

International Science Forum on Computational Toxicology
Research Triangle Park, NC
May 21-23, 2007

**A quantitative understanding of dynamic
cellular processes during detoxification
in human hepatocytes**

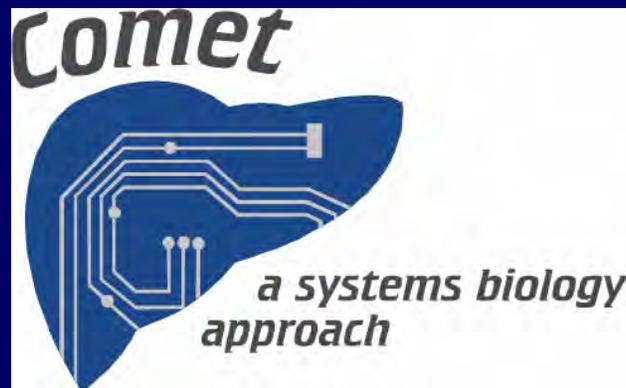


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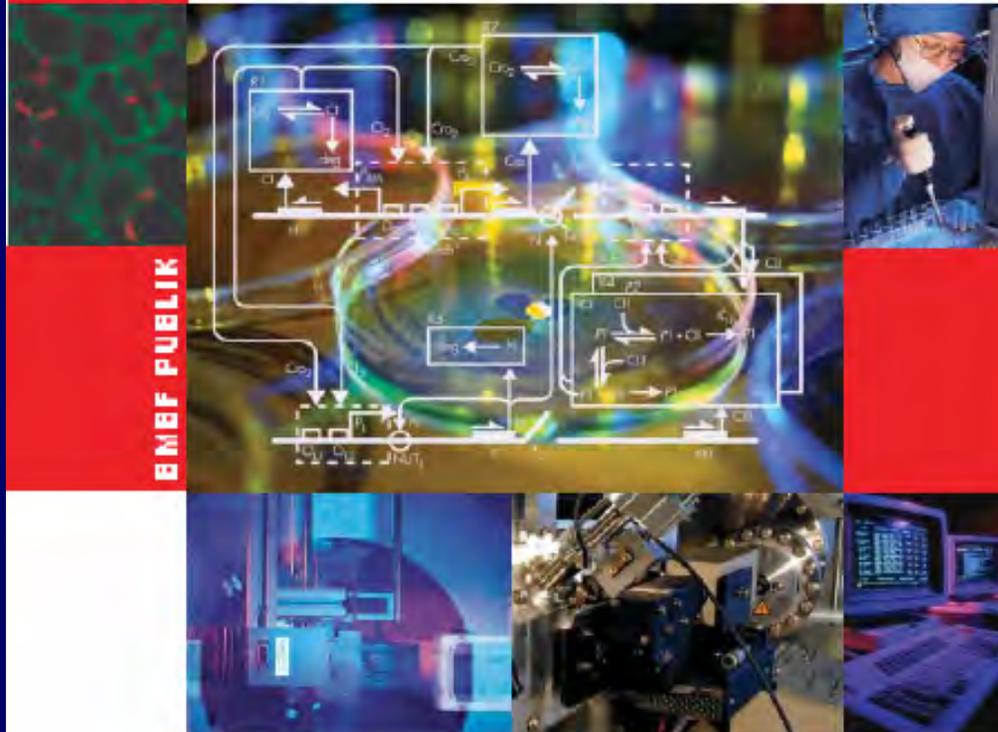
Network Stuttgart

A Systems Biology Approach to Detoxification Processes in Hepatocytes



Systems of Life

Systems Biology

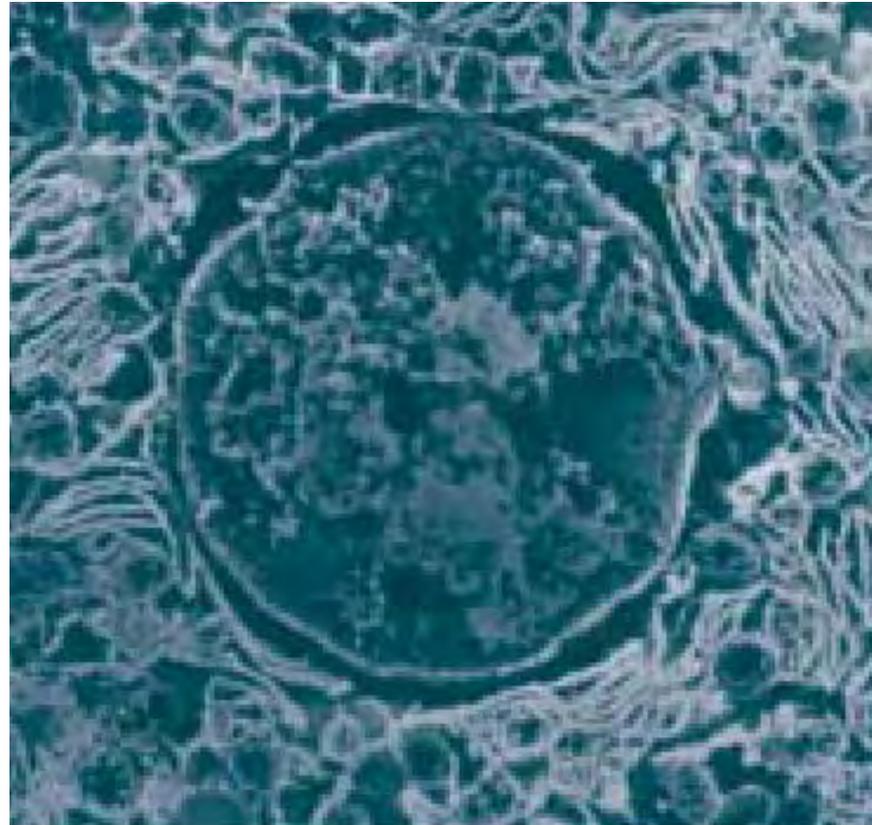


BMRF PUBLIK



Network Systems Biology

HepatoSys



*Research under the Systems of Life –
Systems Biology programme will focus
on the hepatocyte system*

Hepatosys



International Steering Committee

Coordination management

Networks

Platforms

Network Detoxification

Platform Cellbiology

Network Endocytosis

Platform Modeling

Network Iron Regulation

Network Regeneration

DETOXIFICATION

Holistic analysis, mathematical modelling and simulation of the detoxification system of the human hepatocyte

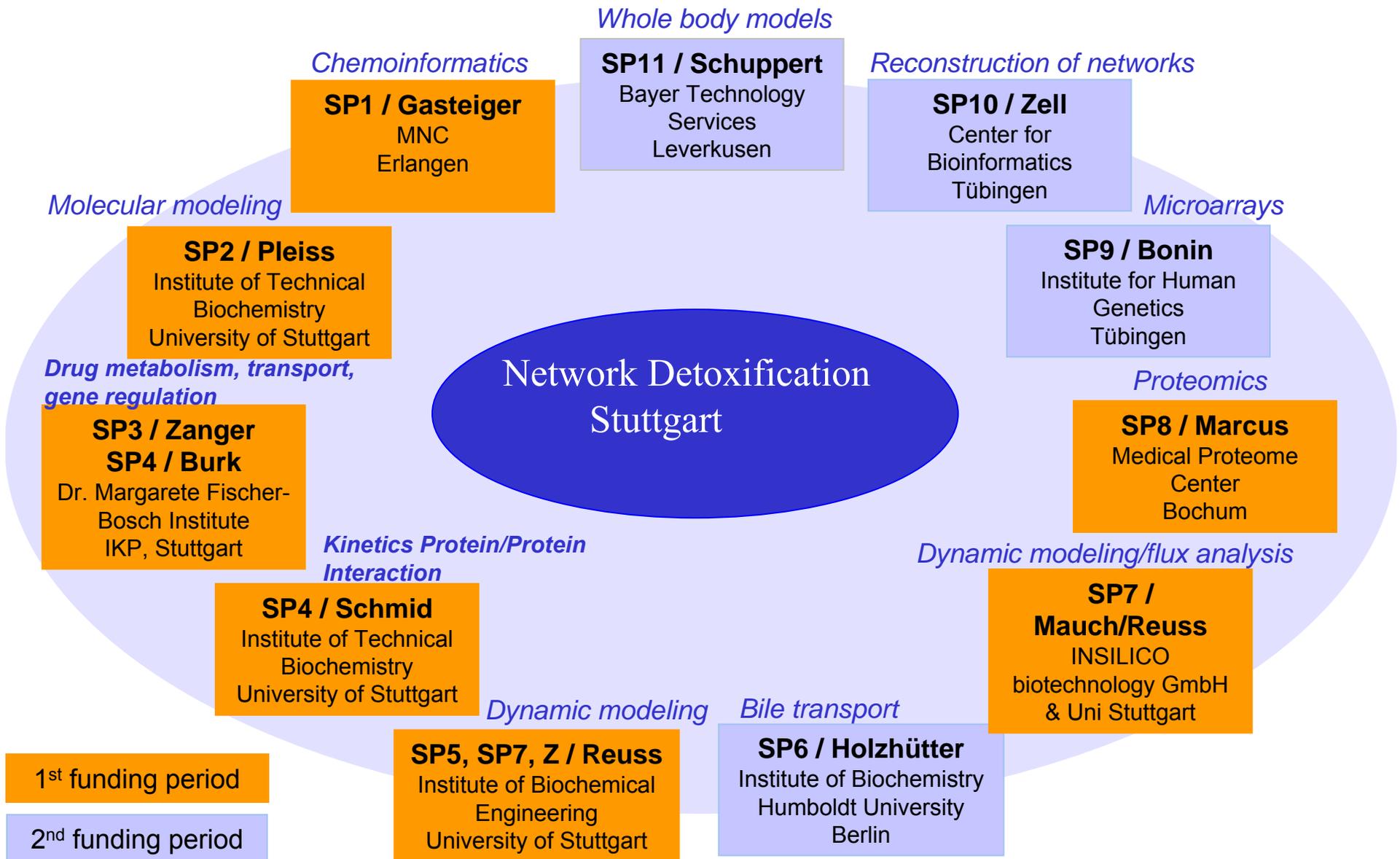
12 research projects: Universities Stuttgart (IBVT, ITB, ISA), Tübingen (Microarray Facility, Centre of Bioinformatics) Erlangen (Chemoinformatics), Bochum (Proteomics), Dr. Margarete – Bosch Institute Clinical Pharmacology, Stuttgart, Charitee/Humboldt-University Berlin, INSILICO biotechnology, Bayer Technology Service)

Chemoinformatic, Molecular Modeling, Dynamic Modeling of Detoxification,

Microarrays, Proteomics, Metabolomics

Modeling and simulation of signaling, regulatory and metabolic networks

Projectpartners Network Detoxification, Stuttgart 2007 - 2009



The diagram features a large, light purple rectangular box with a 3D effect. Inside this box, there are two smaller rectangular boxes side-by-side. The left one is dark blue with white text, and the right one is white with a black border and black text. A blue curved arrow points from the top of the left box to the top of the right box. A white curved arrow points from the bottom of the right box to the bottom of the left box. Below these two boxes, centered, is the text 'Optimal Experimental Design'.

**Experimental
Data**

***In silico* Analysis**

Optimal Experimental Design

**Dynamic modeling and simulation
of the drug metabolizing
P450 network**

By placing the entire integrated network, rather than individual CYPs at the centre of dynamic analysis, a systems behaviour may emerge which differs from the reductionistic kinetics of individual enzymes.

ROBUSTNESS/FRAGILITY

STRUCTURAL AND DYNAMIC ROBUSTNESS

„Robustness enables the system to maintain its **functionality** against **external** and **internal** perturbations“ (*Kitano, 2004*)

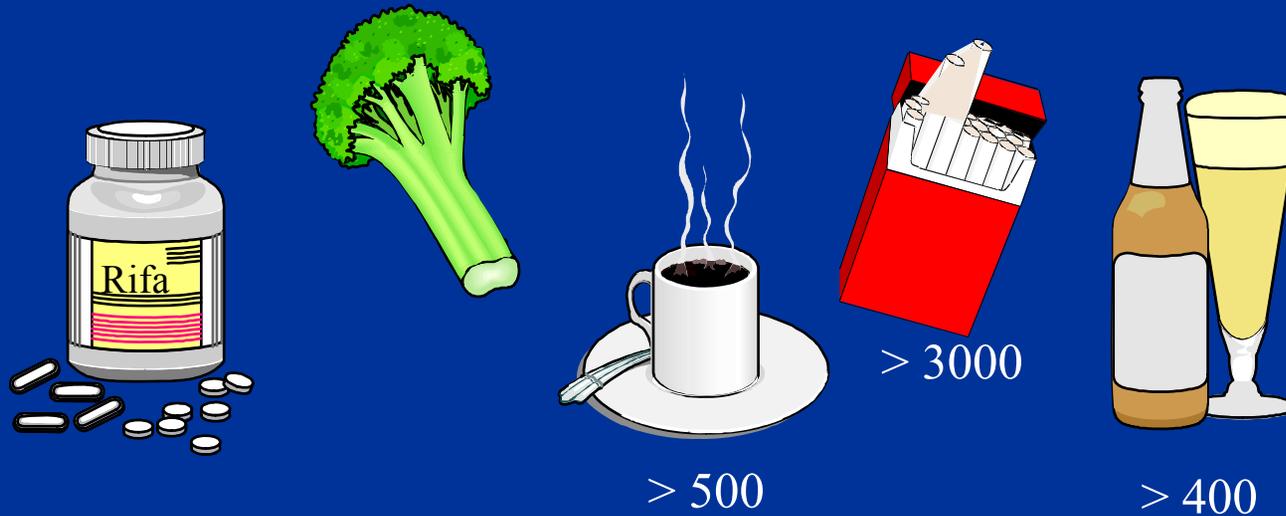


Functionality: Drug detoxification

External perturbation: multitude of substrates (varying concentrations and drugs)

Internal perturbation: different enzyme expression levels; polymorphism, inter-individual variability **→ phenotype plasticity**

Drug Metabolism: First Line of Defense Against Xenobiotics



Biosphere:

~ 3 million substances

Industry:

~ 3000 chemicals/year

Drugs:

~ 6000 (WHO)

Daily intake of xenobiotics: ~ 10.000 substances;

External perturbation

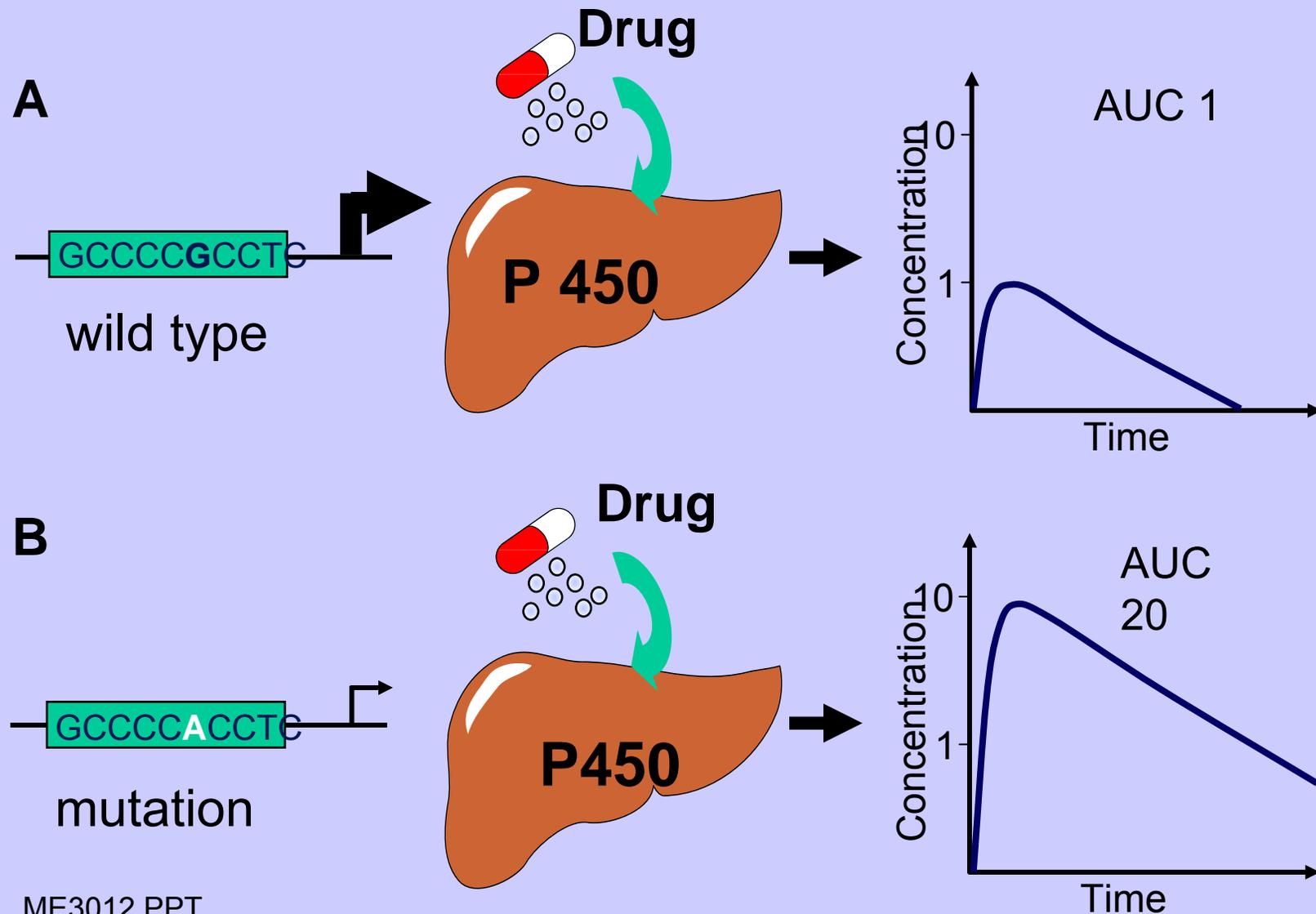
Pharmacogenetics

Individual variability in the efficacy and
toxicity

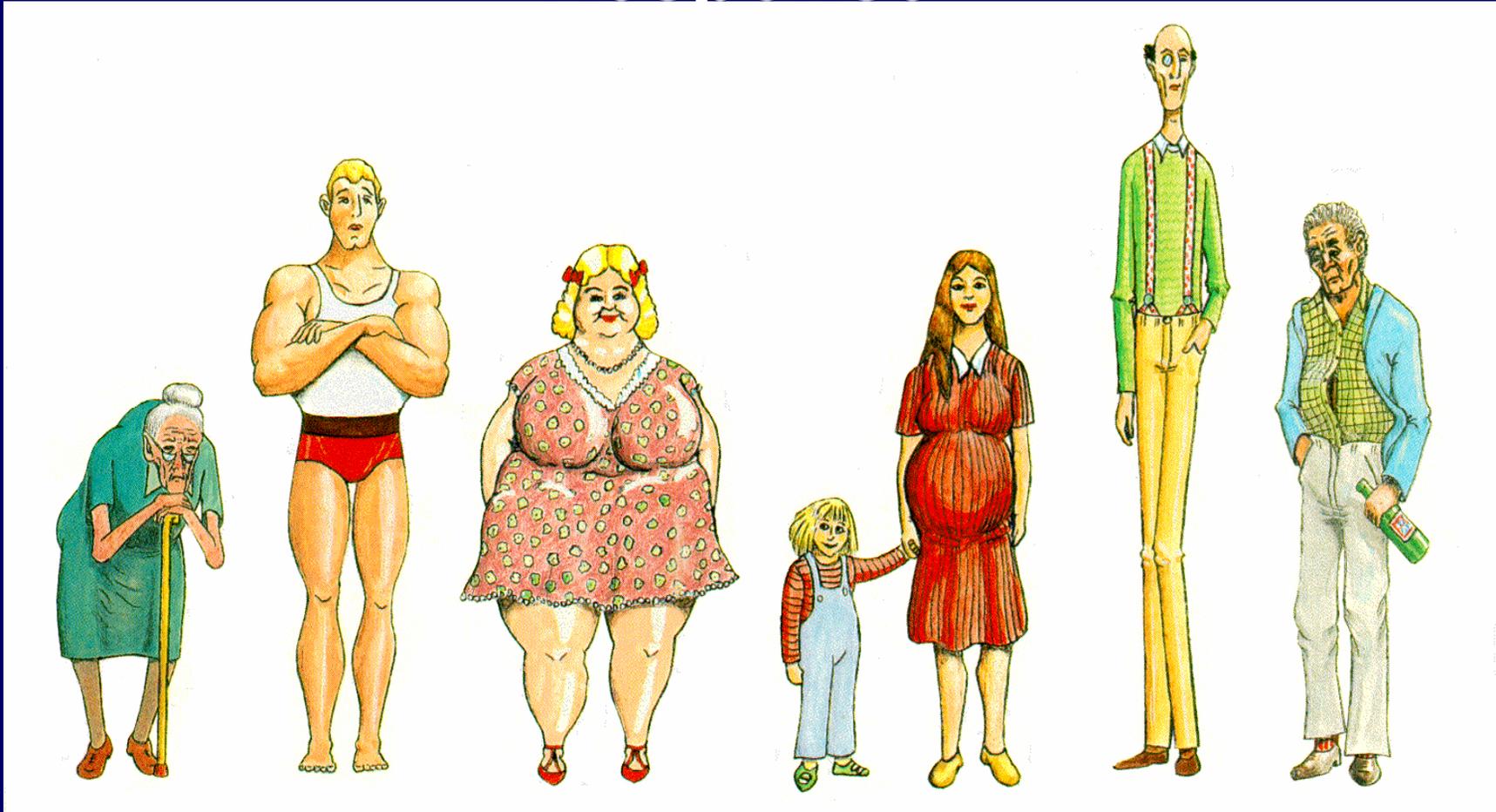
of drugs due to polymorphisms of genes
involved in their disposition and action

Mechanism of Genetic Variability in Drug Response

Same dose but different plasma concentrations



Nongenetic Factors and Drug Response

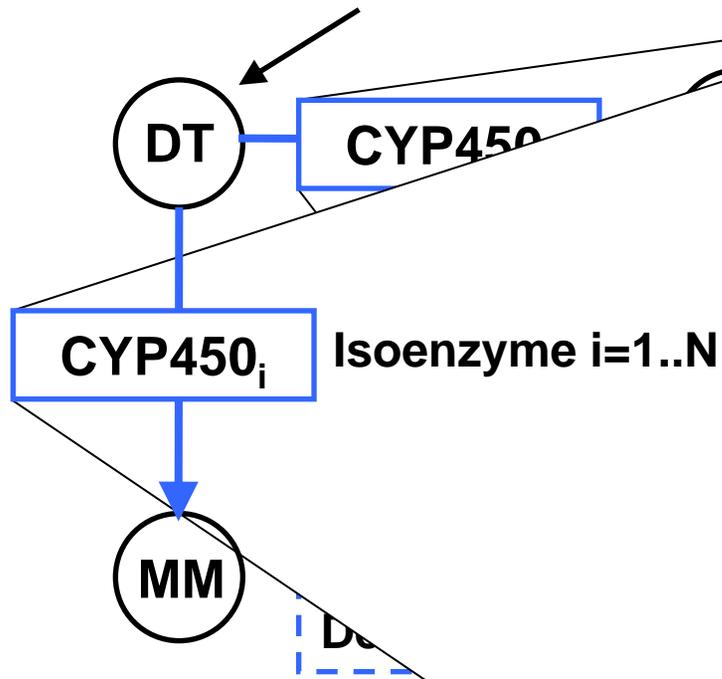


Age, body weight, sex, disease, diet, alcohol, smoking, drugs, hepatic and renal function

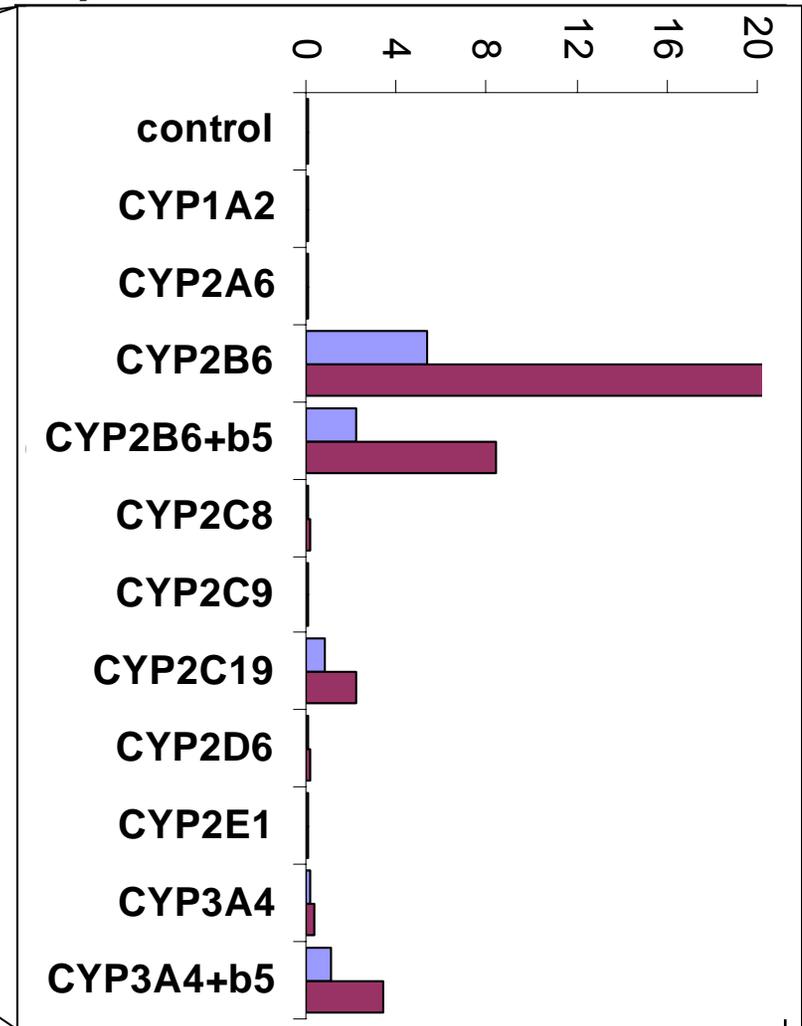


Modeling: Pathway Identification

Substrate Dextromethorphan (DT)

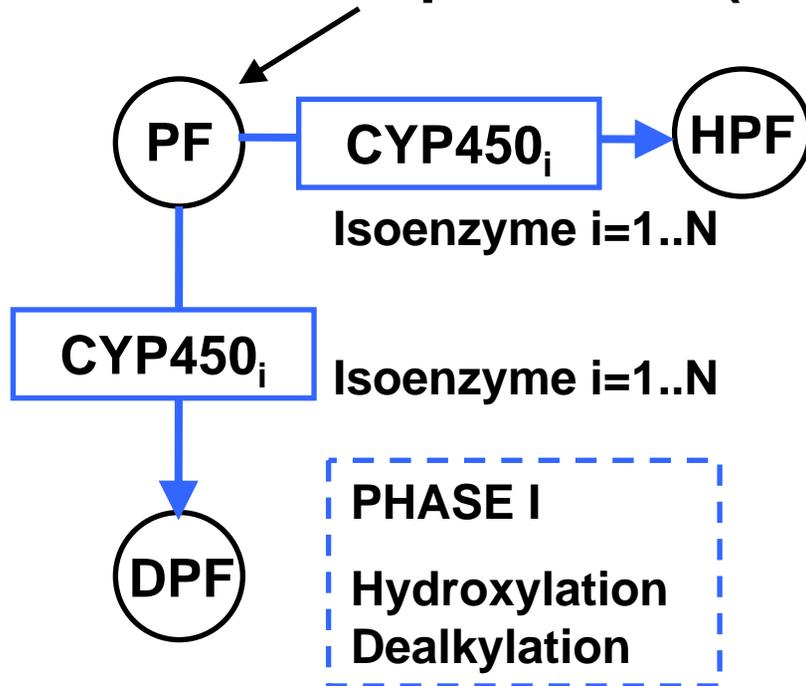


DT: Dextromethorphan
DX: Dextrorphan
MM: Methoxymorphinan



Modeling: Pathway Identification

Substrate Propafenone (PF)



PF: Propafenone
DPF: Desalkyl-Propafenone
HPF: 5-OH-Propafenone

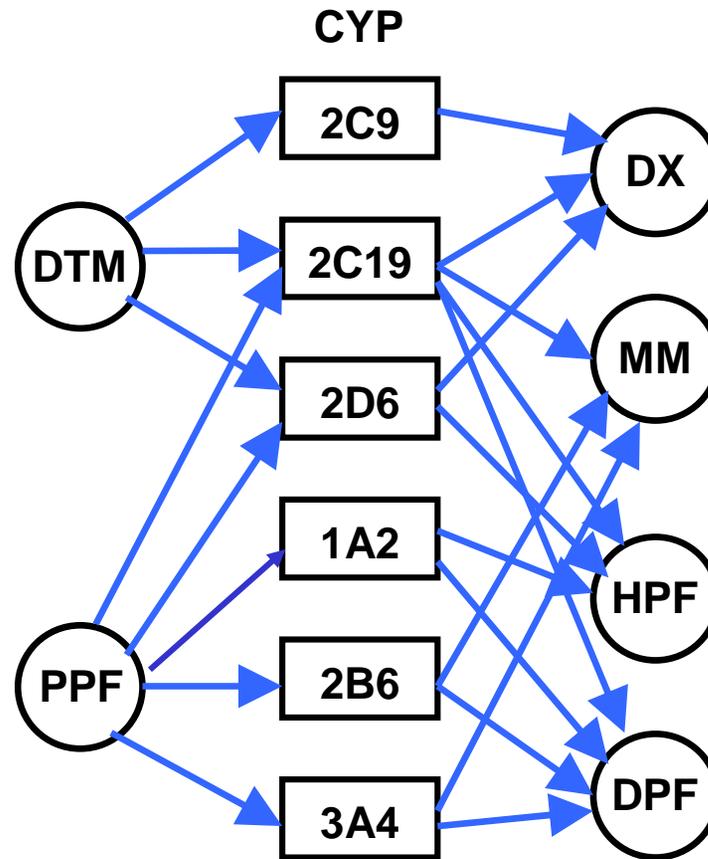
CYP450: Cytochrome P450
monooxygenase

Model Set-up

**Multireactant
isoenzyme system**

Dextromethorphan

Propafenone



Model: Set-up

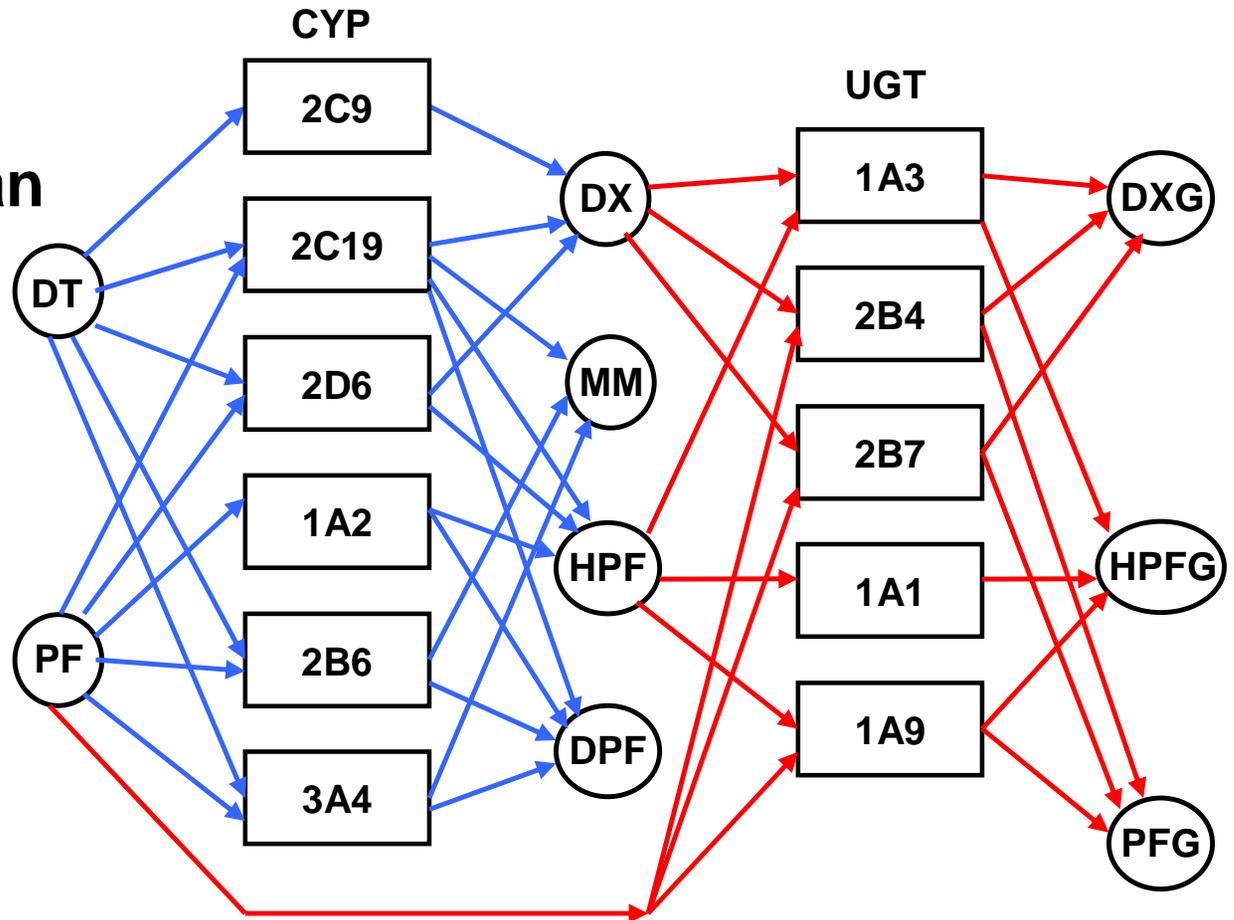
**Multireactant
isoenzyme system**

PHASE I
Dealkylation/
Hydroxylation

PHASE II
Glucuronidation

Dextromethorphan

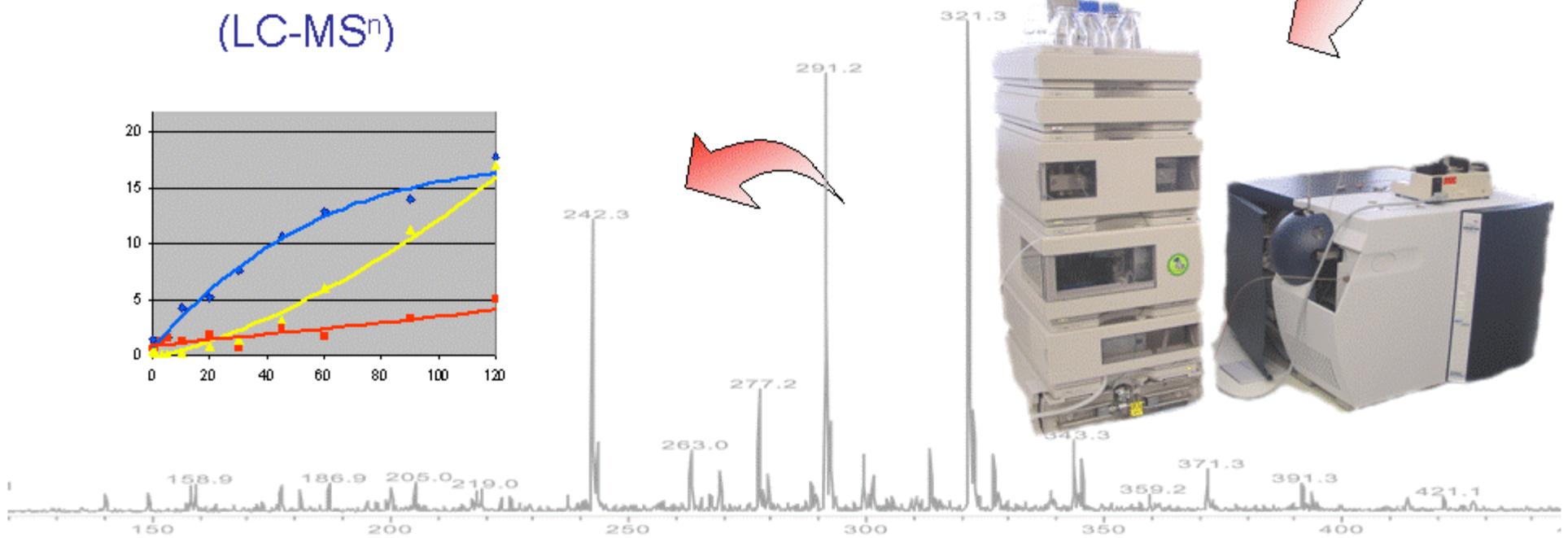
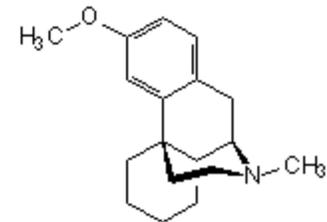
Propafenone



Quantitative LC-MS/MS Analysis of Drugs and Metabolites

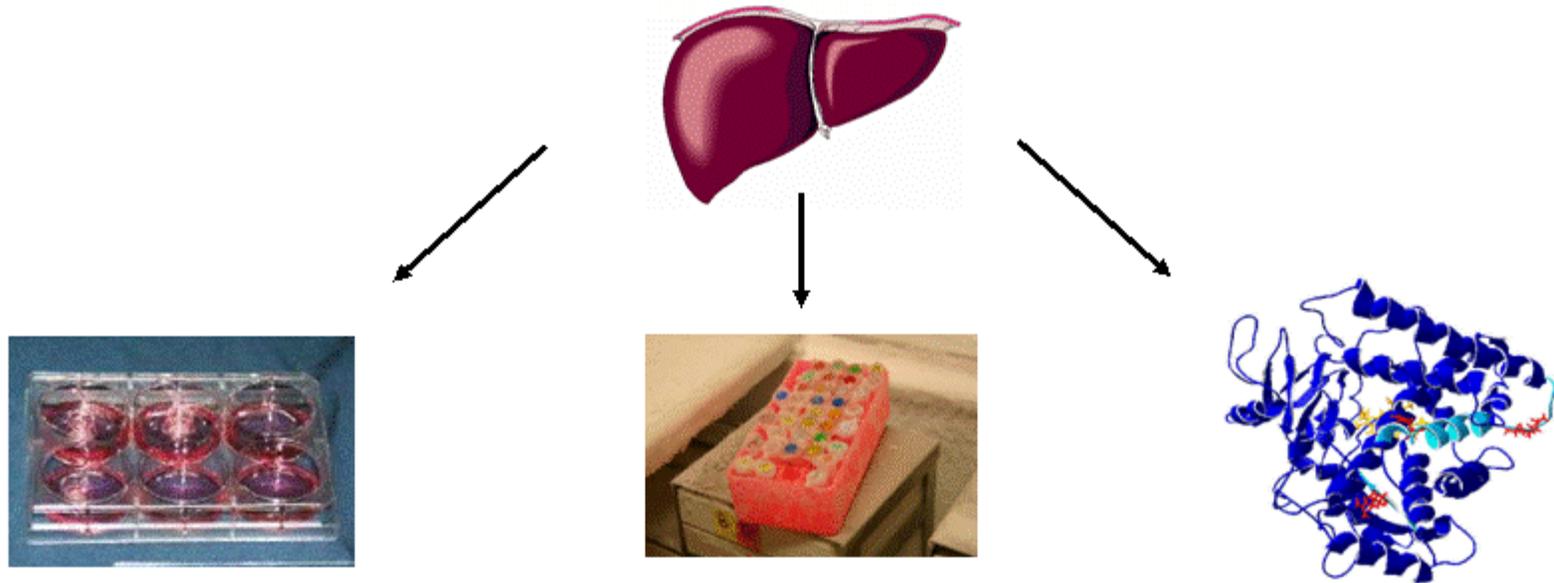
Zanger et al., Clinical Pharmacology, Rober Bosch, Stuttgart

- synthesis of reference compounds and of **stable isotope-labeled analogs**
- method development and validation for quantification from various biological sources
- simultaneous quantitative analysis of drugs and metabolites (phase I / II) by LC-MS/MS
- structure determination of unknown metabolites (LC-MSⁿ)



experimental approaches

complementary approach to human liver

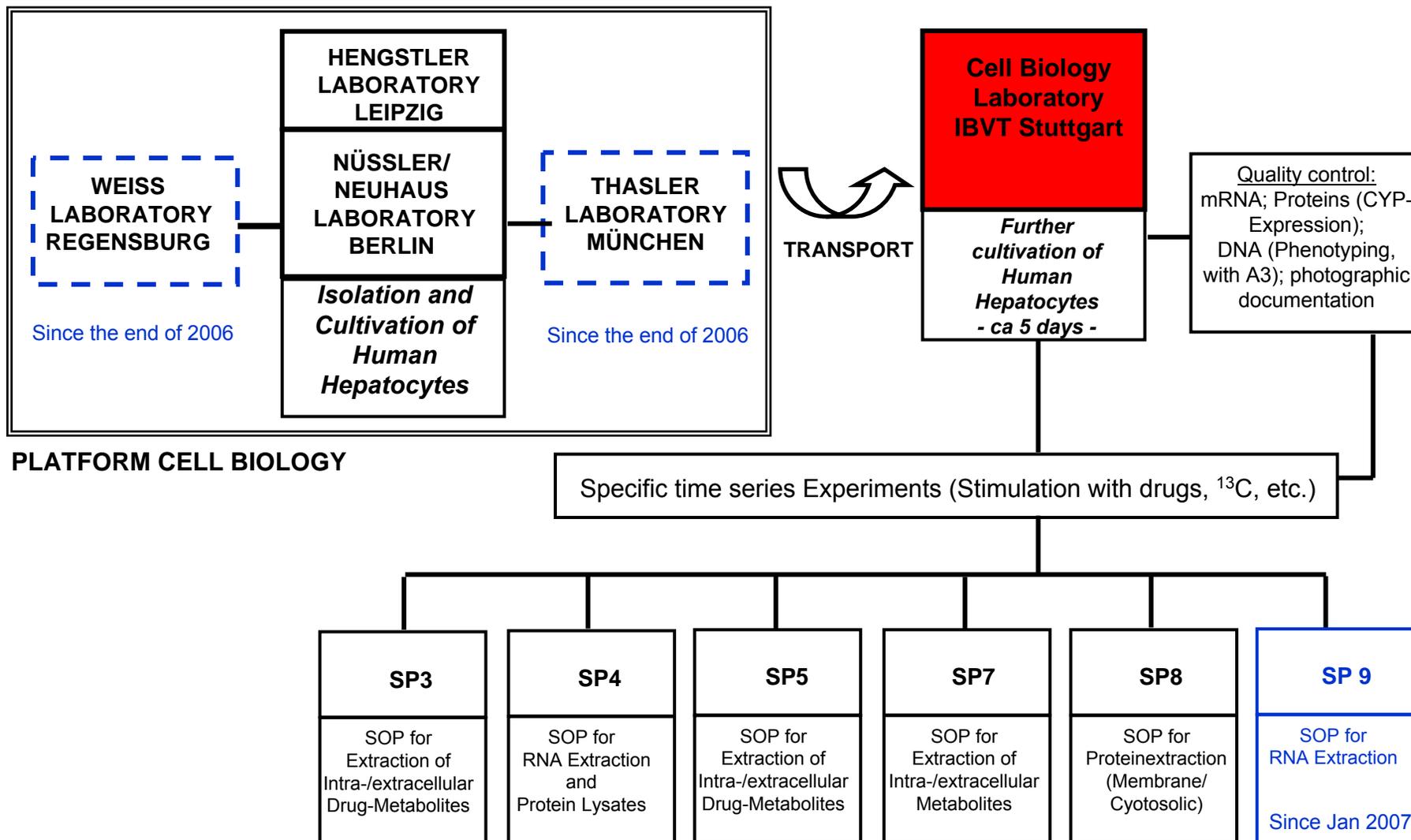


primary hepatocytes

liver tissue bank (N=300)

recombinant systems

Experimental work with primary human hepatocytes - logistics



Mathematical Model

Dynamic Balance Equations

$$\frac{dc_j}{dt} = \sum_i v_{i,j} r_{i,j}(t) = f(c(t), p) \quad i = 1..N \quad \text{Enzymes}, j = \text{Metabolite}$$

Reaction rates: irreversible MM with multiple competitive inhibitions through alternative substrates

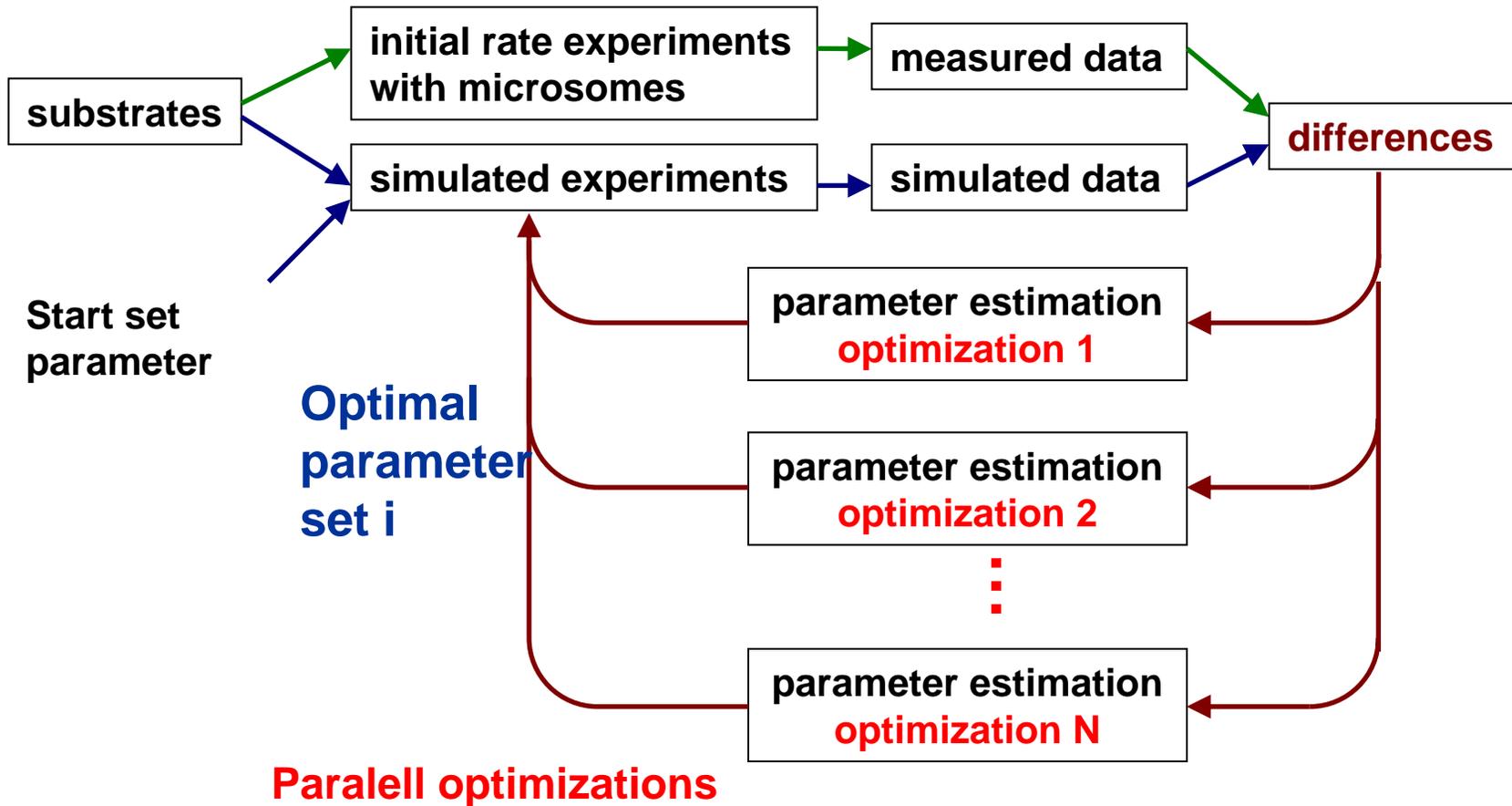
$$r_{S1 \rightarrow P1}^{Enz} = \frac{r_{max, S1 \rightarrow P1}^{Enz} \frac{C_{S1}}{K_{M, S1 \rightarrow P1}^{Enz}}}{1 + \sum_j \frac{C_{S1}}{K_{M, S1 \rightarrow Pj}^{Enz}} + \sum_k \frac{C_{S2}}{K_{M, S2 \rightarrow Pk}^{Enz}}}$$

$$r_{max, S1 \rightarrow P1}^{Enz} = k_{S1 \rightarrow P1}^{Enz} \cdot C^{ENZ}$$

Quantitative Proteomics (labeled peptides, MPC Bochum)

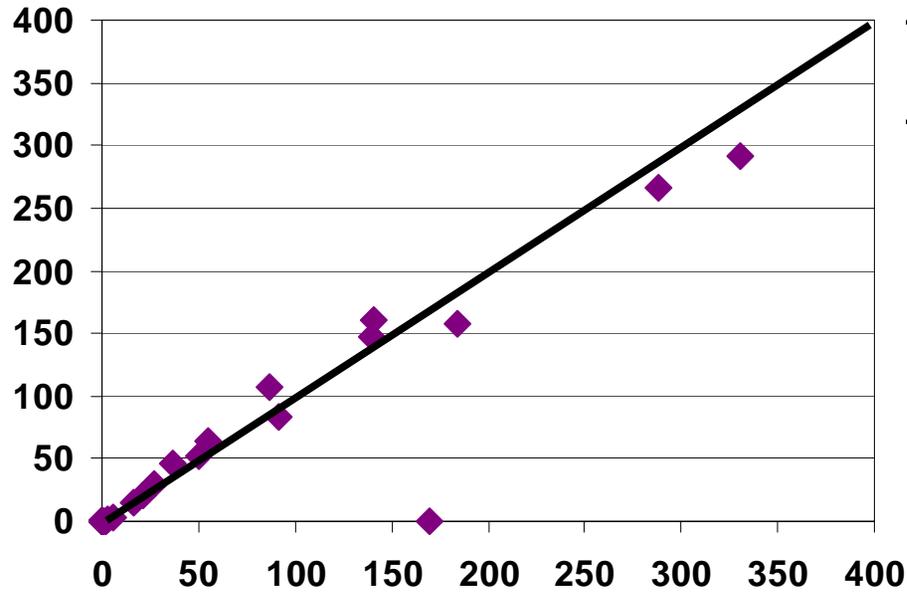
Parameter Identification

Optimization workflow

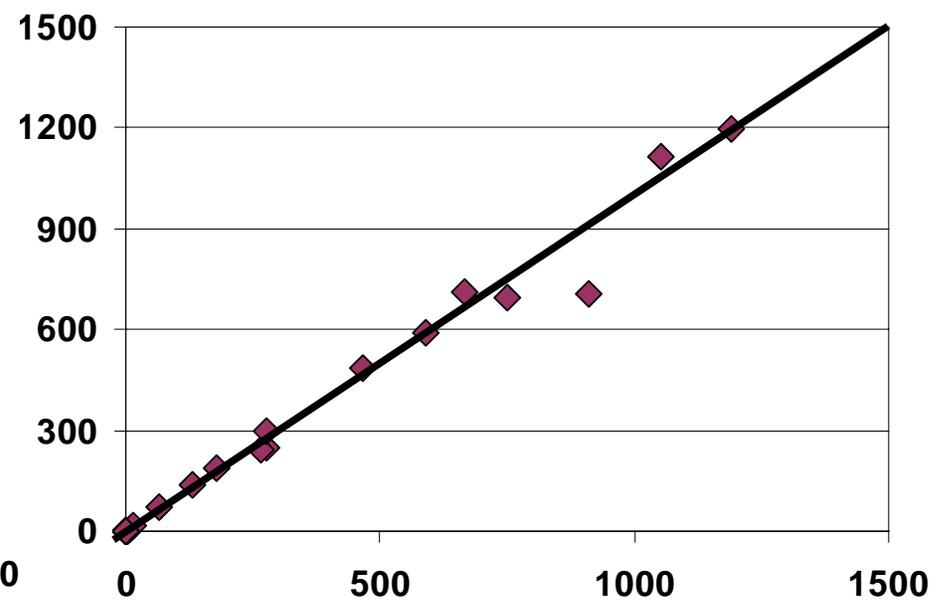


Parameter Identification

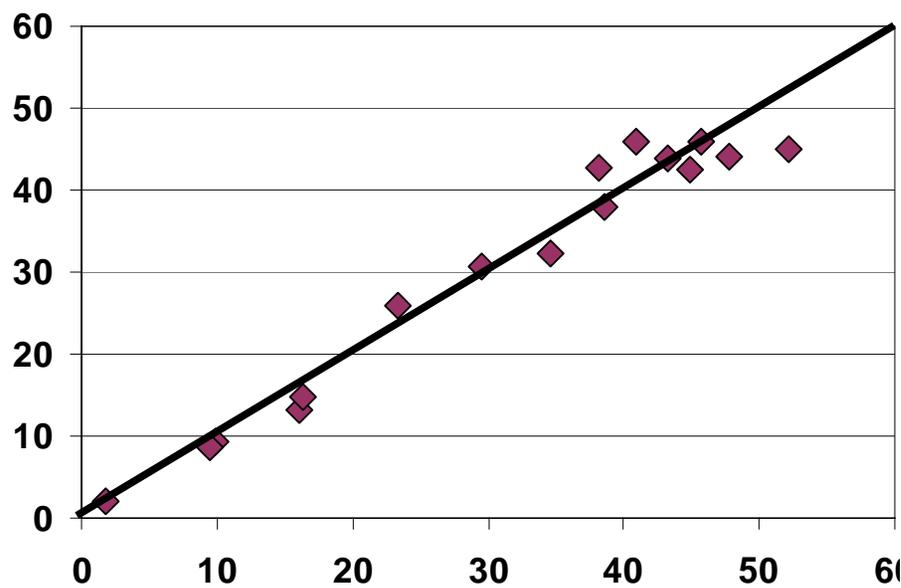
Dextrorphan



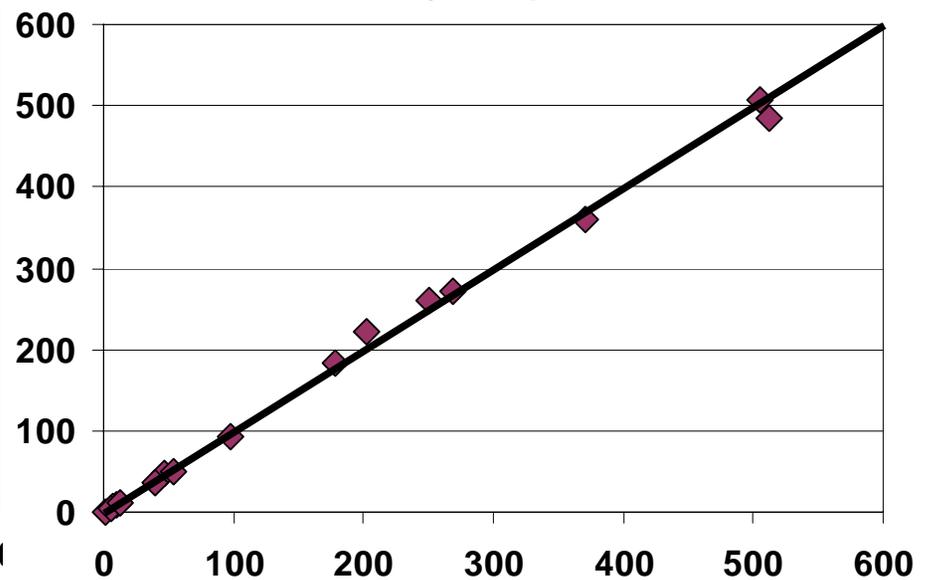
Methoxymorphinan



Hydroxy-Propafenone

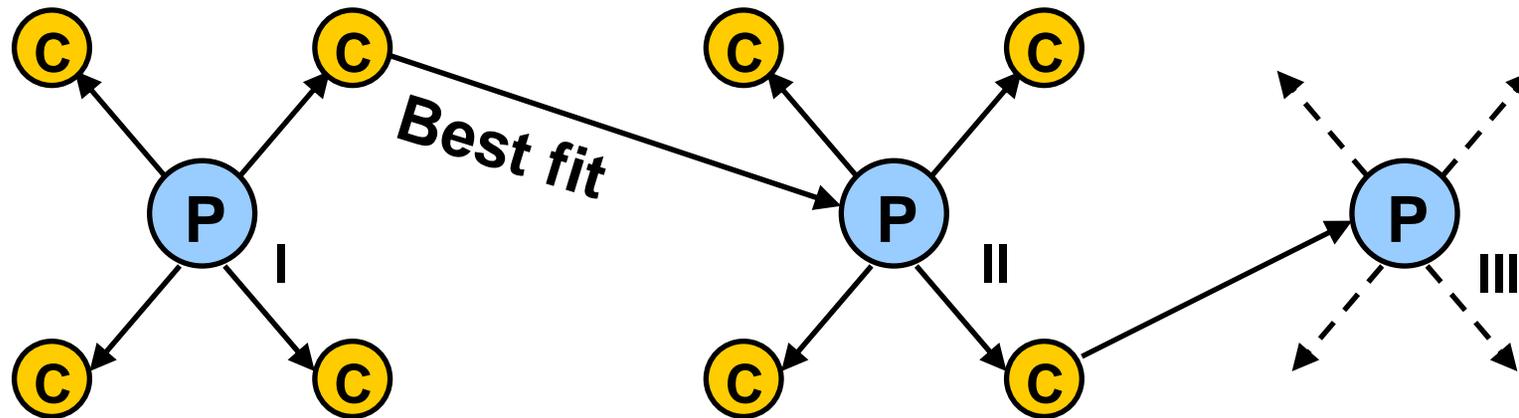


Desalkyl-Propafenone



Parameter Identification

Evolutionary Algorithm (JavaEva¹):
Generation of parameter set,
(c)hildren from (p)arents, via random process

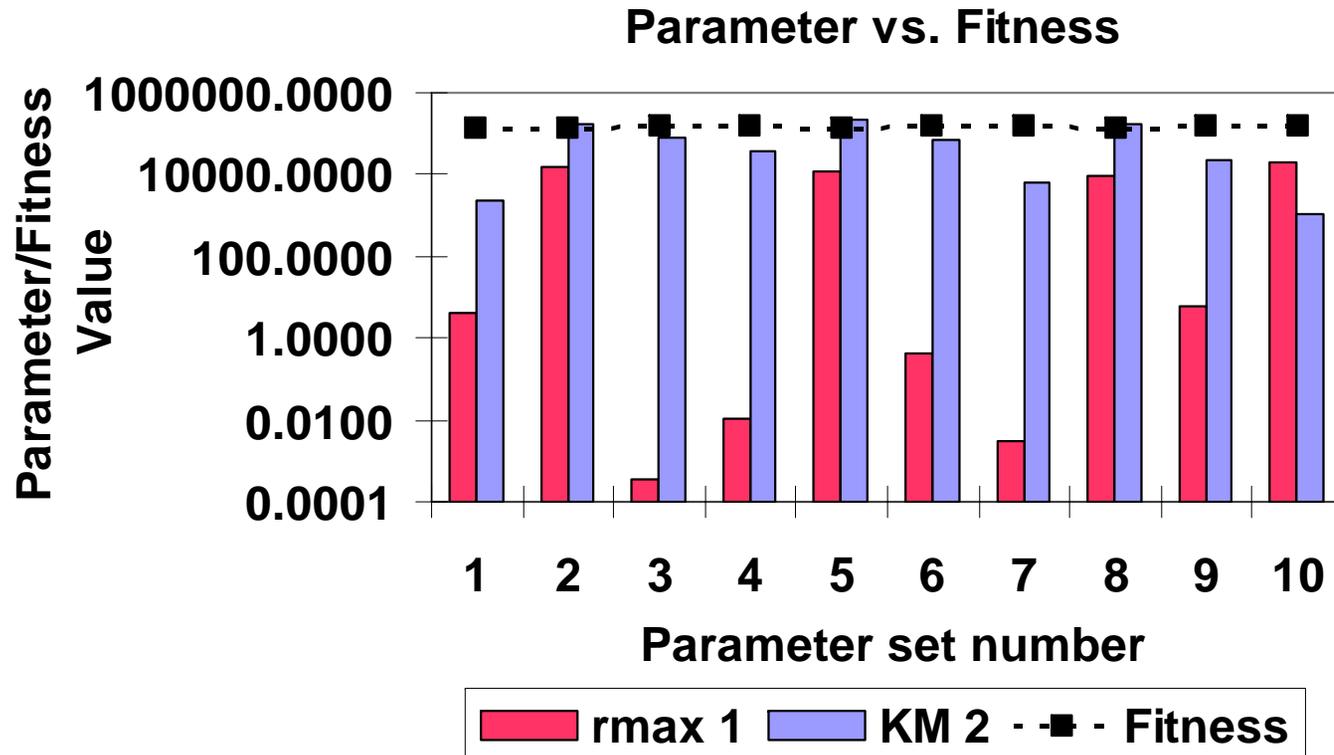


Parallel optimization with different random numbers (variation of parameters)

¹ Prof.A.Zell, Center of Bioinformatics, University of Tübingen

Parameter Quality

Isoenzyme Model



Total average of relative deviation of parameters

82 %

Average Fitness

143705.3

Parameter Quality

Isoenzyme Model

Parameter variation is very high

=> Indication of **Robustness** of the system

➤ **Model reduction feasible?**

Model Reduction

Superposition of
isoenzymes per
reaction, respectively

Number of parameters:

40 -> 22

Number of reactions:

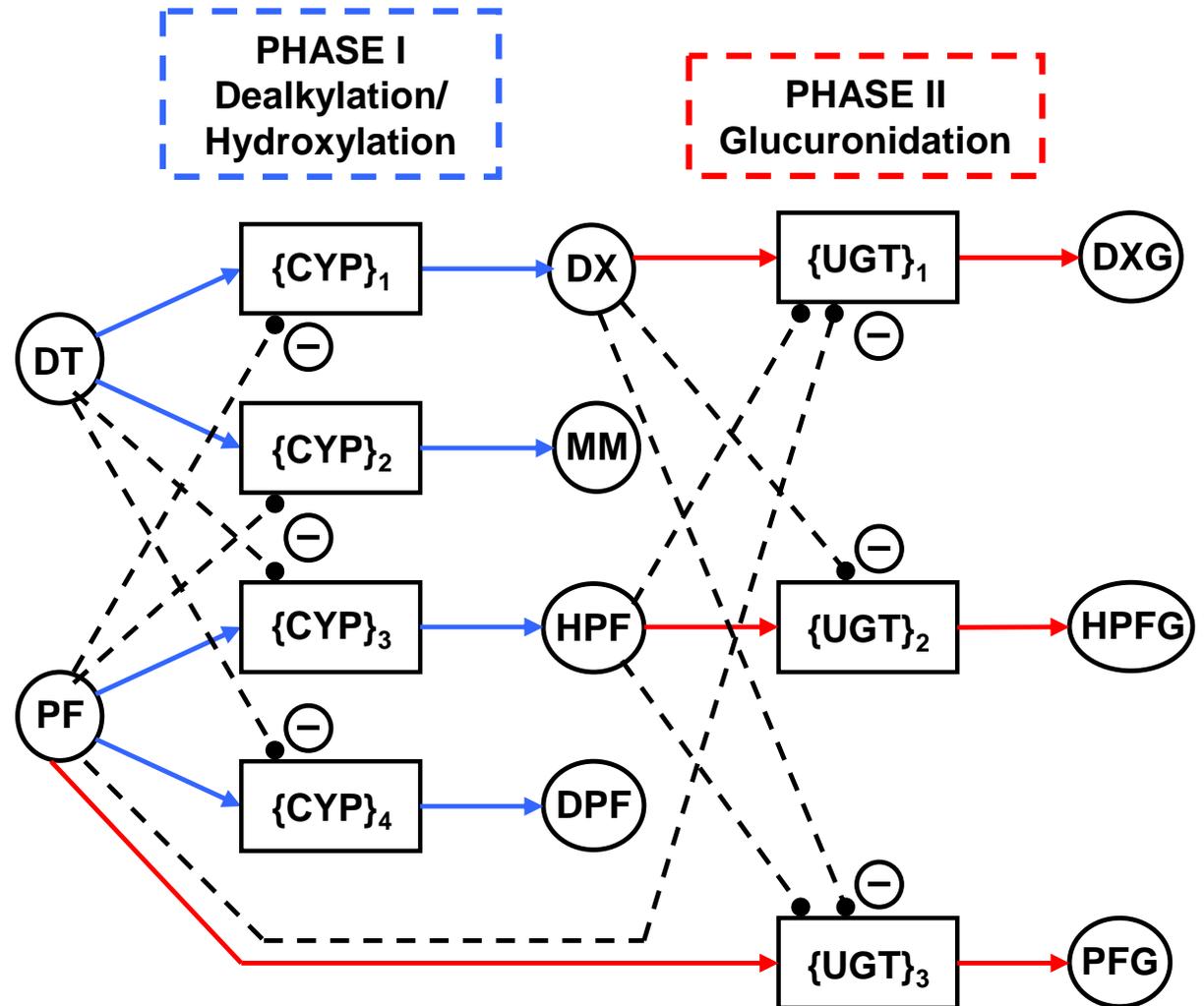
22 -> 7

Total average of relative
deviation of parameters:

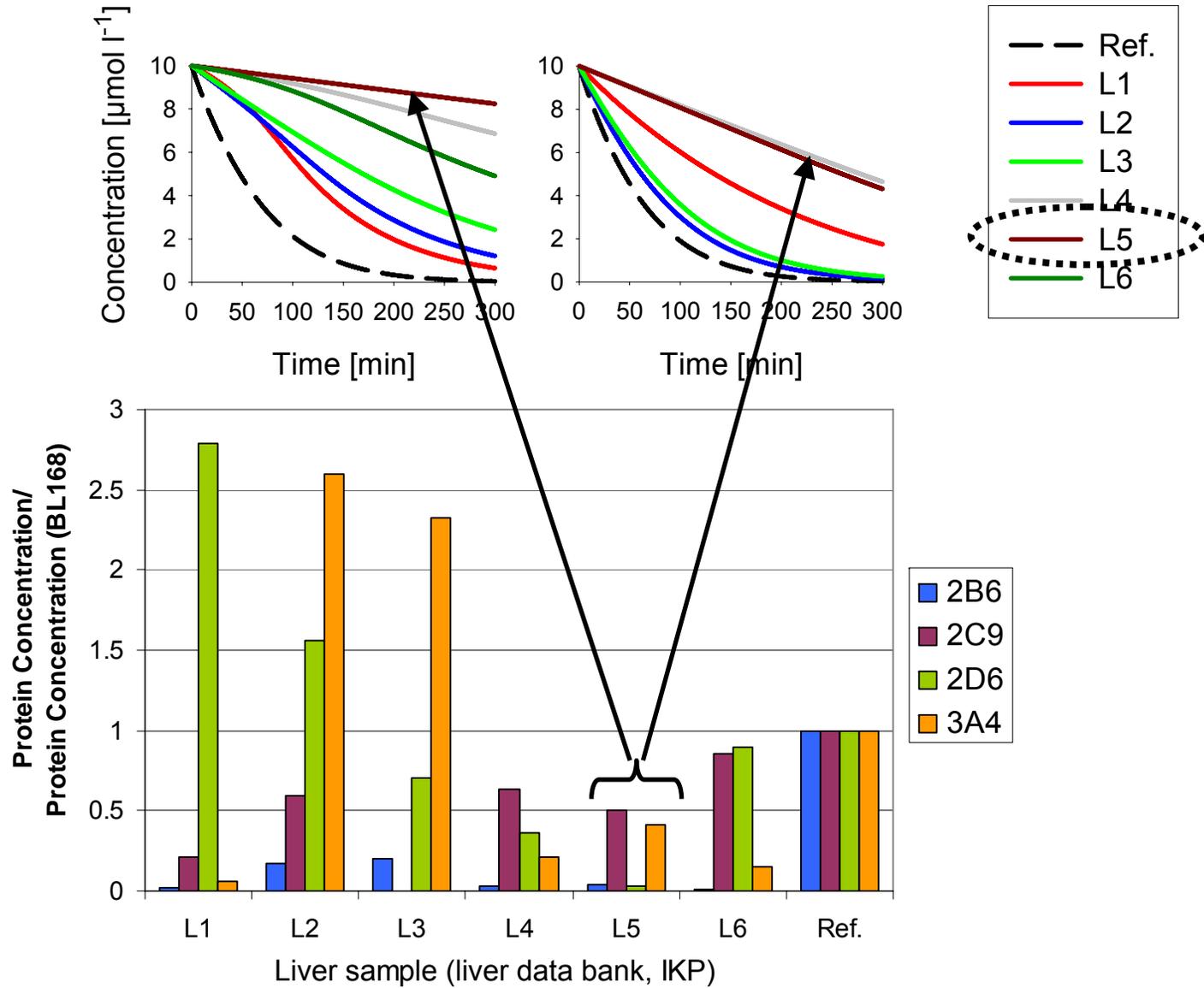
82 % -> 18%

Fitness

143705.3 -> 175253.4



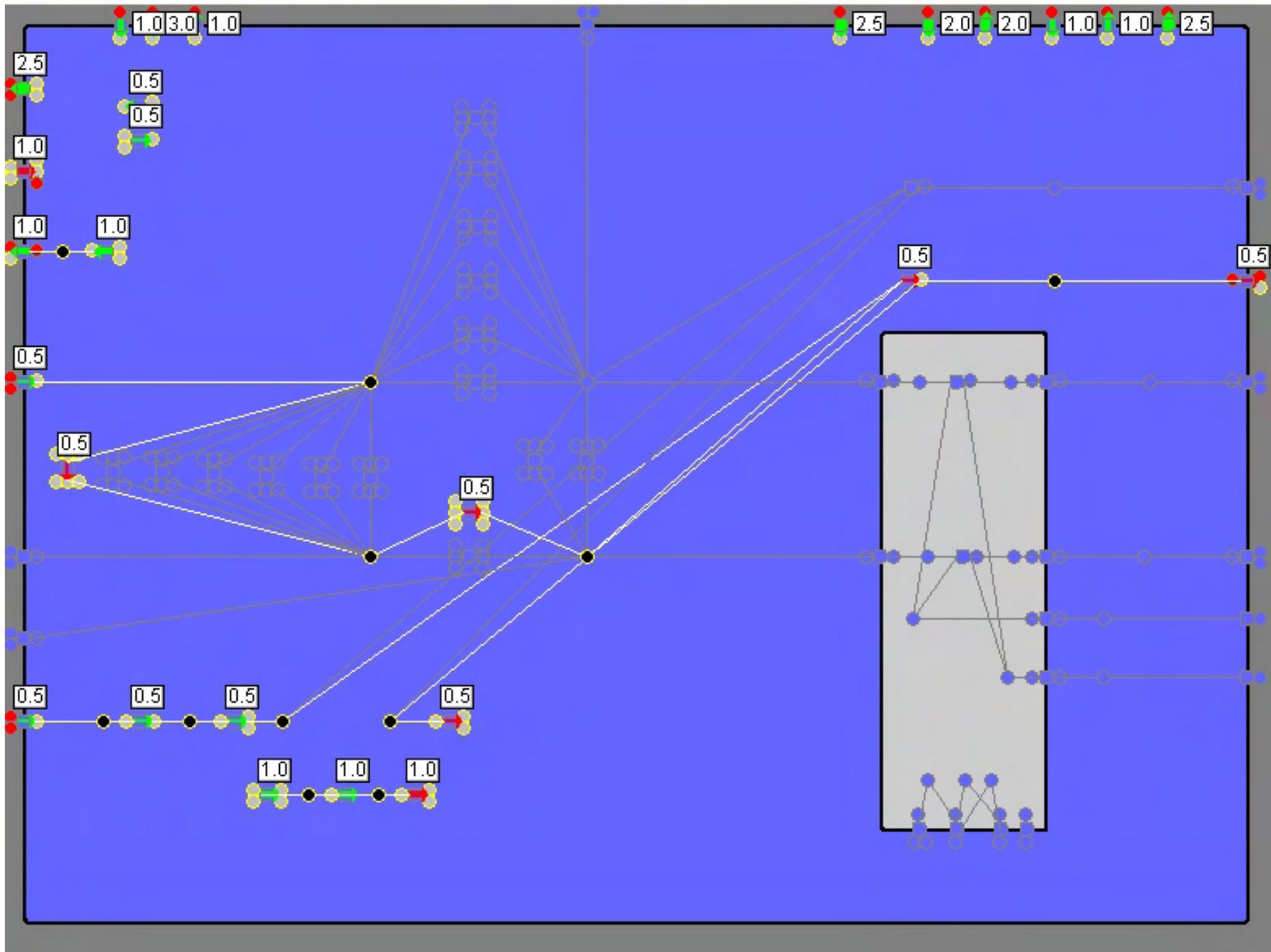
Dynamic Analysis: Drug Detoxification

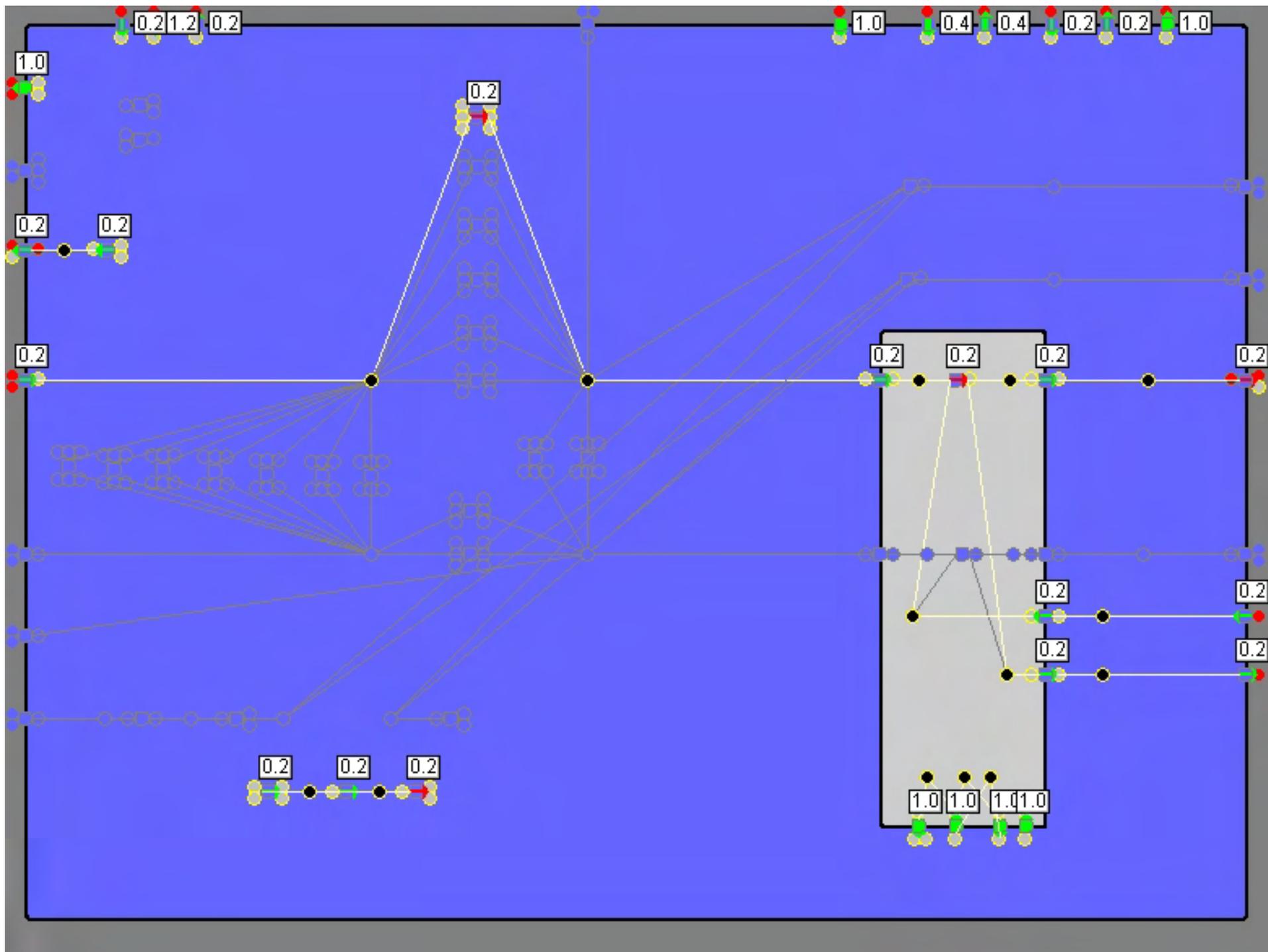


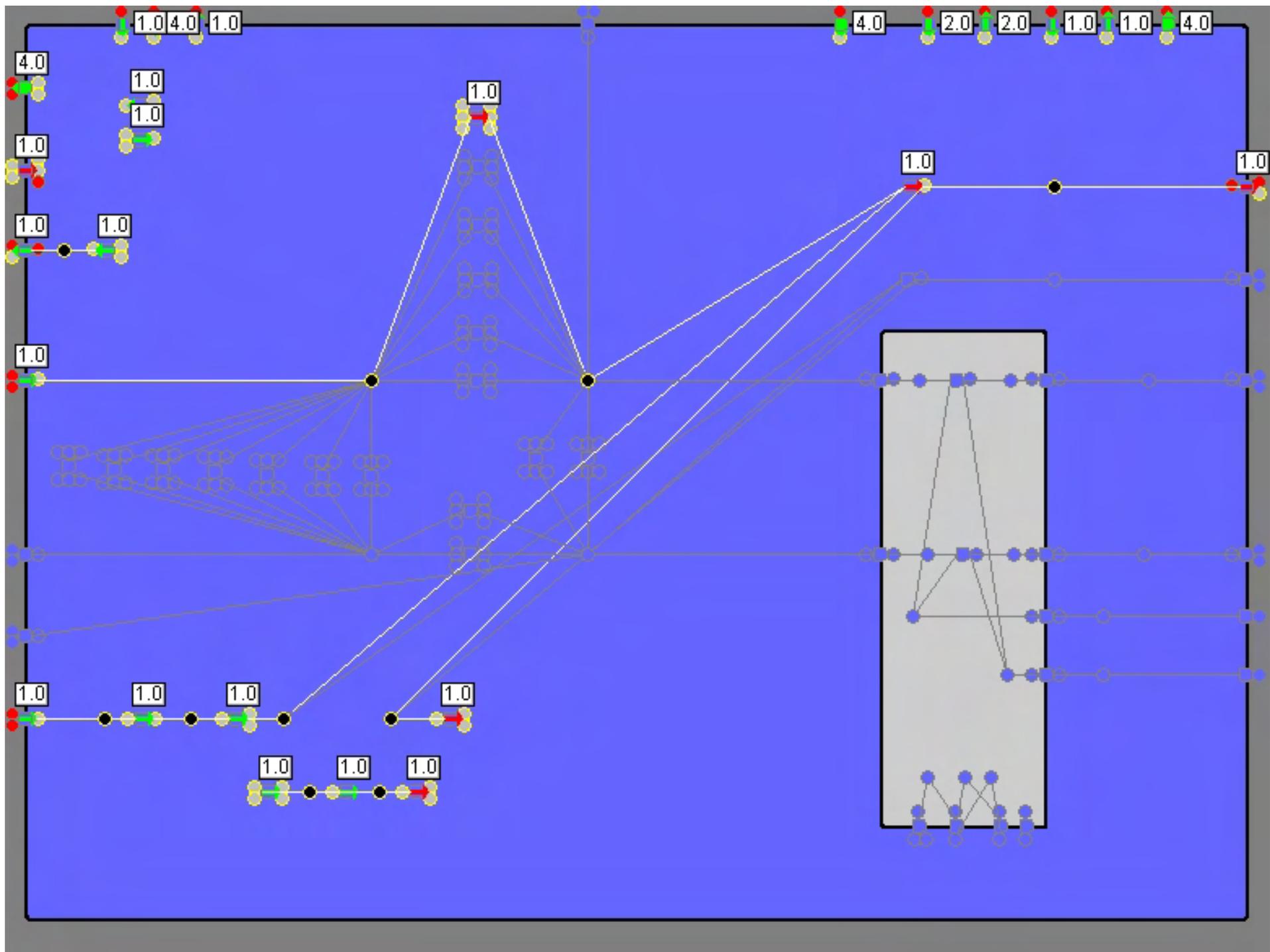
STRUCTURAL ROBUSTNESS

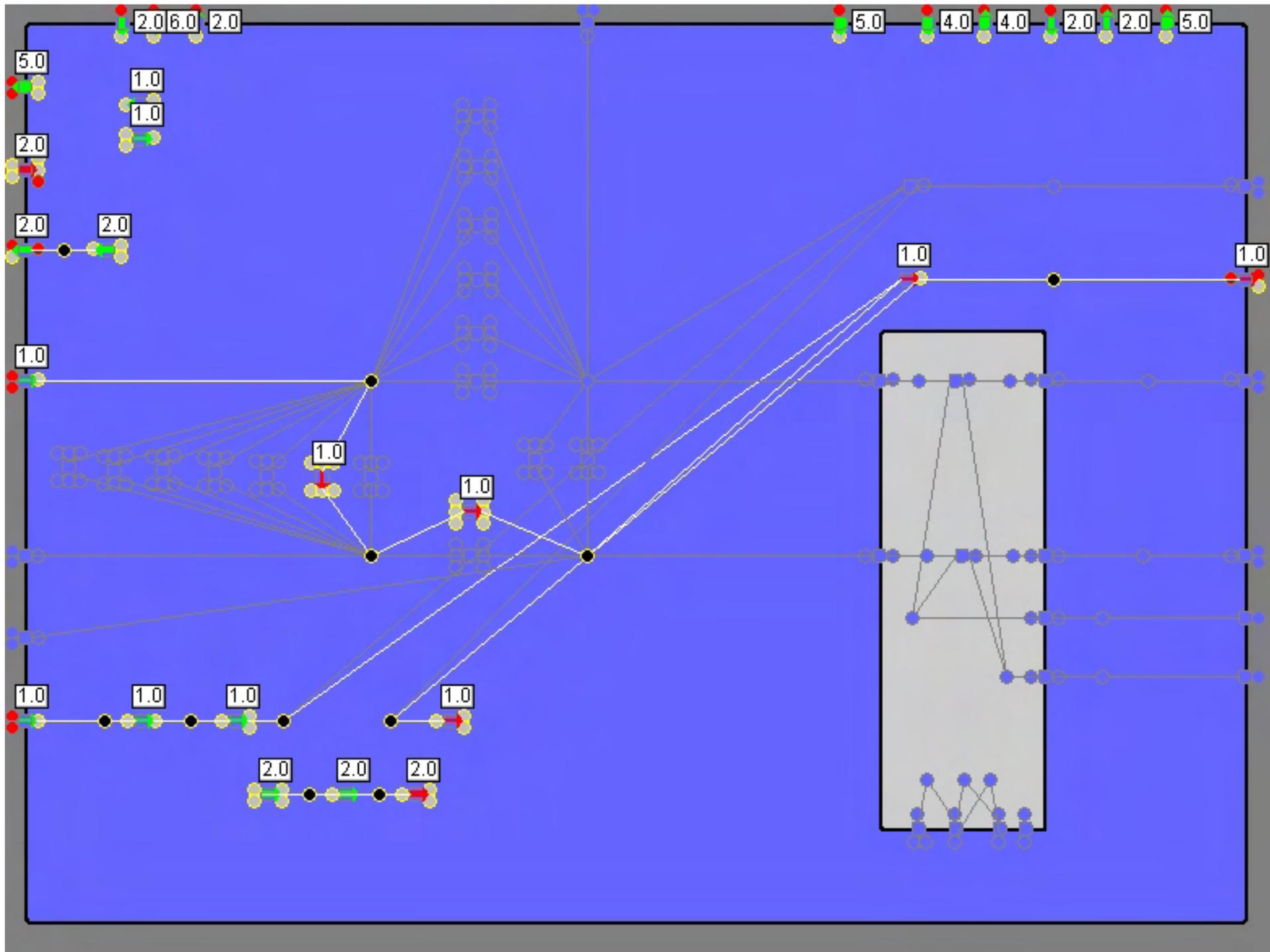
ELEMENTARY FLUX MODES

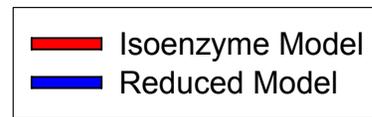
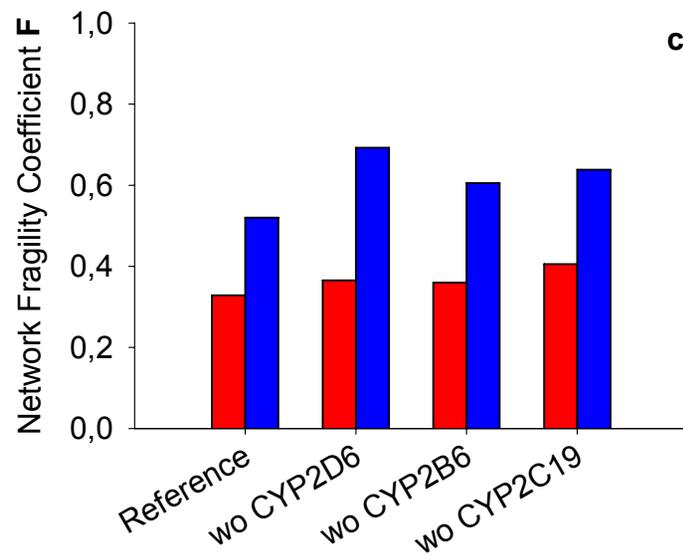
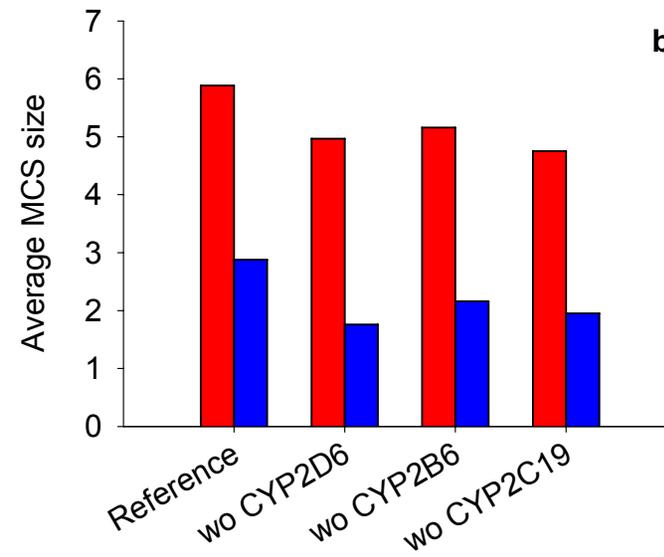
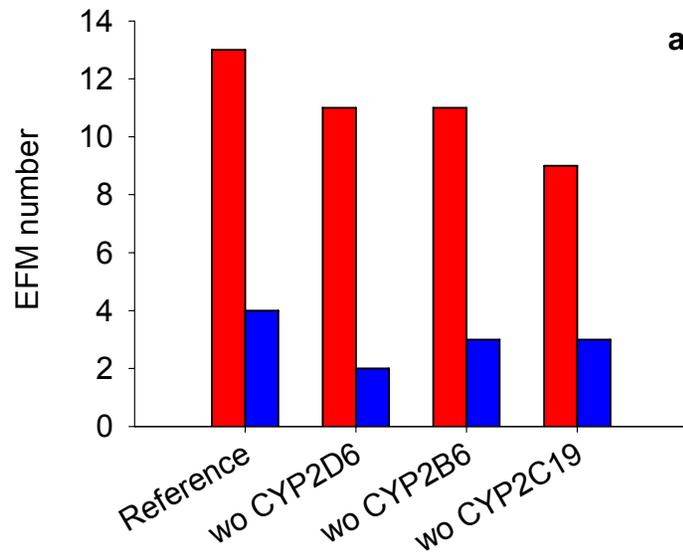
- Fulfil steady state condition
- Fulfil reversibility properties
- Cannot be decomposed into smaller modes (i.e. modes that involve less enzymes)





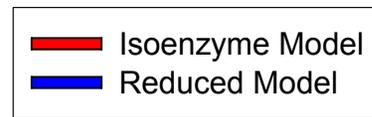
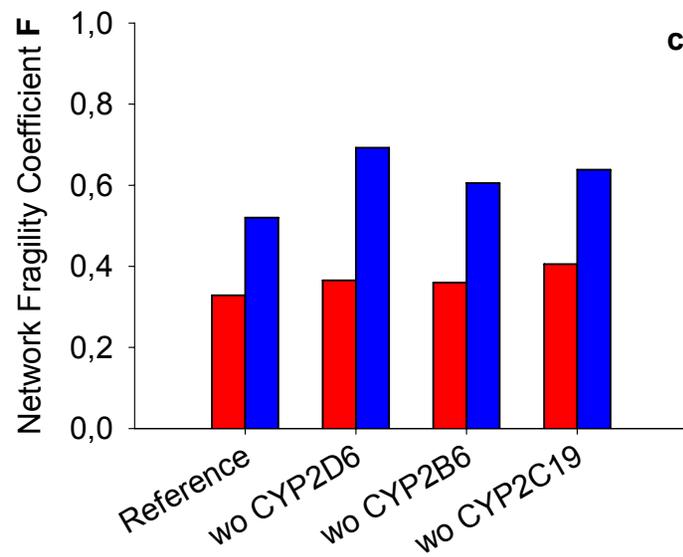
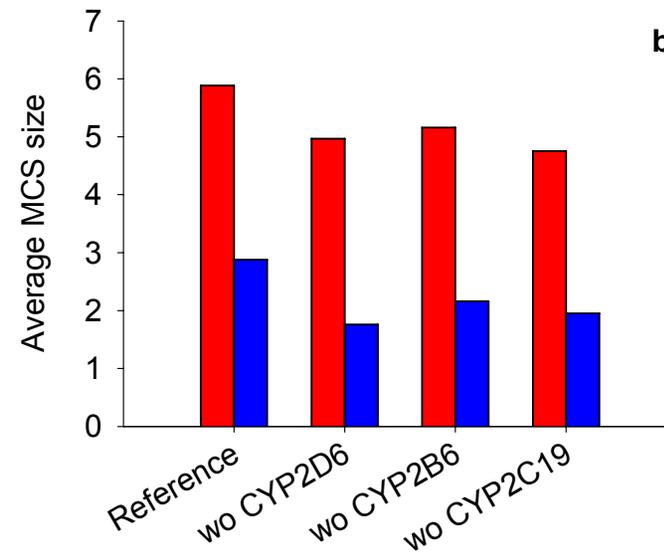
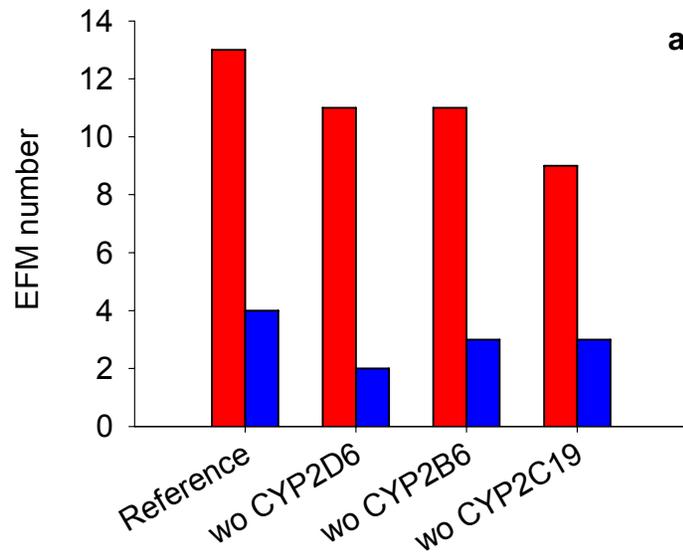






MINIMAL CUT SETS (MCS)

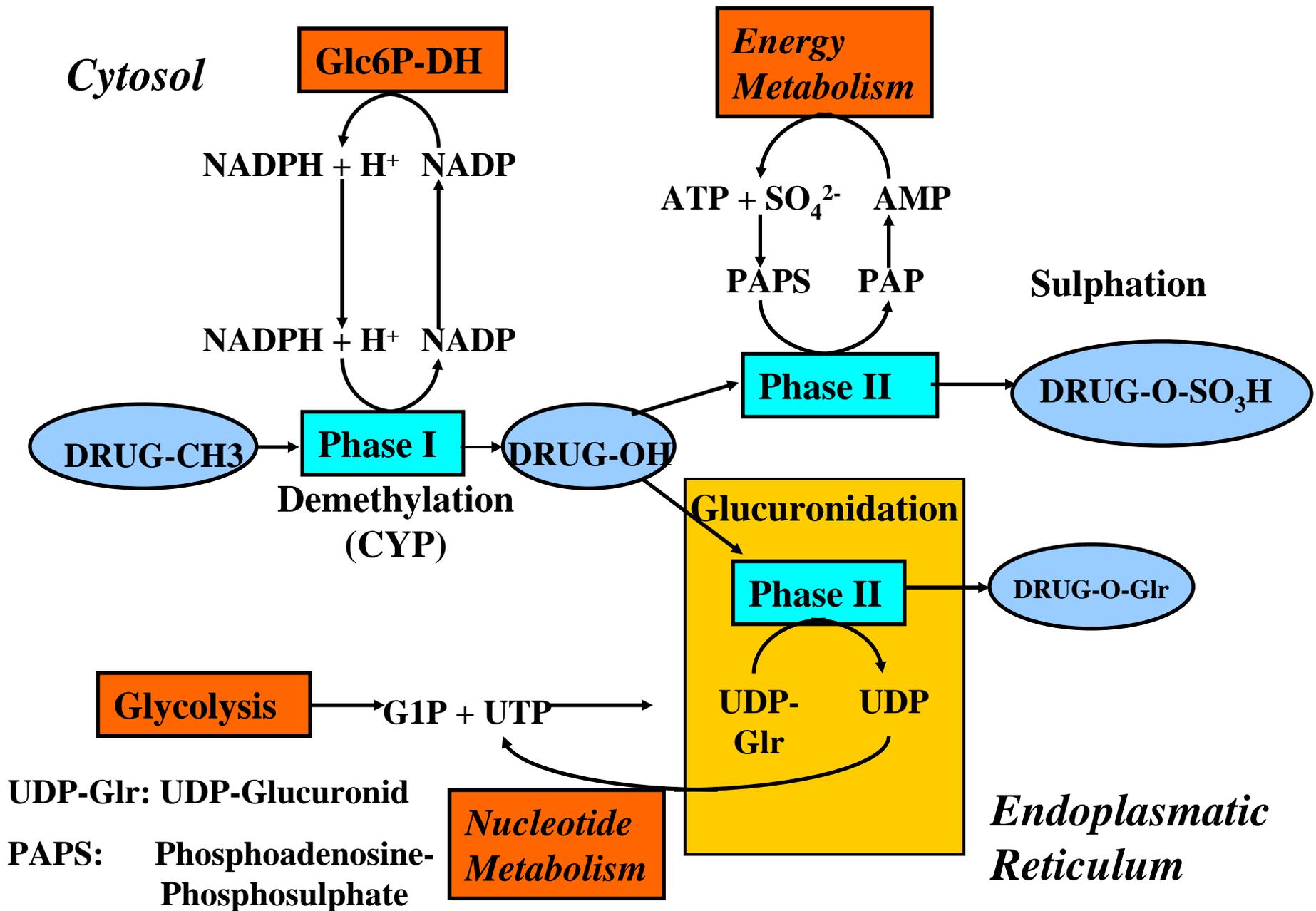
A MCS can be considered as a minimal set of events (loss of reactions) which – if these events occur together - leads to system failure, i.e. that the objective reactions cannot operate in a balanced fashion.

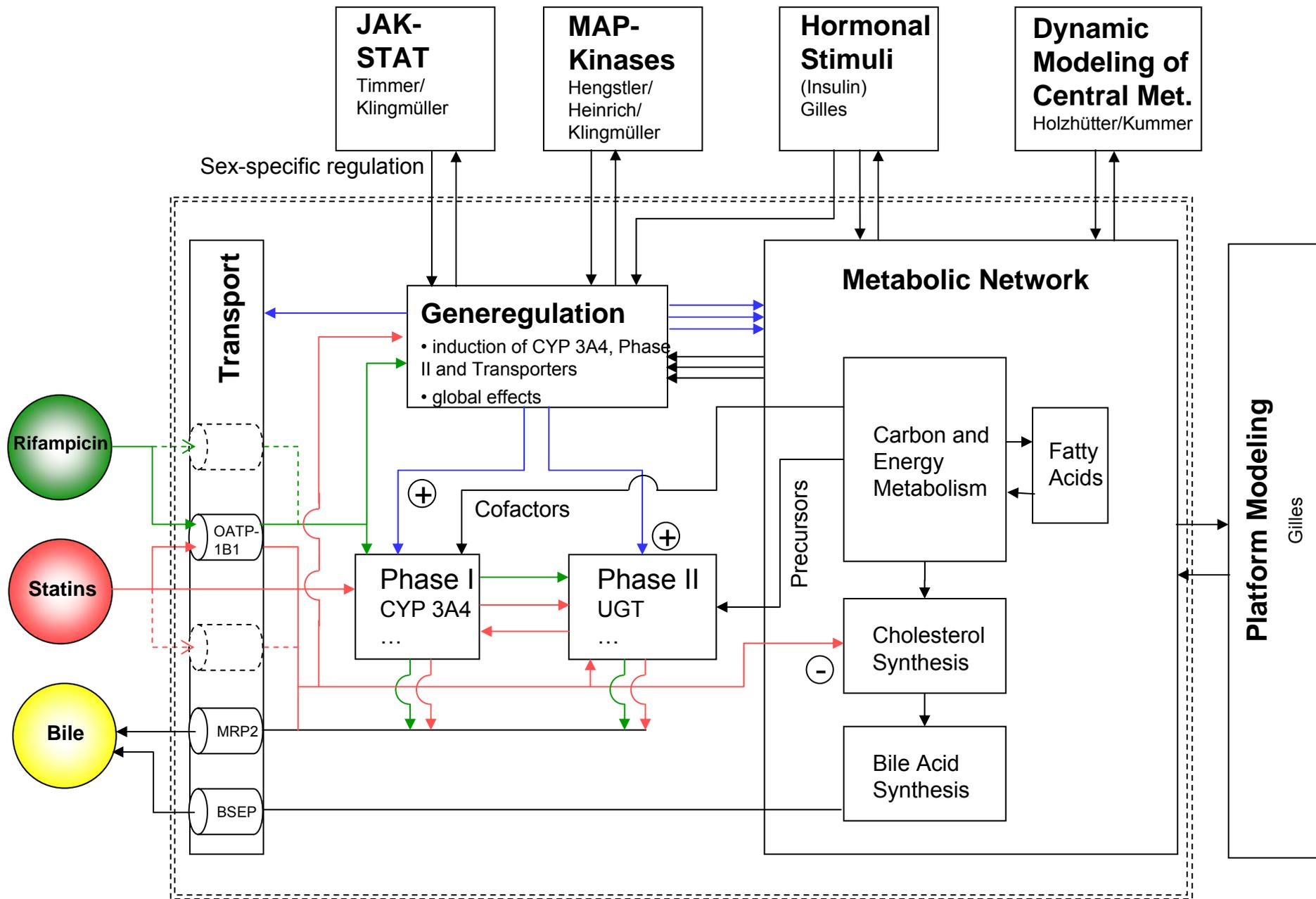


Liver must be extremely robust because of the inter-individual isoenzyme variability (internal perturbations) and multitude of substrates to be tackled by the P450 network (external perturbations)

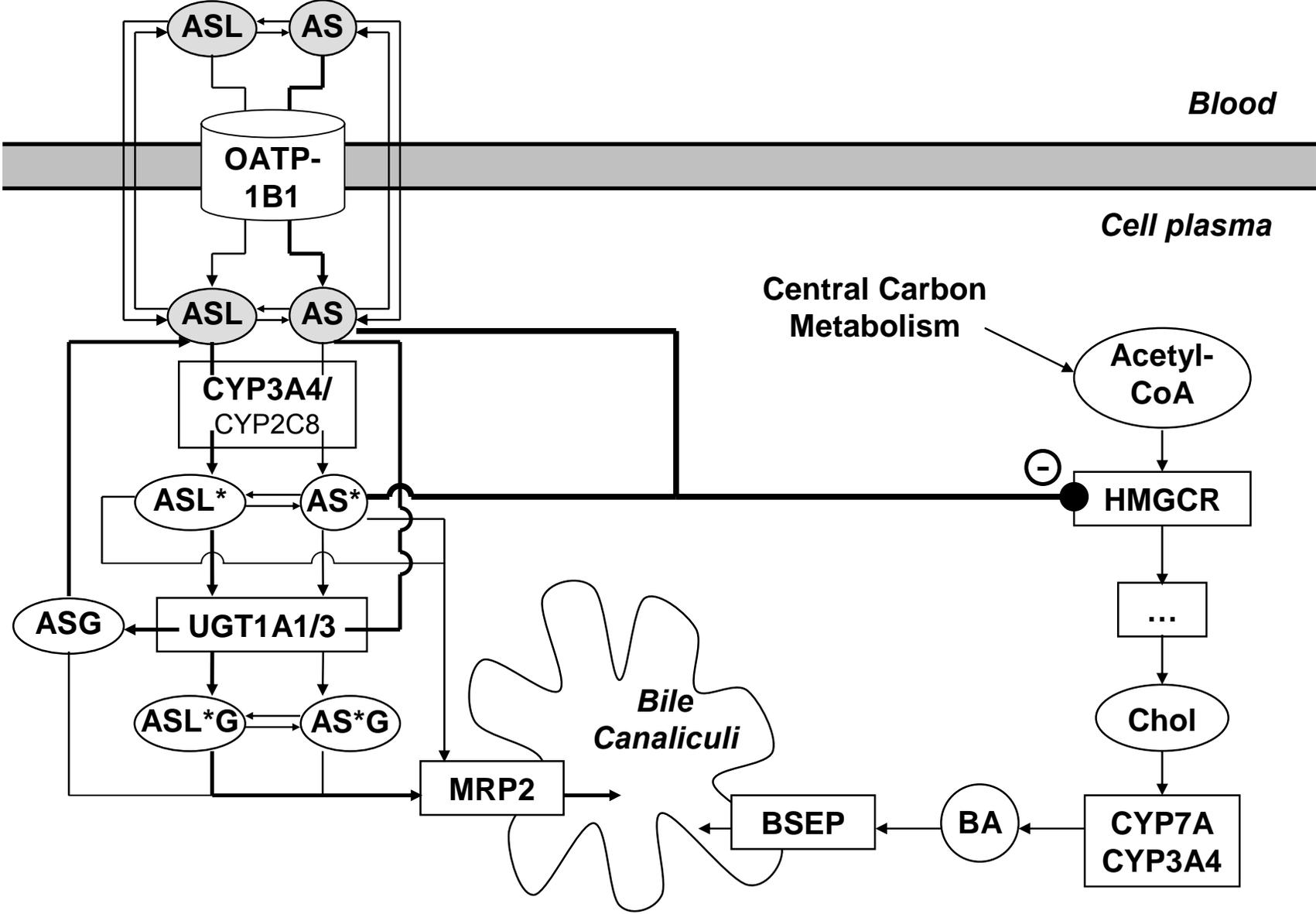
Stand-alone interpretation (kinetic modeling and simulation) of drug metabolism based on single isoenzyme activity (reductionistic pharmacokinetics) does not describe the system as a whole

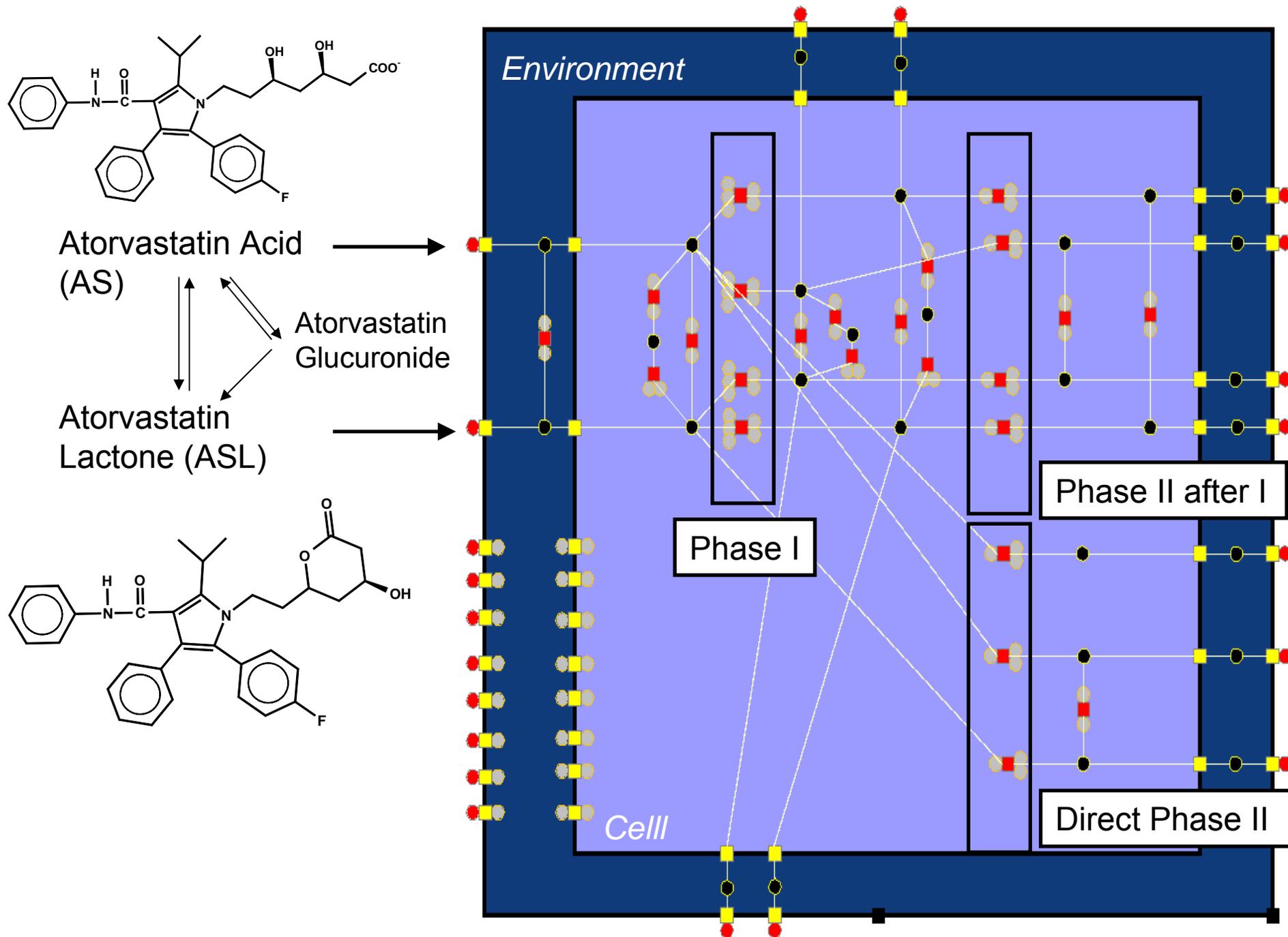
OUTLOOK – ROAD MAP FOR FUTURE DIRECTIONS



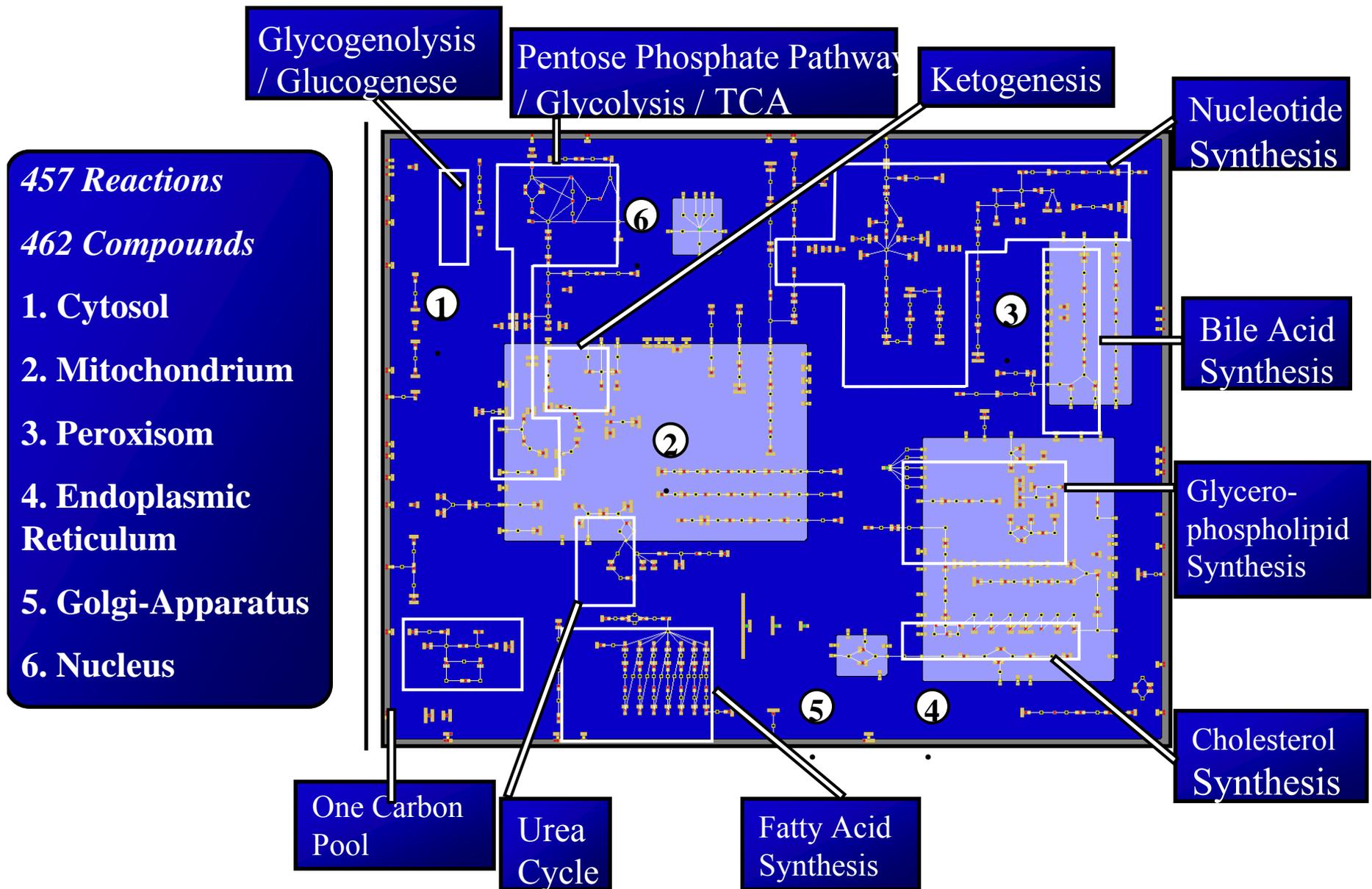


Atorvastatin (Sortis®, Lipitor®) (AS)



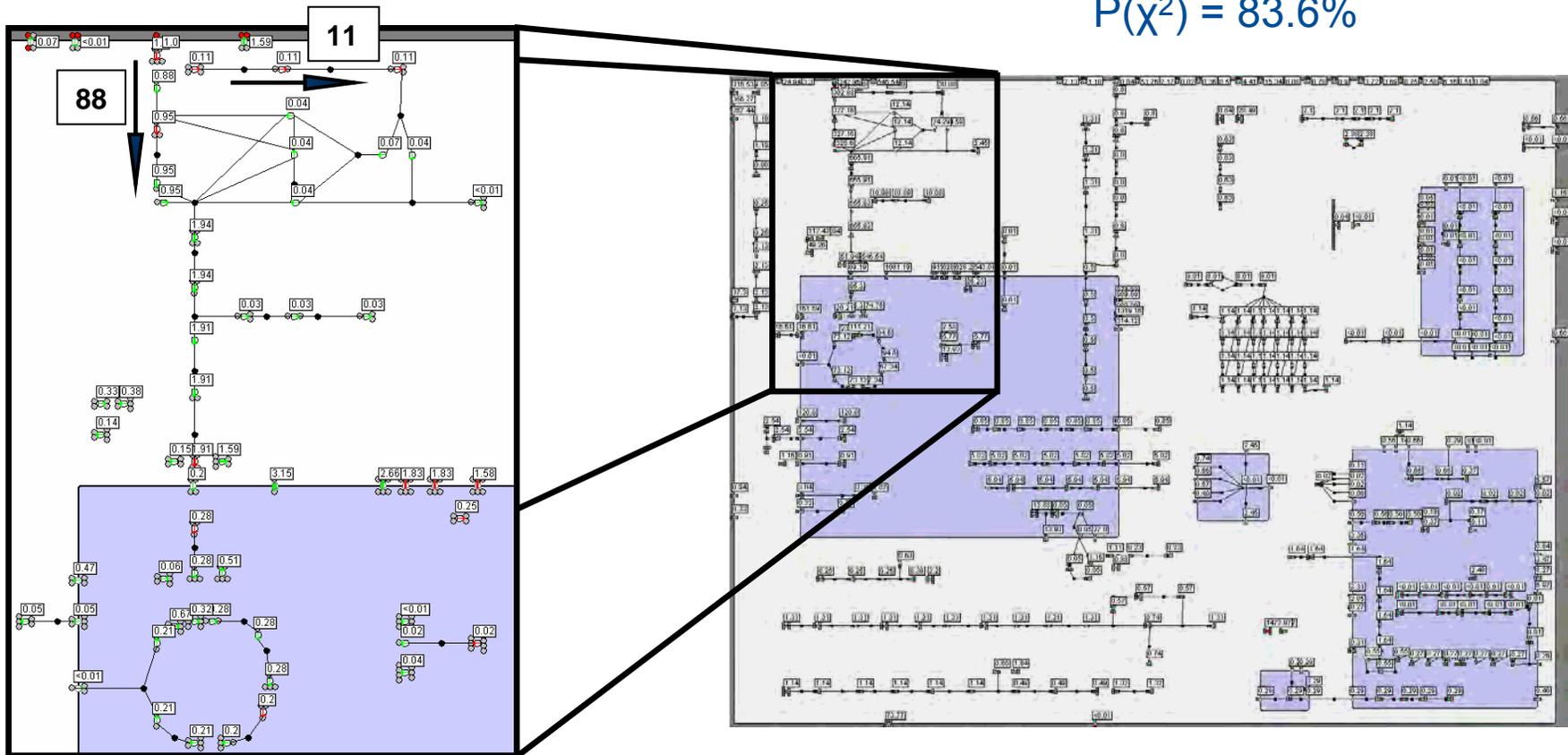


Stoichiometric Model of the Hepatocyte



Metabolic Flux Analysis in HepG2

$P(\chi^2) = 83.6\%$

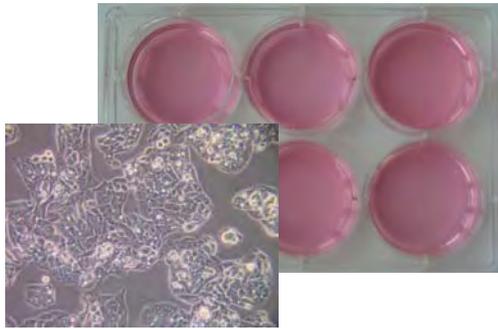
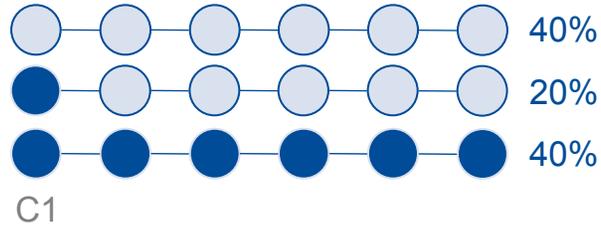
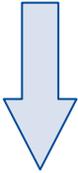


Labeling Experiment

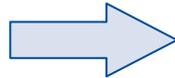
Flux Identification

Flux Distribution

Addition of labeled substrate at time 0



Hepatocytes "cultured" in 6-well plates (-FCS, -Gln) 10^6 cells/ well



Quenching and cell disruption at various time points



Quantification by GC-MS and LC-MS at IKP



- 12 intracellular metabolite concentrations
- 67 mass isotopomers
- 26 extracellular metabolite concentrations



Labeling Experiment

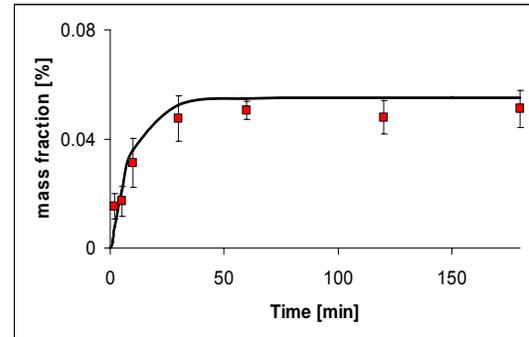
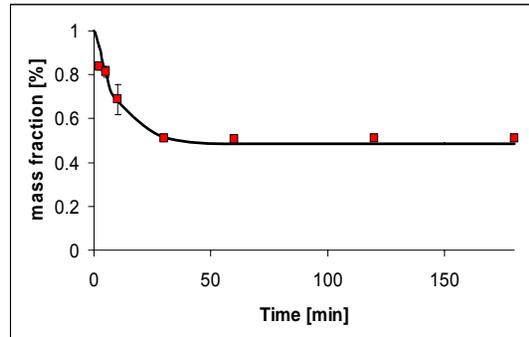
Flux Identification

Flux Distribution

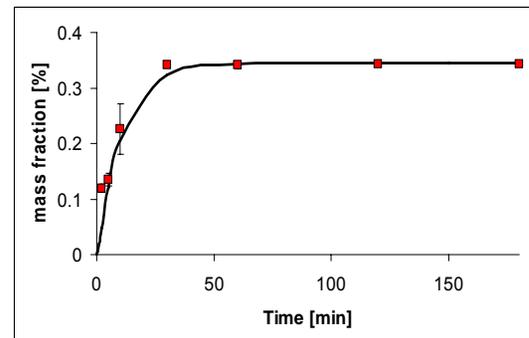
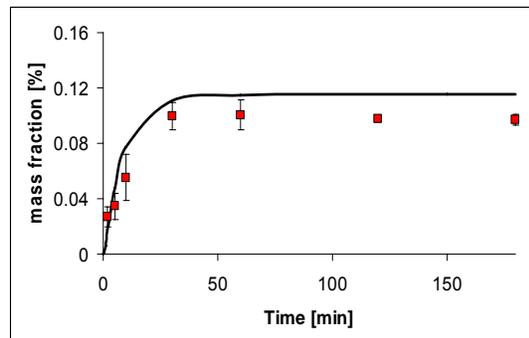
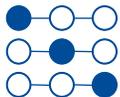
Glycolysis – 3-Phosphoglycerate

■ Experimental data
— Model prediction

m0



m1



Labeling Experiment

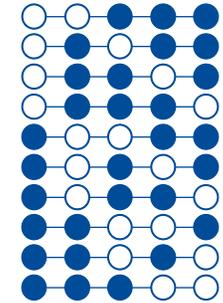
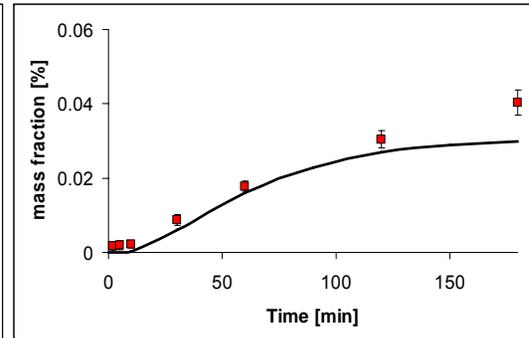
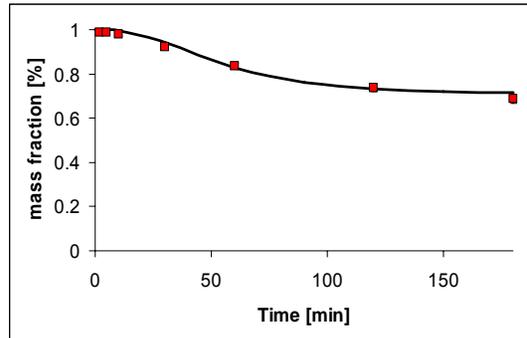
Flux Identification

Flux Distribution

TCA – α -Ketoglutarate

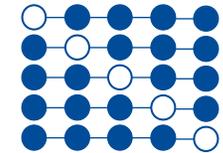
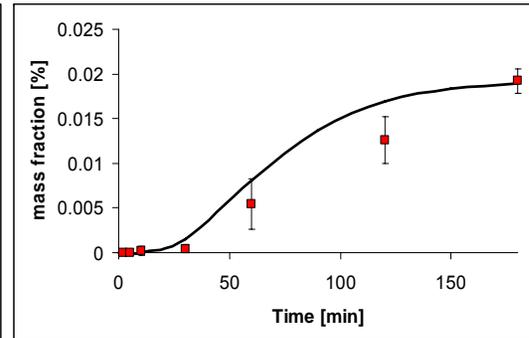
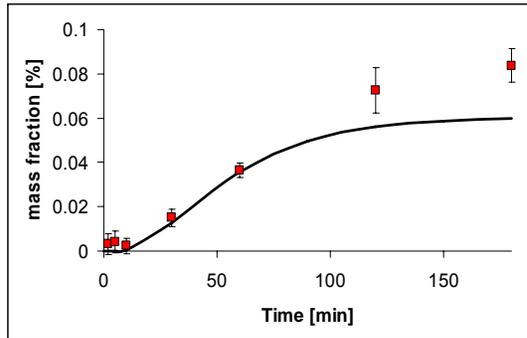
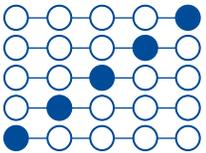
■ Experimental data — Model prediction

m0



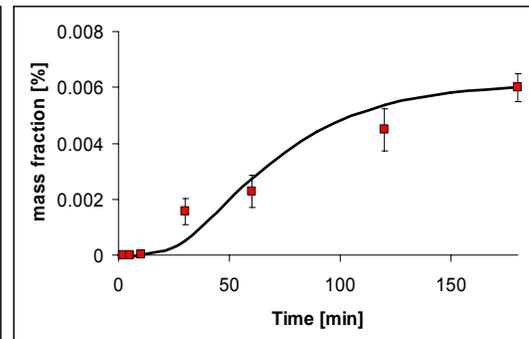
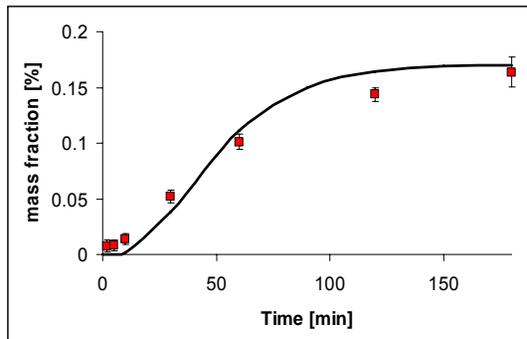
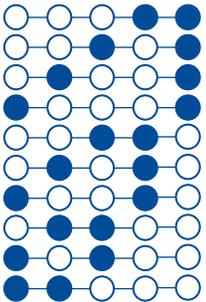
m3

m1

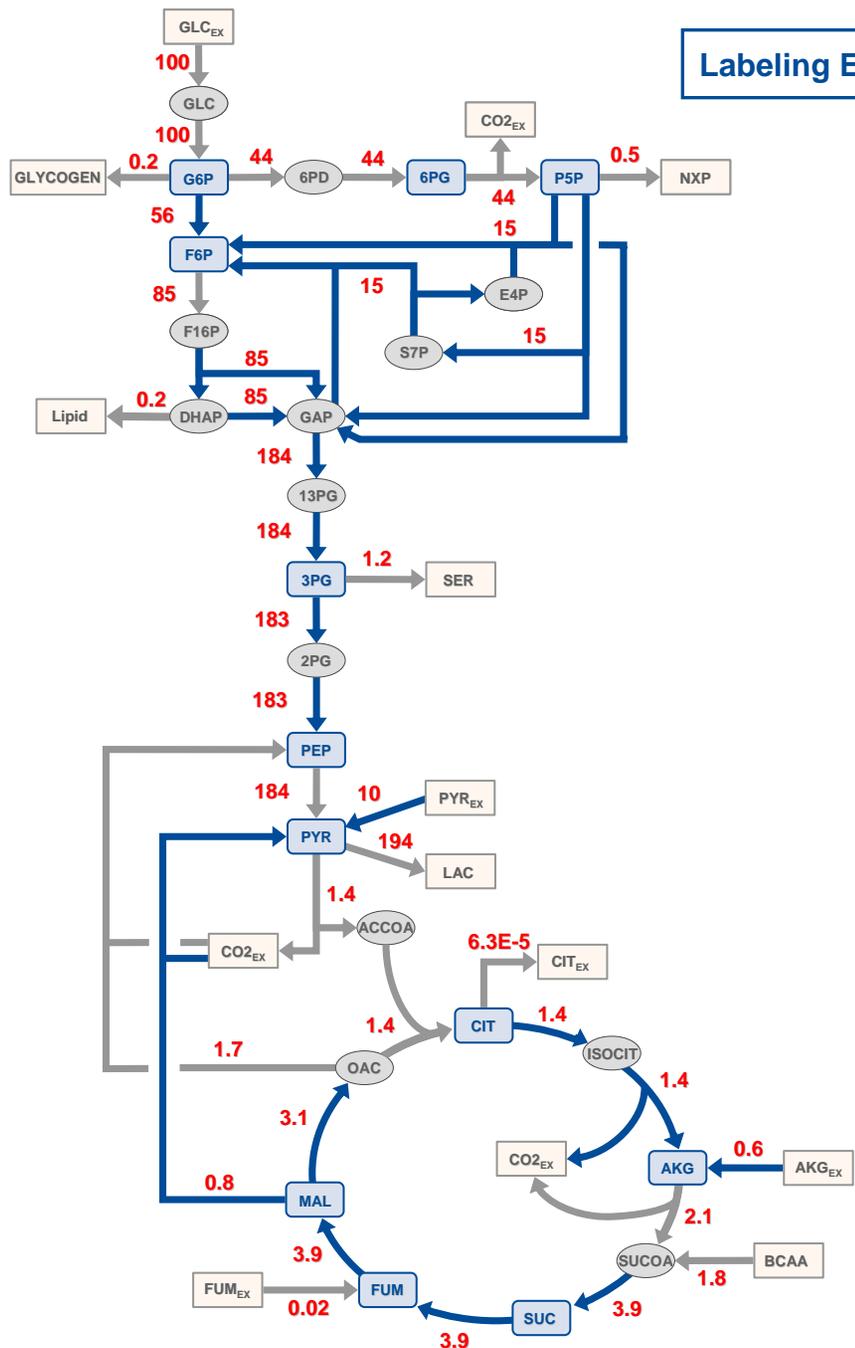


m4

m2

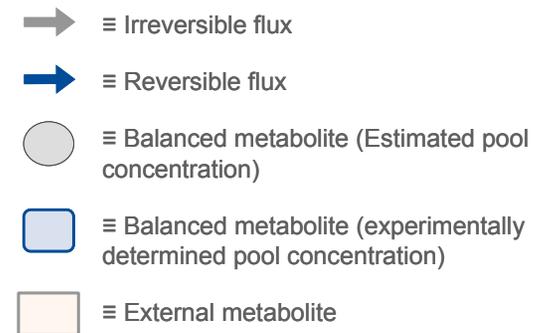


m5



Split Ratio: Glycolysis - PPP

- *Metabolite Balancing*
 - 11% PPP; 88% Glycolysis
 - 5% PPP; 95% Glycolysis
- *Isotopomer Balancing*
 - 44% PPP; 56% Glycolysis



(1) Dynamics of gene regulation of the detoxification system

(Reverse engineering: Unraveling gene-gene interactions from Microarray data)

(2) Quantitative kinetics of drug induced interactions in the nuclear receptor network

(3) Chemoinformatics modeling molecular modeling: from sequence to function

(4) High throughput data: microarrays and proteomics

(5) Hierarchical multi scale modeling:

Integration of single hepatocyte models with whole body (multi organe) models

Acknowledgements:

**Joachim Bucher
Tanja Saussele
Ulrich Zanger**

**Klaus Maier
Klaus Mauch**

Funding:

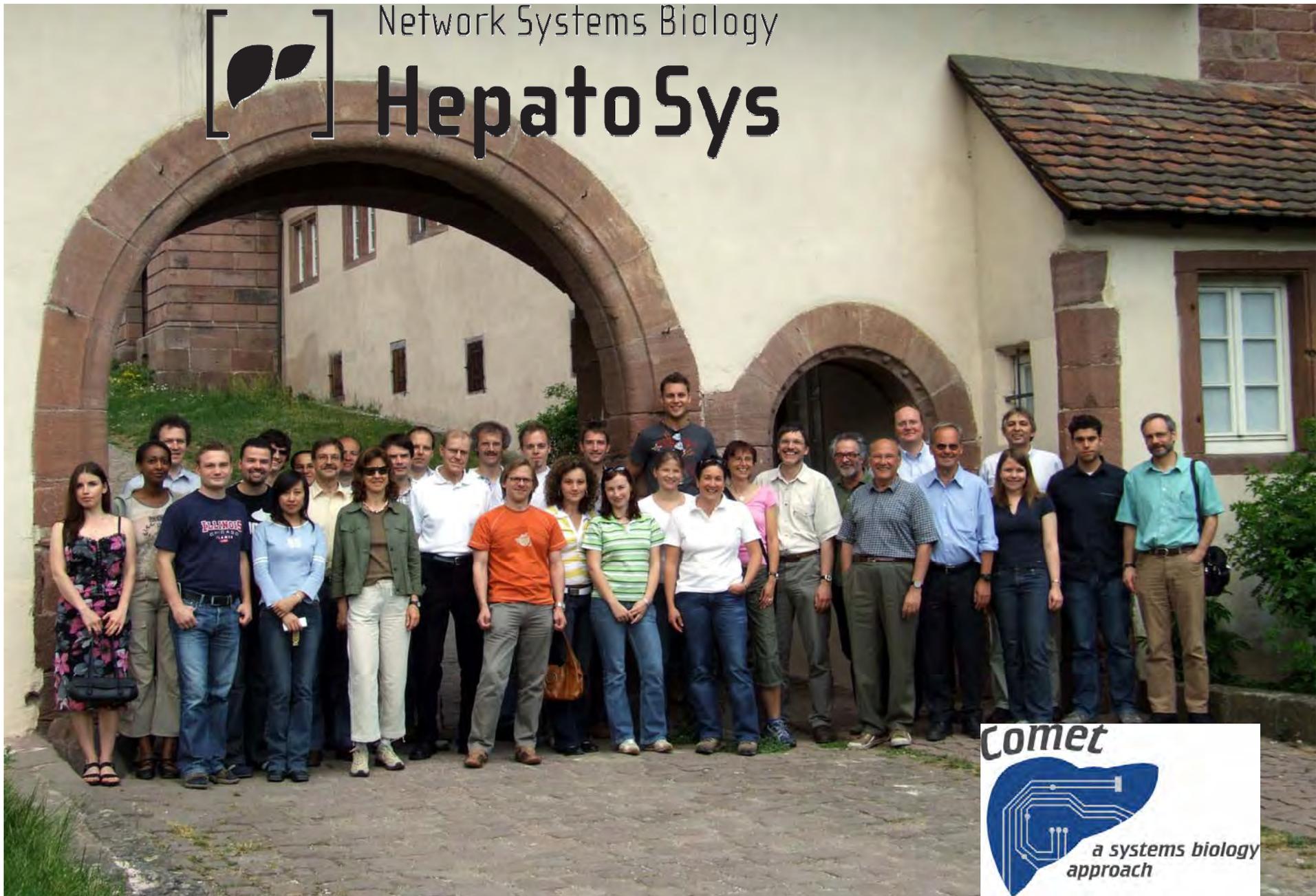
**Federal Ministry of Education and Resarch (BMBF):
Funding Initiative: HepatoSys**





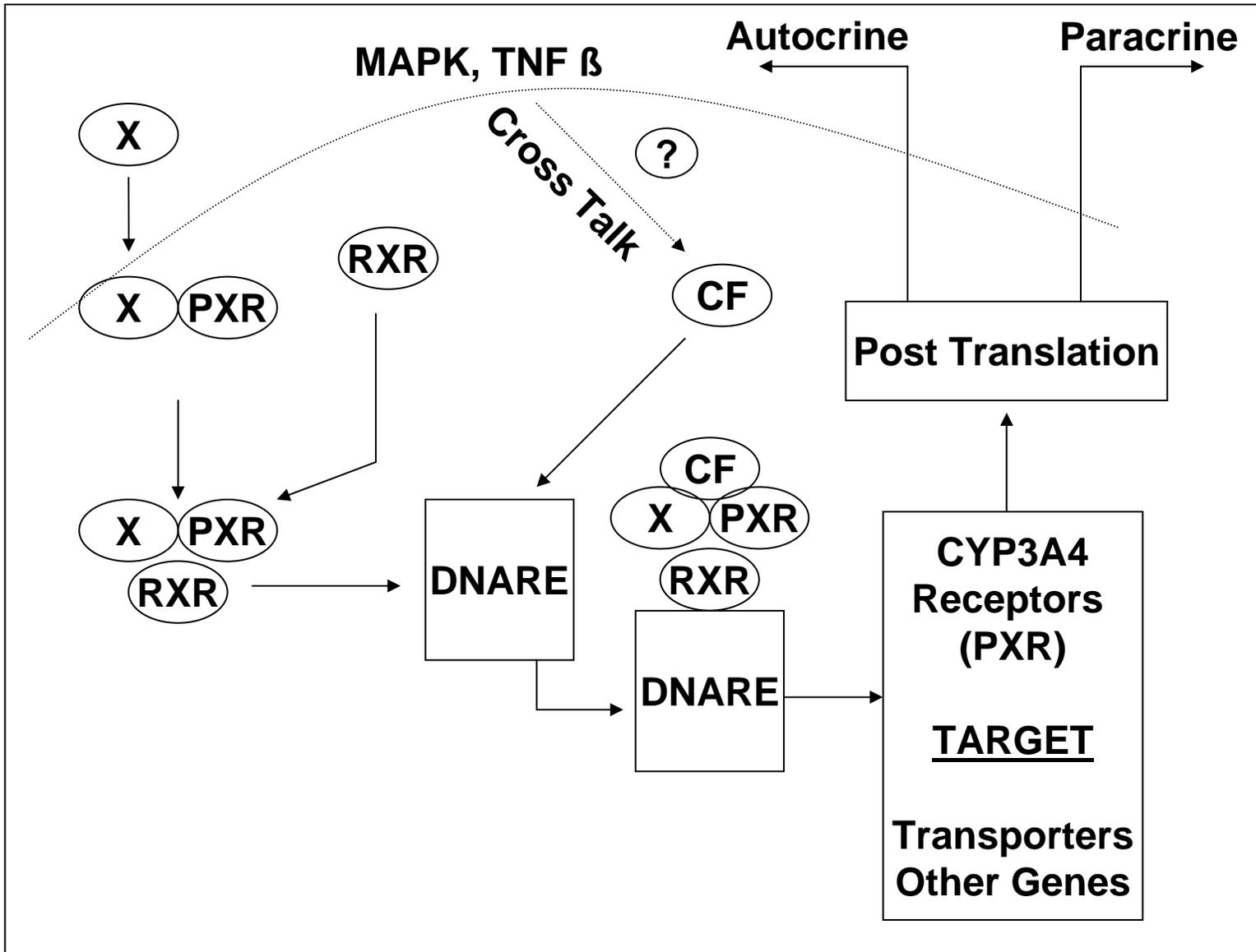
Network Systems Biology

HepatoSys



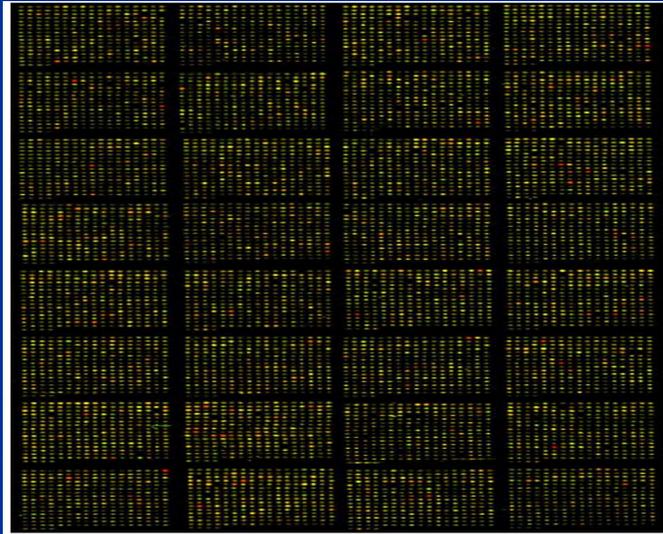
Network Detoxification

Phase I Module



Data Analysis - Work Flow

t_1 t_2 t_3 t_4



**Array Data
(Time Series)**

Cluster
Analysis

Normalization
Calculation of the
mean and sd
Coefficient of
Variation

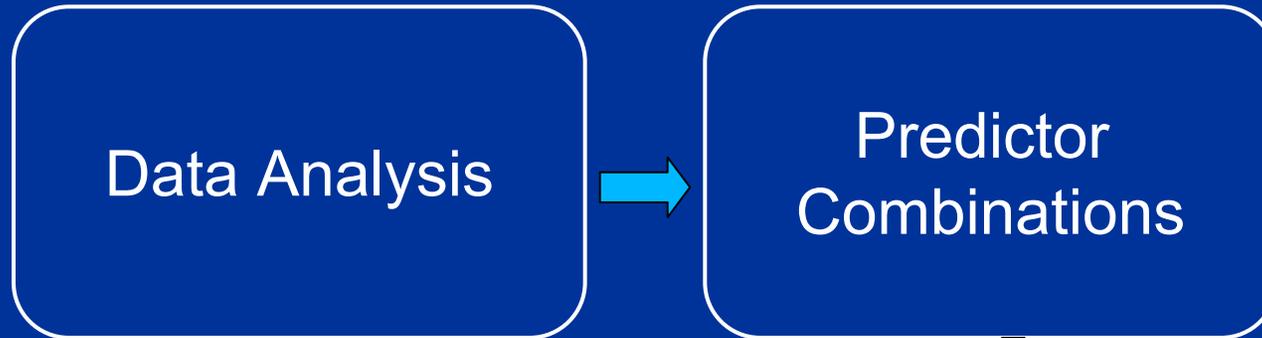
Sorting

Target Genes
(Problem Specific)

Influence

Discretisation (1,0,-1)
Predictor Gene Sets

Methodology



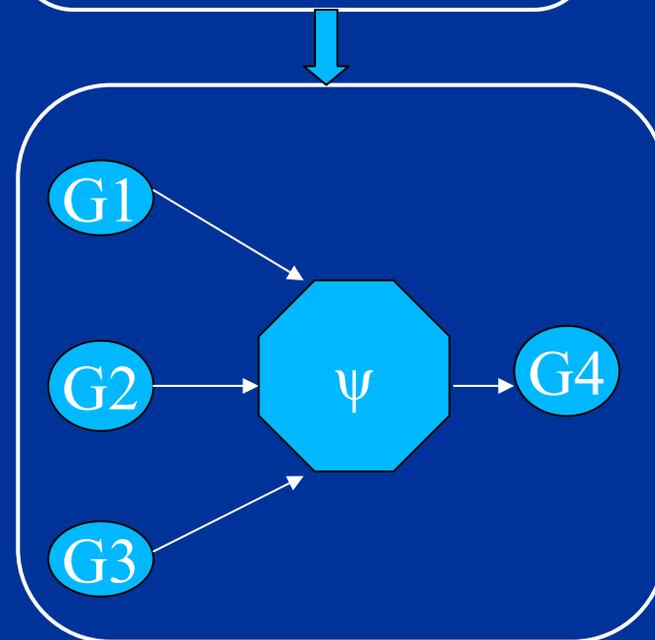
Ψ = Predictor Function

$$\theta = [\epsilon(\Psi_0) - \epsilon(\Psi_{opt})] / \epsilon(\Psi_0)$$

$\epsilon(\Psi)$ – Error Function

θ – Coefficient of Determination

$\theta > \theta_{\text{threshold}} \rightarrow$ prominent



predictor set

CYP3A4 specific Predictors

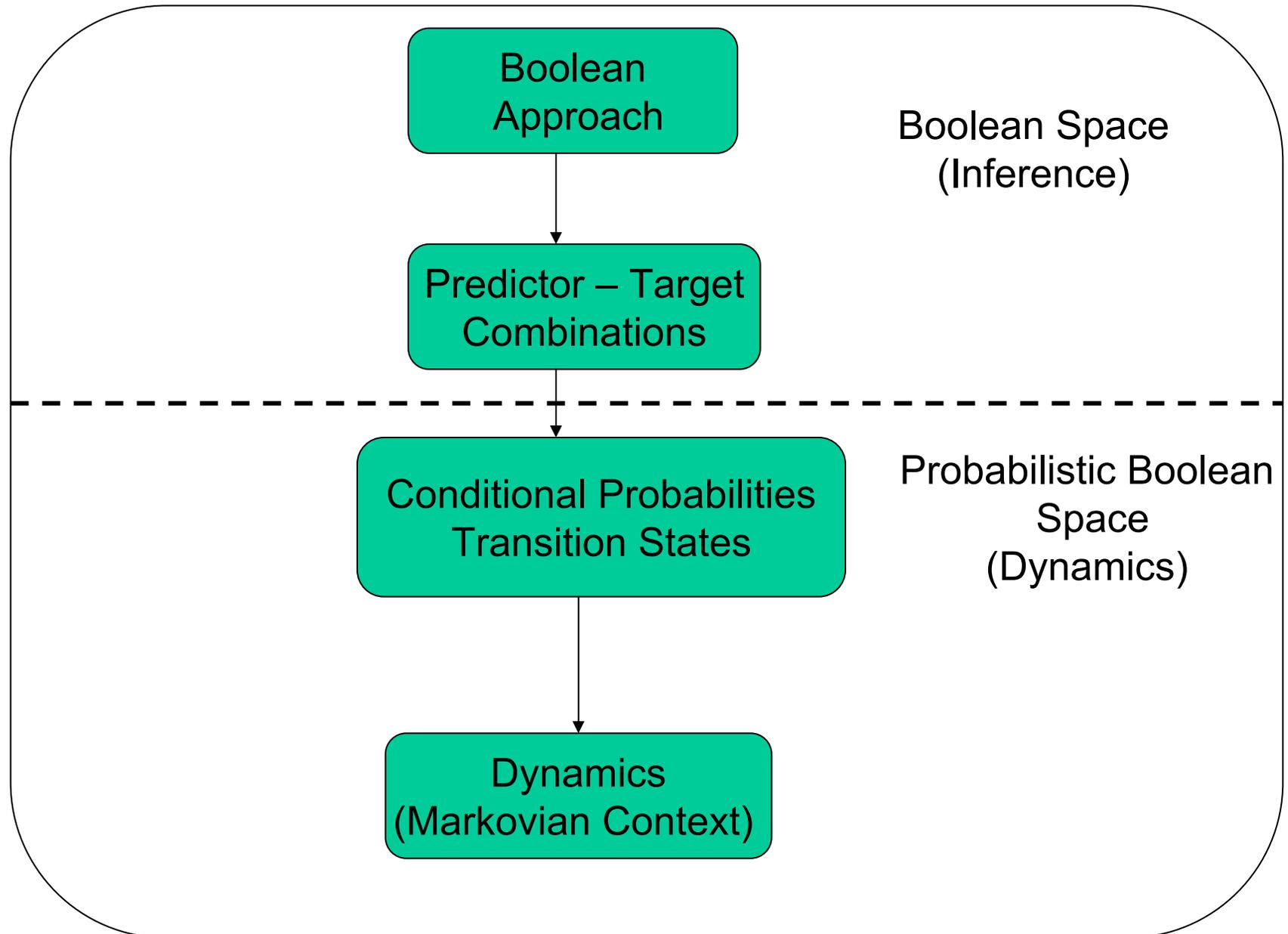
Most Prominent Predictors for CYP3A4 – 37

Predictor	Percentage
PXR	21
MAPK3	19
G6PT	16
CDC14B3	16
RXR	14
HNF 4 – alpha	14
ERK1/2	12
MDR1	9
PRO0786	9
GST2	7
OATP2	5

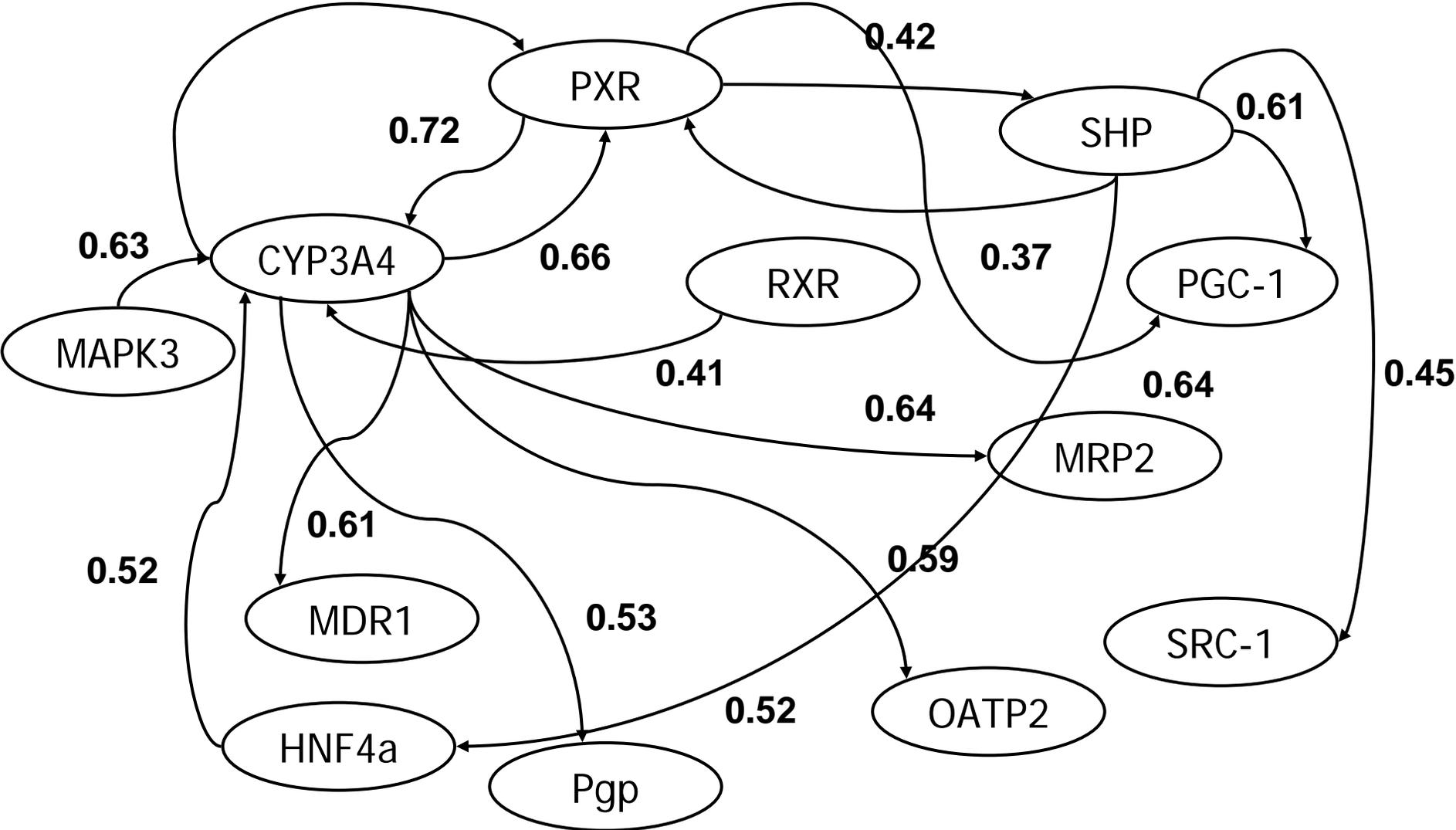
% = No. of times a given predictor was used for a given target

(p48 protein)

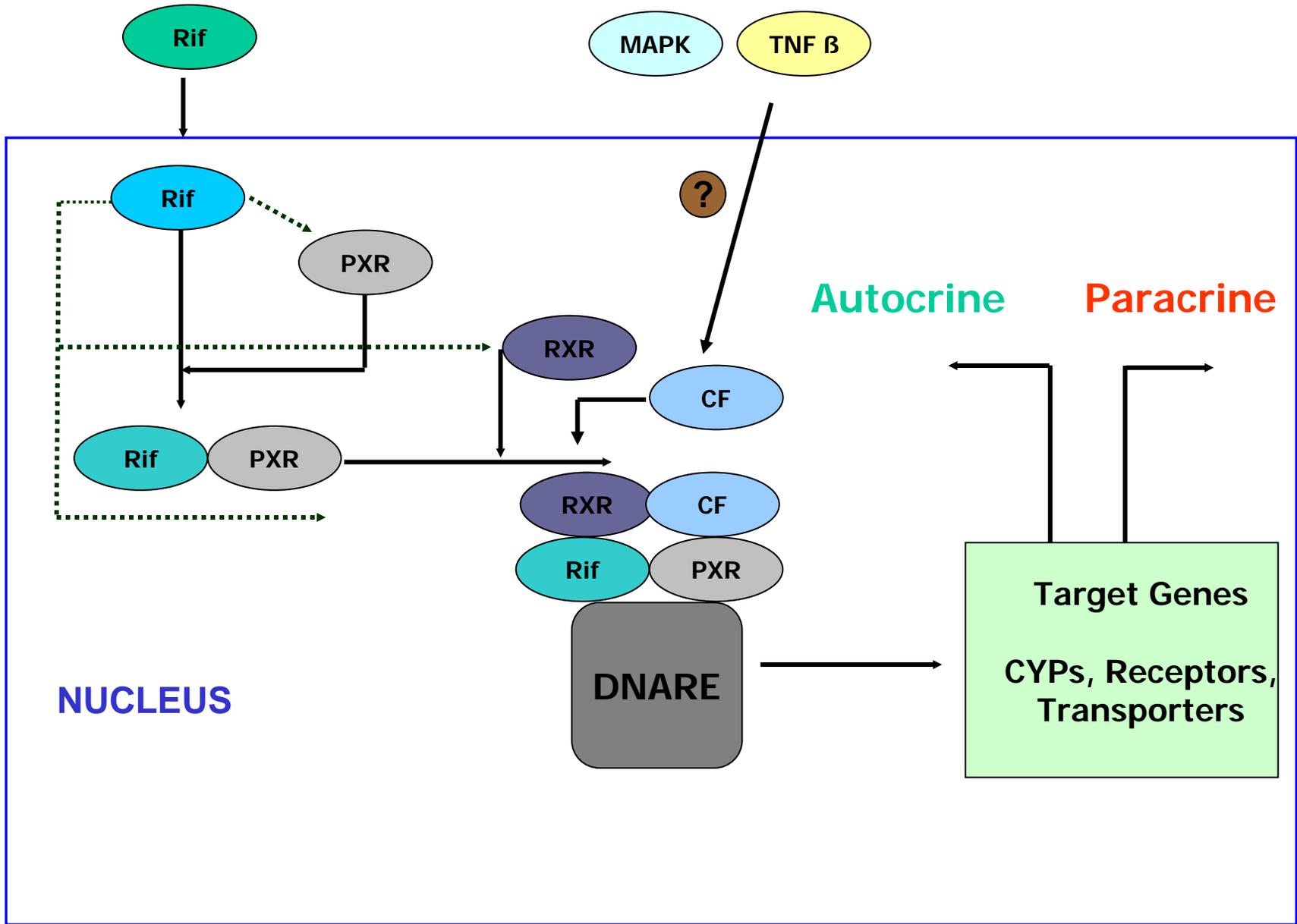
Boolean to Probabilistic Boolean



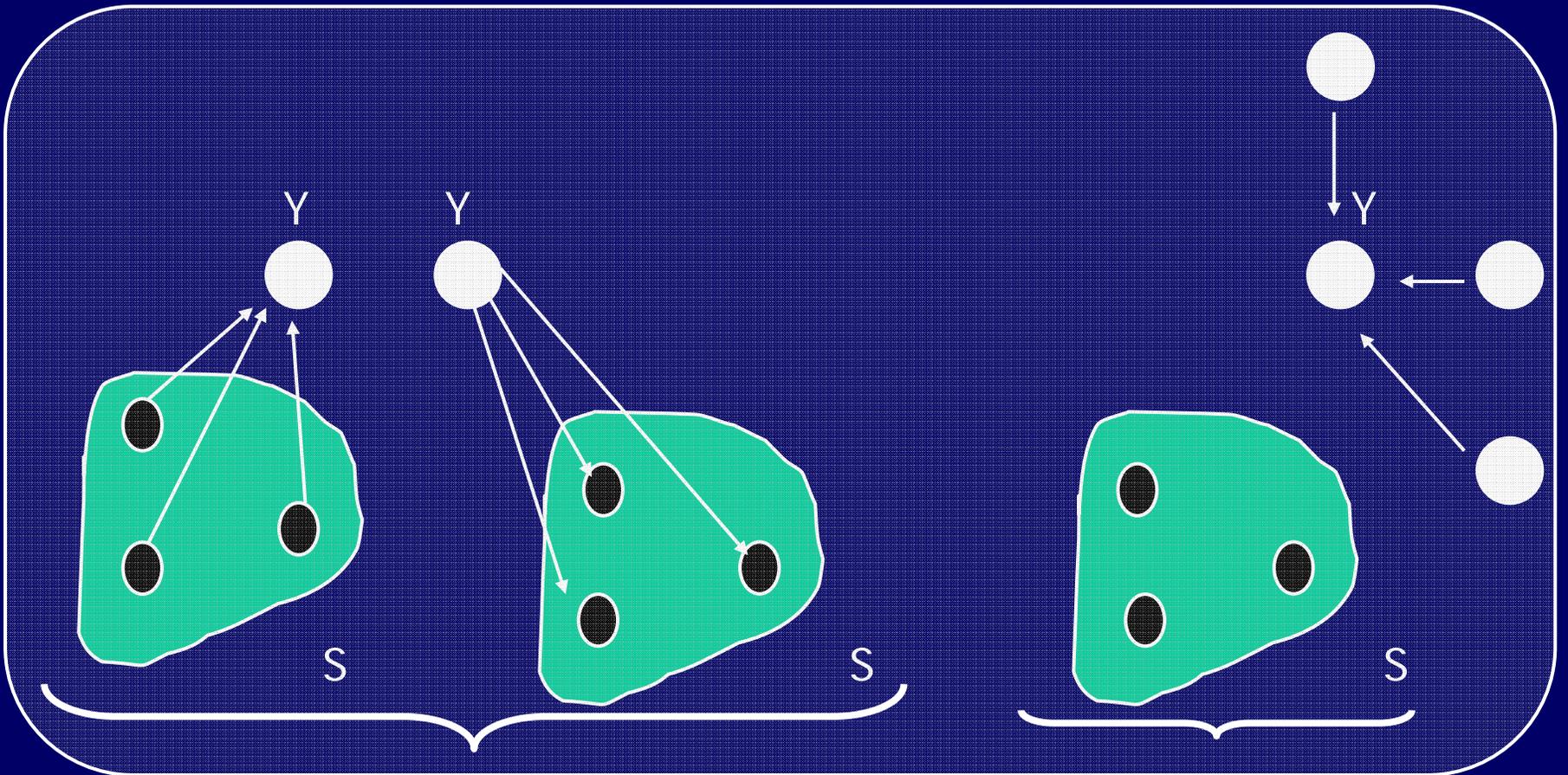
Subnetwork Simulation



0.XX – Connection Strength (Probabilities)

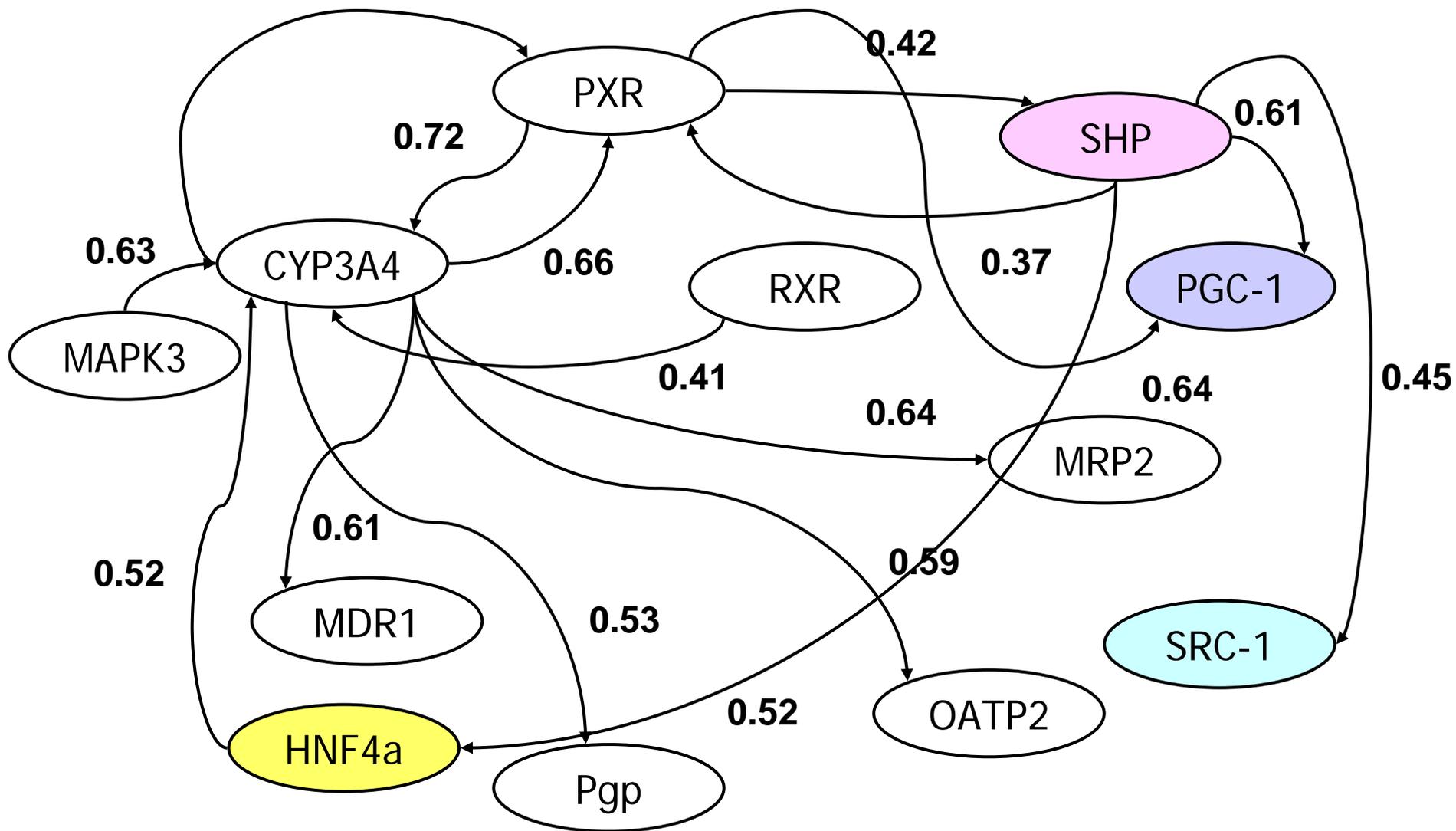


Principle of Autonomy

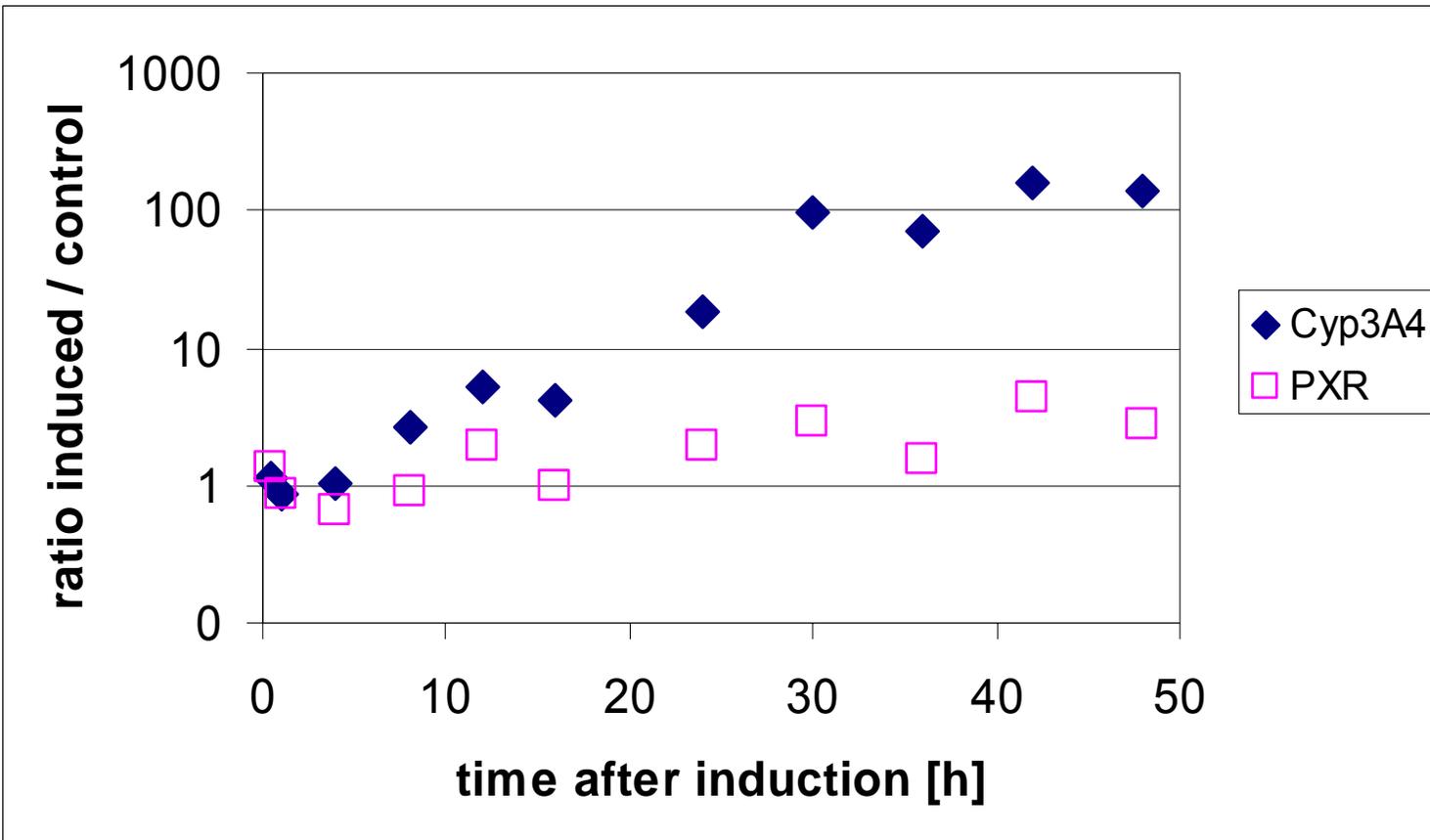


To achieve network autonomy,
both of these strengths of
connections should be high

The sensitivity of Y
from the outside
should be small



0.XX – Connection Strength (Probabilities)



Thomas Reichart, ITB, Uni Stuttgart

From Inference to Dynamics

Inference

Known Pathway

Microarray Data

CoD Validation

Dynamics

Construct Markovian Chain Model

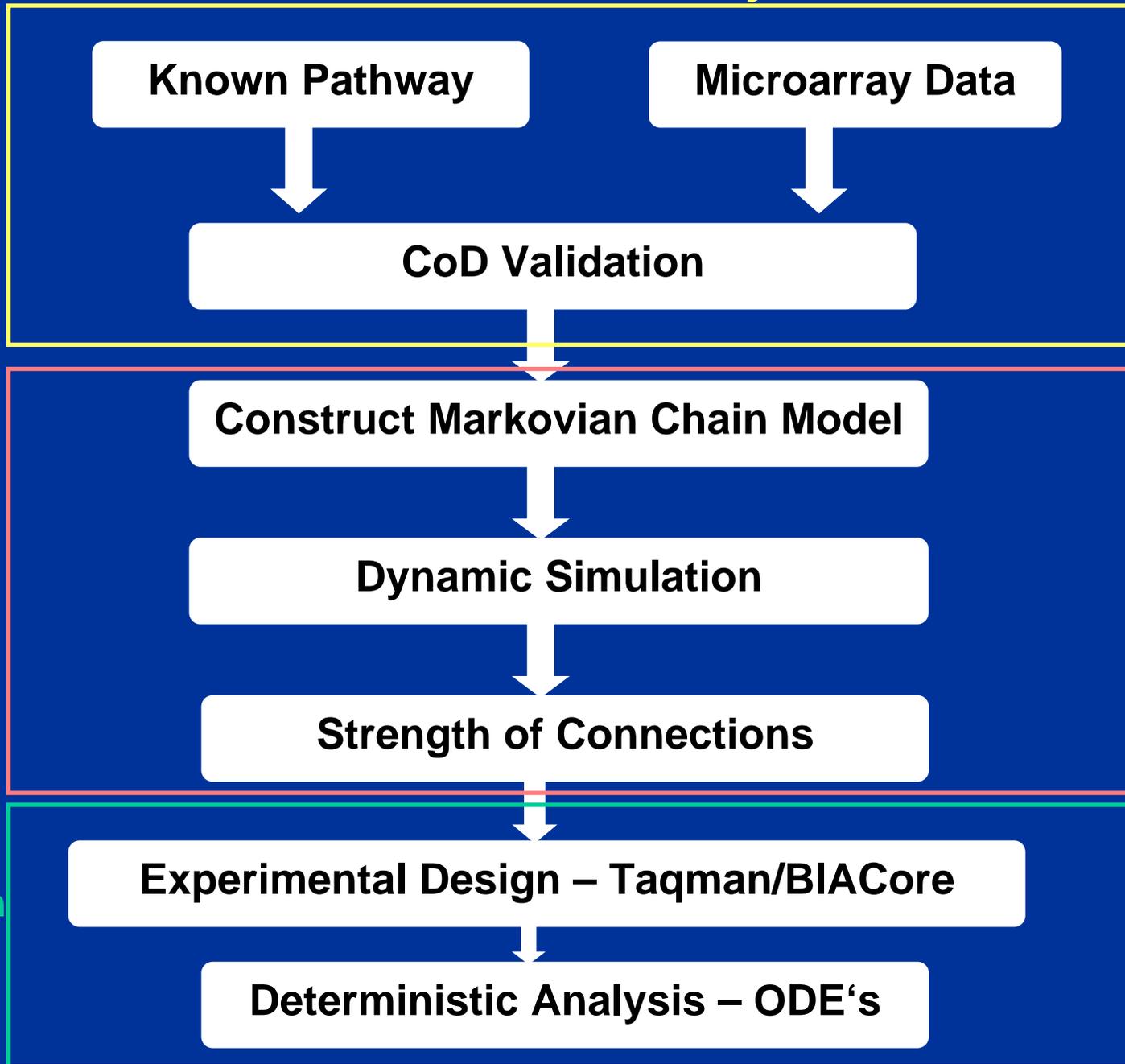
Dynamic Simulation

Strength of Connections

Intervention

Experimental Design – Taqman/BIACore

Deterministic Analysis – ODE's



*** from reverse (inverse) engineering to deterministic modeling**

**•* boolean modeling – probabilistic- boolean modeling
- deterministic modeling**

•* top-down/bottom-up/middle - out