10X	No fungal growth	No
	diffuser			$CFU/in^2 < 3$	
15327S2	Room 15327, supply	4	10X	No fungal growth	No
	diffuser			CFU/in ² < 3	
	Room 15327, supply	4	10X	No fungal growth	No
1532783	diffuser			CFU/in ² < 3	
	Room 15327, supply	4	10X	No fungal growth	No
	diffuser			$CEU/in^2 < 2$	
1532784				$CFO/III^2 < 5$	
1532785	Room 15327, supply	4	10X	No fungal growth	No
	diffuser				
				$CFU/in^2 < 3$	
15327R1	Room 15327, return-light	5	10X	No fungal growth	No
				CFU/in ² < 2	
	Room 15327, return-light	5	10X	No fungal growth	No
15327R2				$CFU/in^2 < 2$	

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
15327R3	Room 15327, return-light	5	10X	1. Penicillium (1) CFU/in ² = 2	No
15327R4	Room 15327, return-light	5	10X	No fungal growth CFU/in ² < 2	No
15212S1	Room 15212, supply diffuser	4	10X	No fungal growth CFU/in ² < 3	No
15212S2	Room 15212, supply diffuser	4	10X	No fungal growth CFU/in ² < 3	No
15212R1	Room 15212, return	5	10X	No fungal growth CFU/in ² < 2	No
15212R2	Room 15212, return	5	10X	No fungal growth CFU/in ² < 2	No

* Colony counts.

*** Toxigenic fungi.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-11R-C</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/9/99

Dates of inoculation: 12/15/99#

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling technique: Vacuum dust sampling

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 12/28/99

Dust samples on MEA and CCA plates

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of
ID		(g)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
15327V1FC	Room 15327, near center of	0.072##	40X-MEA	1. Cladosporium (16*)	No
	room, furniture composite		10X-CCA	2. Alternaria (14)	
				3. Epicoccum (7)	
				4. Paecilomyces (4)	
				5. Aureobasidium (2)	
				6. Bipolaris (2)	
				7. Penicillium (2)	
				$CFU/g = 1.3 \times 10^4$	

15327V2CA Room 15327, near center of	0.100	40X-MEA 1.	Penicillium (21)	Yes (5)
room, carpet		40X-CCA 2.	Aspergillus sp. (3)	CFU/g = 2,000
		3.	Alternaria (2)	
		4.	Epicoccum (1)	
		5.	Paecilomyces (1)	
		6.	Basidiomycetes (1)	
		CF	$U/g = 1.2 \times 10^4$	

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of
ID		(g)	factor	@ 25°C	Stachybotrys chartarum*** on
					CCA @ 25°C
15212V1FC	Room 15212, furniture	0.073##	400X-MEA	1. Alternaria (14)	No
	composite		10X-CCA	2. Cladosporium (6)	
				3. Aureobasidium (1)	
				4. Paecilomyces (1)	
				5. Penicillium (1)	
				6. Rhizopus (1)	
				$CFU/g = 6.6 \times 10^4$	
	Room 15212, carpet	0.100	40X-MEA	1. Cladosporium (8)	No
15212V2CA			10X-CCA	2. Alternaria (3)	
				3. Aspergillus niger**(1)	
				4. Paecilomyces (1)	
				CFU/g = 5,200	

* Colony counts.

** Opportunistic fungi.

*** Toxigenic fungi.

Samples processed upon receipt.

5ml of sterilized distilled water were added instead of 10ml.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA LABORATORY REPORT #NOAA-00-12R

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/10/99

Dates of inoculation: 12/11/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor, Room 15327

Sampling techniques: Wipe sampling

Medium used: Malt extract agrar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: L. Hung

Date characterization completed: 12/20/99

Wipe samples on MEA and CCA plates

Sample	Sampling Location	Area	Dilution factor	Fungi on MEA	Presence of
ID		(in ²)		@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
3-15327-121099-W01	Room 15327, concrete pre-cast	1	10X	 Penicillium (10*) Stachybotrys chartarum*** (1) CFU/in² = 110 	Yes (3) CFU/in ² = 30
3-15327-121099-W02	Room 15327, concrete pre-cast	1	400X-MEA 40X-CCA	 Penicillium (27) Stachybotrys chartarum*** (1) CFU/in² = 1.1 x 10⁴ 	Yes (2) CFU/in ² = 80
3-15327-121099-W03	Room 15327, visible growth on plastic behind removed drywall by the window	1	4,000X	 Stachybotrys chartarum*** (105) Penicillium (40) CFU/in² = 5.8 x 10⁵ 	Yes (>400) CFU/in ² > 1.6 x 10 ⁶
3-15327-121099-W04	Room 15327, control, no visible growth, wipe on plastic behind removed drywall	1	10X	 Penicillium (14) Spororbolomyces (4) Stachybotrys chartarum*** (4) CFU/in² = 220 	Yes (3) CFU/in ² = 30

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
3-15327-121099-W05	Control	NA#	10X	No fungal growth	No

* Colony counts.

*** Toxigenic fungi.

[#] Not applicable.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-13R-A</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/13/99

Dates of inoculation: 12/13/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling techniques: Air (Andersen N-6 sampler) and contact plate samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 12/23/99

(A) Air samples on MEA and CCA plates

Sample ID	Sampling Location	Air	Fungi on MEA	Presence of
		Volume	@ 25º C	<i>Stachybotrys</i> <i>chartarum</i> *** on
15327M1, C1	Room 15327, near	(L) 84.9	1. Alternaria (1*)	CCA @ 25º C Yes (1)
	window		2. <i>Epicoccum</i> (1)	$CFU/m^{3} = 12$
15327M2, C2	Room 15327, center of room	84.9	CFU/m ³ = 24 1. Aspergillus sp. (1) 2. Paecilomyces (1)	Yes (1) CFU/m ³ = 12
			$CFU/m^{3} = 24$	

15212M1, C1 Room 15212, near window	84.9	1.	Aspergillus sp. (2)	No	
		2.	Aureobasidium (1)		
			3.	Cladosporium (1)	
15212M2, C2	Room 15212, center	84.9	CFU 1.	$M/m^3 = 47$ Alternaria (1)	No
			CFU	$M/m^3 = 12$	

Sample ID	Sampling Location	Air		Fungi on MEA	Presence of
		Volume		@ 25º C	Stacnybotrys chartarum*** on
		(L)			CCA @ 25º C
OM1, OC1	Outside bldg. 3	84.9	1. C	Eladosporium (11)	No
			2. P	Penicillium (3)	
			3. A	spergillus sp. (2)	
			4. <i>E</i>	Epicoccum (2)	
			5. E	Basidiomycetes (3)	
			CFU/n	$n^3 = 247$	
OM2, OC2	Outside bldg. 3	28.3	1. C	Cladosporium (6)	No
			2. A	lternaria (1)	
			3. A	spergillus niger** (1)	
			4. <i>P</i>	Paecilomyces (1)	
			5. P	Penicillium (1)	
			6. E	Basidiomycetes (2)	
			CFU/n	$n^3 = 424$	

(B) Contact plate samples on MEA plates

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15327CP1	Room 15327, door to room	No fungal growth
	(entrance/exit)	CFU/plate < 1
15327CP2	Room 15327, wall near window	No fungal growth
		CFU/plate < 1
15327CP3	Room 15327, front of system	No fungal growth
	furniture, 2 nd station	CFU/plate < 1

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	Room 15327, front of black metal	No fungal growth
15327CP4	cabinet	CFU/plate < 1

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15327CP5	Room 15327, top of color laser	1. Penicillium (9)
		2. Alternaria (1)
		3. Aureobasidium (1)
		4. Cladosporium (1)
		CFU/plate = 12
15327CP6	Room 15327, top of grey file cabinet	1. Alternaria (1)
		2. Penicillium (1)
		3. Basidiomycetes (1)
		CFU/plate = 3
15327CP7	Room 15327, top of table in front of	1. Penicillium (3)
	window	2. Cladosporium (1)
		CFU/plate = 4
15327CP8	Room 15327, top of scanner	1. Penicillium (81)
		2. Cladosporium (1)
		CFU/plate = 82
15212CP1	Room 15212, door to room	1. Basidiomycetes (1)
		CFU/plate = 1
15212CP2	Room 15212, wall above coffee pot	1. Chaetomium (3)
		CEU/plate = 3
15212CP3	Room 15212, wall near window	No fungal growth
		CFU/plate < 1
15212CP4	Room 15212, front of shelves	No fungal growth
	opposite desk	CFU/plate < 1
15212CP5	Room 15212, top of desk	1. Rhizopus (1)

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
	,	

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15212CP6	Room 15212, top of computer	1.	Aureobasidium (2)
		2.	Cladosporium (1)
		3.	Epicoccum (1)
		4.	Basidiomycetes (2)
		CF	U/plate = 6
15212CP7	Room 15212, file cabinet of coffee	1.	Chaetomium (2)
	pot	2.	Penicillium (1)
		CFU	U/plate = 3
15212CP8	Room 15212, book shelf (open), front of books	1.	Chaetomium (22)
		2.	Cladosporium (5)
		3.	Alternaria (2)
		4.	Aureobasidium (1)
		5.	Paecilomyces (1)

* Colony counts.

** Opportunistic fungi.

*** Toxigenic fungi.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-13R-B</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/15/99

Dates of inoculation: 12/16/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling technique: Wipe samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 12/28/99

Wipe samples on MEA and CCA plates

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on
1532781	Room 15327 supply		10X	No fungal growth	CCA @ 25 C
1552751	diffuser		107	$CFU/in^2 < 3$	
1532782	Room 15327, supply diffuser	4	10X	No fungal growth $CEU/in^2 < 3$	No
<u> </u>	Room 15327 supply	1	10X	$V_{\rm No}$ fungal growth	No
15327S3	diffuser		10/X	$CFU/in^2 < 3$	NO
	Room 15327, supply diffuser	4	10X	No fungal growth	No
1532784				$CFU/m^2 < 3$	
1532785	Room 15327, supply diffuser	4	10X	No fungal growth	No
				$CFU/in^2 < 3$	
15327R1	Room 15327, return-light	5	10X	No fungal growth	No
<u> </u>				CFU/in ² < 2	
	Room 15327, return-light	5	10X	No fungal growth	No
15327R2				$CFU/in^2 < 2$	

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
15327R3	Room 15327, return-light	5	10X	No fungal growth CFU/in ² < 2	No
15327R4	Room 15327, return-light	5	10X	No fungal growth CFU/in ² < 2	No
1521281	Room 15212, supply diffuser	4	10X	No fungal growth CFU/in ² < 3	No
15212S2	Room 15212, supply diffuser	4	10X	1. Chaetomium (1^*) CFU/in ² = 3	No
15212R1	Room 15212, return	5	10X	No fungal growth CFU/in ² < 2	No
15212R2	Room 15212, return	5	10X	No fungal growth CFU/in ² < 2	No

* Colony counts.

*** Toxigenic fungi.

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USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-13R-C</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/13/99

Dates of inoculation: 12/15/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling technique: Vacuum dust sampling

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 12/28/99

Dust samples on MEA and CCA plates

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of <i>Stachybotrys</i>
ID		(g)	factor	@ 25°C	chartarum*** on CCA @ 25°C
15327V1FC	Room 15327, near	0.053#	40X-MEA	1. Cladosporium (58*)	Yes (1)
	center of room, furniture composite		10X-CCA	2. Alternaria (9)	CFU/g = 94
				3. Epicoccum (4)	
				4. Aureobasidium (2)	
				5. Chaetomium (2)	
				6. Paecilomyces (1)	
				7. Penicillium (1)	
				$CFU/g = 2.9 \times 10^4$	

15327V2CA Room 15327, near	0.100	40X-MEA	1.	Penicillium (33)	Yes (4)
center of room, carpet		10X-CCA	2.	Epicoccum (10)	CFU/g = 400
			3.	Aspergillus niger**(1)	
			4.	Aureobasidium (1)	
			5.	Cladosporium (1)	
			6.	Paecilomyces (1)	
			7.	Pithomyces (1)	
			CFU	$J/g = 1.9 \text{ x } 10^4$	

Sample	Sampling Location	Weight	Dilution		Fungi on MEA	Presence of <i>Stachybotrys</i>
ID		(g)	factor		@ 25°C	chartarum*** on CCA @ 25°C
15212V1FC	Room 15212, furniture	0.020#	40X-MEA	1.	Alternaria (6)	No
	composite		10X-CCA	2.	Aureobasidium (2)	
				3.	Cladosporium (2)	
				4.	Aspergillus niger**(1)	
				5.	Bipolaris (1)	
				6.	Chaetomium (1)	
				7.	Ascomycetes (2)	
				CFU	$U/g = 1.5 \text{ x } 10^4$	
15212V2CA	Room 15212, carpet	0.100	40X-MEA	1.	Cladosporium (24)	No
			10X-CCA	2.	Alternaria (9)	
				3.	<i>Epicoccum</i> (4)	
				4.	Chaetomium (2)	
				5.	Rhizopus (1)	
				CFU	$U/m^3 = 1.6 \ge 10^4$	

* Colony counts.

** Opportunistic fungi.

*** Toxigenic fungi.

[#] 5ml of sterilized distilled water were added instead of 10ml.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-14R</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/10/99

Dates of inoculation: 12/17/99

General location: Silver Spring, MD

Specific location: SSMC-3, 15th floor, Room 15327

Sampling techniques: Wipe and bulk sampling

Medium used: Malt extract agrar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: L. Hung and R. Pickett

Date characterization completed: 12/22/99

(A) Wipe samples on MEA and CCA plates

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
15327-1210-W01	Wipe on black fungal growth on dry wall (interior side)	1	4,000X	1.Stachybotrys chartarum*** (135*)2.Penicillium (35)CFU/in ² = $6.8 \ge 10^5$	Yes
15327-1210-W02	Wipe on black fungal growth on dry wall (interior side)	1	40,000X	1. <i>Penicillium</i> (146) CFU/in ² = 5.8×10^{6}	Yes
15327-1210-W03	Wipe on perforated paper behind the dry wall	1	4,000X	 Stachybotrys chartarum*** (240) Penicillium (1) CFU/in² = 9.6 x 10⁵ 	Yes
15327-1210-W04	Wipe on perforated paper behind the dry wall	1	40,000X	1. Stachybotrys chartarum*** (360) CFU/in ² = 1.4×10^7	Yes
15327-1210-W05	Wipe on perforated plastic material behind the dry wall	1	4,000X	1.Stachybotrys chartarum*** (211)2.Penicillium (1)CFU/in2 = 8.5×10^5	Yes

Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
15327-1210-W06	Wipe on perforated plastic material behind the dry wall	1	4,000X	 Stachybotrys chartarum*** (166) Penicillium (2) 	Yes

(B) Bulk sampling on MEA and CCA plates

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of
ID		(g)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
15327-1210-B01-1	Dry wall sample (interior, brown part)	0.154	40,000X	 Penicillium (64) Stachybotrys chartarum*** (5) CFU/g = 1.8 x 10⁷ 	Yes
15327-1210-B01-2	Dry wall sample (interior, brown part)	0.134	40,000X	 Penicillium (46) Stachybotrys chartarum*** (7) CFU/g = 1.6 x 10⁷ 	Yes
15327-1210-B01-3	Dry wall sample (interior, brown part)	0.113	4,000X	 Penicillium (51) Stachybotrys chartarum*** (9) CFU/g = 2.1 x 10⁶ 	Yes
15327-1210-B02-1	Paper wrapping outside of fiberglass insulation material	0.107	4,000X	 Penicillium (42) Stachybotrys chartarum*** (1) CFU/g = 1.6 x 10⁶ 	Yes
15327-1210-B02-2	Paper wrapping outside of fiberglass insulation material	0.096	4,000X	 Stachybotrys chartarum*** (48) Penicillium (18) CFU/g = 2.8 x 10⁶ 	Yes

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of
ID		(g)	factor	@ 25°C	Stachybotrys
10		(g)		e 25 C	<i>chartarum</i> *** on
					CCA @ 25°C

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15327-1210-B02-3 Paper wrapping	0.123	4,000X	1. Stachybotrys	Yes
outside of fiberglass			chartarum*** (18)	
insulation material			2. Penicillium (10)	
			$CFU/g = 9.1 \times 10^5$	

* Colony counts.

*** Toxigenic fungi.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-15R-A</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/21/99

Dates of inoculation: 12/21/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling techniques: Air (Andersen N-6 sampler) and contact plate samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 1/3/00

(A) Air samples on MEA and CCA plates

Sample ID	Sampling Location	Air		Fungi on MEA	Presence of
		Volume		@ 25º C	Stachybotrys chartarum*** on
		(L)			CCA @ 25º C
15327M1, C1	Room 15327, near window	84.9	1.	Aspergillus sp. (1*)	No
			2.	Cladosporium (1)	
			3.	Penicillium (1)	
			4.	Basidiomycetes (1)	
			CF	$U/m^3 = 47$	

15327M2, C2	Room 15327, center of	84.9	1. Cladosporium (2)	No
	room		2. <i>Penicillium</i> (1)	
			3. Basidiomycetes (1)	
15212M1, C1	Room 15212, near window	84.9	$CFU/m^{3} = 47$ 1. Paecilomyces (2) 2. Chaetomium (1) 2. Do i Victoria (2)	No
15212M2, C2	Room 15212, table in office	84.9	3. Basidiomycetes (2) $CFU/m^3 = 59$ 1. Aspergillus sp. (1)	No
			$CFU/m^3 = 12$	

Sample ID	Sampling Location	Air	Fungi on MEA	Presence of Stachybotrys chartarum*** on
		Volume	@ 25º C	
		(L)		CCA @ 25º C
OM1, OC1	Outside bldg. 3	84.9	1. Cladosporium (7)	No
			2. Basidiomycetes (67)	
OM2, OC2	Outside bldg. 3	28.3	$\begin{array}{l} CFU/m^3 = 872\\ 1. Basidiomycetes (14) \end{array}$	No
SB	Shipping blank	NA [#]	CFU/m ³ = 495 No fungal growth	No

(B) Contact plate samples on MEA plates

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15327CP1	Room 15327, door to room (entrance/exit)	No fungal growth CFU/plate < 1
15327CP2	Room 15327, wall near window	1. <i>Epicoccum</i> (1) CFU/plate = 1
15327CP3	Room 15327, front of system furniture, 2 nd station	 Alternaria (1) Mucor (1) CELUplate - 2
15327CP4	Room 15327, front of black metal cabinet	 Penicillium (5) Stachybotrys chartarum*** (4) CFU/plate = 9

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15327CP5	Room 15327, top of color laser	1. Penicillium (22)
		2. Mucor (1)
		CFU/plate = 23
15327CP6	Room 15327, top of grey file cabinet	1. <i>Chaetomium</i> (1) CFU/plate = 1

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15327CP7	Room 15327, top of table in	1. Penicillium (3)
	front of window	2. Chaetomium (2)
		3 Aspervillus sp (1)
		4 Describer (1)
		4. Paecuomyces (1)
		CFU/plate = 7
15327CP8	Room 15327, top of scanner	No fungal growth
		CFU/plate < 1
15212CP1	Room 15212, door to room	No fungal growth
		CFU/plate < 1
15212CP2	Room 15212, wall above coffee	1. Alternaria (1)
	pot	CFU/plate = 1
15212CP3	Room 15212, wall near window	1. Alternaria (1)
		CFU/plate = 1
15212CP4	Room 15212, front of shelves	1. Basidiomycetes (2)
	opposite desk	CFU/plate = 2
15212CP5	Room 15212, top of desk	1. Cladosporium (1)
		2 Penicillium (1)
15212CDC	D	CFU/plate = 2
15212CP6	Room 15212, top of computer	1. Alternaria (1)
		2. Basidiomycetes (3)
		CFU/plate = 4
15212CP7	Room 15212, file cabinet of	1. Penicillium (3)
	coffee pot	2. Paecilomyces (2)
1		Cru/plate = 5

	Sampling Location	Fungi detected on
Sample ID		MEA @ 25°C
15212CP8	Room 15212, book shelf (open),	1. Chaetomium (20)
	front of books	2. Penicillium (11)
		3. Cladosporium (7)
		CFU/plate = 38

* Colony counts.

*** Toxigenic fungi.

[#] Not applicable.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-15R-B</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/21/99

Dates of inoculation: 12/22/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling technique: Wipe samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 1/3/00

Wipe samples on MEA and CCA plates

FOH	Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID	ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
1	1521281	Room 15212, supply diffuser	4	10X	No fungal growth CFU/in ² < 3	No
2	15212S2	Room 15212, supply diffuser	4	10X	No fungal growth CFU/in ² < 3	No

3	15212R1	Room 15212, return	5	10X	No fungal growth	No
					$CFU/in^2 < 2$	
4	15212R2	Room 15212, return	5	10X	No fungal growth	No
					$CFU/in^2 < 2$	
5	15327S1	Room 15327, supply	4	10X	No fungal growth	No
		diffuser			$CFU/in^2 < 3$	
6	15327S2	Room 15327, supply	4	10X	No fungal growth	No
		diffuser				
					$CFU/in^2 < 3$	
7		Room 15327, supply	4	10X	No fungal growth	No
	1532783	diffuser			$CFU/in^2 < 3$	

FOH	Sample	Sampling Location	Area	Dilution	Fungi on MEA	Presence of
ID	ID		(in ²)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C
		Room 15327, supply	4	10X	No fungal growth	No
8	15327S4	diffuser			$CFU/in^2 < 3$	
9	1532785	Room 15327, supply diffuser	4	10X	No fungal growth CFU/in ² < 3	No
10	15327R1	Room 15327, return	5	10X	No fungal growth CFU/in ² < 2	No
		Room 15327, return	5	10X	No fungal growth	No
11	15327R2				CFU/in ² < 2	
12	15327R3	Room 15327, return	5	10X	No fungal growth CFU/in ² < 2	No
13	15327R4	Room 15327, return	5	10X	No fungal growth CFU/in ² < 2	No

*** Toxigenic fungi.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-15R-C</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 12/21/99

Dates of inoculation: 12/23/99

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling technique: Vacuum dust sampling

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 1/3/00

(A) Dust samples on MEA and CCA plates

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of
ID		(g)	factor	@ 25°C	Stachybotrys chartarum*** on
					CCA @ 25°C
15327FC	Room 15327, furniture	0.037#	40X-MEA	1. Alternaria (14*)	Yes (4)
	composite		40X-CCA	2. Cladosporium (7)	CFU/g = 2,162
				3. Aureobasidium (4)	
				4. Epicoccum (4)	
				5. Bipolaris (3)	
				6. Chaetomium (2)	
				7. Penicillium (1)	
				$CFU/g = 1.9 \text{ x } 10^4$	
15327CA	Room 15327, carpet	0.100	40X-MEA	1. Penicillium (18)	Yes (5)
			40X-CCA	2. Stachybotrys chartarum*** (6)	$CFU/g = 2.0 \times 10^3$
				3. Cladosporium (3)	
				4. Alternaria (2)	
				5. Aureobasidium (2)	
				6. Aspergillus sp. (1)	
				7. Epicoccum (1)	
				CFU/g = 1.3 x 10 ⁴	

Sample	Sampling Location	Weight	Dilution	Fungi on MEA	Presence of
ID		(g)	factor	@ 25°C	Stachybotrys chartarum*** on CCA @ 25°C

15212FC	Room 15212, furniture	0.010#	40X-MEA	1.	Alternaria (5)	No
	composite		10X-CCA	2.	Aspergillus flavus*** (2)	
				3.	Aureobasidium (2)	
				4.	Cladosporium (2)	
				5.	Paecilomyces (1)	
				6.	Basidiomycetes (44)	
				CFU	$J/g = 1.1 \text{ x } 10^5$	

(B) Dust sample on MEA plates by direct plating

Sample Sampling Location		Fungi detected on MEA	
ID		@ 25°C	
15212CA##	Room 15212, carpet	Aureobasidium	
		yeast	

* Colony counts.

*** Toxigenic fungi.

[#] 5ml of sterilized distilled water were added instead of 10ml.

Insufficient amounts of dust for analysis.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-21R</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 1/24/00

Dates of inoculation: 1/24/00 (airs and contact plates) and 1/28/00 (dust)

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling techniques: Air (Andersen N-6 sampler), contact plate, and vacuum dust samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 2/7/00

(A) Air samples on MEA and CCA plates

Sample Sampling Location A		Air	Fungi on MEA	Presence of	
ID		Volume	@ 25º C	<i>Stachybotrys</i> <i>chartarum</i> *** or	
		(L)		CCA @ 25º C	
15212124A1, A2	15 th floor, room 15212,	84.9	No fungal growth	No	
	center		CFU/m ³ < 12		
Field blank	Blank	NA [#]	No fungal growth	No	
15327124A1, A2	15 th floor, room 15327,	84.9	No fungal growth	No	
	center		$CFU/m^3 < 12$		
1240A1, 0A2	Outside bldg. 3	84.9	1. Penicillium (4*)	No	
			$CFU/m^{3} = 47$		
1240A3, OA4	Outside bldg. 3	28.3	1. Penicillium (4)	No	
			$CFU/m^3 = 141$		
SB	Shipping blank	NA	No fungal growth	No	

(B) Contact plate samples on MEA plates

FOH	Sample	Sampling Location	Fungi detected on MEA
ID	ID		@ 25º C
25	15212124CP1	15th floor, room 15212, door to	No fungal growth
		room	CFU/plate < 1
26	15212124CP2	15 th floor, room 15212, wall above	e1. <i>Mucor</i> (2)
		coffee pot	2. Alternaria (1)
27	15212124CP3	15 th floor, room 15212, wall near	CFU/plate = 3 1. <i>Cladosporium</i> (3)
		window	2. Alternaria (2)
			3. <i>Mucor</i> (2)
28	15212124CP4	15 th floor, room 15212, front of	CFU/plate = 7 No fungal growth
		shelves opposite desk	CFU/plate < 1

29	15212124CP5	15 th floor, room 15212, top of desk	<u>,</u> 1.	Chaetomium (2)
			2.	Aspergillus sp. (1)
			3.	Penicillium (1)
30	15212124CP6	15 th floor, room 15212, top of	CFU 1.	l /plate = 4 Cladosporium (1)
		computer	2.	Paecilomyces (1)
			3.	Basidiomycetes (1)
31	15212124CP7	15 th floor, room 15212, top of file with coffee pot	CFU 1.	l/plate = 3 Aspergillus sp. (1)
32	15212124CP8 15 th floor room 15212 one		CFU 1.	//plate = 1 <i>Penicillium</i> (12)
-		of book shelf front of book		
		of book shell, front of book	2.	Cladosporium (8)
		of book shell, from of book	2. 3.	Cladosporium (8) Aspergillus niger** (2)
		of book shell, from of book	2. 3. 4.	Cladosporium (8) Aspergillus niger** (2) Aureobasidium (1)

CFU/plate = 71

FOH	Sample	Sampling Location	Fungi detected on MEA		
ID	ID		@ 25º C		
33	15327124CP1	15th floor, room 15327, door to	No fungal growth		
		room	CFU/plate < 1		
34	15327124CP2	15 th floor, room 15327, wall near	No fungal growth		
		window	CFU/plate < 1		
35	15327124CP3	15 th floor, room 15327, 2 nd	No fungal growth		
26	15005104004	station, front of system furniture	CFU/plate < 1		
36	15327124CP4	15 th floor, room 15327, front of	No fungal growth		
27	15207104CD5	15th Classical States	CFU/plate < 1		
57	13327124CF3	15 th floor, room 15327, top of color laser	No lungar growth		
38	15327124CP6	15th floor room 15227 top of any	CFU/plate < 1		
50	15527124010	file cabinet			
39	15327124CP7	15 th floor room 15327 top of	CFU/plate < 1 No fungal growth		
07	1002/12/01/	table in front of window			
40	15327124CP8	15 th floor, room 15327, top of	1. <i>Cladosporium</i> (1)		
		scanner	2 Panicillium (1)		
			2. <i>Tenicilium</i> (1)		

CFU/plate = 2

(C) Vacuum dust samples on MEA and CCA plates

FOH		Sampling	Weight	Dilution factor	Fungi on	Presence of
ID	Sample ID	Location	(g)		MEA @ 25°C	Stachybotrys chartarum*** on
						CCA @ 25º C
V07	15212124V01	15 th floor, room	0.101	40X-MEA	1. Paecilomyces (4)	No
		15212, carpet		10X-CCA	2. Alternaria (1)	
					3. Chaetomium (1)	
					4. Penicillium (1)	
					5. Trichoderma (1)	
					CFU/g = 3,168	
V08	15212124V02	15 th floor, room	0.041##	40X-MEA	1. Cladosporium (10)	No
		15212, furniture		10X-CCA	2. Alternaria (9)	
					3. Penicillium (3)	
					4. Epicoccum (2)	
					$CFU/g = 1.2 \times 10^4$	
V09	15327124V01	15 th floor, room	0.085	40X-MEA	1. Penicillium (25)	Yes (8)
		15327, carpet		40X-CCA	2. Stachybotrys chartarum*** (7)	CFU/g = 3,765
					3. Aureobasidium (5)	
					4. Cladosporium (3)	
					5. Pithomyces (1)	
					$CFU/g = 1.9 \times 10^4$	

FOH		Sampling	Weight	Dilution factor	Fungi on	Presence of
ID	Sample ID	Location	(g)		MEA @ 25°C	Stachybotrys chartarum*** on CCA @ 25º C

V10	15327124V02 15 th floor, room	0.101##	40X-MEA	1.	Alternaria (16)	No
	15327, furniture		10X-CCA	2.	Epicoccum (8)	
				3.	Penicillium (6)	
				4.	Aspergillus sp. (1)	
				5.	Bipolaris (1)	
				6.	Pithomyces (1)	
				CF	U/g = 6,535	

* Colony counts.

*** Toxigenic fungi.

[#] Not applicable.

5ml of sterilized distilled water were added instead of 10ml.

USPHS DFOH ENVIRONMENTAL MICROBIOLOGY LABORATORY, PHILADELPHIA, PA

LABORATORY REPORT <u>#NOAA-00-23R</u>

Client agency: National Oceanic and Atmospheric Administration, Silver Spring, MD

POIS#/task #: D8H00CO31200 / 9903

Sampling date: 2/1/00

Dates of inoculation: 2/2/00

General location: SSMC-3, Silver Spring, MD

Specific location: 15th floor

Sampling techniques: Wipe samplings

Medium used: Malt extract agar (MEA) and Cellulose Czapek agar (CCA) for fungi

Samples submitted by: J. Sobelman

Date characterization completed: 2/12/00

Wipe samples on MEA and CCA plates

FO	H ID		Sampling Location	Area	Dilution factor	Fungi on	Presence of Stachybotrys
		Sample ID		(in ²)		MEA @ 25°C	<i>chartarum</i> *** on CCA
							@ 25º C
		1521221R1	15 th floor, room	5	40X-MEA	No fungal growth	No
v v	V01		15212, return trougher		10X-CCA	CFU/in ² < 8	

	1521221R2	15 th floor, room	5	40X-MEA	No fungal growth	No
		15212, return trougher		10X-CCA	CFU/in ² < 8	
W02						
W03	1521221S1	15 th floor, room	4	40X-MEA	No fungal growth	No
		15212, supply diffuser		10X-CCA	CFU/in ² < 10	
W04	1521221S2	15 th floor, room	4	40X-MEA	No fungal growth	No
		15212, supply diffuser		10X-CCA	CFU/in ² < 10	
W05	1532721S1	15 th floor, room	4	40X-MEA	No fungal growth	No
		15327, supply diffuser		10X-CCA	CFU/in ² < 10	
W06	1532721S2	15 th floor, room	4	40X-MEA	No fungal growth	No
		15327, supply diffuser		10X-CCA	CFU/in ² < 10	
W07	153272183	15 th floor, room	4	40X-MEA	No fungal growth	No
		15327, supply diffuser		10X-CCA	CFU/in ² < 10	
W08	1532721S4	15 th floor, room	4	40X-MEA	No fungal growth	No
		15327, supply diffuser		10X-CCA	CFU/in ² < 10	

FOH ID		Sampling Location	Area	Dilution factor	Fungi on	Presence of Stachybotrys
	Sample ID		(in ²)		MEA @ 25°C	chartarum*** on CCA
						@ 25° C
W09	153272185	15 th floor, room	4	40X-MEA	No fungal growth	No
		15327, supply diffuser		10X-CCA	CFU/in ² < 10	
W10	1532721R1	15 th floor, room	5	40X-MEA	No fungal growth	No
		15327, return trougher		10X-CCA	CFU/in ² < 8	
W11	1532721R2	15 th floor, room	5	40X-MEA	No fungal growth	No
		15327, return trougher		10X-CCA	CFU/in ² < 8	
W12	1532721R3	15 th floor, room	5	40X-MEA	No fungal growth	No
		15327, return trougher		10X-CCA	CFU/in ² < 8	
W13	1532721R4	15 th floor, room	5	40X-MEA	No fungal growth	No
		15327, return trougher		10X-CCA	CFU/in ² < 8	
Blank	Blank	Field blank	NA [#]	40X-MEA	No fungal growth	No
				10X-CCA		

*** Toxigenic fungi.

[#] Not applicable.

ATTACHMENT C

Results from microscopic examination of air and tape lift samples collected from

rooms 15327 and 15212 of SSMC-3, in December, 1999 and January 2000.

All attachments can be retrieved from the Library located on the Second Floor in SSMC 3

ATTACHMENT D

Revised small-scale drywall removal protocol.

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