

IV. CULTIVARS AND GERM PLASM

USDA-ARS NATIONAL SMALL GRAINS GERMPLASM RESEARCH FACILITY
P.O. Box 307, Aberdeen, ID 83210, USA.
www.ars-grin.gov/npgs

National Small Grains Collection wheat germ plasm evaluations.

H.E. Bockelman, C.A. Erickson, and B.J. Goates.

The USDA-ARS National Small Grains Collection (NSGC) is one of the several components of the National Plant Germplasm System. The NSGC is a working collection in contrast to the base collection at the National Seed Storage Laboratory (NSSL) at Fort Collins, CO. The numbers of accessions in the NSGC are summarized Table 1.

Table 1. USDA-ARS National Small Grains Collection, April, 2001.

Taxonomy	NSGC Accessions
<i>Triticum</i>	54,140
<i>Hordeum</i>	26,971
<i>Avena</i>	21,094
<i>Oryza</i>	17,357
<i>Aegilops</i>	2,213
<i>Secale</i>	2,078
<i>X Triticosecale</i>	1,974
All Species	126,386

The systematic evaluation of wheat accessions in the NSGC and other elite germ plasm continued to be coördinated or conducted by National Small Grains Germplasm Research Facility (NSGGRF) staff at Aberdeen during 2001.

Descriptors appropriate for wheat have been established in collaboration with the Wheat Crop Germplasm Committee. Field evaluation data are recorded on such descriptors as growth habit, number of days from planting to anthesis (heading), plant height, spike or panicle density, lodging, straw breakage, shattering, and awn and glume characteristics, including color. Special nurseries are grown for that purpose at Aberdeen, Idaho and Maricopa, AZ. Disease and insect evaluations are conducted in collaboration and coöperation with ARS and state experiments station specialists.

Data obtained from evaluations of NSGC germ plasm are entered in the Germplasm Resources Information Network (GRIN) system by the NSGGRF staff in coöperation with the ARS National Germplasm Resources Laboratory, Beltsville, MD. GRIN is a database containing the characteristics and availability of all genetic resources included in the National Plant Germplasm System. The Database Manager is J.D. Mowder, Beltsville, Maryland. The NSGGRF staff interacts with the GRIN system in recording NSGC orders (seed requests), entering a variety of data, and conducting information searches. No evaluations have been conducted to date for descriptors such as drought tolerance; salt tolerance; winterhardiness; resistance to *Cephalosporium* stripe, flag smut, leaf blight, loose smut, snow mold, take-all, tan spot, and WSMV; and protein.

Triticum descriptors with data currently on the GRIN system are summarized in Table 2, p. 274.

The authors wish to acknowledge the important contributions of the NSGGRF staff in this effort, with special thanks to Glenda B. Rutger, Scott McNeil, Carol S. Truman, Judy Bradley, Kathy E. Burrup, Kay B. Calzada, Karla Reynolds, and Dave E. Burrup. Mr. Greg Laine is coöordinating the wheat evaluations efforts at Maricopa, AZ.

National Small Grains Collection Activities.

H.E. Bockelman, USDA-ARS, National Small Grains Collection, Aberdeen, ID, USA.

Cultivar name clearance. Breeders in the United States are encouraged to have proposed names for new cultivars checked for duplication. The National Small Grains Collection will be glad to assist you. Send the proposed name to: Harold E. Bockelman, USDA-ARS-NSGC, P.O. Box 307, Aberdeen, ID 83210, Fax 208-397-4165, E-mail to

nsgchb@ars-grin.gov. If desired, more than one name may be submitted, listed in order of preference. This will save considerable time if a conflict is found with the first name. Available records (GRIN, CI/PI cards, variety files, etc.) here at Aberdeen are checked for conflicts with the proposed name. If a conflict is found (previous use of the name for that crop), the breeder is requested to submit a different name. If no conflicts are found, the requested name is forwarded to the Federal Seed Lab, Agricultural Marketing Service where the proposed name is checked against the databases they maintain. The Agricultural Marketing Service does not guarantee that its findings are the final word since there is no single, complete name database. This clearance procedure generally requires about four weeks. Trademark searches should be done by the breeder online at <http://www.uspto.gov>.

Elite germ plasm requested. Breeders are encouraged to consider submitting their elite lines for inclusion in the NSGC. Of special interest are lines that have been in uniform nurseries, but are not to be released as cultivars. Histori-

Table 2. National Small Grains Collection evaluation of disease; insect; and agronomic, taxonomic, and quality data for wheat on the GRIN system, updated February, 2002.

Character	Years	Location	Accessions
DISEASE EVALUATIONS.			
Barley Yellow Dwarf Virus	1985–92	Davis, CA	2,287
Barley Yellow Dwarf Virus	1988–94	Urbana, IL	17,517
Soilborne Mosaic Virus	1985–89	Urbana, IL	6,587
Soilborne Mosaic Virus	2000	Manhattan, KS	4,998
Leaf Rust	1983–89, 91–95	Manhattan, KS	38,751
Leaf Rust ñ Adult	2000	Manhattan, KS	5,000
Stripe Rust – Adult	1984–2001	Mt. Vernon, WA	36,273
Stripe Rust – Adult	1984–2001	Pullman, WA	27,241
Stripe Rust – PST 17	1984–2001	Pullman, WA	18,832
Stripe Rust – PST 20	1984–95	Pullman, WA	12,508
Stripe Rust – PST 25	1984–95	Pullman, WA	1,682
Stripe Rust – PST 27	1984–95	Pullman, WA	14,511
Stripe Rust – PST 29	1984–95	Pullman, WA	14,259
Stripe Rust – PST 37	1984–2001	Pullman, WA	6,146
Stripe Rust – PST 43	1984–2001	Pullman, WA	5,137
Stripe Rust – PST 45	1984–2001	Pullman, WA	6,138
Stripe Rust – PST 78	2000–01	Pullman, WA	1,835
Stem Rust – Adult	1987–94	Rosemount, MN	8,078
Stem Rust – Adult	1987–94	St. Paul, MN	19,141
Stem Rust – HJCS	1987–92	St. Paul, MN	4,342
Stem Rust – QFBS	1987–92	St. Paul, MN	8,639
Stem Rust – QSHS	1987–92	St. Paul, MN	4,455
Stem Rust – RHRS	1987–92	St. Paul, MN	4,312
Stem Rust – RTQQ	1987–92	St. Paul, MN	8,973
Stem Rust – TNMH	1987–92	St. Paul, MN	4,402
Stem Rust – TNMK	1987–92	St. Paul, MN	8,938
Stem Rust – HNLQ	1987–92	St. Paul, MN	4,705
Stem Rust – RKQS	1987–92	St. Paul, MN	4,682
Stem Rust – Genes	1987–92	St. Paul, MN	1,018
Common Bunt – R36	1981–92	Aberdeen, ID ¹	74
Common Bunt – R39	1981–92	Aberdeen, ID ¹	1,422
Common Bunt – R43	1981–92	Aberdeen, ID ¹	318
Common Bunt – T-1	1981–92	Aberdeen, ID ¹	6,301
Common Bunt – Multiple	1981–2000	Aberdeen, ID ¹	17,366
Dwarf Bunt	1978–2001	Aberdeen, ID ²	13,702
<i>Septoria nodorum</i>	1970–78	Bozeman, MT	8,095
Powdery Mildew	1996–2001	Kinston, NC	10,258
Fusarium Head Blight/Scab	1998–2001	Brookings, SD	3,207

Table 2 (continued). National Small Grains Collection evaluation of disease; insect; and agronomic, taxonomic, and quality data for wheat on the GRIN system, updated February, 2001.

Character	Years	Location	Accessions
INSECT EVALUATIONS.			
Hessian Fly – B	1983–94	W. Lafayette, IN	449
Hessian Fly – C	1983–94	W. Lafayette, IN & Manhattan, KS	24,165
Hessian Fly – E	1983–94	W. Lafayette, IN & Manhattan, KS	24,149
Hessian Fly – GP	1983–94	W. Lafayette, IN & Manhattan, KS	14,441
Hessian Fly – L	1983–97	W. Lafayette, IN & Manhattan, KS	8,315
Russian Wheat Aphid (RWA)	1988–95	Stillwater, OK	40,842
Cereal Leaf Beetle	1963–70	Indiana, Michigan	16,347
AGRONOMIC, TAXONOMIC, AND QUALITY EVALUATIONS.			
Growth Habit	1987–01	Aberdeen, ID	52,828
Chromosome Number	1988–91	Columbia, MO	519
Lysine Content	1966–69	Lincoln, NE	10,367
Awn Color	1983–97	Aberdeen, ID & Maricopa, AZ	22,650
Awn Type	1983–97	Aberdeen, ID & Maricopa, AZ	26,561
Glume Color	1983–97	Aberdeen, ID & Maricopa, AZ	22,812
Glume Pubescence	1983–97	Aberdeen, ID & Maricopa, AZ	24,312
Heading Date	1983–94	Aberdeen, ID & Maricopa, AZ	18,365
Heading Date – related to check	1999–2001	Maricopa, AZ	24,968
Kernel Color	1983–94	Aberdeen, ID & Maricopa, AZ	21,319
Kernels/Spike	1983–94	Aberdeen, ID & Maricopa, AZ	3,666
Kernel Weight	1983–94	Aberdeen, ID & Maricopa, AZ	3,669
Leaf Pubescence	1983–94	Aberdeen, ID & Maricopa, AZ	20,888
Plant Height	1983–97	Aberdeen, ID & Maricopa, AZ	21,841
Plant Height – related to check	1999–2001	Maricopa, AZ	24,958
Rachis Length	1995	Maricopa, AZ	2,512
Shattering	1983–94	Aberdeen, ID & Maricopa, AZ	10,637
Spike Density	1983–98	Aberdeen, ID & Maricopa, AZ	15,823
Spikelets/Spike	1995	Maricopa, AZ	2,502
Spike Type	1983–97	Aberdeen, ID & Maricopa, AZ	15,551
Straw Breakage	1983–94	Aberdeen, ID & Maricopa, AZ	16,829
Straw Color	1983–97	Aberdeen, ID & Maricopa, AZ	19,142
Straw Lodging	1983–94	Aberdeen, ID & Maricopa, AZ	23,075
Core			4,523
Market Class			1,411

¹ 1985–86 Pendleton, OR.² Field tests are conducted at Logan, UT, by Aberdeen ARS staff.

cally, uniform nurseries been the testing grounds for the most advanced, elite germ plasm from the various public and private breeding programs. Entries in uniform nurseries and other breeding materials that are never released as cultivars are still of potential value to breeders, pathologists, entomologists, and other researchers. Breeders should submit 200–500 g of *untreated* seed to the NSGC (address: P.O. Box 307, Aberdeen, ID 83210). Seed from outside of the United States should be sent to the USDA Plant Germplasm Quarantine Center (address: Bldg. 580, BARC-East, Beltsville, MD 20705) with enclosed forwarding directions. Provide a description of the germ plasm, including donor (breeder, institution); botanical and common name; cultivar name and/or other identifiers (breeder line or selection number, etc.); pedigree; descriptive information (of important traits and special characteristics); and growth habit. Assignment of a PI number and inclusion in the NSGC makes the germ plasm available for research purposes to bona fide scientists in the U.S. and worldwide. Please note that a different procedure applies if you are obtaining *Crop Science* registration. Follow directions provided by the crop registration committee.

Guidelines for exporting seed. All seed sent to a foreign country should be inspected and receive a phytosanitary certificate. In most cases, a fee payable to APHIS (Animal & Plant Health Inspection Service) is required to cover the cost of the phytosanitary certificate. You may wish to work with APHIS personnel in your state or your State Department of Agriculture to obtain a phytosanitary certificate. Also, please be aware of any import permits and additional declarations that certain importing countries may require to accompany the shipment.

Guidelines for importing seed. Any scientist importing seed should be aware of any restrictions that apply. APHIS personnel can provide current information on applicable restrictions. Of particular importance to wheat researchers are import restrictions related to flag smut and karnal bunt. Presently, some 34 countries have flag smut import restrictions. Six countries currently have karnal bunt import restrictions. *Importation of seed from flag smut and Karnal bunt countries requires a permit from APHIS.* Special handling and grow-out procedures apply to such shipments.

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
615227	<i>aestivum</i> subsp. <i>aestivum</i>	Nagyatadi TF	Hungary	
615229	<i>aestivum</i> subsp. <i>aestivum</i>	Bartweizen	Austria	Upper Austria
615230	<i>aestivum</i> subsp. <i>aestivum</i>	Sipbachzeller	Austria	Upper Austria
615231	<i>aestivum</i> subsp. <i>aestivum</i>	Sipbachzeller	Austria	Styria
615232	<i>aestivum</i> subsp. <i>aestivum</i>	Sipbachzeller	Austria	Upper Austria
615233	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0002	Belgium	Luxembourg
615234	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0003	Belgium	Namur
615235	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0004	Belgium	Luxembourg
615236	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0005	Belgium	Luxembourg
615237	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0006	Belgium	Luxembourg
615238	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0007	Belgium	Luxembourg
615239	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0008	Belgium	Luxembourg
615240	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0009	Belgium	Luxembourg
615241	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0010	Belgium	Luxembourg
615242	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0011	Belgium	Luxembourg
615243	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0012	Belgium	Luxembourg
615244	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0013	Belgium	Luxembourg
615245	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0014	Belgium	Luxembourg
615246	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0015	Belgium	Luxembourg
615247	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0016	Belgium	Luxembourg
615248	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0017	Belgium	Luxembourg
615249	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0018	Belgium	Luxembourg
615250	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0019	Belgium	Luxembourg
615251	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0020	Belgium	Namur
615252	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0021	Belgium	Namur
615253	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0022	Belgium	Luxembourg
615254	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0023	Belgium	Luxembourg
615255	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0024	Belgium	Luxembourg
615256	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0025	Belgium	Luxembourg
615257	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0026	Belgium	Luxembourg
615258	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0027	Belgium	Luxembourg
615259	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0028	Belgium	Luxembourg
615260	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0029	Belgium	Luxembourg
615261	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0030	Belgium	Luxembourg
615262	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0031	Belgium	Luxembourg
615263	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0032	Belgium	Luxembourg
615264	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0033	Belgium	Luxembourg
615265	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0034	Belgium	Luxembourg

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
615266	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0035	Belgium	Luxembourg
615267	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0036	Belgium	Luxembourg
615268	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0037	Belgium	Luxembourg
615269	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0038	Belgium	Luxembourg
615270	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0039	Belgium	Luxembourg
615271	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0040	Belgium	Luxembourg
615272	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0041	Belgium	Luxembourg
615273	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0042	Belgium	Luxembourg
615274	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0043	Belgium	Luxembourg
615275	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0044	Belgium	Luxembourg
615276	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0045	Belgium	Luxembourg
615277	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0046	Belgium	Luxembourg
615278	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0047	Belgium	Namur
615279	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0048	Belgium	Namur
615280	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0049	Belgium	Namur
615281	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0050	Belgium	Namur
615282	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0051f	Belgium	Namur
615283	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0052	Belgium	Namur
615284	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0053	Belgium	Antwerp
615285	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0054	Belgium	Antwerp
615286	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0055	Belgium	Luxembourg
615287	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0056	Belgium	Luxembourg
615288	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0057	Belgium	Luxembourg
615289	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0058	Belgium	Namur
615290	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0059	Belgium	Namur
615291	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0060	Belgium	Namur
615292	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0061	Belgium	Namur
615293	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0062	Belgium	Luxembourg
615294	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0063	Belgium	Luxembourg
615295	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0064	Belgium	Namur
615296	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0065	Belgium	Namur
615297	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0066	Belgium	Namur
615298	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0067	Belgium	Namur
615299	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0068	Belgium	Namur
615300	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0069	Belgium	Namur
615301	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0070	Belgium	Luxembourg
615302	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0071	Belgium	Luxembourg
615303	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0072	Belgium	Luxembourg
615304	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0073	Belgium	Namur
615305	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0074	Belgium	Namur
615306	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0075	Belgium	Namur
615307	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0076	Belgium	Namur
615308	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0077	Belgium	Namur
615309	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0078	Belgium	Luxembourg
615310	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0079	Belgium	Luxembourg
615311	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0080	Belgium	Luxembourg
615312	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0081	Belgium	
615313	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0082	Belgium	
615314	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0083	Belgium	
615315	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0084	Belgium	
615316	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0085	Belgium	

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
615317	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0086	Belgium	
615318	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0087	Belgium	
615319	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0088	Belgium	
615320	<i>aestivum</i> subsp. <i>spelta</i>	GBXTS-0089	Belgium	Luxembourg
615321	<i>aestivum</i> subsp. <i>aestivum</i>	Bai Mong Mai	China	
615322	<i>aestivum</i> subsp. <i>aestivum</i>	Liang Mai	China	
615323	<i>aestivum</i> subsp. <i>aestivum</i>	Ker Yi 26	China	
615324	<i>aestivum</i> subsp. <i>aestivum</i>	Zhong Zho 634	China	
615325	<i>aestivum</i> subsp. <i>aestivum</i>	Yuan Dong 847	China	
615326	<i>aestivum</i> subsp. <i>aestivum</i>	Fong Kong 8	China	
615327	<i>aestivum</i> subsp. <i>aestivum</i>	Dong Shieh 1	China	
615328	<i>aestivum</i> subsp. <i>aestivum</i>	Beijing 837	China	
615329	<i>aestivum</i> subsp. <i>aestivum</i>	Ng Yi 79-5049	China	
615330	<i>aestivum</i> subsp. <i>aestivum</i>	Jing Her 90 Jian 4	China	
615335	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000120	Hungary	Bekes
615336	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000132	Hungary	Szabolcs-Szatmar
615337	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000396	Hungary	Szolnok
615338	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000403	Hungary	Hajdu-Bihar
615339	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000411	Hungary	Veszprem
615340	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000038	Hungary	Zala
615341	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000051	Hungary	Borsod-Abauj-Zempl
615342	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000052	Hungary	Borsod-Abauj-Zempl
615343	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000055	Hungary	Pest
615344	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000057	Hungary	Csongrad
615345	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000059	Hungary	Csongrad
615346	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000060	Hungary	Szabolcs-Szatmar
615347	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000062	Hungary	Heves
615348	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000064	Hungary	Heves
615349	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000065	Hungary	Heves
615350	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000066	Hungary	Heves
615351	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000067	Hungary	Heves
615352	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000069	Hungary	Heves
615353	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000070	Hungary	Heves
615354	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000071	Hungary	Heves
615355	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000140	Hungary	Borsod-Abauj-Zempl
615356	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000248	Hungary	Fejer
615357	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000394	Hungary	Borsod-Abauj-Zempl
615358	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000397	Hungary	Veszprem
615359	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000398	Hungary	Nograd
615360	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000399	Hungary	Nograd
615361	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000400	Hungary	Borsod-Abauj-Zempl
615362	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000401	Hungary	Pest
615363	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000404	Hungary	Bacs-Kiskun
615364	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000405	Hungary	Szabolcs-Szatmar
615365	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000406	Hungary	Somogy
615366	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000408	Hungary	Borsod-Abauj-Zempl
615367	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000409	Hungary	Gyor-Sopron
615368	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000430	Hungary	Szolnok
615369	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000491	Hungary	Gyor-Sopron
615370	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT000412	Hungary	Csongrad
615371	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT001801	Hungary	Szolnok

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
615372	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003312	Hungary	Csongrad
615373	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003313	Hungary	Szabolcs-Szatmar
615374	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003334	Hungary	Bekes
615375	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003418	Hungary	Bekes
615376	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003768	Hungary	Borsod-Abaуй-Zempl
615377	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003782	Hungary	Heves
615378	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003784	Hungary	Csongrad
615379	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003865	Hungary	Bekes
615380	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003874	Hungary	Szolnok
615381	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT003906	Hungary	Bekes
615382	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004126	Hungary	Zala
615383	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT002105	Hungary	Somogy
615384	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT002116	Hungary	Pest
615385	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004129	Hungary	Nograd
615386	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004555	Hungary	Bacs-Kiskun
615387	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004556	Hungary	Nograd
615388	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004759	Hungary	Csongrad
615389	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004795	Hungary	Somogy
615390	<i>aestivum</i> subsp. <i>aestivum</i>	RCAT004797	Hungary	Fejer
615543	<i>aestivum</i> subsp. <i>aestivum</i>	Alsen	United States	North Dakota
615588	<i>aestivum</i> subsp. <i>aestivum</i>	NC99BGTAG11	United States	North Carolina
615589	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1032	United States	Washington
615590	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1033	United States	Washington
615591	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1034	United States	Washington
615592	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1035	United States	Washington
615593	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1037	United States	Washington
615594	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1040	United States	Washington
615595	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1041	United States	Washington
615596	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1042	United States	Washington
615597	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1043	United States	Washington
615598	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1045	United States	Washington
615599	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1051	United States	Washington
615600	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1052	United States	Washington
615601	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1053	United States	Washington
615602	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1054	United States	Washington
615603	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1055	United States	Washington
615604	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1058	United States	Washington
615605	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1060	United States	Washington
615606	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1061	United States	Washington
615607	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1063	United States	Washington
615608	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1065	United States	Washington
615609	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1067	United States	Washington
615610	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1068	United States	Washington
615611	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1069	United States	Washington
615612	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1070	United States	Washington
615613	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1073	United States	Washington
615614	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1076	United States	Washington
615615	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1077	United States	Washington
615616	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1078	United States	Washington
615617	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1079	United States	Washington
615618	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1083	United States	Washington

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
615619	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1086	United States	Washington
615620	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1087	United States	Washington
615621	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1088	United States	Washington
615622	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1089	United States	Washington
615623	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1091	United States	Washington
615624	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1094	United States	Washington
615625	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1095	United States	Washington
615626	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1096	United States	Washington
615627	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1097	United States	Washington
615628	<i>aestivum</i> subsp. <i>aestivum</i>	96ARS 1099	United States	Washington
615629	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 131	United States	Washington
615630	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 140	United States	Washington
615631	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 148	United States	Washington
615632	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 149	United States	Washington
615633	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 155	United States	Washington
615634	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 135	United States	Washington
615635	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 136	United States	Washington
615636	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 138	United States	Washington
615637	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 159	United States	Washington
615638	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 160	United States	Washington
615639	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 169	United States	Washington
615640	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 170	United States	Washington
615641	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 172	United States	Washington
615642	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 178	United States	Washington
615643	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 179	United States	Washington
615644	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 165	United States	Washington
615645	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 168	United States	Washington
615646	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 181	United States	Washington
615647	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 182	United States	Washington
615648	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 192	United States	Washington
615649	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 195	United States	Washington
615650	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 196	United States	Washington
615651	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 214	United States	Washington
615652	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 216	United States	Washington
615653	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 227	United States	Washington
615654	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 199	United States	Washington
615655	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 200	United States	Washington
615656	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 202	United States	Washington
615657	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 204	United States	Washington
615658	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 205	United States	Washington
615659	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 197	United States	Washington
615660	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 198	United States	Washington
615661	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 219	United States	Washington
615662	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 225	United States	Washington
615663	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 230	United States	Washington
615664	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 209	United States	Washington
615665	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 210	United States	Washington
615666	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 211	United States	Washington
615667	<i>aestivum</i> subsp. <i>aestivum</i>	97ARS 212	United States	Washington
616491	<i>aestivum</i> subsp. <i>aestivum</i>	WQL6WTNS	United States	Washington
616492	<i>aestivum</i> subsp. <i>aestivum</i>	WQL6WTNH	United States	Washington

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
617032	<i>aestivum</i> subsp. <i>aestivum</i>	Lakin	United States	Kansas
617033	<i>aestivum</i> subsp. <i>aestivum</i>	Stanton	United States	Kansas
617053	<i>aestivum</i>	Sisson	United States	Virginia
617054	<i>aestivum</i>	Century II	United States	Virginia
617055	<i>aestivum</i>	USG 3209	United States	Virginia
617061	<i>aestivum</i> subsp. <i>aestivum</i>	NW97S343	United States	Nebraska
617062	<i>aestivum</i> subsp. <i>aestivum</i>	NW96S016	United States	Nebraska
617063	<i>aestivum</i> subsp. <i>aestivum</i>	NW97S154	United States	Nebraska
617064	<i>aestivum</i> subsp. <i>aestivum</i>	N95L11881	United States	Nebraska
617065	<i>aestivum</i> subsp. <i>aestivum</i>	97L9520	United States	Nebraska
617066	<i>aestivum</i> subsp. <i>aestivum</i>	97L9521	United States	Nebraska
617067	<i>aestivum</i> subsp. <i>aestivum</i>	97L9522	United States	Nebraska
617068	<i>aestivum</i> subsp. <i>aestivum</i>	97L9531	United States	Nebraska
617069	<i>aestivum</i> subsp. <i>aestivum</i>	96MD7413-58	United States	Nebraska
617070	<i>aestivum</i> subsp. <i>aestivum</i>	96MD7413-36	United States	Nebraska
617071	<i>aestivum</i> subsp. <i>aestivum</i>	96MD7110-71	United States	Nebraska
617072	<i>aestivum</i> subsp. <i>aestivum</i>	Macon	United States	Washington
617073	<i>aestivum</i> subsp. <i>aestivum</i>	Tara	United States	Washington
618588	<i>aestivum</i>	766	United States	Virginia
618611	<i>aestivum</i>	Coker 9025	United States	
618734	<i>aestivum</i> subsp. <i>aestivum</i>	Iona	United States	Idaho
619051	<i>aestivum</i>	38247	United States	Virginia
619052	<i>aestivum</i>	38158	United States	Virginia
619072	<i>aestivum</i>	Granite	Germany	
619086	<i>aestivum</i> subsp. <i>aestivum</i>	Explorer	United States	Montana
619089	<i>aestivum</i>	NuFrontier	United States	
619098	<i>aestivum</i> subsp. <i>aestivum</i>	Wahoo	United States	Nebraska
619166	<i>aestivum</i> subsp. <i>aestivum</i>	BigSky	United States	Montana
619167	<i>aestivum</i> subsp. <i>aestivum</i>	NuSky	United States	Montana
619197	<i>aestivum</i>	Mitchell	United States	
619198	<i>aestivum</i>	NuHorizon	United States	
619199	<i>aestivum</i>	Dumas	United States	
619200	<i>aestivum</i>	Charter	United States	
619231	<i>aestivum</i> subsp. <i>aestivum</i>	N96L9970	United States	Nebraska
619354	<i>aestivum</i> subsp. <i>aestivum</i>	99ID388	United States	Nebraska
619355	<i>aestivum</i> subsp. <i>aestivum</i>	99ID389	United States	Nebraska
619356	<i>aestivum</i> subsp. <i>aestivum</i>	99ID435	United States	Nebraska
619357	<i>aestivum</i> subsp. <i>aestivum</i>	99ID450	United States	Nebraska
619358	<i>aestivum</i> subsp. <i>aestivum</i>	99ID454	United States	Nebraska
619359	<i>aestivum</i> subsp. <i>aestivum</i>	99ID477	United States	Nebraska
619360	<i>aestivum</i> subsp. <i>aestivum</i>	99ID484	United States	Nebraska
619361	<i>aestivum</i> subsp. <i>aestivum</i>	99ID490	United States	Nebraska
619362	<i>aestivum</i> subsp. <i>aestivum</i>	99ID496	United States	Nebraska
619363	<i>aestivum</i> subsp. <i>aestivum</i>	99ID498	United States	Nebraska
619364	<i>aestivum</i> subsp. <i>aestivum</i>	99ID510	United States	Nebraska
619365	<i>aestivum</i> subsp. <i>aestivum</i>	99ID516	United States	Nebraska
619366	<i>aestivum</i> subsp. <i>aestivum</i>	99ID520	United States	Nebraska
619367	<i>aestivum</i> subsp. <i>aestivum</i>	99ID524	United States	Nebraska
619368	<i>aestivum</i> subsp. <i>aestivum</i>	99ID529	United States	Nebraska
619369	<i>aestivum</i> subsp. <i>aestivum</i>	99ID536	United States	Nebraska
619370	<i>aestivum</i> subsp. <i>aestivum</i>	99ID546	United States	Nebraska

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
619371	<i>aestivum</i> subsp. <i>aestivum</i>	99ID548	United States	Nebraska
619372	<i>aestivum</i> subsp. <i>aestivum</i>	99ID554	United States	Nebraska
619373	<i>aestivum</i> subsp. <i>aestivum</i>	99ID569	United States	Nebraska
619374	<i>aestivum</i> subsp. <i>aestivum</i>	99ID590	United States	Nebraska
619375	<i>aestivum</i> subsp. <i>aestivum</i>	99ID594	United States	Nebraska
619376	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008068	United States	Nebraska
619377	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008069	United States	Nebraska
619378	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008070	United States	Nebraska
619379	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008075	United States	Nebraska
619380	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008093	United States	Nebraska
619381	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008094	United States	Nebraska
619382	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008106	United States	Nebraska
619383	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008109	United States	Nebraska
619384	<i>aestivum</i> subsp. <i>aestivum</i>	NWX008118	United States	Nebraska
619397	<i>aestivum</i> subsp. <i>aestivum</i>	Buck Pronto	Netherlands	
619419	<i>aestivum</i> subsp. <i>aestivum</i>	Declo	United States	
619426	<i>aestivum</i>	Keystone	United States	
619428	<i>aestivum</i>	Bonus	United States	
619608	<i>aestivum</i>	Hanna	United States	
619609	<i>aestivum</i>	Knudson	United States	
619610	<i>aestivum</i>	CDC Falcon	United States	
619611	<i>aestivum</i>	25R78	United States	
619612	<i>aestivum</i>	25R42	United States	
619613	<i>aestivum</i>	25R23	United States	
619633	<i>aestivum</i>	Nahuatl F2000	Mexico	Mexico
619634	<i>aestivum</i>	Tlaxcala F2000	Mexico	Mexico
619635	<i>aestivum</i>	Juchi F2000	Mexico	Mexico
620628	<i>aestivum</i> subsp. <i>aestivum</i>	A93324S-76kbr	United States	Idaho
620629	<i>aestivum</i> subsp. <i>aestivum</i>	IDO513	United States	Idaho
620630	<i>aestivum</i> subsp. <i>aestivum</i>	IDO517	United States	Idaho
620631	<i>aestivum</i> subsp. <i>aestivum</i>	IDO526	United States	Idaho
620632	<i>aestivum</i> subsp. <i>aestivum</i>	Gary	United States	Idaho
620633	<i>aestivum</i> subsp. <i>aestivum</i>	IDO556	United States	Idaho
620634	<i>aestivum</i> subsp. <i>aestivum</i>	IDO563	United States	Idaho
620635	<i>aestivum</i> subsp. <i>aestivum</i>	IDO569	United States	Idaho
620636	<i>aestivum</i> subsp. <i>aestivum</i>	IDO571	United States	Idaho
620637	<i>aestivum</i> subsp. <i>aestivum</i>	IDO576	United States	Idaho
620638	<i>aestivum</i> subsp. <i>aestivum</i>	IDO580	United States	Idaho
620641	<i>aestivum</i> subsp. <i>aestivum</i>	3870118	United States	Oregon
620642	<i>aestivum</i> subsp. <i>aestivum</i>	3900394	United States	Oregon
620643	<i>aestivum</i> subsp. <i>aestivum</i>	3930396	United States	Oregon
620644	<i>aestivum</i> subsp. <i>aestivum</i>	3930153	United States	Oregon
620645	<i>aestivum</i> subsp. <i>aestivum</i>	900327	United States	Oregon
620646	<i>aestivum</i> subsp. <i>aestivum</i>	889294	United States	Oregon
620647	<i>aestivum</i> subsp. <i>aestivum</i>	3930324	United States	Oregon
620648	<i>aestivum</i> subsp. <i>aestivum</i>	899210	United States	Oregon
620649	<i>aestivum</i> subsp. <i>aestivum</i>	3880045	United States	Oregon
620650	<i>aestivum</i> subsp. <i>aestivum</i>	910102	United States	Oregon
620651	<i>aestivum</i> subsp. <i>aestivum</i>	880494	United States	Oregon
620652	<i>aestivum</i> subsp. <i>aestivum</i>	3900438	United States	Oregon
620653	<i>aestivum</i> subsp. <i>aestivum</i>	3880039	United States	Oregon

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
620660	<i>aestivum</i> subsp. <i>aestivum</i>	3930264	United States	Oregon
620661	<i>aestivum</i> subsp. <i>aestivum</i>	910147	United States	Oregon
620662	<i>aestivum</i> subsp. <i>aestivum</i>	908294	United States	Oregon
620663	<i>aestivum</i> subsp. <i>aestivum</i>	900438	United States	Oregon
620664	<i>aestivum</i> subsp. <i>aestivum</i>	861555	United States	Oregon
620665	<i>aestivum</i> subsp. <i>aestivum</i>	890074	United States	Oregon
620666	<i>aestivum</i> subsp. <i>aestivum</i>	3950719	United States	Oregon
620667	<i>aestivum</i> subsp. <i>aestivum</i>	3950803	United States	Oregon
620668	<i>aestivum</i> subsp. <i>aestivum</i>	3950804	United States	Oregon
620669	<i>aestivum</i> subsp. <i>aestivum</i>	3930267	United States	Oregon
620670	<i>aestivum</i> subsp. <i>aestivum</i>	3920022	United States	Oregon
620671	<i>aestivum</i> subsp. <i>aestivum</i>	3870552	United States	Oregon
620672	<i>aestivum</i> subsp. <i>aestivum</i>	3930326	United States	Oregon
620673	<i>aestivum</i> subsp. <i>aestivum</i>	3930330	United States	Oregon
620674	<i>aestivum</i> subsp. <i>aestivum</i>	908237	United States	Oregon
620675	<i>aestivum</i> subsp. <i>aestivum</i>	899407	United States	Oregon
620676	<i>aestivum</i> subsp. <i>aestivum</i>	908134	United States	Oregon
620677	<i>aestivum</i> subsp. <i>aestivum</i>	910379	United States	Oregon
620678	<i>aestivum</i> subsp. <i>aestivum</i>	2626	United States	Oregon
620679	<i>aestivum</i> subsp. <i>aestivum</i>	3870115	United States	Oregon
620680	<i>aestivum</i> subsp. <i>aestivum</i>	3870634	United States	Oregon
620681	<i>aestivum</i> subsp. <i>aestivum</i>	3930387	United States	Oregon
620682	<i>aestivum</i> subsp. <i>aestivum</i>	3930257	United States	Oregon
620683	<i>aestivum</i> subsp. <i>aestivum</i>	3950865	United States	Oregon
620684	<i>aestivum</i> subsp. <i>aestivum</i>	899302	United States	Oregon
620685	<i>aestivum</i> subsp. <i>aestivum</i>	1922	United States	Oregon
620686	<i>aestivum</i> subsp. <i>aestivum</i>	3870538	United States	Oregon
620687	<i>aestivum</i> subsp. <i>aestivum</i>	3930055	United States	Oregon
620688	<i>aestivum</i> subsp. <i>aestivum</i>	920548	United States	Oregon
620689	<i>aestivum</i> subsp. <i>aestivum</i>	3920068	United States	Oregon
620690	<i>aestivum</i> subsp. <i>aestivum</i>	900327	United States	Oregon
620691	<i>aestivum</i> subsp. <i>aestivum</i>	900192	United States	Oregon
620692	<i>aestivum</i> subsp. <i>aestivum</i>	900381	United States	Oregon
620693	<i>aestivum</i> subsp. <i>aestivum</i>	908289	United States	Oregon
620694	<i>aestivum</i> subsp. <i>aestivum</i>	908164	United States	Oregon
620695	<i>aestivum</i> subsp. <i>aestivum</i>	889303	United States	Oregon
620696	<i>aestivum</i> subsp. <i>aestivum</i>	899511	United States	Oregon
620697	<i>aestivum</i> subsp. <i>aestivum</i>	908127	United States	Oregon
620698	<i>aestivum</i> subsp. <i>aestivum</i>	910231	United States	Oregon
620699	<i>aestivum</i> subsp. <i>aestivum</i>	900031	United States	Oregon
620700	<i>aestivum</i> subsp. <i>aestivum</i>	908110	United States	Oregon
620701	<i>aestivum</i> subsp. <i>aestivum</i>	910262	United States	Oregon
620702	<i>aestivum</i> subsp. <i>aestivum</i>	3940160	United States	Oregon
620703	<i>aestivum</i> subsp. <i>aestivum</i>	3940263	United States	Oregon
620704	<i>aestivum</i> subsp. <i>aestivum</i>	3950710	United States	Oregon
620705	<i>aestivum</i> subsp. <i>aestivum</i>	3950712	United States	Oregon
620706	<i>aestivum</i> subsp. <i>aestivum</i>	3950727	United States	Oregon
620707	<i>aestivum</i> subsp. <i>aestivum</i>	3950728	United States	Oregon
620708	<i>aestivum</i> subsp. <i>aestivum</i>	3950730	United States	Oregon
620709	<i>aestivum</i> subsp. <i>aestivum</i>	3950733	United States	Oregon
620710	<i>aestivum</i> subsp. <i>aestivum</i>	3950750	United States	Oregon

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
620711	<i>aestivum</i> subsp. <i>aestivum</i>	OR9630057	United States	Oregon
620712	<i>aestivum</i> subsp. <i>aestivum</i>	OR9630058	United States	Oregon
620713	<i>aestivum</i> subsp. <i>aestivum</i>	OR9630062	United States	Oregon
620714	<i>aestivum</i> subsp. <i>aestivum</i>	OR9630064	United States	Oregon
620715	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951041	United States	Oregon
620716	<i>aestivum</i> subsp. <i>aestivum</i>	928132	United States	Oregon
620717	<i>aestivum</i> subsp. <i>aestivum</i>	940005	United States	Oregon
620718	<i>aestivum</i> subsp. <i>aestivum</i>	940043	United States	Oregon
620719	<i>aestivum</i> subsp. <i>aestivum</i>	940214	United States	Oregon
620720	<i>aestivum</i> subsp. <i>aestivum</i>	940228	United States	Oregon
620721	<i>aestivum</i> subsp. <i>aestivum</i>	940078	United States	Oregon
620722	<i>aestivum</i> subsp. <i>aestivum</i>	940079	United States	Oregon
620723	<i>aestivum</i> subsp. <i>aestivum</i>	940447	United States	Oregon
620724	<i>aestivum</i> subsp. <i>aestivum</i>	940099	United States	Oregon
620725	<i>aestivum</i> subsp. <i>aestivum</i>	940105	United States	Oregon
620726	<i>aestivum</i> subsp. <i>aestivum</i>	940205	United States	Oregon
620727	<i>aestivum</i> subsp. <i>aestivum</i>	940719	United States	Oregon
620728	<i>aestivum</i> subsp. <i>aestivum</i>	942732	United States	Oregon
620729	<i>aestivum</i> subsp. <i>aestivum</i>	942742	United States	Oregon
620730	<i>aestivum</i> subsp. <i>aestivum</i>	942969	United States	Oregon
620731	<i>aestivum</i> subsp. <i>aestivum</i>	943065	United States	Oregon
620732	<i>aestivum</i> subsp. <i>aestivum</i>	943075	United States	Oregon
620733	<i>aestivum</i> subsp. <i>aestivum</i>	943302	United States	Oregon
620734	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-739	United States	Oregon
620735	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-740	United States	Oregon
620736	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-740	United States	Oregon
620737	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-761	United States	Oregon
620738	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-761	United States	Oregon
620739	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-761	United States	Oregon
620740	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-767	United States	Oregon
620741	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-939	United States	Oregon
620742	<i>aestivum</i> subsp. <i>aestivum</i>	SEG95-997	United States	Oregon
620743	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951007	United States	Oregon
620744	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951032	United States	Oregon
620745	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951036	United States	Oregon
620746	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951047	United States	Oregon
620747	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951104	United States	Oregon
620748	<i>aestivum</i> subsp. <i>aestivum</i>	SEG951128	United States	Oregon
620749	<i>aestivum</i> subsp. <i>aestivum</i>	908127	United States	Oregon
620750	<i>aestivum</i> subsp. <i>aestivum</i>	910227	United States	Oregon
620751	<i>aestivum</i> subsp. <i>aestivum</i>	910231	United States	Oregon
620752	<i>aestivum</i> subsp. <i>aestivum</i>	910263	United States	Oregon
620753	<i>aestivum</i> subsp. <i>aestivum</i>	920070	United States	Oregon
620754	<i>aestivum</i> subsp. <i>aestivum</i>	920450	United States	Oregon
620755	<i>aestivum</i> subsp. <i>aestivum</i>	941102	United States	Oregon
620756	<i>aestivum</i> subsp. <i>aestivum</i>	943202	United States	Oregon
620757	<i>aestivum</i> subsp. <i>aestivum</i>	940205	United States	Oregon
620758	<i>aestivum</i> subsp. <i>aestivum</i>	941379	United States	Oregon
620759	<i>aestivum</i> subsp. <i>aestivum</i>	OR9800949	United States	Oregon
620766	<i>aestivum</i> subsp. <i>aestivum</i>	Avalanche	United States	Colorado

PI Assignments in *Triticum* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
620884	<i>turgidum</i> subsp. <i>durum</i> <i>turgidum</i> subsp. <i>dicoccum</i>	to	Iran	
628270	<i>turgidum</i> subsp. <i>turanicum</i> <i>turgidum</i> subsp. <i>turgidum</i>			
628343	<i>aestivum</i>	91426E39	United States	
628344	<i>aestivum</i>	901146E15	United States	
628363	<i>aestivum</i>	Eldon	United States	
628368	<i>turgidum</i> subsp. <i>durum</i>	Orita	United States	
628369	<i>turgidum</i> subsp. <i>durum</i>	Alamo	United States	
628640	<i>aestivum</i> subsp. <i>aestivum</i>	Finch	United States	Washington
628641	<i>aestivum</i> subsp. <i>aestivum</i>	Chukar	United States	Washington
628644	<i>aestivum</i> subsp. <i>aestivum</i>	UCRBW01-1	United States	California
628645	<i>aestivum</i> subsp. <i>aestivum</i>	UCRBW01-2	United States	California
628646	<i>aestivum</i> subsp. <i>aestivum</i>	UCRBW01-3	United States	California
628647	<i>aestivum</i> subsp. <i>aestivum</i>	UCRBW01-4	United States	California
628648	<i>aestivum</i> subsp. <i>aestivum</i>	UCRBW01-5	United States	California
628649	<i>aestivum</i> subsp. <i>aestivum</i>	UCRBW01-6	United States	California
628650	<i>aestivum</i> subsp. <i>aestivum</i>	UCRD01-1	United States	California
628651	<i>turgidum</i> subsp. <i>durum</i>	UCRD01-2	United States	California
628652	<i>turgidum</i> subsp. <i>durum</i>	UCRD01-3	United States	California
628653	<i>turgidum</i> subsp. <i>durum</i>	UCRD01-4	United States	California
628654	<i>turgidum</i> subsp. <i>durum</i>	UCRD01-5	United States	California
628655	<i>turgidum</i> subsp. <i>durum</i>	UCRD01-6	United States	California
628987	<i>aestivum</i> subsp. <i>aestivum</i>	Residence	Netherlands	
628988	<i>aestivum</i> subsp. <i>aestivum</i>	Semper	Netherlands	

PI Assignments in *Secale* and *X Triticosecale* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
615331	<i>cereale</i> subsp. <i>cereale</i>	RCAT013928	Hungary	Bacs-Kiskun
615332	<i>sylvestre</i>	RCAT042415	Hungary	Pest
615333	<i>sylvestre</i>	RCAT042416	Hungary	Pest
615334	<i>sylvestre</i>	RCAT042417	Hungary	Pest
615414	<i>X Triticosecale</i> sp.	Triticale 64	Hungary	
615529	<i>X Triticosecale</i> sp.	Alzo	Poland	
618640	<i>cereale</i> subsp. <i>cereale</i>	17583	Portugal	
618641	<i>cereale</i> subsp. <i>cereale</i>	17584	Portugal	
618642	<i>cereale</i> subsp. <i>cereale</i>	17585	Portugal	
618643	<i>cereale</i> subsp. <i>cereale</i>	17592	Portugal	
618644	<i>cereale</i> subsp. <i>cereale</i>	17537	Portugal	
618645	<i>cereale</i> subsp. <i>cereale</i>	17600	Portugal	
618646	<i>cereale</i> subsp. <i>cereale</i>	17542	Portugal	
618647	<i>cereale</i> subsp. <i>cereale</i>	17545	Portugal	
618648	<i>cereale</i> subsp. <i>cereale</i>	17546	Portugal	
618649	<i>cereale</i> subsp. <i>cereale</i>	17547	Portugal	

PI Assignments in *Secale* and *X Triticosecale* from January 2001–February 2002.

PI number	Taxon	Cultivar name or Identification number	Country	State/Province
618650	<i>cereale</i> subsp. <i>cereale</i>	17548	Portugal	
618651	<i>cereale</i> subsp. <i>cereale</i>	17602	Portugal	
618652	<i>cereale</i> subsp. <i>cereale</i>	17603	Portugal	
618653	<i>cereale</i> subsp. <i>cereale</i>	17605	Portugal	
618654	<i>cereale</i> subsp. <i>cereale</i>	17564	Portugal	
618655	<i>cereale</i> subsp. <i>cereale</i>	17574	Portugal	
618656	<i>cereale</i> subsp. <i>cereale</i>	17683	Portugal	
618657	<i>cereale</i> subsp. <i>cereale</i>	17694	Portugal	
618658	<i>cereale</i> subsp. <i>cereale</i>	17695	Portugal	
618659	<i>cereale</i> subsp. <i>cereale</i>	17697	Portugal	
618660	<i>cereale</i> subsp. <i>cereale</i>	Kuckucks Riesengebirg	Germany	
618661	<i>cereale</i> subsp. <i>cereale</i>	Osterreich	Austria	
618662	<i>cereale</i> subsp. <i>afghanicum</i>	30225	Armenia	
618663	<i>cereale</i> subsp. <i>ancestrale</i>	14357	Turkey	
618664	<i>cereale</i> subsp. <i>ancestrale</i>	17791	Sweden	
618665	<i>cereale</i> subsp. <i>ancestrale</i>	24263	Poland	
618666	<i>cereale</i> subsp. <i>ancestrale</i>	30226	Turkey	
618667	<i>cereale</i> subsp. <i>dighoricum</i>	17785	Sweden	
618668	<i>cereale</i> subsp. <i>dighoricum</i>	30224	Russian Federation	
618669	<i>cereale</i> subsp. <i>rigidum</i>	14193	Turkey	
618670	<i>cereale</i> subsp. <i>segetale</i>	7520	Azerbaijan	
618671	<i>cereale</i> subsp. <i>segetale</i>	14372	Turkey	
618672	<i>cereale</i> subsp. <i>segetale</i>	24269	Poland	
618673	<i>cereale</i> subsp. <i>segetale</i>	1782/94	Turkey	
618674	<i>sylvestre</i>	838/96	Ukraine	
618675	<i>sylvestre</i>	839/96	Bulgaria	
618676	<i>sylvestre</i>	14185	Poland	
618677	<i>sylvestre</i>	30227	Ukraine	
618678	<i>vavilovii</i>	30228	Armenia	
618678	<i>vavilovii</i>	833/96	Georgia	
618678	<i>vavilovii</i>	1783/94	Turkey	
618678	<i>vavilovii</i>	1784/94	Turkey	
618678	<i>vavilovii</i>	14197	Poland	
619184	<i>cereale</i> subsp. <i>segetale</i>	SD 83-3	United States	South Dakota
620762	<i>X Triticosecale</i> sp.	TCLF-AN-31	Mexico	Coahuila
620763	<i>X Triticosecale</i> sp.	TCLF-AN-34	Mexico	Coahuila
628642	<i>cereale</i> subsp. <i>cereale</i>	UCRR1-2001	United States	California
628643	<i>cereale</i> subsp. <i>cereale</i>	UCRR2-2001	United States	California
628656	<i>X Triticosecale</i> sp.	UCRTCL1-2001	United States	California
628657	<i>X Triticosecale</i> sp.	UCRTCL2-2001	United States	California
628658	<i>X Triticosecale</i> sp.	UCRTCL3-2001	United States	California

V. CATALOGUE OF GENE SYMBOLS FOR WHEAT: 2002 SUPPLEMENT

R.A. McIntosh¹, K.M. Devos², J. Dubcovsky³, and W.J. Rogers⁴.

¹Plant Breeding Institute, The University of Sydney, 107 Cobbitty Road, Cobbitty, N.S.W., Australia, 2570.

²John Innes Centre, Norwich Research Park, Colney, Norwich, Norfolk, NR4 7UH, U.K.

³Department of Agronomy and Range Science, University of California, Davis, 95616 CA, U.S.A.

⁴Catedra de Genetica y Fitotecnia, Universidad Nacional del Centro de la Provincia de Buenos Aires, 7300 Azul, Argentina.

The most recent edition of the Catalog appeared in the Proceedings of the 9th International Wheat Genetics Symposium Vol. 5 (Slinkard AE ed, University Extension Press, University of Saskatchewan, Saskatoon, Canada). A modified version is displayed on the Graingenes Website <http://wheat.pw.usda.gov/>

The 2000 and 2001 supplements are included in *Annual Wheat Newsletters* and *Wheat Information Service* and are listed in the Graingenes Website. The present Supplement will be offered to editors/curators for similar listing.

Revisions.**10. Laboratory Designators for DNA markers**

barc Cregan, P.

USDA-ARS

Bdge 006, HH-19 BARC-WE

Beltsville, MD 20705-2350

U.S.A.

cnl Sorrells, M.

Department of Plant Breeding & Biometry

Cornell University

252 Emerson Hall

Ithaca, NY 14853

U.S.A.

mes12@cornell.edu

gdm Röder, M.S.

(Gatersleben D-genome microsatellite*)

Institut für Pflanzengenetik und Kulturpflanzenforschung (IPK)

Corrensstr. 3

D-06466 Gatersleben

Germany

roder@mendel.ipk-gatersleben.de

isc Luigi Cattivelli

Istituto Sperimentale Cerealicoltura *

Via S. Protaso, 302

Fiorenzuola d'Arca (PC)

I-29017 Italy

l.cattivelli@iol.it

unl Gill, K.
 Department of Agronomy
 362H Plant Science
 P.O. Box 830915
 University of Nebraska
 Lincoln NE68583-0915USA
 U.S.A.
 kgill@unl.edu

uaz Helentjaris, T.
 (University of Arizona*)
 Pioneer Hi-Bred International
 7250 N.W. 62nd Avenue
 Johnston, IA 50131
 U.S.A.

ucg Hasselkorn, R.
 Department of Molecular Genetics and Cell Biology
 University of Chicago*
 Chicago, IL 60637
 U.S.A.
 rh01@midway.uchicago.edu

Gross Morphology : Spike Characteristics

5. Elongated glume

PI. Revise:

PI. [*P* {911}; *Eg* {922}; *P-A^{pol}1* {0254}; *P-A^{pet}1* {0254}]. 7AL {922, 1547}, 7A, or 7B (based on linkage of 0.2 with a gene for red coleoptile {922}).

ii: Saratovskaya 29*8//Novsibirskaya 67*2/*T. polonicum* {922}.

itv: P-LD222 = LD222*11/*T. turgidum* var *polonicum* {1546,1547}.

tv: *T. polonicum* {0254}; *T. petropavlovskyi* {0254}.

ma: *Xgwm260*-7A (S) – 2.3 cM – *P1* – 5.6 cM – *Xgwm1083*-7A (L) {0254};

Xgwm890-7A – 2.1 cM – *P1* {0254}.

Add at the end of the 'Elongated glume' section:

Note: The loci determining elongated glumes in the *T. turanicum* and *T. durum* conv. *falcatum* are not homoeologous to the *P* loci in the centromeric region of the group-7 chromosomes {0254}.

Alkylresocinols Content in Grain

Ar1. {0250} 5AL {0281}. High alkylresocinols content is dominant {0281}.

tv: Langdon {0281}.

ar1 {0281}. **tv:** Ardente {0281}. This cultivar has a low content compared to all tested durum and common wheats {0281}.

Aluminium Tolerance

Alt2. **ma:** Add: 'Alt2 cosegregated with *Xbcd1230*-4D and fell within the interval *Xgdm125*-4D – 4.8 cM – *Alt2* – 1.1 cM – *Xpsr914*-4D {0248}.'

Anthocyanin Pigmentation

3. Red/purple coleoptiles

Replace the previous entries with:

There is an orthologous gene series on the short arms of homoeologous group 7. The 'a' alleles confer red coleoptiles.

Rc-A1a {0250}.	[Rc1, R {401}].	7A {769,1293}, 7AS {0250}.	s: CS*6/Hope 7A {1293}.
	v: Hope <i>Rc-B1</i>		
	ma: <i>Rc-A1</i> (distal) – 11.9 cM – <i>Xgwm913-7A</i> {0250}.		
Rc-B1a {0250}.	[Rc2, R2 {401}].	7B {742}, 7BS {401, 769, 0250}	s: CS*6/Hope 7B {769}.
	v: Hope <i>Rc-A1</i> .		
	ma: <i>Xgwm263-7B</i> – 26.1 cM – <i>Rc-B1</i> – 11.0 cM – <i>Xgwm1184</i> {0250}.		
Rc-D1a {0250}.	[Rc3].	7D {596}, 7DS {1241, 1444, 0250}.	
	v: Mironovskaya 808 {1444}; Tetra Canthatch/ <i>Ae. squarrosa</i> var. <i>strangulata</i> RL 5271, RL 5404 {1240}; Tetra Canthatch/ <i>Ae. squarrosa</i> var. <i>meyeri</i> RL 5289, RL 5406 {1240}; Sears' <i>T. dicoccoides/Ae. squarrosa</i> = Sears' Synthetic {596}.		
	ma: <i>Rc-D1</i> (distal) – 3 cM – <i>Xpsr108-7D</i> {180}; <i>Xgwm44-7D</i> – 6.4 cM – <i>Rc-D1</i> – 13.7 cM – <i>Xgwm111-7D</i> {0250}.		

Tahir & Tsunewaki {1453} reported that *T. spelta* var. *duhamelianum* carries genes promoting pigmentation on chromosomes 7A and 7D and genes suppressing pigmentation on 2A, 2B, 2D, 3B, and 6A. Sutka {1444} reported a fourth factor in chromosome 6B and suppressors in 2A, 2B, 2D, 4B, and 6A.

Awnedness

1. Dominant Inhibitors

1.2. Tipped 1

B1 Revise '5AL {1293}.' to '5AL {1293,0242}.'

DNA Markers

Group 1S

Amendments:

Xabg500-1A. Revise the first column to 'Xabg500-1A {280}⁵, 1B,D {0252}¹.'

Xbcd446-1A. Add '(1BL)' in the last column.

Xbcd1124-1A,B. Revise the first column to 'Xbcd1124-1A {280}⁵, 1B {1529}¹, 1D {0252}¹.'

Xbcd1706-1A,B. Revise the first column to 'Xbcd1796-1A {280}⁵, 1B {1529}¹, 1D {0252}¹.'

Xcd099-1B,D. Revise the first column to 'Xcd099-1A {0252}¹, 1B {154}¹, 1D {1529}⁴.'

Xcd0388-1B,D. Revise the first column to 'Xcd0388-1B.I [{1529,0252}]¹, 1D {1529}⁴,' add '[Xcd0388-1B {1529}, Xcd0388a-1B {0252}]' in the second column, and revise the last column to '(1BL, 2B, 3D, 4A,D, 5A,B, A,D).'

Xcd0534-1B. Revise the last column to '(3A, 6A,B,D, 7A).'

Xcd0580-1A. Revise the first column to 'Xcd0580-1A {280}^{1,3,5}, {1529}¹, 1D {0242}.'

Xcd0658-1A,B,D. Revise the first column to 'Xcd0658-1A {280}^{3,5}, {0252}¹, 1B,D {445}¹.'

Xcd01173-1A,B,D. Revise the first column to 'Xcd01173-1A {280}^{3,5}, {0252}¹, 1B {1529}¹, 1D {445}¹.'

Xcd01188-1A,I,B,I,D. Revise the first column to 'Xcd01188-1A.I [{280}]^{3,5}, [{0252}]¹, 1B.I [{1529}]¹, 1D {445}¹,' add '[Xcd01188-1A.{280}^{3,5}, {0252}¹, 1B {1529}¹]' in the second column, and add '(1AL,BL)' in the last column.

Xcd01340-1B. Revise the first column to 'Xcd01340-1A {0252}, 1B {1529}, 1D {0252}' and add '(1BL)' to the last column.

Xcmwg645-1A,2. Revise the first column to 'Xcmwg645-1A.I {280}^{3,5}, [{0252}]¹, 1B.I, D.I [{0252}]¹' and add '[Xcmwg645a-1A,B,D {0252}]' in the second column.

Xgwm18-1B. Add '(4B)' in the last column.

Xgwm33-1A. Revise the first column to 'Xgwm33-1A {1226}¹, 1B {0270}².' and add '(1BL)' in the last column.

Xgwm136-1A. Revise the first column to 'Xgwm136-1A {9929},{0269}².'

Xgwm273-1B. Revise the first column to 'Xgwm273-1B {9929},{0270}².'

Xgwm413-1B. Revise the first column to 'Xgwm413-1B {9929},{0270}².'

XksuE19-1A,B,D. Revise the last column to '(6D, 7B).'

XksuF43-1B,1,2. Revise the first column to 'XksuF43-1A {252}, 1B,1,2 {1529}, 1D {0252}'.'

Xmwg60-1A. Revise the first column to 'Xmwg60-1A {280}^{1,5}, 1B,D {0250}]¹.'

Xmwg68-1A,B. Revise the first column to 'Xmwg68-1A {280}⁵, {0252}¹, 1B {1529}¹, 1D {0252}'.'

Xmwg837-1B.1,D. Revise the first column to ‘*Xmwg837-1A* [{0252}], *1B.1,D* {1529}.’ and add ‘[*Xmwg837a-1A* {0252}]’ in the second column.

Xmwg938-1B,D. Revise the first column to ‘*Xmwg938-1A* [{0252}], *1B.1* [{1529, 0252}], *1D* {1529}’, add ‘[*Xmwg938a-1A,B* {0252}, *Xmwg938-1B* {1529}]’ in the second column, and revise the last column to ‘(1BL, 7A).’

Xmwg2021-1A.2,,2. Revise the first column to ‘*Xmwg2021-1A.1* {280}^{3,5}, *1A.2* {280}¹, *1B.1,D* [{0252}]’, add ‘[*Xmwg2021a-1B,D* {0252}]’ in the second column, and revise the last column to ‘(1BL, 2A, 3A).’

Xmwg2048-1A. Revise the first column to ‘*Xmwg2048-1A* {282}³, {0252}¹, *1B,D* {0252}¹’.

Xmwg2083-1A. Revise the first column to ‘*Xmwg2083-1A* {280}⁵, [{0250}]¹, *1B.1,D* [{0252}]¹’, add ‘[*Xmwg2083a-1B,D* {0252}]’ in the second column, and add ‘(1BL)’ in the last column.

Xmwg2245-1D. Revise the first column to ‘*Xmwg2245-1A,B* {0252}, *1D* {0135}’.

Xpsr596-1A,B,D. Add ‘(2B, 3A).’ in the last column.

Xrz244-1A. Revise the first column to ‘*Xrz244-1A* {1529}, *1B,D* {0252}’.

Xsfr2(Lrk10)-1A. Revise the first column to ‘*Xsfr2(Lrk10)-1A* [{356, 0252}], *1B,D* [{0252}]’ and revise the second column to ‘[*Lrk10* {356}, *XLrk10-1A,B,D* {0252}]’.

Xutv1391-1I: Revise the first column to ‘*Xutv1391-1A* {9959}², *1B* {0269}²’.

Add:

<i>Xabg53-1A,B,D</i> {0252}.		ABG53.	
<i>Xabg59-1B,D</i> {0252}.		ABG59.	
<i>Xabg74-1A,B,D</i> {0252}.		ABG74.	
<i>Xabg494-1A,B,D</i> {0252}.		ABG494.	
<i>Xbcd368-1A</i> {0242}.		BCD368.	
<i>Xbcd371-1B</i> {0275}.		BCD371.	
<i>Xbcd762-1A.2,B.2,D</i> [{0252}].	[<i>Xbcd762a-1A,B,D</i> {0252}].	BCD762.	(1AL,BL)
<i>Xbcd1340-1A,B,D</i> {0252}.		BCD1340.	
<i>Xcdol27-1A,B.1,D</i> [{0252}].	[<i>Xcdol27a-1A,B,D</i> {0252}].	CDO127.	(1B, 3A).
<i>Xcdol27-1B.2</i> [{0252}].	[<i>Xcdol27b-1B</i> {0252}].	CDO127.	(1A,B,D, 3A).
<i>Xcdol580-1A,B.1,D</i> [{0252}].	[<i>Xcdol580a-1A,B,D</i> {0252}].	CDO580.	(1B).
<i>Xcdol580-1B.2</i> [{0252}].	[<i>Xcdol580b-1B</i> {0252}].	CDO580.	(1A,B,D).
<i>Xcdol618-1B</i> {0269} ² .		CDO618.	(1A,B,D).
<i>Xcdol373-1B</i> {0269} ² .		CDO1373.	(1BL).
<i>Xcdol423-1A,B,D</i> {0252}.		CDO1423.	
<i>Xmwg835-1A,B.1,D</i> [{0252}].	[<i>Xmwg835a-1A,B,D</i> {0252}].	MWG835.	(1BL, 2A, 5A).
<i>Xmwg913-1B.1</i> [{0252}].	[<i>Xmwg913a-1B</i> {0252}].	MWG913.	(1BS).
<i>Xmwg913-1B.2</i> [{0252}].	[<i>Xmwg913b-1B</i> {0252}].	MWG913.	(1BS).
<i>Xmwg2056-1A,B,D</i> [{0252}].	[<i>Xmwg2056a-1A,B,D</i> {0252}].	MWG2056.	(1BL).
<i>Xmwg2148-1A,B.1,D</i> [{0252}].	[<i>Xmwg2148a-1A,B,D</i> {0252}].	MWG2148.	(1BL).
<i>Xmwg2197-1A,B</i> {0252}.		MWG2197.	
<i>Xsun18-1B</i> {0256}.		SUN 18F/SUN18R.	
<i>Xuaz299-1B</i> {0269} ² .		UAZ299.	
<i>Xutv1366-1A.1</i> {0269} ² .	[<i>Xutv1366d-1A</i> {0269} ²].	UTV1366.	
<i>Xutv1366-1A.2</i> {0269} ² .	[<i>Xutv1366c-1A</i> {0269} ²].	UTV1366.	
<i>Xwmc24-1A</i> [{0242}].	[<i>Xwmc024-1A</i> {0242}].	WMC 24F/WMC 24R.	(2A).
<i>Xwmc147-1D</i> {0242}.		WMC 147F/WMC 147R.	
<i>Xwmc336-1D</i> {0242}.		WMC 336F/WMC 336R.	
<i>Xwmc432-1D</i> {0242}.		WMC 432F/WMC 432R.	

Group 1L

Amendments:

Xabc151. Revise the first column to ‘*Xabc151-1B* {0252}¹, *ID* {1529}⁴.’*Xabg452.* Revise the first column to ‘*Xabg452-1A* {1529}¹, {280}^{1,3,5}, *IB,D* {0252}¹.’*Xbcd22-1A.* Revise the first column to ‘*Xbcd22-1A* {280}⁵, *IB* {0242, 0252}¹, *ID* {0252}¹.’ and revise the last column to ‘(3A,D).’*Xbcd207-1A.* Revise the first column to ‘*Xbcd207-1A* {280}⁵, *IB* {0274}¹.’*Xbcd310-1B.* Revise the first column to ‘*Xbcd310-1A* {0242}, *IB* {445}.’*Xbcd265-1A,B,D.* Revise the last column to ‘(2B, 4B,D, 5A).’*Xbcd762-1A,B.* Revise the first column to ‘*Xbcd762-1A.1* [{280}]⁵, *IB.1* [{1529,0252}]¹.’, add ‘[*Xbcd762-1A* {280}⁵, *Xbcd762-1B* {1529}¹, *Xbcd762b-1B* {0252}¹.]’ in the second column, and add ‘(1AS,BS,DS).’ in the last column.*Xbcd808-1A.1, .2.* Revise the first column to ‘*Xbcd808-1A.1* {1529}¹, {280}^{1,3,5}, *IA.2* {1529}, *IB* {0242}.’*Xbcd921-1A,D.* Revise the first column to ‘*Xbcd921-1A* {280}⁵, *IB* {0252}¹, *ID* {445}¹.’*Xcd0105-1A.* Revise the first column to ‘*Xcd0105-1A* {280}^{1,5}, *IB* {0275}¹.’*Xcd0393-1A,B.* Revise the first column to ‘*Xcd0393-1A* {1529}¹, {280}³, *IB* {154}, *ID* {0242}.’*Xcd0346-1B.* Revise the last column to ‘(2B, 5D).’*Xcd0473-1A.* Revise the first column to ‘*Xcd0473-1A* {1529}, *IB* {0242}.’*Xcd01160-1A.* Revise the first column to ‘*Xcd01160-1A* {1529}, *IB* {0275}.’*Xcd01396-1A.* Revise the first column to ‘*Xcd01396-1A* {280}⁵, *IB* {0275}¹.’*Xcmwg645-1A.2.* Revise the first column to ‘*Xcmwg645-1A.2* {280}⁵, *IB.2* [{1529,0250}]¹.’, add [*Xmwg645* {280,1529}, *Xcmwg645b-1B* {0252}].’ in the second column, and revise the last column to ‘(1AS,BS,DS, 5A).’*Xcmwg758-1A,B.* Revise the first column to ‘*Xcmwg758-1A* {280}^{1,3,5}, *IB* {1529}¹, *ID* {0252}.’*Xgwm124-1B.* Revise the first column to ‘*Xgwm124-1B* {9929}¹, {0270}^{1,2}.’*Xgwm153-1B.* Revise the first column to ‘*Xgwm153-1B* {9929}¹, {0270}^{1,2}.’*Xgwm268-1B.* Revise the first column to ‘*Xgwm268-1B* {9929}¹, {0270}^{1,2}.’*Xgwm403-1B.* Revise the first column to ‘*Xgwm403-1B* {9929}¹, {0270}^{1,2}.’*Xgwm498-1B.* Revise the first column to ‘*Xgwm498-1B* {9929}, {0270}^{1,2}.’*XksuA1-1B.* Revise the last column to ‘(2D, 3B, 5B, 7D).’*XksuD49-1B,D.* Revise the first column to ‘*XksuD49-1A* {0242}, *IB* {728}, *ID* {448}^{1,4}, {1529}¹.’*Xmwg837-1B.2.* Add ‘(1AS,BS,DS).’ in the last column.*Xwg180-1A.* Revise the last column to ‘(4B, 7BS,L).’

Add:

<i>Xbcd372-1B,D</i> {0252}.		BCD372.	(3A,D).
<i>Xbcd402-1D</i> {0242}.		BCD402.	(4A, 5A,4B,D).
<i>Xbcd446-1B</i> {0275}.		BCD446.	(1AS).
<i>Xbcd1495-1B</i> {0269} ² .		BCD1495.	(6B).
<i>Xcd0388-1B.2</i> [{0252}].	[<i>Xcd0388b-1B</i> {0252}].	CDO388.	(1BS,DS, 2B, 3D, 4A,D, 5A,B, 6A,D).
<i>Xcd0583-1B</i> [{0242}].	[<i>Xcd0583a-1B</i> {0242}].	CDO583	(3B).
<i>Xcd01340-1B</i> {0269} ² .		CDO1340.	(1AS,BS,DS).
<i>Xcd01373-1B</i> {0269} ² .		CDO1373.	(1BS).
<i>Xmwg539-1A,B,D</i> {0252}.		MWG539.	(7D).
<i>Xmwg584-1A,B,D</i> {0252}.		MWG584.	(3A, 4A ^m , 5D).
<i>Xmwg835-1B.2</i> [{0252}].	[<i>Xmwg835b-1B</i> {0252}].	MWG835.	(1AS,BS,DS, 2A, 5A).
<i>Xmwg896-1A,B,D</i> {0252}.		MWG896.	
<i>Xmwg938-1B.2</i> [{0252}].	[<i>Xmwg938b-1B</i> {0252}].	MWG938.	(1AS,BS,DS, 7A).
<i>Xmwg2021-1B.2</i> [{0252}].	[<i>Xmwg2021b-1B</i> {0252}].	MWG2021.	(1AS,BS,DS, 2A, 3A).
<i>Xmwg2056-1B.2</i> [{0252}].	[<i>Xmwg2056b-1B</i> {0252}].	MWG2056.	(1AS,BS,DS).
<i>Xmwg2083-1B.2</i> [{0252}].	[<i>Xmwg2083b-1B</i> {0252}].	MWG2083.	(1AS,BS,DS).
<i>Xmwg2148-1B.2</i> [{0252}].	[<i>Xmwg2148b-1B</i> {0252}].	MWG2148.	(1AS,BS,DS, 1BL).
<i>Xmwg2148-1B.3</i> [{0252}].	[<i>Xmwg2148c-1B</i> {0252}].	MWG2148.	(1AS,BS,DS, 1BL)
<i>Xpsr305-1B</i> {0242}.		PSR305.	(3A,B,D).
<i>Xuaz243-1B</i> {0269} ² .		UAZ243.	
<i>Xutv135-1A</i> {0269} ² .		UTV135.	(3BS, 4B).

<i>Xutv1441-1A</i> {0269} ² .	UTV1441.	(3BL, 4B).
<i>Xwmc304-1A</i> {0242}.	WMC 304F/WMC 304R.	
<i>Xwmc312-1A</i> {0242}.	WMC 312F/WMC 312R.	
<i>Xwmc373-1B</i> [{0242}].	[<i>Xwmc373-1-1B</i> {0242}].	WMC 373F/WMC 373R.
<i>Xwmc429-1D</i> {0242}.		WMC 429F/WMC 429R.

Group 1

Amendments:

Xcmwg758-1D. Revise the last column to '(1AL,BL,DL).'*Xwg232-1A*. Revise the last column to '(4A,B, 5A,B,D, 6B, 7A,B).'*Xwpg501(Pdi)-1B*. Add reference 0263 in the first column, i.e. '{0064,0263}'.

Add:

<i>Xbcd175-1A</i> {0242}.	BCD175.	
<i>Xbcd1072-1A,B,D</i> {0252}.	BCD1072.	
<i>Xsun19-1B</i> {0156}.	SUN 19F/SUN 19R.	
<i>Xwmc84-1A</i> [{0242}].	[<i>Xwmc084-1A</i> {0242}].	WMC 84F/WMC 84R.
<i>Xwmc406-1B</i> {0242}.		WMC 406F/WMC 406R.

Group 2S

Amendments:

Xabg378-2A. Revise the last column to '(6A,D, 7A,4A).'*Xbcd152-2A,B*. Add '(6B)' to the last column.*Xbcd348-2A.1.,2,B,D*. Add '(4A)' to the last column.*Xbcd718-2A,D*. Revise the first column to '*Xbcd718-2A* {1060}¹, *2B* {0269}², *ID* {1060}¹. '.*Xcd01090-2A*. Revise the first column to '*Xcd01090-2A* {1060}, *2B* {0269}². '.*Xfba38-2D*. Revise the first column to '*Xfba38-2B* {0242}, *2D* {1060}. ' †*Xfbbl85-2B*. Revise the last column to '(3B, 6B).'*Xfba349-2D*. Add '(7A)' to the last column.*Xgwm129-2B*. Revise the last column to '(4D, 5A).'*Xpsr801(Rbcs)-2A,B,D*. To the note added to this listing in the 2001 Supplement add the reference 0271, i.e. '{0149, 0271}'.

Add:

<i>Xbcd175-2D</i> {0242}.	BCD175.	
<i>Xbcd221-2B</i> {0269} ² .	BCD221.	(4B, 6B).
<i>Xcnl9(Pdc)-2B</i> [{0269}].	[<i>Pdc-2B</i> {0269}].	Pdc.
<i>Xpsr596-2B</i> {0242}.	PSR596.	(1A,B,D, 3A).
<i>Xsun17-2D</i> {0256}.	SUN 17F/SUN 17R.	
<i>Xwmc25-2B</i> [{0242}].	[<i>Xwmc025.2-2B</i> {0242}].	WMC 25F/WMC 25R.
<i>Xwmc111-2D</i> {0242}.		(2D).
<i>Xwmc112-2D</i> {0242}.	WMC 111F/WMC 111R.	
<i>Xwmc154-2B</i> {0242}.	WMC 112F/WMC 112R.	
<i>Xwmc314-2B</i> {0242}.	WMC 154F/WMC 154R.	
		WMC 314F/WMC 314R.

Group 2L

Amendments:

Xabc451-2A. Revise the first column to '*Xabc451-2A* {282}³, *2B* [{0242}], *2D* {0242}.' and add '[*Xabc451a-2B* {0242}]' in the second column. †*Xbcd135-2B,D*. Add '(4A, 5D)' in the last column.*Xbcd266-2D*. Revise the first column to '*Xbcd266-2A* {0242}, *2B* {0164}, *2D* {864}. '.*Xbcd292-2A,D*. Revise the first column to '*Xbcd292-2A* {1060}, *2B* {0242}¹, {0269}², *2D* {864}¹. '.

Xbcd410-2A,D. Revise the first column to ‘*Xbcd410-2A* {1060}, 2B [{0242}], 2D {1060}.’ and add ‘[*Xbcd410d-2B* {0242}].’ in the second column.

Xgwm526-2B. Add ‘(2A).’ in the last column.

XksuF43-2D. Revise the last column to ‘(1A,B,D, 4D, 5D, 6D).’.

Xmwg835-2A. Revise the last column to ‘(1A,B,D, 5A).’.

Xmwg950-2B. Revise the first column to ‘*Xmwg950-2B* {1060}, 2D {0242}.’.

Xmwg2021-2A. Revise the last column to ‘(1A,B,D, 3A).’.

Xutv861-2B. Revise the first column to ‘*Xutv861-2A* {0269}², 2B {9959}².’.

Xwg184-2D. Revise the last column to ‘(3D, 4D, 5A).’.

Add:

<i>Xabc165-2D.2</i> [{0242}].	[<i>Xabc165b-2D</i> {0242}]	ABC165.	(2D).
<i>Xbcd265-2B</i> [{0242}].	[<i>Xbcd265c-2B</i> {0242}].	BCD265.	(1A,B,D, 4B,D, 5A).
<i>Xbcd512-2B</i> {0242}.		BCD512.	
<i>Xcd0346-2A</i> {0269} ² .		CDO346.	(1B, 5D).
<i>Xcd0365-2B</i> {0269} ² .		CDO365.	(6B).
<i>Xcd0669-2B</i> [{0242}].	[<i>Xcd0669a-2B</i> {0242}].	CDO669.	(4A,B,D, 7A).
<i>Xgwm526-2A</i> [{0253}].	[<i>Xgwm526-2A.2</i> {0253}].	WMS F526/WMS R526.	(2B).
<i>Xisc14(Cor)-2A</i> [{0246}] ³ .	[<i>Cor14b</i> {0246}].	COR14b.	
<i>XksuA1-2D</i> [{0242}].	[<i>XksuA1b-2D</i> {0242}].	pTksuA1.	(1B, 3B, 5B, 7D).
<i>XksuF37-2B</i> {0242}.		pTksuF37.	(6A,B,D).
<i>Xksu931(Chi4)-2D</i> [{0266}] ⁴ .	[<i>Xksu931(Cht4)</i> {0266}].	SM383.	
<i>Xksu932(Chi7)-2D</i> [{0266}] ⁴ .	[<i>Xksu932(Cht7)</i> {0266}].	SM194.	
<i>Xpsp3045-2A</i> {0253}.		PSP3045F/PSP3045R.	(5B, 7D).
<i>Xpsr370-2B,D</i> [{0242}].	[<i>Xpsr370b-2B, Xpsr370a-2D</i> {0242}].	PSR370.	(5A,B,D).
<i>Xsun11-2B</i> {0256},[{0242}].	[<i>Xsunm11-2B</i> {0242}].	SUN 11F/SUN 11R.	
<i>Xsun21-2B</i> {0256}.		SUN 21F/SUN 21R.	
<i>Xsun22-2B</i> {0256}.		SUN 22F/SUN 22R.	
<i>Xwmc339-2B</i> {0242}.		WMC 339F/WMC 339R.	
<i>Xwmc360-2B</i> {0242}.		WMC 360F/WMC 360R.	

Group 2

Amendments:

Xabg356-2D. Revise the first column to ‘*Xabg356-2B* {0242}, 2D {9926}⁴.’.

Xwmc24-2A. Add ‘(1A).’ in the last column.

Xwmc25-2D. Add ‘(2B).’ in the last column.

Xwmc149-2B. Add ‘(5B).’ in the last column.

Add:

<i>Xabc162-2A</i> {0242}.		ABC162.	
<i>Xabc165-2D.1</i> [{0242}].	[<i>Xabc165a-2D</i> {0242}].	ABC165.	(2DL).
<i>Xcd0366-2B,D</i> {0242}.		CDO366.	
<i>Xcd0665-2A</i> {0242}.		CDO665.	(4A).
<i>Xgwm271-2B</i> [{0242}].	[<i>Xgwm271a-2B</i> {0242}].	WMS F271/WMS R271.	(5B, 5D).
<i>XksuE7-2B</i> {0242}.		pTksuE7	(7D).
<i>Xstm773-2B</i> {0242}.		STM 773F/STM 773R.	
<i>Xwmc18-2D</i> [{0242}].	[<i>Xwmc018-2D</i> {0242}].	WMC 18F/WMC 18R.	
<i>Xwmc35-2B</i> [{0242}].	[<i>Xwmc035a-2B</i> {0242}].	WMC 35F/WMC 35R.	(4B).
<i>Xwmc190-2D</i> {0242}.		WMC 190F/WMC 190R.	
<i>Xwmc198-2A</i> {0242}.		WMC 198F/WMC 198R.	
<i>Xwmc445-2B</i> {0242}.		WMC 445F/WMC 445R.	

Group 3S

Amendments:

Xabg471-3A,B. Add '(6B).' to the last column.*Xcdo395-3A,D.* Revise the first column to 'Xcdo395-3A {1061}¹, 3B {0242}¹, 3D {9926}⁴.'*Xfbb185-3B.* Revise the last column to '(2B, 6B).'*Xgwm369-3A.* Revise the first column to 'Xgwm369-3A {9929}¹, {0269}².'*Xgwm389-3B.* Revise the first column to 'Xgwm389-3B {9929}¹, {0269}².'*XksuA1-3B.* Revise the last column to '(1B, 2D, 5B, 7D).'*XksuB8-3D.* Revise the first column to 'XksuB8-3A,B {0242}, 3D {448}.' †*Xmwg584-3A.* Revise the last column to '(1A,B,D, 4A^m, 5D).'*Xmwg2021-3A.* Revise the last column to '(1A,B,D, 2A).'*Xpsr305-3A,B,D.* Add '(1B)' to the last column.*Xsfr2(Lrk10)-3B,D.* Revise the last column to '(1A,B,D).'

Add:

Xpsr311-3A {0242}.

PSR311.

(7A,B,D).

Xutv135-3B {0269}².

UTV135.

(1A, 4B).

Xwmc11-3A [{0242}].[*Xwmc011-3A* {0242}].

WMC 11F/WMC 11R.

Xwmc43-3B [{0242}].[*Xwmc043-3B* {0242}].

WMC 43F/WMC 43R.

Group 3L

Amendments:

Xbcd115-3A,D. Revise the first column to 'Xbcd115-3A {1061}, 3B {0269}², 3D [{862}].'*Xbcd22-3D.* Revise the first column to 'Xbcd22-3A {0242}, 3D {1061}.' and revise the last column to '(1A,B,D).'*Xbcd195-3B.* Revise the first column to 'Xbcd195-3A {0269}², 3B {0078}¹.'*Xbcd372-3A,D.* Add '(1B,D)' to the last column.*Xbcd451-3A,D.* Revise the first column to 'Xbcd451-3A {1061}, 3B {0242}, 3D [{862}].'*Xcdo105-3B.* Revise the last column to '(1A,B).'*Xcdo482-3A,D.* Revise the first column to 'Xcdo482-3A {1061}, 3B {0242}, 3D {862}.'*Xcdo583-3B.* Add '(1B)' to the last column.*Xcdo718-3B.* Revise the first column to 'Xcdo718-3A {0242}, 3B {1061}.'*Xfba175-3A.* Add '(6B)' in the last column.*Xgwm155-3A.* Revise the first column to 'Xgwm155-3A {9929}¹, {0269}².'*Xgwm299-3B.* Revise the first column to 'Xgwm299-3B {9929}¹, {0269}².'*XksuG59-3A,D.* Revise the first column to 'XksuG59-3A {282}³, 3B {0242}¹, 3D {448}⁴, {1061}¹.'*Xutv416-3A.* Revise the first column to 'Xutv416-3A {9959}², 3B {0269}².'*Xutv560-3A.* Revise the first column to 'Xutv560-3A {9959}², 3B, 1., 2 [{0269}]².' and add '[Xutv560a,b-3B {0269}]' in the second column.

Add:

Xabg75-3B {0242}.

ABG75.

Xbcd941-3A {0175}.

BCD941.

Xcdo534-3A {0269}².

CDO534.

(1B, 6A,B,D, 7A).

Xksu933(Glb3)-3B,D {0266}.

SM289.

Xksu934(Glb3)-3D {0266}.

SM638.

Xmwg2153-3A {0269}².

MWG2153.

Xpsr596-3A,B [{0242}].[*Xpsr596b-3A, Xpsr596a-3B* {0242}].

PSR596.

(1A,B,D, 2B).

Xpsr604-3B [{0242}].[*Xpsr604-3B* {0242}].

PSR604.

(7A,4A,7D).

Xsun23-3A {0256}.

SUN 23F/SUN 23R.

Xucg2(Acc-2)-3A,B,D [{0265}].[*Xucg2-3A,B,D* {0265}].

UCG2.

(5D).

Xutv601-3A {0269}².

UTV601.

Xutv920-3A {0269}².

UTV920.

Xutv1151-3A {0269}².

UTV1151.

(6A).

Xutv1371-3B {0269}².

UTV1371.

(1AL, 4B).

<i>Xutv1441-3B</i> {0269} ² .	UTV1441.
<i>Xutv1474-3A</i> {0269} ² .	UTV1474.
<i>Xwmc169-3A</i> {153,0238}.	WMC 169F/WMC 169R.
<i>Xwmc236-3B</i> {0242}.	WMC 236F/WMC 236R.
<i>Xwmc334-3B</i> {0242}.	WMC 334F/WMC 334R.
<i>Xwmc428-3A</i> {0242}.	WMC 428F/WMC 428R.

Group 3

Amendments:

Xwg184-3D. Revise the last column to '(2D, 4D, 5A).'.*Xwmc169-3A*. Delete (moved to 3L).

Add:

<i>Xwmc50-3A</i> [{0242}].	[<i>Xwmc050-3A</i> {0242}].	WMC 50F/WMC 50R.
<i>Xwmc375-3D</i> {0242}.		WMC 375F/WMC 375R.
<i>Xwmc379-3A</i> {0238}.		WMC 379F/WMC 379R.

Group 4S (4AL:4BS:4DS)

Amendments:

Xbcd265-4B,D. Revise the last column to '(1A,B,D, 2B, 5A).'.*Xbcd583-4A*. Revise the last column to '(1A,B)'.*Xbcd402-4A*. Revise the last column to '(1D, 5A,4B,D)'.*Xcd0669-4A,B,D*. Add '(2B, 4A^m)' in the last column.*Xcd0795-4B*. Revise the first column to '*Xcd0795-4A* {0242}, 4B {1059}'.*Xgwm18-4B*. Add '(1B)' in the last column.*Xwg184-4D*. Revise the last column to '(2D, 3D, 5A)'.

Add:

<i>Xcd0949-4B</i> {0269} ² .	CDO949.	(4DL).
<i>Xcn110(Lpx-B1)-4B</i> [{0269}] ² .		
[Loxmjt].	LOXMJT.	
<i>Xfba363-4B</i> {0242}.	FBA363.	(7A).
<i>Xutv434.1-4A.1,.2</i> [{0269}] ² . <i>[Xutv434a,b-4A</i> {0269}] ² .	UTV434.	(4A).
<i>Xwg232-4B</i> {0269} ² .	WG232.	(1A, 4A, 5A,B,D, 6B, 7A,B).
<i>Xwmc141-4B</i> {0242}.	WMC 141F/WMC 141R.	

4A^mS

Amendments:

Xcd066-4A. Revise the last column to '(2B, 4A,B,D)'.*Xmwg584-4A.1*. Revise the last column to '(1A,B,D, 3A, 4A^mL, 5D)'.*Xwg622-4A*. Revise the last column to '(4AL,BS,DS, 6A)'.**Group 4L (4AS:4BL:4DL)**

Amendments:

Xcd01395-4B. Revise the first column to '*Xcd01395-4B* {1008}, 4D {0248}'.*Xwg622-4A,B,D*. Add '(6A)' in the last column.

Add:

<i>Xbcd1230-4D</i> {0248}.	BCD1230.
----------------------------	----------

4A^mL

Amendments:

Xabg463-4A. Revise the last column to '(4D, 5A, 5B).*Xmwg584-4A.2.* Revise the last column to '(1A,B,D, 3A, 4A^mS, 5D).**Group 5AL:4BL:4DL**

Amendments:

Xbcd402-5A,4B,D. Revise the last column to '(1D, 4A).'*Xcd0949-4D.* Add '(7B)' in the last column.*Xutv434-4A.1.* Revise the last column to '(4AS, 4AL).'**Group 4**

Amendments:

Xabg397-4D. Add '(5A)' in the last column.*Xabg463-4D.* Revise the last column to '(4A, 5B, 5D).'*XksuF43-4D.1.* Revise the last column to '(1AB,D, 2D, 4D, 5D, 6D).'*XksuF43-4D.2.* Revise the last column to '(1A,B,D, 2D, 4D, 5D, 6D).'*Xwg232-4A.* Revise the last column to '(1A, 4B, 5A,B,D, 6B, 7A,B).'*Xwmc35-4B.* Add '(2B)' in the last column.*Xwpdg501(Pdi)-4A,B,D.* Add reference 0263 in the first column, i.e. '{0064, 0263}'.

Add:

<i>Xbcd221-4B</i> {0269} ² .	BCD221.	(2B, 6B).
<i>Xbcd1975-4A</i> {0269} ² .	BCD1975.	(7D).
<i>Xbcd348-4A</i> {0269} ² .	BCD348.	(2A,B,D).
<i>Xcd0414-4A</i> {0269} ² .	CDO414.	(7B).
<i>Xcd0949-4A</i> [{0242}].	[<i>Xcd0949b-4A</i> {0242}].	CDO949.
<i>Xgwm129-4D</i> {0242}.		WMS F129/WMS R129.
<i>Xgwm613-4A</i> {0269} ² .		WMS F613/WMS R613.
<i>Xstm91-4D</i> {0242}.		STM 91F/STM 91R.
<i>Xutv135-4B</i> {0269} ² .	UTV135.	(1A, 3B).
<i>Xutv434-4A.3</i> [{0269}] ² .	[<i>Xutv434d-4A</i> {0269}] ² .	UTV434.
<i>Xutv1136-4A.1,.2</i> [{0269}] ² .	[<i>Xutv1136a,c-4A</i> {0269}] ² .	UTV1136.
<i>Xutv1441-4B</i> {0269} ² .		UTV1441.
<i>Xwg180-4B</i> [{0242}].	[<i>Xwg180a-4B</i> {0242}].	WG180.
<i>Xwmc47-4B</i> [{0242}].	[<i>Xwmc047-4B</i> {0242}].	WMC 47F/WMC 47R.
<i>Xwmc48-4A,B,D</i> [{0242}].	[<i>Xwmc048a-4A, Xwmc048c-4B,</i> <i>Xwmc048b-4D</i> {0242}].	WMC 48F/WMC 48R.

Group 5S

Amendments:

Xabg873-5B. Revise the first column to '*Xbcd873-5B* {1059}, 5D {0242}.' and add '(7A,D)' in the last column.*Xbcd207-5A.* Revise the last column to '(1A,B).'*Xgwm129-5A.* Revise the last column to '(2B, 4D).'*Xgwm443-5B.* Add '(5A)' in the last column.*Xmwg835-5A.* Revise the last column to '(1A,B,D, 2A).'

Add:

<i>Xabg397-5A</i> {0242}.	ABG397.	(4D).
<i>Xcd0465-5A</i> {0269} ² .	CDO465.	(5AL,BL,DL).
<i>Xgdm68-5D</i> {0242}.	DMS 68F/DMS 68R.	(5A,B, 5DL).
<i>Xgwm443-5A</i> {0242}.	WMS F443/WMS R443.	(5B).
<i>Xpsp3045-5B</i> [{0253}].	[<i>Xpsp3045-5B.2</i> {0253}].	PSP3045F/PSP3045R.
<i>Xwg184-5A</i> {0242}.	WG184.	(2D, 3D, 4D).

<i>Xwmc149-5B</i> {0242}.	WMC 149F/WMC 149R.	(2B).
<i>Xwmc159-5A</i> {0242}.	WMC 159F/WMC 159R.	
<i>Xwmc233-5D</i> {0242}.	WMC 233F/WMC 233R.	

Group 5L

Amendments:

Xabg473-5A,B,D. Revise the last column to '(6A,B).'
Xbcd265-5A. Revise the last column to '(1A,B,D, 2B, 4B,D).'.
Xcd0346-5D. Revise the last column to '(1B, 2B).'.
Xcd0465-5A,B,D. Add '(5AS)' to the last column.
Xcd01090-5A. Revise the last column to '(1A,B)'.
Xgdm68-5D. Revise the last column to '(5A,B, 5DS).'
XksuA1-5B. Revise the last column to '(1B, 2D, 3B, 7D)'.
Xgwm271-5D. Add '(2B, 5B)' in the last column.
Xpsr370-5A,B,D. Add '(2B)' to the last column.
Xwg232-5A,B,D. Revise the last column to '(1A, 4A,B, 6B, 7A,B)'.

Add:

<i>Xabg463-5B</i> {0242}.	ABG463.	(4A,D, 5D).
<i>Xcd0775-5B</i> {0269} ² .	CDO775.	(7A,B,D).
<i>Xcn111(Lpx-B2)-5B</i> [{0269}] ² . [<i>Lox11-1</i> {0269}] ² .	LOX11-1.	
<i>Xgwm271-5B</i> [{0242}].	WMS F271/WMS R271.	(2B, 5D).
<i>Xstm652-5B</i> {0242}.	STM 652F/STM 652R.	
<i>XsunG5-5B</i> [{0242}].	SUN G5F/SUN G5R.	
<i>Xucg2(Acc-2)-5D</i> [{0265}].	UCG2.	(3A,B,D).
<i>Xunl1-5B</i> {0247}.	UNL1.	
<i>Xunl2-5B</i> {0247}.	UNL2	
<i>Xunl3-5B</i> {0247}.	UNL3.	
<i>Xutv497-5A</i> {0269} ² .	UTV497.	
<i>Xutv1435-5A</i> {0269} ² .	UTV1435.	
<i>Xwmc28-5B</i> [{0242}].	WMC 28F/WMC 28R.	
<i>Xwmc235-5B</i> {0242}.	WMC 235F/WMC 235R.	
<i>Xwmc376-5B</i> {0242}.	WMC 376F/WMC 376R.	

4AL:5BL:5DL

Add:

<i>Xcd0506-4A</i> {0242}.	CDO506.	
A <i>Xcd0506-5D</i> locus has been reported in {1059} in the 7BS:5BL:5DL category. It is possible that this is a misclassification and that <i>Xcd0506-4A</i> and <i>Xcd0506-5D</i> are homoeologous.		
<i>Xwmc161-4A</i> [{0242}].	[<i>Xwmc161a-4A</i> {0242}].	WMC 161F/WMC 161R.
<i>Xwmc258-4A</i> {0242}.		WMC 258F/WMC 258R.

Group 5

Amendments:

Xabg463-5D.1,.2. Revise the last column to '(4A,D, 5B)'.
Xcmwg645-5A. Revise the last column to '(1A,B,D)'.
Xgdm68-5A,B. Revise the last column to '(5DS, 5DL)'.
XksuF43-5D.1,.2. Revise the last column to '(1A,B,D, 2D, 4D, 6D)'.
Xmwg584-5D. Revise the last column to '(1A,B,D, 3A, 4A).
Xwg232-5A.1,B. Revise the last column to '(1A, 4A,B, 6B, 7A,B)'.
Xwg232-5A.2. Revise the last column to '(1A, 4A,B, 6B, 7A,B)'.
Xwg420-5D.1,.2. Revise the last column to '(7A,B,D)'.

Add:

<i>Xabg3-5A,D</i> [{0242}].	[<i>Xabg3b-5A</i> {0242}].	ABG3.
<i>Xbcd135-5D</i> {0242}.		BCD135.
<i>Xgwm304-5A</i> {0242}.		WMS F304/WMS R304.
<i>Xstm286-5B</i> {0242}.		STM 286F/STM 286R.
<i>Xstm337-5A</i> [{0242}].	[<i>Xstm337a-5A</i> {0242}].	STM 337F/STM 337R.
<i>Xwmc96-5A</i> [{0242}].	[<i>Xwmc096-5A</i> {0242}].	WMC 96F/WMC 96R.
<i>Xwmc110-5A</i> {0242}.		WMC 110F/WMC 110R.

Group 6S

Amendments:

Xabg378-6A,D. Revise the last column to '(2A, 7A,4A).'. *Xbcd1398-6D.* Revise the first column to '*Xbcd1398-6B* {0242}, 6D {900}.'. *Xbcd1821-6A,D.* Revise the first column to '*Xbcd1821-6A* {900}, 6B {0244}, 6D {900}.'. *Xpdo270-6A,D.* Revise the first column to '*Xpdo270-6A* {900}¹, 6B {0269}², 6D {900}¹.'. *Xpdo365-6B.* Add '(2B)' to the last column. *Xpdo534-6A,B,D.* Revise the last column to '(1B, 3A, 7A)'. *Xfba307-6A,D.* Revise the first column to '*Xfba307-6A* {900}, 6B {0244}, 6D {900}.'. *Xfba381-6B,D,2.* Revise the first column to '*Xfba381-6B,2,D,2* [{0081}]]', add '[*Xfba381-6B* {0081}]' in the second column and revise the last column to '(6BL,DL)'. *Xfbb194-6A.* Add '(4A)' to the last column. *Xgwm132-6B.* Add '(6D)' to the last column. *Xgwm613-6B.* Add '(4A)' to the last column. *Xgwm644-6B.* Revise the first column to '*Xgwm644-6B* {9929}¹, {0269}²'. *XksuF43-6D.* Revise the last column to '(1A,B,D, 2D, 4D, 5D)'. *Xmwg573-6A,2,B,D.* Revise the last column to '(6AL, 6BL)'. *Xpsr546-6A.* Revise the first column to '*Xpsr546-6A,1* [{9927}]²', add [*Xpsr546-6A* {9927}]².' in the second column, and revise the third column to '(6AL, 6BL,DL)'.

Add:

<i>Xabg471-6B</i> {0269} ² .		ABG471.	(3A,B).
<i>Xbarc101-6B</i> {0175}.		BARC F101/BARC R101 {0239}.	
<i>Xbcd152-6B</i> {0269} ² .		BCD152.	(2A,B).
<i>Xbcd1299-6B</i> {0269} ² .		BCD1299.	
<i>Xfba175-6B,1</i> [{0244}].	[<i>Xfba175a-6B</i> {0244}].	FBA175.	(3A, 6BL).
<i>Xgwm132-6D</i> {0242}.		WMS F132/WMS R132.	(6B).
<i>Xpsr119-6A</i> [{0242}].	[<i>Xpsr119a-6A</i> {0242}].	PSR119.	(7A,4A,7D).
<i>Xutv1151-6A,1,2</i> [{0269}] ² .	[<i>Xutv1151a,b-6A</i> {0269}] ² .	UTV1151.	(3A).
<i>Xwmc104-6B</i> {0032, 0276}		WMC F104/WMC R104 {0037}.	

Group 6L

Amendments:

Xabc175-6A,D. Revise the first column to '*Xabc175-6A* {9927}², {0081}¹, 6B [{0242}]¹, 6D {900}¹.' and add '[*Xabc175a-6B* {0242}]' in the second column. *Xabg473-6B.* Revise the first column to '*Xabg473-6A* {0242}, 6B {900}.'. *Xabg652-6A.* Add '(7A)' in the last column. *Xfba381-6D.* Revise the first column to '*Xfba381-6B,1* [{0244}], 6D,1 [{900}]]', revise the second column to '[*Xfba381-6B* {0244}, 6D {900}]', and add '(6BS,DS)' in the last column. *Xgwm427-6A.* Revise the first column to '*Xgwm427-6B* {9929}¹, {0269}².'. *XksuF37-6A,B.* Add '(2B, 6D)' in the last column. *Xmwg573-6A,1.* Revise the last column to '(6AS,BS,DS, 6BL)'. *Xpsr546-6B,D.* Revise the last column to '(6AS, 6AL)'.

Add:

<i>Xbcd279-6B</i> {0269} ² .	BCD279.
---	---------

<i>Xcd0686-6B</i> {0269} ² .		CDO686.	(7B).
<i>Xfba175-6B.2</i> [{0244}].	[<i>Xfba175b-6B</i> {0244}].	FBA175.	(3A, 6BS).
<i>Xfbb185-6B</i> [{0242}].	[<i>Xfbb185c-6B</i> {0242}].	FBB185.	(2B, 3B).
<i>Xmwg573-6B</i> {0242}.		MWG573.	(6AS,BS,DS, 6AL).
<i>Xpsr546-6A.2</i> [{0242}].	[<i>Xpsr546a-6A</i> {0242}].	PSR546.	(6AS, 6BL,DL).
<i>Xsun5-6D</i> [{0242}].	[<i>XsunM5b-6D</i> {0242}].	SUN 5F/SUN 5R.	
<i>Xutv1136-6A</i> {0269} ² .		UTV1136.	
<i>Xwg622-6A</i> {0242}.		WG622.	(4A,B,D).
<i>Xwmc163-6A</i> {0242}.		WMC 163F/WMC 163R.	

Group 6

Amendments:

- Xabc451-6D.* Revise the last column to '(2A,B,D).'.
Xbcd221-6B. Add '(2B, 4B).' to the last column.
Xbcd1299-6B. Add '(6B).' to the last column.
Xbcd1495-6B. Add '(1B).' to the last column.
XksuE19-6D. Revise the last column to '(1A,B,D, 7B).'.
XksuF37-6D. Add '(2B, 6A,B).' to the last column.
Xwmc76-6B. Add '(7B).' to the last column.
Xwmc104-6B. Delete (entry moved to 6S).
Xwmg573-6D. Revise the last column to '(6AS,BS,DS, 6AL, 6BL).'.

Add:

<i>Xwg232-6B</i> [{0242}].	[<i>Xwg232b-6B</i> {0242}].	WG232.	(1A, 4A,B, 5A,B,D, 7A,B).
<i>Xwmc416-6D</i> {0242}.		WMC 416F/WMC 416R.	

Group 7S

Amendments:

- Xabc158-7A.* Revise the first column to '*Xabc158-7A* {1059}, 7B {0242}.'.
Xabc465-7A,D. Revise the first column to '*Xabc465-7A* {282}³, 7B [{0242}]¹, 7D {9926}⁴.', add '[*Xabc465a-7B* {0242}].' in the second column and add '(4A).' in the last column.
Xbcd310-7B. Revise the last column to '(1A,B).'.
Xcd0534-7A. Revise the last column to '(1B, 3A, 6A,B,D).'.
Xcd01395-7A. Revise the last column to '(4B,D).'.
Xfba363-7A. Add '(4B).' in the last column.
Xgwm60-7A. Delete (entry moved to 7AS:4AL:7DS).
Xgwm537-7B. Revise the first column to '*Xgwm537-7B* {9929},{0242}.' and remove the sentence 'Whether *Xgwm537-7B.* belongs to the 7S arm group or the 7BS:5BL:5DL arm group is uncertain.'.
Xgwm631-7A. Delete (entry moved to 7L).
Xwg180-7B. Revise the last column to '(1A, 4B, 7BL).'.

Add:

<i>Xgwm111-7D</i> {9929, 0211}.	WMS F111/WMS R111.	(4A, 7BL).
<i>Xgwm255-7B</i> {0250}.	WMS F255/WMS R255.	
<i>Xgwm263-7B</i> {0250}.	WMS F263/WMS R263.	
<i>Xgwm890-7A</i> {0254}.	WMS F890/WMS R890.	
<i>Xgwm913-7A</i> {0254}.	WMS F913/WMS R913.	
<i>Xgwm1002-7D</i> {0250}.	WMS F1002/WMS R1002.	
<i>Xgwm1065-7A</i> {0254}.	WMS F1065/ WMS R1065.	
<i>Xgwm1173-7B</i> {0250}.	WMS F1173/WMS R1173.	
<i>Xgwm1184-7B</i> {0250}.	WMS F1184/WMS R1184.	
<i>Xgwm1220-7D</i> {0250}.	WMS F1220/WMS R1220.	

It is not known whether *Xwmc1220-7A* belongs to group 7S or 7AS:4AL:7DS.

Xsun16-7B {0256}, [{0242}].[*XsunM16-7B* {0242}]. SUN 16F/SUN 16R.*Xutv621-7B* {0269}².

UTV621.

Xwmc17-7A [{0242}].[*Xwmc017-7A* {0242}]. WMC 17F/WMC 17R.It is not known whether *Xwmc17-7A* belongs to group 7S or 7AS:4AL:7DS.*Xwmc76-7B* [{0242}].[*Xwmc076-7B* {0242}]. WMC 76F/WMC 76R.

(6B).

Xwmc83-7A {0153}, [{0242}].

WMC 83F/WMC 83R {0161}.

Xwmc283-7A {0242}.

WMC 283F/WMC 283R.

Xwmc338-7B {0242}.

WMC 338F/WMC 338R.

It is not known whether *Xwmc338-7B* belongs to group 7S or 7BS:5BL:5DL.*Xwmc405-7A,D* [{0242}]. [*Xwmc405a-7A* {0242}]. WMC 450F/WMC 405R.**7AS:4AL:7DS**

Amendments:

Xabg378-7A. Revise the first column to ‘*Xabg378-7A* {282}³, 4A [{0242}]¹.’ and add ‘[*Xabg378b-4A* {0242}].’ in the second column.*Xabg704-7A*. Revise the first column to ‘*Xabg704-7A* {282}³, 4A {0242}¹, 7D [{0242}]¹.’ and add ‘[*Xabg704c-7D* {0242}].’ in the second column.*Xbcd129-7D*. Revise the first column to ‘*Xbcd129-7A* {0242}, 7D [{1059}] {1057}.’*Xbcd1975-7D*. Add ‘(4A).’ in the last column.*Xcd665-4A*. Add ‘(2A).’ in the last column.*Xpsr604-7A,4A,7D*. Add ‘(3B).’ in the last column.*Xutv434-4A.2*. Revise the last column to ‘(4AS, 4AL).’.

Add:

Xabc465-4A [{0242}]. [*Xabc465b-4A* {0242}]. ABC465. (7A,B,D).*Xabg75-7A,D* [{0242}]. [*Xabg75b-7A, Xabg75a-7D* {0242}]. ABG75.*Xbcd135-7A, 4A* [{0242}]. [*Xbcd135a-7A, Xbcd135b-4A* {0242}]. BCD135. (2B,D, 5D).*Xbcd873-7A,D* [{0242}]. [*Xbcd873a-7A, Xbcd873b-7D* {0242}]. BCD873. (5B,D).*Xfbb194-4A* {0242}. FBB194. (6A).*Xgwm60-7A* {724,0250}. WMS F60/WMS R60.*Xstm271-7A* {0242}. STM 271F/STM 271R.It is not known whether *Xstm271-7A* belongs to group 7AS:4AL:7DS or 7S.*Xwmc262-4A* {0242}. WMC 262F/WMC 262R.*Xwmc313-4A* {0242}. WMC 313F/WMC 313R.**Group 7L**

Amendments:

Xcd414-7B. Add ‘(4A).’ to the last column.*Xcd686-7B*. Add ‘(6B).’ to the last column.*Xcd775-7A,B,D*. Add ‘(5B).’ to the last column.*Xgwm111-7B,D*. Revise the first column to ‘*Xgwm111-7B* [{0031}].’ and revise the last column to ‘(4A, 7DS).’.*Xgwm332-7A*. Revise the first column to ‘*Xgwm332-7A.3* [{9929}].’, add ‘[*Xgwm332-7A* {9929}].’ in the second column, and add ‘(7AS).’ to the last column.*XksuA1-7D*. Revise the last column to ‘(1B, 2D, 3B, 5B).’.*Xmwg938-7A*. Revise the last column to ‘(1A,B,D).’.*Xpsr311-7A,B,D*. Add ‘(3A).’ to the last column.*Xutv1110-7A*. Revise the first column to ‘*Xutv1110-7A* {9959}², 7B {0269}².’.*Xutv1267-7A*. Revise the first column to ‘*Xutv1267-7A* {9959}², 7B {0269}².’.*Xwg180-7B*. Revise the last column to ‘(1A, 4B, 7BS).’.*Xwg232-7A*. Revise the first column to *Xwg232-7A* [{154}], 7B [{0242}], add ‘[*Xwg232a-7B* {0242}]’ in the second column, and revise the last column to ‘(1A, 4A,B, 5A,B,D, 6B).’.

Xwg420-7A,D. Revise the first column to 'Xwg420-7A {282}³, 7B {0242}¹, 7D {1059}¹.

Add:

<i>Xabg652</i> -7A {0242}.	ABG652.	(6A).
<i>Xfba349</i> -7A {0242}.	FBA349.	(2D).
<i>Xgwm332</i> -7A.1 [{0269}] ² . [Xgwm332a-7A {0269}] ² .	WMS F332/WMS R332.	(7AL).
<i>Xgwm332</i> -7A.2 [{0269}] ² . [Xgwm332b-7A {0269}] ² .	WMS F332/WMS R332.	(7AL).
<i>Xgwm631</i> -7A.{0178,0254}.	WMS F631/WMS R631.	
<i>Xgwm698</i> -7A {0254}.	WMS F698/WMS R698.	
<i>Xgwm748</i> -7A {0254}.	WMS F748/WMS R748.	
<i>Xgwm767</i> -7B {0250}.	WMS F767/WMS R767.	
<i>Xgwm870</i> -7A {0254}.	WMS F870/WMS R870.	
<i>Xgwm871</i> -7B {0250}.	WMS F871/WMS R871.	
<i>Xgwm897</i> -7B {0250}.	WMS F897/WMS R897.	
<i>Xgwm963</i> -7B {0250}.	WMS F963/WMS R963.	
<i>Xgwm1044</i> -7D {0250}.	WMS F1044/WMS R1044.	
<i>Xgwm1061</i> -7A {0254}.	WMS F1061/WMS R1061.	
<i>Xgwm1066</i> -7A {0254}.	WMS F1066/WMS R1066.	
<i>Xgwm1085</i> -7B {0250}.	WMS F1085/WMS R1085.	
<i>Xgwm1083</i> -7A {0254}.	WMS F1083/WMS R1083.	
<i>XksuE19</i> -7B {0242}.	pTksuE19.	(1A,B,D, 6D).
<i>Xrz508</i> -7A.1 [{0269}] ² .	RZ508.	(7AL, 7B).
<i>Xrz508</i> -7A.2 [{0269}] ² .	RZ508.	(7AL, 7B).
<i>Xutv507</i> -7B {0269}] ² .	UTV507.	
<i>Xutv1521</i> -7A {0269}] ² .	UTV1521.	
<i>Xwmc14</i> -7D [{0242}].	WMC 14F/WMC 14R.	
<i>Xwmc116</i> -7A {0242}.	WMC 116F/WMC 116R.	
<i>Xwmc157</i> -7D {0242}.	WMC 157F/WMC 157R.	
<i>Xwmc247</i> -7A {0242}.	WMC 247F/WMC 247R.	
<i>Xwmc346</i> -7A {0242}.	WMC 346F/WMC 346R.	

Group 7

Add:

<i>Xbcd410</i> -7D.	Revise the last column to '(2A,B,D)'.
<i>Xbcd707</i> -7D.	Revise the first column to 'Xbcd707-7B {0242}, 7D {1059}'.
<i>XksuE7</i> -7D.	Add '(2B)' in the last column.
<i>Xmwg539</i> -7D.	Add '(1A,B,D)' in the last column.
<i>Xpsp3045</i> -7D.	Add '(2A, 5B)' in the last column.
<i>Xwg232</i> -7A.1.	Revise the last column to '(1A, 4A,B, 5A,B,D, 6B, 7B)'.
<i>Xwg232</i> -7A.2.	Revise the last column to '(1A, 4A,B, 5A,B,D, 6B, 7B)'.
<i>Xwmc47</i> -7A.	Add '(4B)' in the last column.
<i>Xwmc83</i> -7A.	Delete (the entry has been moved to group 7S).

Add:

<i>Xcd949</i> -7B {0242}.	CDO949.	(4D).
<i>Xstm337</i> -7B [{0242}].	STM 337F/STM 337R.	(5A).
<i>Xstm764</i> -7A [{0242}].	STM 764F/STM 764R.	
<i>Xwmc94</i> -7D {0242}.	WMC 94F/WMC 94R.	
<i>Xwmc121</i> -7D {0242}.	WMC 121F/WMC 121R.	
<i>Xwmc364</i> -7B {0242}.	WMC 364F/WMC 364R.	
<i>Xwmc402</i> -7B {0242}.	WMC 402F/WMC 402R.	

Dormancy (Seed)

Cross AC Domain/Haryutaka: one major QTL in chromosome 4AL and two lesser possibly homoeologous QTLS in 4BL and 4DL {0226}.

Ear emergence

QEet.ipk-2D 2DS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. Lateness was contributed by W-7984 {0255}.

ma: Associated with *Xfba400-2D* and *Xcdol379* {0255}.

QEet.ipk-2D coincides with a QTL for flowering time, *QFlt.ipk-2D*. Both QTLs are likely to correspond to *Ppd-D1* {0255}

QEet.ipk-5D {0255}.

5DL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. Lateness was contributed by W-7984 {0255}.

ma: Associated with *Xbcd450-5D* {0255}.

QEet.ipk-5D coincides with a QTL for flowering time, *QFlt.ipk-5D*. Both QTLs are likely to correspond to *Vrn-D1* {0255}.

Flowering time*QFlt.ipk-3A* {0255}.

3AL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. Lateness was contributed by W-7984 {0255}.

ma: Associated with *Xbcd451* {0255}.

Frost resistance

Responses to cold exposure and their genetics are reviewed in {0020,0274}.

Fr1. **ma:** *Fr1* mapped 2 cM proximal to *Xwg644-5A* and *Vrn-A1* {0291} and was flanked by deletion points 0.67 and 0.68 {0292}.

Fr2 {0291}. 5DL {0291}. **s:** CS*7/Cheyenne 5D {0291}.

ma: *Fr2* mapped 10 cM proximal to *Vrn-D1* {0291}.

QWin.ipk-6A 6AS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. Winter hardiness was contributed by W-7984 {0255}.

ma: Associated with *Xfba85* and *Xpsr10(Gli-2)* {0255}.

Gluem colour**1. Red (brown/bronze)**

Rg2.

QRg.ipk-1D {0255}.

1DS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. The glume colour was contributed by W-7984 {0255}.

ma: Associated with *Gli-D1* {0255}.

This QTL coincides with a QTL for awn colour, *QRaw.ipk-1D* {0255}.

7. Awn colour*QRaw.ipk-1A* {0255}.

1AS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. The awn colour was contributed by W-7984 {0255}.

ma: Associated with *Gli-A1* {0255}.

QRaw.ipk-1D {0255}.

1DS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}. The awn colour was contributed by W-7984 {0255}.

ma: Associated with *Gli-D1* {0255}.

Grain hardness / Endosperm texture

Add at end of section:

QTL: Ten QTLs for kernel hardness (54 % of the variation) were mapped in a cross 'Forno'/'Oberkulmer' spelt {0280}.

Grain Quality Parameters**1. Sedimentation value**

QTL: QTL associated with *Glu-1* on chromosome arms 1AL and 1DL and *Gli-1/Glu-3* on 1BS were detected in RSLs from the cross Cheyenne (high quality) x CS (low quality) {0251}. Cultivar Cheyenne contributed the higher SDS sedimentation values {0251}. The QTL on 1AL coincided with a QTL for bread loaf volume {0251}. The QTL on 1DL and 1BS coincided with QTL for bread mixing time {0251}.

4. Milling yield

QTL: A QTL associated with *Pinb* on chromosome arm 5DS was detected in RILs from the cross NY6432-18 x Clark's Cream {0241}. Cultivar Clark's Cream contributed the higher flour yield allele {0241}. This QTL coincided with QTL for hardness, hydration traits (dough water absorption, damaged starch and alkaline water retention capacity (AWRC), and baked product traits (cookie diameter and cookie top grain) {0241}.

5. Alveograph dough strength W

Add at the end of section:

QTL: Ten QTLs for W (39 % of the variation), nine QTLs for P (48% of the variation) and seven QTLs for P:L (38% of the variation) were mapped in 'Forno'/'Oberkulmer' spelt {0280}.

6. Mixograph peak time (new category)

QTL: A QTL associated with *Glu-Dy1* on chromosome arm 1DL was detected in RILs from the cross NY6432-18 x Clark's Cream {0241}. Cultivar Clark's Cream contributed the higher mixograph peak time allele {0241}. This QTL coincided with a QTL for bread mixing time {0241}.

Height

Rht-B1. Add at end of section: The line XN004, earlier considered to have *Rht21* {0230}, was shown to carry an allele at the *Rht-B1* locus {0231}.

Rht-D1. Add at end of section: The line XN004, earlier considered to have *Rht21* {0230}, was shown to carry an allele at the *Rht-D1* locus {0231}.

Various common wheat and durum NIL pairs differing at the *Rht-A1* or *Rht-D1* loci are listed in {02102}.

Rht8a. Integrate alphabetically in the v: section:

Hope {0243}; Marquis {0243}; Michigan Amber {0243}.

Rht8b. Integrate alphabetically in the v: section:

Arthur {0243}; Carsten V {0243}; Diakovchanka {0243}; Odom {0243}; Oasis {0243}; Purdue Abe {0243}; Salzmünder Bartweizen 14/44 {0243}; Tp114/65 {0243}; Wiskonsin 245 C/11226 {0243};

Rht8c Integrate alphabetically in the v: section:

Al'batros odesskii {0243}; Arthur 71 {0243}; Donskaya polukarlikovaya {0243}; Erythrospermum 127 {0243}; Erythrospermum 1072 {0243}; Erythrospermum 272-87 {0243}; Erythrospermum 949-38 {0243}; Fakir {0243}; Fedorovka {0243}; Kaloyan {0243}; Khar'kovskaya 50 {0243}; Khar'kovskaya 93 {0243}; Khersonskaya 86 {0243}; Mv 03-89 {0243}; Mv 06-88 {0243}; Mv 17{0243}; Obrii {0243}; Odesskaya 51 {0243}; Odesskaya 117 {0243}; Odesskaya 132 {0243}; Odesskaya krasnokolosaya {0243}; Odesskaya polukarlikovaya {0243}; Roazon {0243}; Simvol odesskii {0243}; Sivka {0243}; Stremok {0243}; Tira {0243}; Ukrainka odesskaya {0243}; Vympel {0243}; Yubileinaya 75 {0243}; Zolotava {0243}.

At the end of the list {1999 Suppl.} add: 'Although CS carries a 192 bp fragment, sequencing showed it was a different allele than other genotypes with *Rht8c* {02103}.'

Rht8g. Associated with a 196-bp fragment of WMS 261 [{0243}]. v: Mirleben {0243}.

Rht8h. Associated with a 206-bp fragment of WMS 261 [{0243}]. v: Weihenstephan M1 {0243}.

Rht21 {0230}. 2DL {0230}. v: XN004 {0230}.

The existence of this gene could not be confirmed {0231}.

QHt.ipk-4A {0255}.

4AL {0255}. v: Opata/W-7984 (ITMI) RI mapping population {0255}. The height is contributed by Opata {0255}.

ma: Associated with *Xmwg549*, *Xabg390* and *Xbcd1670* {0255}.

QHt.ipk-4A coincides with QTLs for ear length (*QEl.ipk-4A*), grain number (*QGnu.ipk-4A*), and grain weight per ear (*QGwe.ipk-4A*) {0255}.

QHt.ipk-6A {0255}.

6A {0255}. v: Opata/W-7984 (ITMI) RI mapping population {0255}. The height is contributed by W-7984 {0255}.

ma: Associated with *Xcdo29* and *Xfba234* {0255}.

QHt.ipk-6A coincides with QTLs for peduncle length (*QPdl.ipk-6A*) and ear length (*QEl.ipk-6A*) {0255}

Leaf erectness (new category)

QLer.ipk-2A {0255}.

2AS {0255}. v: Opata/W-7984 (ITMI) RI mapping population {0255}. The erect leave... phenotype was contributed by Opata {0255}.

ma: Associated with *Xbcd348* {0255}.

Note: Mutants lacking ligules are known to have erect leaves. However, the QTL for leaf erectness reported here is not related to liguleless mutants {0255}.

Male sterility

ms1d {0290}.

v: Mutant FS2 {0290}.

ms1e {0290}.

v: Mutant FS3 {0290}.

ms1f {0290}.

v: Mutant FS24 {0290}.

ms3.

ma: *Xwg341-5A* – 0.8cM – *ms3* – cent {0289}. *Xcdo-677-5A* and *Xbcd1130-5A* also cosegregated with *Xwg341-5A* but were located in a different region in the physical map {0289}.

ms4 {0293}.

4DS {0293}. v: Konzak's male sterile.

Dominant allele for sterility, distinguished from *ms2* on the basis of different degrees of recombination with the 4D centromere.

ms5 {0290}.

3A {0290}. v: Mutant FS20 {0290}.

Meiotic characters

2. Pairing homoeologous

Ph1.

On a new line following the *ph1c* entry add: 'Several *ph1* mutants are described in {0219}.'.

ma: Add: PCR-based assays for presence and absence of *Ph1* have been described {0214, 0217, 9965}. The *Ph1* factor(s) was restricted to a region flanked by *Xrgc846-5B* and *Xpsr150-5B* {0219}.

Nucleolus organiser regions

Add at the end of descriptive paragraph and before allele descriptions:

'Deletion mapping divided the *Nor-B1* in a proximal subregion *Nor-B1p* (short repeat) and a distal subregion *Nor-B1d* (long repeat) {0275}'.

Proteins**1. Grain protein content**

QTL: Nine QTLs (51 % of the variation) were mapped in cross ‘Forno’/‘Oberkulmer’ spelt {0280}.

QGpc.ndsu-6Bb. Add at the end of the **ma:** section: {0244} reports the location of this QTL in the 4 cM interval flanked by *Xmwg79-6B* and *Xcd0365-6B*.

QTL: A QTL for grain and flour protein content, contributed by CS, was associated with *XTri-ID/Centromere* in a RSL population from the cross Cheyenne (high quality wheat) x CS (low quality wheat) {0251}.

3. Endosperm storage proteins**3.1 Glutenins*****Glu-A1***

Add:

Glu-A1u [{02106}]. 2^{*B} {02106}. v: Bánkúti 1201.

At the end of the *Glu-A1* section, just before the entry for the *Glu-B1* locus, add the paragraph:

‘The allele designated *Glu-A1u* above and *Glu-A1-Iu* in the appropriate list below encodes a high molecular weight glutenin subunit (denominated 2^{*B}) that is identical to subunit 2* apart from one amino acid difference involving the exchange of serine for cysteine (which itself is due to a C to G point mutation at the 1181-bp point of the coding region of 2*). The authors of {02106} suggest that the additional cysteine residue facilitates the formation of further disulphide bonds (cf. the 1Dx5 subunit) which might lead to an improvement in gluten quality characters.’

Glu-D1

Add:

Glu-D1al [{02107}]. 2.2* {02107}. v: MG315.

At the end of the *Glu-D1* section, just before the entry for the *Glu-1-1* and *Glu-1-2*, add the paragraphs:

‘The subunit 2.2* encoded by *Glu-D1al* above and *Glu-D1-1m* in the appropriate list below has an unusually high M_r; comparison of its N-terminal sequence and amino acid composition with those of subunit 2 (encoded by *Glu-D1-1a*) indicates that its greater M_r could be due to the presence of a larger central repetitive domain, although further evidence suggests that this does not affect the conformational properties of the subunit {02107}.

The alleles designated *Glu-D1w* (encoding ‘subunits’ 2 (or 2^t denoting its origin in the *Ae. tauschii* genome) +T1+T2), *Glu-D1ae* (encoding 2.1 (or 2.1^t)+T1+T2), *Glu-D1af* (encoding 3 (or 3^t)+T1+T2) and *Glu-D1ag* (encoding 1.5 (or 1.5^t)+T1+T2) share the component T1 that was originally classified as a HMW glutenin. However, it has since been shown {02108} that this protein is soluble in aqueous ethanol, casting doubt upon this classification. More recently, it has been shown {02109}, from one- and two-dimensional gel electrophoresis based upon SDS-PAGE and A-PAGE, and from N-terminal sequencing, that this protein is an ω-gliadin of unusually low electrophoretic mobility in SDS-PAGE, encoded by a locus located on the short arm of chromosome 1D, though distant (13.18 cM) from the principal gliadin encoding locus on 1D, *Gli-D1*, and 40.20 cM from the high molecular weight encoding locus, *Glu-D1*. The authors named the locus *Gli-D1* (see below, section ‘3.2 Gliadins’).

Glu-A1-1

Add:

Glu-A1-1u [{02106}]. 2^{*B} {02106}. v: Bánkúti 1201.

Glu-D1-1

Add:

Glu-D1-1m [{02107}]. 2.2* {02107}. v: MG315.

Just before the entry for *Glu-E3*, add the following phrase to the previous paragraph (which, following the amendments made in the 2000 Supplement, reads: ‘In {00111}, in a study of bread and durum wheats from Portugal, the authors used the nomenclature system described in {00113} for the LMW subunits in bread wheat, and that described in {00114} for the LMW subunits in durum wheat.’): ‘The latter system, updated according to {02110}, is reproduced here:

Follow this with the following new entries:

Glu-B2

Add:

<i>Glu-B2a</i> {00114}.	12 {00114}.	v: Mexicali, <i>T. durum</i> .
<i>Glu-B2b</i> {00114}.	Null {00114}.	v: Langdon, <i>T. durum</i> .

Add at the end of the sentence ‘The *Glu-3* loci are defined as the cluster of LMW glutenin genes previously considered a component of the compound *Gli-1* loci.’

‘More than 30 LMW glutenin complete genes, partial genes or pseudogenes have been sequenced from *Triticum* species (reviewed in {0245}).

Glu-A3

Add:

<i>Glu-A3a</i> {00114}.	6 {00114}.	v: Mexicali, <i>T. durum</i> .
<i>Glu-A3b</i> {00114}.	5 {00114}.	v: Langdon, <i>T. durum</i> .
<i>Glu-A3c</i> {00114}.	6+10 {00114}.	v: Cocorit, <i>T. durum</i> .
<i>Glu-A3d</i> {00114}.	6+11 {00114}.	v: Alaga, <i>T. durum</i> .
<i>Glu-A3e</i> {00114}.	11 {00114}.	v: Blatfort, <i>T. durum</i> .
<i>Glu-A3f</i> {00114}.	6+11+20 {00114}.	v: Clarofino, <i>T. durum</i> .
<i>Glu-A3g</i> {00114}.	6+10+20 {00114}.	v: Claro de Balazote, <i>T. durum</i> .
<i>Glu-A3h</i> {00114}.	null {00114}.	v: Jiloca, <i>T. durum</i> .
<i>Glu-A3i</i> {02110}.	8*+11 {02110}.	v: Mourisco Fino, <i>T. durum</i> .

Glu-B3

Add:

<i>Glu-B3a</i> {00114}.	2+4+15+19 {00114}.	v: Mexicali, <i>T. durum</i> .
<i>Glu-B3b</i> {00114}.	8+9+13+16 {00114}.	v: Langdon, <i>T. durum</i> .
<i>Glu-B3c</i> {00114}.	2+4+14+15+19 {00114}.	v: Jiloca, <i>T. durum</i> .
<i>Glu-B3d</i> {00114}.	2+4+15+17+19 {00114}.	v: Mundial, <i>T. durum</i> .
<i>Glu-B3e</i> {00114}.	2+4+15+16+18 {00114}.	v: Granja Badajoz, <i>T. durum</i> .
<i>Glu-B3f</i> {00114}.	2+4+15+17 {00114}.	v: Ardente, <i>T. durum</i> .
<i>Glu-B3g</i> {00114}.	2+4+15+16 {00114}.	v: Claro de Balazote, <i>T. durum</i> .
<i>Glu-B3h</i> {00114}.	1+3+14+18 {00114}.	v: Alaga, <i>T. durum</i> .
<i>Glu-B3i</i> {00114}.	7+8+14+18 {00114}.	v: Blatfort, <i>T. durum</i> .
<i>Glu-B3j</i> {02110}.	4+6*+15+19 {02110}.	v: Mourisco Fino, <i>T. durum</i> .
<i>Glu-B3k</i> {02110}.	8+9+13+16+19 {02110}.	v: Faísca, <i>T. durum</i> .

At end of the Glutenins section, just before the heading ‘3.2 Gliadins’, add the paragraph:

‘The following loci, *Glu-D4* and *Glu-D5*, encoding low molecular weight subunits of glutenin (30-32 kDa) have been described in {02111}; the proteins encoded by them were first observed earlier {02114, 02115}, and the former was later tentatively assigned the symbol *Glu-4* {02116}, before its chromosomal location was established and the locus definitively named as *Glu-D4* in {02111}. While this locus is located on chromosome 1D (in accordance with the position on the group 1 chromosomes of the remaining glutenin encoding loci found to date), the locus *Glu-D5* is located on chromosome 7D. In SDS-PAGE, the proteins from both loci are detected only in the presence of 4-vinylpyridine added to the sample extract. Their amino acid composition does not match that of the major prolamin groups; nonetheless, they classify as glutenins based upon solubility, immunological behaviour and N-terminal amino acid sequence (the latter suggesting an evolutionary link with the major (B and C) low molecular weight glutenin subunits).’

Then add the following entries:

<i>Glu-D4</i> {02111}.	1D {02111}.	su: CS/Langdon 1D(1A); CS/Langdon 1D(1B) {02111}.
<i>Glu-D4a</i> {02111}.		v: J 24.
<i>Glu-D4b</i> {02111}.		v: PBW 154.
<i>Glu-D4c</i> {02111}.	null allele.	v: NI 4.
<i>Glu-D5</i> {02111}.	7D {02111}.	su: CS/Langdon 7D(7A); CS/Langdon 7D(7B) {02111}.
<i>Glu-D5a</i> {02111}.		v: PBW 154.
<i>Glu-D5b</i> {02111}.	null allele	v: K 68.

Continue with the following paragraph:

'A collection of 173 *Triticum tauschii* accessions have been analysed for low molecular weight glutenin subunits by SDS-PAGE {02112}. 33 different patterns for B-subunits and 43 for C-subunits were identified, some of which were of identical electrophoretic mobility to those observed in bread wheat. Also observed were subunits with the same mobilities as the D-subunits and as the subunits encoded by the *Glu-D4* and *Glu-D5* loci. This variation represents a source of novel germplasm of potential value for breeding programmes aimed at improving the D-genome of bread wheat in the context of bread-making quality.'

3.2. Gliadins

Add at the end of the section, just before the heading '3.3 Other endosperm storage proteins', add the paragraph:

'A locus designated *Gli-DT1* controlling an ω -gliadin of *Ae. tauschii* has been mapped on the short arm of chromosome 1D between loci *Gli-D1* (strictly *Gli-D'1*) and *Glu-D1* (strictly *Glu-D'1*), 13.18 cM proximal to the former and 40.20 cM from the latter {02109}. The only ω -gliadin to date identified as being encoded by this locus, namely T1, is of unusually low electrophoretic mobility in SDS-PAGE gels and was formally thought to be a high molecular weight glutenin encoded by the *Glu-D'1* locus of *Ae. tauschii* (see note following the *Glu-D1* list in section '3.1 Glutenins'). The authors speculate that, due to their similar relative map positions, the loci *Gli-A4*, *Gli-D4*, *Gli-R3*, *Gli-S'4* and this locus, *Gli-DT1*, form a series of '*Gli-4*' orthologous loci. However, this should be interpreted in the light of the above discussion on *Gli-A3* and *Gli-A4*.'

Then add the entry:

Gli-DT1 {02109}. 1DS {02109}.
Gli-DT1a [{02109}]. T1.

dv: AUS18913 *Ae. tauschii*; L/18913 (synthetic 6X).
dv: AUS18913 *Ae. tauschii*; L/18913 (synthetic 6X).

Follow this entry with the following paragraph:

'Four new classes of low molecular weight proteins related to gliadins, though not sufficiently similar to be classified as such, have been reported in {02113}. One of the classes has no close association to previously described wheat endosperm proteins.'

5. Other proteins

5.6 Waxy proteins

At end of preamble add: 'Partial genomic clones of various diploid, tetraploid, and hexaploid wheats have been sequenced {0278,0279}.'

Wx-d1e {0234}. **v:** Tanikei A6599-4 {0234}. Relative to Kanto 107, Tanikei A6599-4 carries an alanine to threonine substitution at position 258 in the mature protein {0234}.

5.8. Puroindolines

Revised section: Puroindolines a and b are the major components of friabilin, a protein complex that is associated with grain texture (see 'Grain Hardness'). Hard wheats result from unique changes in the puroindoline amino acid sequence or, currently, four null forms {0295} of the completely linked genes (max. map distance 4.3 cM) {452}.

Pina-A^m1 {0083} 5A^mS {0083}. **dv:** *T. monococcum* DV92, G3116 {0083}

In *T. monococcum* *Pina-A^m1* is completely linked to *Gsp-A^m1* {0083}.

Pina-D1 5DS {452}. **v:** CS

Pina-D1a {452}.

v: Aurelio *Pinb-D1a* {0249}; Bellevue {0249}; Bezostaja *Pinb-D1b* {0249}; Bilancia *Pinb-D1a* {0249}; Bolero *Pinb-D1a* {0249}; Brasilia *Pinb-D1b* {0249}; Centauro *Pinb-D1a* {0249}; Cerere *Pinb-D1b* {0249}; Chinese Spring *Pinb-D1a* {452,0249}; Colfiorito *Pinb-D1b* {0249}; Cologna 21 *Pinb-D1b* {0249}; Courtot {0249}; David *Pinb-D1b* {0249}; Democrat *Pinb-D1b* {0249}; Etruria *Pinb-D1b* {0249}; Fortuna {0249}; Francia *Pinb-D1b* {0249}; Galaxie 0249; Gemini *Pinb-D1b* {0249}; Genio *Pinb-D1b* {0249}; Gladio *Pinb-D1b* {0249}; Heron {1035}; Lampo *Pinb-D1a* {0249}; Leone *Pinb-D1a* {0249}; Leopardi *Pinb-D1a* {0249}; Libero *Pinb-D1a* {0249}; Livio *Pinb-D1a* {0249}; Marberg *Pinb-D1b* {0249}; Mentana *Pinb-D1a* {0249}; Mieti *Pinb-D1b* {0249}; Mosè *Pinb-D1a* {0249}; Neviana *Pinb-D1a* {0249}; Newana *Pinb-D1b* {0249}; Oscar *Pinb-D1a* {0249}; Pandas *Pinb-D1b* {0249}; Pascal *Pinb-D1b* {0249}; Sagittario *Pinb-D1b* {0249};

Salgemma *Pinb-D1b* {0249}; Saliente *Pinb-D1b* {0249}; Salmone *Pinb-D1b* {0249}; Serena *Pinb-D1a* {0249}; Serio *Pinb-D1b* {0249}; Soissons {0249}; Veda *Pinb-D1b* {0249}; Zena *Pinb-D1b* {0249}.'

Pinb-D1a is present in all soft hexaploid wheats and possibly all hard hexaploid wheats carrying a hardness mutation in puroindoline b {452,1035,0082, 0204}.

Pinb-D1b {1035}. Null allele

v: Amidon *Pinb-D1a* {0249}; Barra *Pinb-D1a* {0249}; Butte 86 {1035}; Ciano *Pinb-D1a* {0249}; Dorico *Pinb-D1a* {0249}; Eridano {0249}; Falcon {1035}; Fortuna (USA) *Pinb-D1a* {0249}; Glenman *Pinb-D1a* {0249}; Golia *Pinb-D1a* {0249}; Guadalupe *Pinb-D1a* {0249}; Inia 66 *Pinb-D1a* {0249}; Jecora *Pinb-D1a* {0249}; Indice *Pinb-D1a* {0249}; Kalyansona {0249}; Manital *Pinb-D1a* {0249}; Mendoz *Pinb-D1a* {0249}; Padus *Pinb-D1a* {0249}; Prinqual *Pinb-D1a* {0249}; Sibilia *Pinb-D1a* {0249}; Super X {0249}; Yecora Rojo {0204}.

i: Gamanya sel. {0298,0203}; Heron/7*Falcon sel. {0298, 0203}.

Present only in some hard hexaploid wheats. *Pinb-D1b* is associated with harder texture than *Pinb-D1b*{0177, 0206}.

Pinb-A^mI {0083}. 5A^mS {0083}. **dv:** *T. monococcum* DV92, G3116 {0083}.

In *T. monococcum* *Pinb-A^mI* is 0.1 cM proximal to *Pinb-A^mI* and both loci are less than 36 kb apart.

Pinb-D1 5DS {452}

v: CS.

Pinb-D1a {452}.

v: Amidon *Pina-D1b* {0249}; Aurelio *Pina-D1a* {0249}; Barra *Pina-D1b* {0249}; Bilancia *Pina-D1a* {0249}; Bolero *Pina-D1a* {0249}; Centauro *Pina-D1a* {0249}; Chinese Spring *Pina-D1a* {452,0249}; Ciano *Pina-D1b* {0249}; Dorico *Pina-D1b* {0249}; Fortuna (USA) *Pina-D1b* {0249}; Glenman *Pina-D1b* {0249}; Golia *Pina-D1b* {0249}; Guadalupe *Pina-D1b* {0249}; Hill 81 {452}; Inia 66 *Pina-D1b* {0249}; Jecora *Pina-D1b* {0249}; Idice *Pina-D1b* {0249}; Lampo *Pina-D1a* {0249}; Leone *Pina-D1a* {0249}; Leopardo *Pina-D1a* {0249}; Libero *Pina-D1a* {0249}; Livio *Pina-D1a* {0249}; Manital *Pina-D1b* {0249}; Mendoz *Pina-D1b* {0249}; Mentana *Pina-D1a* {0249}; Mos^È *Pina-D1a* {0249}; Neviano *Pina-D1a* {0249}; Oscar *Pina-D1a* {0249}; Padus *Pina-D1b* {0249}; Prinqual *Pina-D1b* {0249}; Serena *Pina-D1a* {0249}; Sibilia *Pina-D1b* {0249}.

Pinb-D1a is present in all soft hexaploid wheats and possibly all hard hexaploid wheats carrying the *Pina-D1b* mutation {452,1035,0082,0204}.

Pinb-D1b {452}. 5DS {452}.

i: Paha*2//Early Blackhull/5*Paha {02111,0203}; Early Blackhull der./5*Nugaines sel. {0298,0203}.

s: CS*7/Cheyenne 5D {452}.

v: Bezostaya *Pina-D1a* {0249}; Brasilia *Pina-D1a* {0249}; Cerere *Pina-D1a* {0249}; Colfiorito *Pina-D1a* {0249}; Cologna 21 *Pina-D1a* {0249}; David *Pina-D1a* {0249}; Democrat *Pina-D1a* {0249}; Etruria *Pina-D1a* {0249}; Francia *Pina-D1a* {0249}; Gemini *Pina-D1a* {0249}; Genio *Pina-D1a* {0249}; Gladio *Pina-D1a* {0249}; Marberg *Pina-D1a* {0249}; Mieti *Pina-D1a* {0249}; Newana *Pina-D1a* {0249}; Pandas *Pina-D1a* {0249}; Pascal *Pina-D1a* {0249}; Sagittario *Pina-D1a* {0249}; Salgemma *Pina-D1a* {0249}; Saliente *Pina-D1a* {0249}; Salmone *Pina-D1a* {0249}; Serio *Pina-D1a* {0249}; Thatcher {0204}; Veda *Pina-D1a* {0249}; Wanser {452}; Zena *Pina-D1a* {0249}; hard component of Turkey {0204}.

Pinb-D1b is a “loss-of-function” mutation resulting from the replacement of a glycine by a serine at position 46 {452}.

Pinb-D1c {0082}. **v:** Avle {0082}; Reno {0082}; Tjalve {0082}; Bjorke {0082}; Portal {0082}.

Pinb-D1c is a “loss-of-function” mutation resulting from the replacement of a leucine by a proline at position 60 {0082}.

Pinb-D1d {0082}. **v:** Bercy {0082}; Mjolner {0082}.

Pinb-D1d is a “loss-of-function” mutation resulting from the replacement of a tryptophan by a arginine at position 44 {0082}.

Pinb-D1e {0204}. **v:** Gehun {0204}; Canadian Red {0204}; Chiefkan {0204}.

Pinb-D1e is a “loss-of-function” mutation resulting from the replacement of a tryptophan by a stop codon at position 39 {0204}.

Pinb-D1f {0204}. **v:** the hard component of Utac {0204}.

Pinb-D1f is a “loss-of-function” mutation resulting from the replacement of a tryptophan by a stop codon at position 44 {0204}.

Pinb-D1g {0204}. **v:** Andrews {0204}.

Pinb-D1g is a “loss-of-function” mutation resulting from the replacement of a cysteine by a stop codon at position 56 {0204}.

Pinb-D1b, *Pinb-D1c*, *Pinb-D1d*, *Pinb-D1e*, *Pinb-D1f*, or *Pinb-D1g* are present in hard hexaploid wheats not carrying the *Pina-D1b* (null) mutation {452,1035,0082,0204}.

Wheats with *Pinb-D1b* were slightly softer and a little superior to those with *Pina-D1b* in milling and bread-making characteristics although there was considerable overlap {0206}. Transgenic rice with the *Pina-D1a* and *Pinb-D1a* alleles possessed softer grain {0207}.

Genotypes for a selection of North American wheats are given in {0204}.

5.9. Histone H1 proteins

<i>HstH1-A1</i> {0215}.	5AL {0215}.	v: CS {0215}.
<i>HstH1-B1</i> {0215}.	5BL {0215}.	v: CS {0215}.
<i>HstH1-D1</i> {0215}.	5DL {0215}.	v: CS {0215}.
<i>HstH1-D1a</i> {0215}.		v: CS {0215}; 18 others {0215}.
<i>HstH1-D1b</i> {0215}.		v: Grekum 114 {0215}; Kirgizsky Karlik {0215}.
<i>HstH1-A2</i> {0215}.	5AL {0215}.	v: CS {0215}.
<i>HstH1-A2a</i> {0215}.		v: CS {0215}.
<i>HstH1-A2b</i> {0215}.	Null allele {0215}.	v: Mara {0215}; 10 others {0215}.
<i>HstH1-B2</i> {0215}.	5BL {0215}.	v: CS {0215}.
<i>HstH1-B2a</i> {0215}.		v: CS {0215}; 19 others {0215}.
<i>HstH1-B2b</i> {0215}.		v: Excelsior {0215}.
<i>HstH1-D2</i> {0215}.	5DL {0215}.	v: CS {0215}.

The relationship of this gene series with a *Hst-A1*, *Hst-B1*, *Hst-D1* series in group 5 chromosomes {0216} based on DNA hybridization studies was not established.

Response to tissue culture

Add at the end of the section:

<i>QGpp.kvl-2A</i> {0253}.	2AL {0253}.	v: Ciano/Walter DH mapping population {0253}. The green plant percentage was contributed by Ciano {0253}.
	ma:	Associated with <i>Xpsp3045-2A</i> {0253}.
<i>QGpp.kvl-2B.1</i> {0253}.	2BL {0253}.	v: Ciano/Walter DH mapping population {0253}. The green plant percentage was contributed by Ciano {0253}.
	ma:	Associated with <i>Xgwm388-2B</i> {0253}.
<i>QGpp.kvl-2B.2</i> {0253}.	2BL {0253}.	v: Ciano/Walter DH mapping population {0253}. The green plant percentage was contributed by Ciano {0253}.
	ma:	Associated with AFLP markers {0253}.
<i>QGpp.kvl-2A</i> {0253}.	2AL {0253}.	v: Ciano/Walter DH mapping population {0253}. The green plant percentage was contributed by Ciano {0253}.
	ma:	Associated with <i>Xpsp3045-2A</i> {0253}.

Response to vernalization

Vrn-B1. Vrn2. 5BL or 7BL. Add to reference {635}, i.e. {635,0282}.

In the final paragraph include reference 0202 with the first reference, i.e. {1173, 0202}.

Yellow berry tolerance

QTL: A QTL for yellow berry tolerance, contributed by RS111, was associated with *Xgwm190* and *Xgwm174* on chromosome 5D in a RIL population from RS111/CS {0237}. A tolerance QTL contributed by CS, the susceptible parent, was detected on 6B {0237}.

Yield components**1000-grain weight**

QTgw.ipk-5A {0255}. 5AL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
The higher yielding allele is contributed by W-7984 {0255}.

ma: Associated with *Xfba351* and *Xcdol312* {0255}.

QTL: QTLs for grain size were identified on chromosome arms 1DS, 2DL, and 6BL in a RIL population from RS111/CS {0236}.

QTL: Eight QTLs for 1,000-kernel weight (54 % of the variation) were mapped in ‘Forno’/‘Oberkulmer’ spelt {0280}.

Kernel number per spike

QGnu.ipk-4A {0255}. 4AL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
Higher kernel number was contributed by Opata {0255}.

ma: Associated with *Xmwg549*, *Xabg390* and *Xbcd1670* {0255}.

QGnu.ipk-4A coincides with QTL for height (*QHt.ipk-4A*), spike length (*XEl.ipk-4A*), and grain weight per ear (*QGwe.ipk-4A*) {0255}.

Spike length

QEl.ipk-1B {0255}. 1BL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
Longer ear was contributed by Opata {0255}.

ma: Associated with *Xbcd388* and *Xwg605* {0255}.

QEl.ipk-4A {0255}. 4AL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
Longer ear was contributed by Opata {0255}.

ma: Associated with *Xmwg549*, *Xabg390* and *Xbcd1670* {0255}.

This QTL is likely to be a pleiotropic effect of the gene underlying the height QTL, *QHt.ipk-4A* {0255}.

QEl.ipk-5A {0255}. 5AL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
Longer ear was contributed by W-7984 {0255}.

ma: Associated with *Xmwg522* {0255}.

Grain weight/ear

QGwe.ipk-2D {0255}. 2DS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
Higher grain weight was contributed by Opata {0255}.

ma: Associated with *Xcdol379* and *Xbcd1970* {0255}.

QGwe.ipk-4A {0255}. 4AL {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.
Higher grain weight was contributed by Opata {0255}.

ma: Associated with *Xmwg549*, *Xabg390* and *Xbcd1670* {0255}.

QGwe.ipk-4A coincides with QTL for height (*QHt.ipk-4A*), spike length (*XEl.ipk-4A*) and grain number (*QGnu.ipk-4A*) {0255}.

Pathogenic Disease/Pest Reaction**Reaction to Barley Yellow Dwarf Virus**

Bdv2.

tr: TC14 {059,0201}.

v: TC14*2/Hartog {0225}; TC14/2*Spear {0201}; TC14/2*Tatiara {0225}.

ma: Complete association with *Xpsr129-7D*, *Xpsr548-7D*, *XksuD2-7D*, *XcslH8I-7D*, and *Xgwm37-7D* selected as a diagnostic marker {0225}.

Reaction to *Diuraphis noxia*

Dn1. 7DS {0211}. **i:** Betta-Dn1 {0211}; Karee-Dn1 {0211}; Tugela-Dn1 {0211}.

ma: $Xgwm111\text{-}7D_{210}$ - 3.20 ± 0.20 cM - *Dn1* {0211}.

Dn2. 7DS {0211}. **i:** Betta-Dn2 {0211}; Karee-Dn2 {0211}; Tugela-Dn2 {0211}.

ma: $Xgwm111\text{-}7D_{200}$ - 3.05 ± 0.18 cM - *Dn2* {0211}.

Dn4. **v:** Halt {0209}.

Dn5. 7DS {0211}. **i:** Betta-Dn1 {0211}.

ma: $Xgwm111\text{-}7D_{220}$ - < 3.20 cM - *Dn5* {0211}.

Dn8 {0211}. 7DS {0211}. **i:** Karee-Dn8.

v: PI 294994 *Dn5Dn9* {0211}.

ma: $Xgwm635\text{-}7D_{100}$ - < 3.20 cM - *Dn8* {0211}.

Dn9 {0211}. 1DL {0211}. **i:** Betta-Dn9.

v: PI 294994 *Dn5Dn8* {0211}.

ma: $Xgwm642\text{-}7D_{180}$ - < 3.20 cM - *Dn9* {0211}.

Dnx {0211}. 7DS {0211}. **v:** PI 220127 {0211}.

ma: $Xgwm111\text{-}7D_{210}$ - 1.52 ± 0.15 cM - *Dnx* {0211}.

Dnx was considered to be located at a locus different from *Dn1*, *Dn2*, or *Dn5* {0211}, which were likely to be identical or allelic.

Reaction to *Erysiphe graminis*

Pm4b. **ma:** *Pm4b* - 4.8cM - *Xgbx3119b* {0272}.

Pm5a {0257}. **Pm5**{787}. **v:** Add: 'Galaxie' {0257}; Kutulukskaya {0257}; Lambros {0257}; Navid {0257}; Pagode {0257}; Regina {0257}; Sicco (0257); Tarasque {0257}; Zolotistaya {0257}.'

Pm5b {0257}. **Mli** {540,558}. **v:** Add: Cucurova {0257}; Fruhprobst {0257}; Kirkpinar-79 {0257}; Kontrast {0257}; Ilona {0257}; Nadadores {0257}; Siete Cerros {0257}; Una {0257}; Wettiness {0257};

Pm5c {0257}. 7B {0257}. **v:** *T. sphaerococcum* cv. Kolandi {0257}.

Pm5d {0257}. 7B {0257}. **i:** IGV 1-455 = CI 10904/7*Prins {0257}; CI 10904/7*Starke {0257}.

Pm5e {0258}. **mlfz** {0259}. **v:** Fuzhuang 30 {0258}.

ma: $Xgwm1267\text{-}7B$ - 6.6cM - *Pm5e* - 12.6cM - *Xubc405*₆₂₈ {0258}.

Pm8. **ma:** A STS marker distinguishes *Pm17* from *Pm8* {0286}.

Pm17. T1BL·1RS.
ma: A STS marker distinguishes *Pm17* from *Pm8* {0286}.

Pm30. **ma:** *Pm30* - 5.6 cM - *Xgwm159-5B* {0163}.

Mlxbd {0259}. 7B {0259}. **v:** Xiaobaidong {0258}.

Add to genotype list: '{02104}' (Hungarian wheats).'

QTL: Several QTLs were detected in two 'RE714/Hardi' populations when tested at two growth stages and with different cultures over three years. The most persistent band effective QTL was located in the vicinity of *Xgwm174-5D* {0272}. Three QTLs, *QPm.vt-1B*, *QPm.vt-2A*, and *QPm.vt-2B*, with additive gene action, accounted for 50 % of the variation in a population developed from Becker/Massey {0284}.

QPm.ipk-2B {0255}. 2BS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.

Resistance was contributed by Opata {0255}.

ma: Associated with *Xcdo405* and *Xmwg950* {0255}.

QPm.ipk-4B {0255}. 4B {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.

Resistance was contributed by W-7984 {0255}.

ma: Associated with *Xcdo795* and *Xbcd1262* {0255}.

QPm.ipk-7D {0255}. 7DS {0255}. **v:** Opata/W-7984 (ITMI) RI mapping population {0255}.

Resistance was contributed by Opata {0255}.

ma: Associated with *Xwg834* and *Xbcd1872* {0255}.

Reaction to *Fusarium graminearum*

QFhs.ndsu-3B {9925,0175}.

3BS {9925}. **v:** Sumai 3 {9925,0175}.

ma: Associated with *Xbcd907-3B.2* (LOD>3) {9925} and microsatellite markers *Xgwm533* and *Xgwm493* {0175}. This QTL explained 41.6 % of the variation in the cross Sumai3/Stoa {0175}.

QTL: Two additional QTL for resistance to *Fusarium graminearum* were identified in the cross 'Sumai3/Stoa' {0175}. The QTL on 4BS was associated with *Xwg909* and the QTL on 6BS was associated with *Xbarc101* and *Xbcd1383* {0175}. The QTL associated with markers *Xgwm493/Xgwm533* (explaining 24.8 % of the variation), and *Xbarc101/Xbcd1383* were also identified in a RIL population from the cross 'ND2603/Butte 86' {0175}. In addition, one QTL on chromosome 3AL associated with *Xbcd941* and one on chromosome 6AS associated with *XksuH4* were identified in RILs from the cross 'ND2603/Butte 86' {0175}.

The resistance QTL on chromosome 3BS associated with *Xgwm493* and *Xgwm533* was also identified in a DH population of the cross 'CM-82036 (a Sumai 3 derivative) x Remus' {0240}. Additional QTL in this cross were detected on chromosome 5A, associated with *Xgwm293* and *Xgwm304*, and possibly on 1B, associated with *Glu-B1* {0240}.

For review see {0283}.

Reaction to *Heterodera avenae*

Cre8 {0220}. **CreF** {0012,0138}. 6B {0220}. [On basis of linkage with *Xbcd1* and *Xcdo347*].

v: Barunga {0220}; Festiguay {0012,0220}; Frame {0138, 0220}; Molineaux {0220}.

ma: Associated with a unique allele when probed with CDO367, which hybridizes to group 7L {1059}.

Reaction to *Mycosphaerella graminicola*

Stb5. **ma:** *Rc3* - 6.6cM- *Stb5* - 7.2cM - *Xgwm44/Centromere* {0186}.

Reaction to *Phaeosphaeria nodorum*

SnbTM. **ma:** *UBC521*₆₅₀ – 15 cM – *SnbTM* – 13.1 cM – *RC37*₅₁₀ {0212}. *UBC521*₆₅₀ was converted to a SCAR marker {0212}.

Reaction to *Pseudocercosporella herpotrichoides*

Pch1. 7A {0224}. **tv:** Five recombinant lines {0224}.

Reaction to *Puccinia graminis tritici*

Sr22. **ma:** Add: 'See also {0158}.'

Sr26. **ma:** Can be detected with several RFLP probes {0138}.

Sr31. T1BL·1RS: **v:** Cougar {0267}; Rawhide (heterogeneous) {0267}.

Sr36.

v: GK KincsQ {0235}.

- Sr38* {062}. Derived from *Ae. ventricosa*. See Reaction to *P. recondita tritici Lr37* and *P. striiformis tritici Yr17* for details.
 v: Moisson derivatives Mx12 and Mx22 also carry *Sr38* {0213}.

Reaction to *Puccinia recondita**Lr10.* v: Scout 66 {02101}.

Lrk10. A receptor-like kinase. The locus *Xsf1(Lrk10)-1A*, detected by the probe *Lrk10*, is completely linked with *Lr10* in chromosome 1AS {356}. *Lrk10* encodes a receptor-like kinase with extracellular and kinase domains {0297}. Using probe pLrk10-A, developed from the extracellular domain, six homologues were found in chromosomes 1A (1), 1B (3), and 1D (2) as well as group-1 chromosomes of *T. monococcum*, *Ae. tauschii*, and barley {0296, 0294}. Probes based on the kinase domain identified further homologues in chromosomes 3AS and 3BS as well as the corresponding regions in rice and maize {0294}. Both orthologous and paralogous evolution were suggested.

- Lr11.* v: Karl 92 *Lr3 Lr10* {02101}.
- Lr12.* v: AC Domain *Lr10 Lr34*{0228}.
- Lr13.* v: Hereward {0288}; Moulin {0288}; Pastiche {0288}. BH1146 *Lr34* {0268}.
- Lr16.* v: Arapahoe {02101}; Brule {02101}; Millenium {02101}; Redland {02101}; Vista {02101}.
- Lr17b.* 2A {1350}. v: Brock {0260}; Tarso {0229}; Norman {1350}.
- Lr19.* 7BL. v: 4 further derivatives of 88M22-149 {0232}.
 ma: An STS marker closely linked and distal to *Lr19* was developed from an AFLP {0273}.
- Lr21.* v: McKenzie {0228}; WGRC2 = 'TA1649/3* Wichita' {0299}; WGRC7 = 'Wichita/TA1649//2* Wichita' {0299}.
 dv: *Aegilops squarrosa* accessions: RL5289 = TA1599 {1241}; TA1649 {0299}; TA1691 {0299}; TA2378 {0299}; TA2470 {0299}; TA2483 {0299}; TA2495 {0299}; TA2527 {0299}; TA2528 {0299}.
 ma: All members of the *Lr21* family carry a STS derivative of *ksuD14-1D* that has a resistance gene analogue structure {0299}.
- Lr23.* v: Cranbrook {02119}.
- Lr26.* T1BL·1RS: v: Cougar {0267}; Rawhide (heterogeneous) {0267}.
Lr34. v: Westphal 12 {0268}; BH1146 *Lr13* {0268}.
- Lr37* {062}. 2AS {062}. Derived from *Ae. ventricosa*.
VPM1 and derivatives: 2AS {062} = T2AL·2AS-2N^vS {0213}.
 i: Tc*8/VPM1 {316}; Various NILs listed in {0213}.
 v: Hyak {021}; Madsen {020}; Rendezvous {062}; VPM1 {062}. See also Reaction to *P. striiformis tritici Yr17*.
- Moisson derivatives: *Lr* {113}. 2AS = T2AL·2AS-2N^vS {113}.
 ad: Moisson + 6N^v = 6N^vS.6N^vL-2N^vS or 6N^vL.6N^vS-2N^vS {0009}.
 v: Mx12 {0213}; Mx22 {0213}.
 ma: (relevant to both groups of derivatives). PCR primers designed from marker csVrga1D3' {0183} producing a 383 bp product allows detection of the 2N^vS segment {0213}. See also: Reaction to *P. striiformis tritici Yr17*.

Lr37 can be recognised in seedling tests at low temperatures (17°C) and is effective in adult plants under field conditions.

Lr39 {02100}. 2DS {02100}. Derived from *Aegilops tauschii* {02100}.
v: TA4186 = TA1675*2/Wichita {02100}.
dv: *Aegilops tauschii* TA 1675 {02100}.
ma: 10.7 cM distal to *Xgwm210-2D* {02100}.

Lr41. **v:** Thunderbolt {02100}.

Lr50 {0221}. 2BL {0221} [Based on linkage with SSR markers].
v: WGR36 = TAM107*3/TA870/Wichita {0221}.
tv: *T. armeniacum* TA870 {0221}.

LrTm {0277}. **dv:** *T. monococcum*.
ma: Linked to microsatellite locus *Xgwm136* {0277}.

LrTr {0227}. **v:** *Aegilops triuncialis* derivatives {0227}.
ad: WL711 BC2F5 addition lines {0227}.
al: *Aegilops triuncialis* Acc. 3549 {0227}.
ma: Lines with *LtTr* possessed a homologue of *Xgwm368-4B* {0227}.

Genotype lists: Australian wheats {0288}, European wheats {0229, 0260, 0288}.

Reaction to *Puccinia striiformis tritici*

Yr9. T1BL·1RS: **v:** Cougar {0267}; Rawhide (heterogeneous) {0267}.

Yr10. YrVav {0262}. **v:** QLD709 = Janz*2/*T. vavilovii* {0262}.
tv: *T. vavilovii* AUS 22498 {0262}.
ma: A SCAR marker was described in {0261}. QLD709 and *T. spelta* 415, both with white glumes, failed to amplify the SCAR sequence, but both carried unique alleles at the *Gli-B1* and *Xpsp3000* loci {0262}. These differed from the Moro source of *Yr10*. *Yr10* – 1.5 ± 0.9 cM – *Gli-B1* – 1.1 ± 0.8 cM – *Xpsp3000* {0262}.

Yr17 {062}. 2AS {062}. See reaction to *P. recondita tritici* *Lr37* for details.
v: Genotype list in {02105}.
v: Arche {0044}, Balthazar {0044}, Brigadier {0044}, Cordial {0044}, Eureka {0044}, Hussar {0044}, Lynx {0044}, Pernel {0044}, Renan {0044}.
ma: *Yr17* was closely linked to the SCAR marker SC-Y15, developed from RAPD marker OP-Y15₅₈₀, and to *Xpsr150-2N^v* {0044}.

Yr26. 1BS {0285}. The earlier reported location of T6AL-6VS {617} is not correct.
v: Wheat-Haynaldia villosa lines R43, R55, R64, and R77 {0285}.
tv: *T. turgidum* Gamma 80-1.
ma: *Yr26* – 1.9 cM – *Xgwm11-1B/Xgwm18-1B* {0285}.

Yr28. **ma:** Linkage with *Xmwg634-4D* {1377}.

YrH52.

QTL: In the ITMI mapping population, QTLs were found in 2BS, 7DS, and possibly 5A, 3D, and 6D {0287}. In Camp Remy/Michigan Amber, QTLs were found in 2AL and 2BL {0287}.

Reaction to *Pyrenophora tritici repentis*

2. Resistance to chlorosis induction

QTsc.ndsu-1A. Add {0040,0264}' to the references for QTL and the marker association.

QTsc.ndsu-4A. Add to: **v:** In W-7976/Trenton resistance was contributed by W-7976 {0264}.

ma: Add: 'In W-7976/Trenton there was association with Xwg622-4A{0264} and minor QTLs in chromosomes 1AL, 7DS, 5AL, and 3BL were associated with resistance in adult plants {0264}'.

Reaction to *Sitodiplosis mosellana*

Insect pest: Orange blossom wheat midge, Wheat midge

Sm1 {0218}. 2B {0218}. **v:** Augusta {0218}; Blueboy {0218}; Caldwell {0218}; Clark {0218}; FL302 {0218}; Howell {0218} Knox 62 {0218}; Mono {0218}; Seneca {0218}.
ma: Linked to a SCAR marker {0223}.

Reaction to *Ustilago tritici*

Add: Resistance to race 19 was associated with chromosome 6A of Cadet, Kota, Thatcher, and TD18 {0208}. In the case of Cadet, resistance was localized to 6AS {0208}.

Resistance to colonization by *Eriophyes tulipae*

Curl mite colonization

Cmc1. **v:** Norstar derivative {0222}.

Cmc3 {0222}. 1A = T1AL·1RS. **v:** Amigo; TAM107. KS96GRC40 *Cmc4* {0222}.

Cmc4 {0222}. 6DS {0222}. **v:** KS96WRC40 *Cmc3* {0222}.

dv: *Aeg. Tauschii* (accession no {0222}).

Genetic Linkages

To the references in the first paragraph in the 2001 Supplement, add: '187'.

Chromosome 4D

4DS

ms4 – centromere I {0293}.

Summary Table 1

Add:

<i>Ar</i>	Alkylresocinols content of grain
<i>Eet</i>	Ear emergence time
<i>El</i>	Ear length
<i>Flt</i>	Flowering time
<i>Gnu</i>	Grain number
<i>Gpp</i>	Green plant percentage
<i>Gwe</i>	Grain weight/ear
<i>Ler</i>	Leaf erectness
<i>Pdc</i>	Pyruvate decarboxylase
<i>Pdl</i>	Peduncle length
<i>Raw</i>	Red awn colour
<i>Tgw</i>	1000-grain weight
<i>Win</i>	Winter hardiness

References

Amendments.

1350. Singh D, Park RF, Bariana HS & McIntosh 2001 Chromosome location and linkage studies of leaf rust resistance gene *Lr17b* in wheat cultivar Harrier. *Plant Breeding* 120: 7-12.
0107. Jahier J, AbElard P, Tonguy AM, Dédryver F, Rivoal R, Khatkar R & Bariana HS 2001 The *Aegilops ventricosa* segment on chromosome 2AS of the wheat cultivar 'VPM1' carries the cereal cyst nematode gene *Cre5*. *Plant Breeding* 120: 125-128.
0117. Sharifloo MR, Hassani ME & Sharp PJ 2001 A PCR-based DNA marker for detection of mutant and normal alleles of the *Wx-D1* gene of wheat. *Plant Breeding* 120: 121-124.
0138. Ogbonnaya FC, Seah S, Delibes A, Jahier J, López-Braña I, Eastwood RF & Lagudah ES 2001 Molecular-genetic characterization of a new nematode resistance gene in wheat. *Theoretical & Applied Genetics* 102: 623-629.
0163. 2002. *Euphytica* 123: 21-29.
0175. Anderson JA, Stack RW, Liu S, Waldron BL, Fjeld AD, Coyne C, Moreno-Sevilla B, Mitchell Fetch J, Song QJ, Cregan PB & Frohberg RC 2001 DNA markers for Fusarium head blight resistance QTLs in two wheat populations. *Theoretical & Applied Genetics* 102: 1164-1168.
0186. Arraino LS, Worland, Ellerbrook C & Brown JKM 2001 Chromosomal location of a gene for resistance to septoria tritici blotch (*Mycosphaerella graminicola*) in a hexaploid wheat 'Synthetic 6X'. *Theoretical & Applied Genetics* 103: 758-764.
0188. McIntosh RA, Devos KM, Dubcovsky J & Rogers J 2001 Catalogue of gene symbols for wheat: 2001 Supplement. *Annual Wheat Newsletter* 47: 333-354.
0197. Liu ZY, Sun QX, Ni ZF, Nevo E & Yang TM 2002 Molecular characterization of a novel powdery mildew resistance gene *Pm30* in wheat originating from wild emmer. *Euphytica* 123: 21-29.

New.

0201. Ayala L, van Ginkel M, Khairallah M, Keller B & Henry M 2001 Expression of *Thinopyrum intermedium*-derived barley yellow dwarf virus resistance in elite bread wheat backgrounds. *Phytopathology* 91: 55-62.
0202. Koöner J & Pánková K 1999 Impact of homoeologous group 5 chromosomes with different *vrn* loci on leaf size and tillering. *Czech Journal of Genetics & Plant Breeding* 35: 65-72.
0203. Morris CF, King GE, Allan RE & Simeone MC 2001 Identification and characterization of near-isogenic hard and soft hexaploid wheats. *Crop Science* 41: 211-217.
0204. Morris CF, Lillemo M, Simeone MC, Giroux MJ, Babb SL & Kidwell KK 2001 Prevalence of puroindoline grain hardness genotypes among historically significant North American spring and winter wheats. *Crop Science* 218-228.
0205. Lillemo M & Morris CF 2000 A leucine to proline mutation in puroindoline b is frequently present in hard wheats from Northern Europe. *Theoretical & Applied Genetics* 100: 1100-1107.
0206. Martin JM, Frohberg RC, Morris CF, Talbert LE & Giroux MJ 2001 Milling and bread baking traits associated with puroindoline sequence type in hard red spring wheat. *Crop Science* 41: 228-234.
0207. Krishnamurthy K & Giroux MJ 2001 Expression of wheat puroindolime genes in transgenic rice enhances grain softness. *Nature Biotechnology* 19: 162-166.
0208. Knox RE & Howes NK 1994 A monoclonal antibody chromosome marker analysis used to locate a loose smut resistance gene in wheat chromosome 6A. *Theoretical & Applied Genetics* 89: 787-793.
0209. Quick JS, Ellis GE, Normann RM, Stramberger JA, Shanahan JF, Peairs FB, Rudolph JB & Lorenz K 1996 Registration of 'Halt' wheat. *Crop Science* 36: 210.
0210. Toit F du 1989 Inheritance of resistance in two *Triticum aestivum* lines to Russian wheat aphid (Homoptera: Aphidae). *Journal of Economic Entomology* 82: 1251-1253.
0211. Liu XM, Smith CM, Gill BS & Tolmay V 2001 Microsatellite markers linked to six Russian wheat aphid resistance genes in wheat. *Theoretical & Applied Genetics* 102: 504-510.
0212. Cao W, Hughes GR, Ma H & Dong Z 2001 Identification of molecular markers for resistance to *Septoria nodorum* blotch in durum wheat. *Theoretical & Applied Genetics* 102: 551-554.
0213. Seah S, Bariana H, Jahier J, Sivasithamparam K & Lagudah ES 2001 The introgressed segment carrying rust resistance genes *Yr17*, *Lr37* and *Sr38* in wheat can be assayed by a cloned disease resistance gene-like sequence. *Theoretical & Applied Genetics* 102: 600-605.

0214. Gill KS & Gill BS 1996 A PCR-based screening assay of *Ph1*, the chromosome pairing regulator gene of wheat. *Crop Science* 36: 719-722.
0215. Dudnikov AJ, Gorel FL & Berdnikov VA 2001 Chromosomal location of histone *H1* genes in common wheat. *Cereal Research Communications*. In press.
0216. Nasuda S, Liu Y, Sakamoto A, Nakayama T, Iwabuchi M & Tsunewaki K 1993 Chromosomal locations of the genes for histones and a histone-binding protein family HBP-1 in common wheat. *Plant Molecular Biology* 22: 603-614.
0217. Segal G, Liu B, Vega JM, Abbo S, Rodova M & Feldman M 1997 Identification of a chromosome-specific probe that maps within the *Ph1* deletions in common and durum wheat. *Theoretical & Applied Genetics* 94: 968-970.
0218. McKenzie Lamb Aung Wise Barker & Orfert 2002 Inheritance of resistance to wheat midge, *Sitodiplosis mosellana*, in spring wheat. Manuscript.
0219. Roberts MA, Reader SM, Dalgliesh C, Miller TE, Foote TN, Fish LJ, Snape TW & Moore G 1999 Induction and characterization of *ph1* wheat mutants. *Genetics* 153: 1909-1918.
0220. Williams K 2001 Personal communication.
0221. Brown-Guedira G 2001 Personal communication.
0222. Brown-Guedira G 2001 Personal communication.
0224. Huguet-Robert V, Dedryver F, Röder MS, Korzun V, Abéard P, Tanguy AM, Jaudeau B & Jahier J 2001 Isolation of a chromosomally engineered durum wheat line carrying the *Aegilops ventricosa* *Pch1* gene for resistance to eyespot. *Genome* 44: 345-349.
0225. Ayala L, Henry M, González-de-León D, Van Ginkel M, Mujeeb-Kazi A, Keller B & Khairallah M 2001 A diagnostic molecular marker allowing the study of *Th. intermedium*- derived resistance to BYDV in bread wheat segregating populations. *Theoretical & Applied Genetics* 102: 942-949.
0226. Kato K, Nakamura W, Tabiki T & Miura H 2001 Detection of loci controlling seed dormancy on group 4 chromosomes of wheat and comparative mapping with rice and barley genomes. *Theoretical & Applied Genetics* 291: 980-985.
0227. Aghaee-Sarbarzeh M, Harjit-Singh & Dhaliwal HS 2001 A microsatellite marker linked to leaf rust resistance transferred from *Aegilops triuncalis* into hexaploid wheat. *Plant Breeding* 120: 259-261.
0228. Kolmer JA 2001 Physiologic specialization of *Puccinia tritica* in Canada in 1998. *Plant Disease* 85: 155-158.
0229. Park RF, Goyeau H, Felsenstein FG, Bartoö P & Zeller FJ 2001 Regional phenotypic diversity of *Puccinia triticina* and wheat host resistance in western Europe, 1995. *Euphytica* 122: 113-127.
0230. Yang TZ, Zhang XK, Liu HW & Wang ZH 1998 Chromosomal arm location of a dominant dwarfing gene *Rht21* in XN004 of common wheat. In: Proceedings of the 8th International Wheat Genetics Symposium, Beijing, 1993 (Li ZS & Xin Zy eds): 839-842.
0231. Börner A & Worland AJ 2001 Does the Chinese dwarf wheat variety 'XN004' carry *Rht21*? *Cereal Research Communications* (In press).
0232. Marais GF, Marais AS & Groenewald JZ 2000 Evaluation and reduction of *Lr19-149*, a recombinant form of the *Lr19* translocation of wheat. *Euphytica* 121: 289-295.
0233. Seo YW, Jang CS & Johnson JW 2001 Development of AFLP and STS markers for identifying wheat-rye translocations possessing 2RL. *Euphytica* 121: 279-287.
0234. Yanagisawa T, Kiribuchi-Otobe C & Yoshida H 2001 An alanine to threonine change in the *Wx-D1* protein reduces GBSS I activity in a waxy wheat mutant. *Euphytica* 121: 209-214.
0235. Csecz M, Bartos P & Mesterházy Á 2001 Identification of stem rust resistance gene *Sr36* in the wheat cultivar GK KincQ and its derivatives. *Cereal Research Communications* 29: 267-273.
0236. Ammiraju JSS, Dholakia BB, Santra DK, Singh H, Lagu MD, Tamhankar SA, Dhaliwal HS, Rao VS, Gupta VS & Ranjekar PK 2001 Identification of inter simple sequence repeat (ISSR) markers associated with seed size in wheat. *Theoretical & Applied Genetics* 102: 726-732.
0237. Ammiraju JSS, Dholakia BB, Jawdekar G, Santra DK, Gupta VS, Röder MS, Singh H, Lagu MD, Dhaliwal HS, Rao VS, & Ranjekar PK 2001 Inheritance and identification of DNA markers associated with yellow berry tolerance in wheat (*Triticum aestivum* L.). *Euphytica*. In press.
0238. Harker N, Rampling LR, Shariflou MR, Hayden MJ, Holton TA, Morell MK, Sharp PJ, Henry RJ, & Edwards KJ 2001 Microsatellites as markers for Australian wheat improvement. *Australian Journal of Agricultural Research* 52: 1121-1130.
0239. Cregan P 2002 Personal Communication.

0240. Buerstmayr H, Lemmens M, Hartl L, Doldi L, Steiner B, Stierschneider M & Ruckenbauer P 2001 Molecular mapping of QTLs for Fusarium head blight resistance in spring wheat. I. Resistance to fungal spread (Type II resistance). *Theoretical & Applied Genetics*. In press.
0241. Campbell KG, Finney PL, Bergman CJ, Gualberto DG, Anderson JA, Giroux MJ, Siritunga D, Zhu JQ, Gendre F, Roue C, Verel A & Sorrells ME 2001 Quantitative trait loci associated with milling and baking quality in a soft x hard wheat cross. *Crop Science* 41: 1275-1285.
0242. Chalmers KJ, Campbell AW, Kretschmer J, Karakousis A, Henschke PH, Pierens S, Harker N, Pallotta M, Cornish GB, Sharifloo MR, Rampling LR, McLauchlan A, Daggard G, Sharp PJ, Holton TA, Sutherland MW, Appels R & Langridge P 2001 Construction of three linkage maps in bread wheat, *Triticum aestivum*. *Australian Journal of Agricultural Research* 52: 1089-1119.
0243. Chebotar SV, Korzun VN & Sivolap YM 2001 Allele distribution at locus WMS261 marking the dwarfing gene Rht8 in common wheat cultivars of southern Ukraine. *Russian Journal of Genetics* 37: 894-898.
0244. Chee PW, Elias EM, Anderson JA & Kianian SF 2001 Evaluation of a high grain protein QTL from *Triticum turgidum* L. var. *dicoccoides* in an adapted durum wheat background. *Crop Science* 41: 295-301.
0245. Cloutier S, Rampitsch C, Penner GA & Lukow OM 2001 Cloning and expression of a LMW-i glutenin gene. *Journal of Cereal Science* 33: 143-154.
0246. Galiba G, Kerepesi I, Vagujfalvi A, Kocsy G, Cattivelli L, Dubcovsky J, Snape JW & Sutka J 2001 Mapping of genes involved in glutathione, carbohydrate and COR14b cold induced protein accumulation during cold hardening in wheat. *Euphytica* 119: 173-177.
0247. Gill KS & Sandhu D 2001 Candidate-gene cloning and targeted marker enrichment of wheat chromosomal regions using RNA fingerprinting - differential display. *Genome* 44: 633-639.
0248. Rodriguez Milla MA & Gustafson JP 2001 Genetic and physical characterization of chromosome 4DL in wheat. *Genome* 44: 883-892.
0249. Corona V, Gazza L, Boggini G & Pogna NE 2001 Variation in friabilin composition as determined by A-PAGE fractionation and PCR amplification, and its relationship to grain hardness in bread wheat. *Journal of Cereal Science* 34: 243-250.
0250. Khlestkina EK, Pestsova EG, Röder MS & Börner A 2001 Molecular mapping, phenotypic expression and geographical distribution of genes determining anthocyanin pigmentation of coleoptiles in wheat (*Triticum aestivum* L.). *Theoretical & Applied Genetics*. In press.
0251. Rousset M, Brabant P, Kota RS, Dubcovsky J & Dvorak J 2001 Use of recombinant substitution lines for gene mapping and QTL analysis of bread making quality in wheat. *Euphytica* 119: 81-87.
0252. Sandhu D, Champoux JA, Bondareva SN & Gill KS 2001 Identification and physical localization of useful genes and markers to a major gene-rich region on wheat group 1S chromosomes. *Genetics* 157: 1735-1747.
0253. Torp AM, Hansen AL & Andersen SB 2001 Chromosomal regions associated with green plant regeneration in wheat (*Triticum aestivum* L.) anther culture. *Euphytica* 119: 377-387.
0254. Wang H-J, Huang XQ, Röder MS & Börner A 2001 Genetic mapping of loci determining long glumes in the genus *Triticum*. *Euphytica*. In press.
0255. Börner A, Schumann E, Fürste A, Cöster H, Leithold B, Röder MS & Weber WE 2001 Mapping of quantitative trait loci determining agronomic important characters in hexaploid wheat (*Triticum aestivum* L.). *Theoretical and Applied Genetics*. In press.
0256. Delibes A 2002 Personal communication.
0257. Hsam SLK, Huang XQ & Zeller 2001 Chromosomal location of genes for resistance to powdery mildew in common wheat (*Triticum aestivum* L. em. Thell.). Alleles at the *Pm5* locus. *Theoretical & Applied Genetics* 102: 127-133.
0258. Huang XQ, Wang LX, Xu MX & Röder M 2002 Microsatellite mapping of the wheat powdery mildew resistance gene *Pm5e* in common wheat (*Triticum aestivum* L.). Personal communication.
0259. Huang XQ, Hsam SLK & Zeller 2000 Chromosomal location of two novel genes for resistance to powdery mildew in Chinese landraces (*Triticum aestivum* L. em. Thell.). *Journal of Genetics & Breeding* 54: 311-317.
0260. Singh D, Park RF & McIntosh RA 2001 Postulation of leaf (brown) rust resistance genes in 70 wheat cultivars grown in the United Kingdom. *Euphytica* 120: 205-215.
0261. Frick MM, Hucl R, Nykiforuk CL, Conner RL, Kuzik A & Laroche A 1998 Molecular characterisation of a wheat stripe rust resistance gene in Moro wheat. In: *Proceedings 9th International Wheat Genetics Symposium*, Saskatoon, Canada (Slinkard AE ed.) Vol 3 pp 181-182.
0262. Bariana HS, Brown GN, Ahmed NU, Khatkar S, Conner RL, Wellings CR, Haley S, Sharp PJ & Laroche A 2002 Characterisation of *Triticum vavilovii*-derived stripe rust resistance using genetic, cytogenetic and molecular analyses and its marker-assisted selection. *Theoretical & Applied Genetics* 104: 315-320.

0263. Ciaffi M, Paolacci AR, Dominici L, Tanzarella OA & Porceddu E 2001 Molecular characterization of gene sequences coding for protein disulphide isomerase (PDI) in durum wheat (*Triticum turgidum* ssp. *durum*). *Gene* 265: 147-156.
0264. Effertz RJ, Anderson JA & Franci LJ 2001 Restriction fragment length polymorphism mapping of resistance to two races of *Pyrenophora tritici repens* in adult and seedling wheat. *Phytopathology* 91: 572-578.
0265. Faris J, Sirikhachornkit A, Haselkorn R, Gill BS, Gornicki 2001 Chromosome mapping and phylogenetic analysis of the cytosolic acetyl-CoA carboxylase loci in wheat. *Molecular Biology & Evolution* 18: 1720-1733.
0266. Li WL, Faris JD, Muthukrishnan S, Liu DJ, Chen PD & Gill BS 2001 Isolation and characterization of novel cDNA clones of acidic chitinases and beta-1,3-glucanases from wheat spikes infected by *Fusarium graminearum*. *Theoretical & Applied Genetics* 102: 353-362.
0267. Baenziger PS, Shelton DR, Shipman MJ & Graybosch RA 2001 Breeding for end-use quality: Reflections on the Nebraska experience. *Euphytica* 119: 95-100.
0268. Kolmer JA & Liu JQ 2001 Simple inheritance of partial resistance to leaf rust in two wheat cultivars. *Plant Pathology* 50: 546-551.
0269. Nachit MM, Elouafi I, Pagnotta MA, El Saleh A, Iacono E, Labhilili M, Asbati A, Azrak M, Hazzam H, Benschoter D, Khairellah M, Ribaut JM, Tanzarella OA, Porceddu E & Sorrells ME 2001 Molecular linkage map for an intraspecific recombinant inbred population of durum wheat (*Triticum turgidum* L. var. *durum*). *Theoretical & Applied Genetics* 102: 177-186.
0270. Peng JH, Fahima T, Röder MS, Huang QY, Dahan A, Li YC, Grama A & Nevo E 2000 High-density molecular map of chromosome region harboring stripe-rust resistance genes *YrH52* and *Yr15* derived from wild emmer wheat, *Triticum dicoccoides*. *Genetica* 109: 199-210.
0271. Sasanuma T 2001 Characterization of the rbcS multigene family in wheat: subfamily classification, determination of chromosomal location and evolutionary analysis. *Molecular Genetics & Genomics* 265: 161-171.
0272. Chantret N, Mingeot D, Sourdille P, Bernard M, Jacquemin JM & Doussinault G 2001 A major QTL for powdery mildew resistance is stable over time and at two development stages in winter wheat. *Theoretical & Applied Genetics* 103: 962-971.
0273. Prins R, Groenewald JZ, Marais GF, Snape JW & Koebner RMD 2001 AFLP and STS tagging of *Lr19*, a gene conferring resistance to leaf rust in wheat. *Theoretical & Applied Genetics* 103: 618-624.
0274. Sutka J 2001 Genes for frost resistance in wheat. *Euphytica* 119: 167-172.
0275. Tsujimoto H, Yamada T, Hasegawa K, Usami N, Kojima T, Endo TR, Ogiwara Y & Sasakuma T 2001 Large-scale selection of lines with deletions in chromosome 1B in wheat and applications for fine deletion mapping. *Genome* 44: 501-508.
0276. Varshney RK, Prasad M, Roy JK, Röder MS, Balyan HS, Gupta PK 2001 Integrated physical maps of 2DL, 6BS and 7DL carrying loci for grain protein content and pre-harvest sprouting tolerance in bread wheat. *Cereal Research Communications* 29: 33-40.
0277. Vasu K, Harjit-Singh, Singh S, Chhuneja P & Dhaliwal HS 2001 Microsatellite marker linked to a leaf rust resistance gene from *Triticum monococcum* L. transferred to bread wheat. *Journal of Plant Biochemistry & Biotechnology* 10: 127-132.
0278. Yan L & Bhave M 2000 Sequences of the waxy loci of wheat: Utility in analysis of waxy proteins and developing molecular markers. *Biochemical Genetics* 38: 391-411.
0279. Yan LL & Bhave M 2001 Characterization of waxy proteins and waxy genes of *Triticum timopheevii* and *T. zhukovskyi* and implications for evolution of wheat. *Genome* 44: 582-588.
0280. Zanetti S, Winzeler M, Feuillet C, Keller B & Messmer M 2001 Genetic analysis of bread-making quality in wheat and spelt. *Plant Breeding* 120: 13-19.
0281. Snape JW 2002 Personal communication.
0282. Iwaki K, Nakagawa K & Kato K 2001 The possible candidate for *Vrn-B1* in wheat, as revealed by monosomic analysis of *Vrn* genes carried by Triple Dirk (B), the former *Vrn2*. *Wheat Information Service* 92: 9-11.
0283. Kolb FL, Bai GH, Muehlbuer GJ, Anderson JA, Smith KP & Fedak G 2001 Host plant resistance genes for Fusarium head blight: mapping and manipulation with molecular markers. *Crop Science* 41: 611-619.
0284. Liu SX, Griffey CA & Saghai-Marsoof MA 2001 Identification of molecular markers associated with adult plant resistance to powdery mildew in common wheat cultivar Massey. *Crop Science* 41: 1268-1275.
0285. Ma JX, Zhou RG, Dong YS, Wang LF, Wang XM & Jia JZ 2001 Molecular mapping and detection of the yellow rust resistance gene *Yr26* in wheat transferred from *Triticum turgidum* L. using microsatellite markers. *Euphytica* 120: 219-226.

0286. Mohle V, Hsam SLK, Zeller FJ & Wenzel G 2001 An STS marker distinguishing the rye-derived powdery mildew resistance alleles at the *Pm8/Pm17* locus of common wheat. *Plant Breeding* 120: 448-450.
0287. Boukhatem N, Baret PV, Mingeot D & Jacquemin JM 2002. Quantitative trait loci for resistance against yellow rust in two wheat-derived inbred wheat line populations. *Theoretical & Applied Genetics* 104: 111-115.
0288. Singh D, Park RF & McIntosh RA 2001 Inheritance of seedling and adult plant resistance of selected Australian spring and English winter wheat varieties. *Plant Breeding* 120: 503-507.
0289. Qi LL & Gill BS 2001 High-density physical maps reveal the dominant gene *Ms3* is located in a genomic region of low recombination in wheat and is not amenable to map-based cloning. *Theoretical & Applied Genetics* 103: 998-1006.
0290. Klindworth DL, Williams ND & Maan SS 2002 Chromosomal location of genetic male sterility genes in four mutants of hexaploid wheat. *Crop Science* (in press).
0291. Snape JW, Semikhodskii A, Fish L, Sarma RN, Quarrie SA, Galiba G & Sutka J 1997 Mapping frost tolerance loci in wheat and comparative mapping with other cereals. *Acta Agronomica Hungarica* 45: 268-270.
0292. Sutka J, Galiba G, Vagujfalvi A, Gill BS & Snape JW 1999 Physical mapping of the *Vrn-A1* and *Fr1* genes on chromosome 5A of wheat using deletion lines. *Theoretical & Applied Genetics* 99: 199-202.
0293. Maan SS & Kianian SF 2001 Third dominant male sterility gene in common wheat. *Wheat Information Service* 93: 27-31.
0294. Feuillet C, Penger A, Gellner K, Mast A & Keller B 2001 Molecular evolution of receptor-like kinase genes in hexaploid wheat. Independent evolution of orthologs after polyploidization and mechanisms of local rearrangements at paralogous loci. *Plant Physiology* 125: 1304-1313.
- 0295 Morris CF 2002 Puroindolines: the molecular genetic basis of wheat grain hardness. *Plant Molecular Biology* (in press)
0296. Feuillet C & Keller B 1999 High gene density is conserved at syntenic loci of small and large grass genomes. *Proceedings of the National Academy of Sciences U.S.A.* 96: 8265-8270.
0297. Feuillet C, Reuzeau C, Kjellbom P & Keller B 1998 Molecular characterization of a new type of receptor-like kinase (wlrk) gene family in wheat. *Plant Molecular Biology* 37: 943-953.
0298. Morris CF & Allan RE 2001 Registration of hard and soft near-isogenic lines of hexaploid wheat genetic stocks. *Crop Science* 41: 935-936.
0299. Huang L & Gill BS 2001 An RGA-like marker detects all known *Lr21* leaf rust resistance gene family members in *Aegilops tauschii* and wheat. *Theoretical & Applied Genetics* 103: 1007-1013.
0300. Raupp WJ, Sukhwinder-Singh, Brown-Guerdira & Gill BS 2001 Cytogenetic and molecular mapping of the leaf rust resistance gene *Lr39* in wheat. *Theoretical & Applied Genetics* 102: 347-352.
0301. Watkins JE, Schimelfenigk J & Baenziger PS 2001 Virulence of *Puccinia triticina* on wheat in Nebraska during 1997 and 1998. *Plant Disease* 85: 159-164.
0302. Singh RP, Huerta-Espino J, Rajaram S & Crossa J 2001 Grain yield and other traits of tall and dwarf isolines of modern bread and durum wheats. *Euphytica* 119: 241-244.
0303. Worland AJ, Sayers EJ & Korzun V 2001 Allelic variation at the dwarfing gene *Rht8* locus and its significance in international breeding programs. *Euphytica* 119: 155-159.
0304. Szunics L, Szunics Lu, Vida G, Bedö Z & Svec M 2001 Dynamics of changes in the races and virulences of wheat powdery mildew in Hungary between 1971 and 1999. *Euphytica* 119: 143-147.
0305. Robert O, Dedryver F, Leconte M, Rolland B & de Vallavieille-Pope C 2000 Combination of resistance tests and molecular tests to postulate the yellow rust resistance gene *Yr17* in bread wheat lines. *Plant Breeding* 119: 467-472.
0306. Juhász A, Tamás L, Karsai I, Vida G, Láng L & Bedö Z 2001 Identification, cloning and characterisation of a HMW-glutenin gene from an old Hungarian wheat variety, Bánkáti 1201. *Euphytica* 119: 75-79.
0307. Buonocore F, Hickman DR, Caporale C, Porceddu E, Lafiandra D, Tatham AS & Shewry PR 1996 Characterisation of a novel high M_r glutenin subunit encoded by chromosome 1D of bread wheat. *Journal of Cereal Science* 23: 55-60.
0308. Mackie AM, Lagudah ES, Sharp PJ & Lafiandra D 1996 Molecular and biochemical characterisation of HMW glutenin subunits from *Triticum tauschii* and the D genome of hexaploid wheat. *Journal of Cereal Science* 2: 213-225.
0309. Gianibelli MC, Lagudah ES, Wrigley CW & MacRitchie F 2002 Biochemical and genetic characterization of a monomeric storage protein (T1) with an unusually high molecular weight in *Triticum tauschii*. *Theoretical and Applied Genetics* 104: 497-504.

-
- 02110. Brites C & Carrillo JM 2000 Inheritance of gliadin and glutenin proteins in four durum wheat crosses. *Cereal Research Communications* 28: 239-246.
 - 02111. Sreeramulu G & Singh NK 1997 Genetic and biochemical characterization of novel low molecular weight glutenin subunits in wheat (*Triticum aestivum* L.). *Genome* 40: 41-48.
 - 02112. Gianibelli MC, Wrigley CW & MacRitchie F 2002 Polymorphism of low M_r glutenin subunits in *Triticum tauschii*. *Journal of Cereal Science* (in press; currently available online at <http://www.idealibrary.com>, ref. doi: 10.1006/jcrs.2001.0424).
 - 02113. Anderson OD, Hsia CC, Adalsteins AE, Lew EJ-L & Kasarda DD 2001 Identification of several new classes of low-molecular-weight wheat gliadin-related proteins and genes . *Theoretical and Applied Genetics* 103: 307-315.
 - 02114. Singh NK, Donovan GR & MacRitchie F 1990 Use of sonication and size-exclusion high performance liquid chromatography in the study of wheat flour proteins II. Relative quantity of glutenin as a measure of bread-making quality. *Cereal Chemistry* 67: 161-170.
 - 02115. Singh NK, Shepherd KW & Cornish GB 1991 A simplified SDS-PAGE procedure for separating LMW subunits of glutenin. *Journal of Cereal Science* 14: 203-208.
 - 02116. Sreeramulu G, Vishnuvardhan D & Singh NK 1994 Seed storage protein profiles of seven Indian wheat varieties (*Triticum aestivum* L.). *Journal of Plant Biochemistry and Biotechnology* 3: 47-51.

VI. ABBREVIATIONS USED IN THIS VOLUME.
PLANT DISEASES, PESTS, AND PATHOGENS:

BYDV = barley yellow dwarf virus
BMV = barley mosaic virus
CCN = cereal cyst nematode, *Heterodera avenae*
FHB = Fusarium head blight
RWA = Russian wheat aphid
SBMV = soilborne mosaic virus
SLB = Septoria leaf blotch
WSBMV = wheat soilborne mosaic virus
WSMV = wheat streak mosaic virus
WSSMV = wheat spindle streak mosaic virus
E. graminis* f.sp. *tritici = *Erysiphe graminis* f.sp. *tritici* = the powdery mildew fungus
F. graminearum = *Fusarium graminearum* = head scab fungus
F. nivale = *Fusarium nivale* = snow mold fungus
H. avenae = *Heterodera avenae* = cereal cyst nematode
P. recondita* f.sp. *tritici = *Puccinia recondita* f.sp. *tritici* = leaf rust fungus
P. striiformis* f.sp. *tritici = *Puccinia striiformis* f.sp. *tritici* = strip rust fungus
P. graminis = *Polymyxa graminis* = wheat soilborne mosaic virus vector
R. cerealis = *Rhizoctonia cerealis* = sharp eyespot
R. solani = *Rhizoctonia solani* = Rhizoctonia root rot
R. padi = *Rhonopalosiphum padi* = bird cherry-oat aphid
S. tritici = *Septoria tritici* = Septoria leaf spot fungus
S. graminearum = *Schizaphus graminearum* = greenbug
St. nodorum = *Stagonospora nodorum* = Stagonospora glume blotch
T. indica = *Tilletia indica* = Karnal bunt fungus

SCIENTIFIC NAMES AND SYNONYMS OF GRASS SPECIES (NOTE: CLASSIFICATION ACCORDING TO VAN SLAGEREN, 1994):

A. strigosa = *Avena strigosa*
Ae. cylindrica = *Aegilops cylindrica* = *Triticum cylindricum*
Ae. geniculata = *Aegilops geniculata* = *Aegilops ovata* = *Triticum ovatum*
Ae. speltoides = *Aegilops speltoides* = *Triticum speltoides*
Ae. tauschii = *Aegilops tauschii* = *Aegilops squarrosa* = *Triticum tauschii*
Ae. triuncialis = *Aegilops triuncialis* = *Triticum triunciale*
Ae. umbellulata = *Aegilops umbellulata* = *Triticum umbellulatum*
Ae. peregrina = *Aegilops peregrina* = *Aegilops variabilis* = *Triticum peregrinum*
Ae. ventricosa = *Aegilops ventricosa* = *Triticum ventricosum*
S. cereale = *Secale cereale* = rye
T. aestivum = *Triticum aestivum* = hexaploid, bread, or common wheat
T. monococcum* subsp. *aegilopoides = *Triticum boeoticum*
T. dicoccon = *Triticum dicoccon* = *T. dicoccum*
T. durum = *Triticum durum* = durum, pasta, or macaroni wheat
T. macha = *Triticum macha*
T. militinae = *Triticum militinae*
T. spelta = *Triticum spelta*
T. timopheevii* subsp. *timopheevii = *Triticum timopheevii*
T. timopheevii* subsp. *armeniacum = *Triticum araraticum* = *T. araraticum*
T. turgidum* subsp. *dicoccoides = *Triticum dicoccoides* = wild emmer wheat
T. turgidum* subsp. *dicoccum = *Triticum dicoccum*
T. urartu = *Triticum urartu*
Th. bessarabicum = *Thinopyrum bessarabicum*

SCIENTIFIC JOURNALS AND PUBLICATIONS:

Agron Abstr = Agronomy Abstracts
Ann Wheat Newslet = Annual Wheat Newsletter
Cereal Res Commun = Cereal Research Communications
Curr Biol = Current Biology
Eur J Plant Path = European Journal of Plant Pathology
Int J Plant Sci = International Journal of Plant Science
J Cereal Sci = Journal of Cereal Science
J Hered = Journal of Heredity
J Phytopath = Journal of Phytopathology
J Plant Phys = Journal of Plant Physiology
Mol Gen Genet = Molecular and General Genetics
PAG = Plant and Animal Genome (abstracts from meetings)
Plant Breed = Plant Breeding
Plant, Cell and Envir = Plant, Cell and Environment
Plant Cell Rep = Plant Cell Reporter
Plant Physiol = Plant Physiology
Sci Agric Sinica = Scientia Agricultura Sinica
Theor Appl Genet = Theoretical and Applied Genetics
Wheat Inf Serv = Wheat Information Service

UNITS OF MEASUREMENT:

bp = base pairs
bu = bushels
cM = centimorgan
ha = hectares
T = tons
m³ = cubic meters
μ = micron
me = milli-equivalents
mmt = million metric tons
mt = metric tons
Q = quintals

MISCELLANEOUS TERMS:

Al = aluminum
AFLP = amplified fragment length polymorphism
ANOVA = analysis of variance
A-PAGE = acid polyacrylamide gel electrophoresis
AUDPC = area under the disease progress curve
BW = bread wheat
CHA = chemical hybridizing agent
CMS = cytoplasmic male sterile
CPS = Canadian Prairie spring wheat
DH = doubled haploid
DON = deoxynivalenol
ELISA = enzyme-linked immunosorbent assay
EMS = ethyl methanesulfonate
EST = expressed sequence tag
FAWWON = Facultative and Winter Wheat Observation Nursery
GA = gibberellic acid
GM = genetically modified
HPLC = high pressure liquid chromatography

HMW = high-molecular weight (glutenins)

HRSW = hard red spring wheat

HRRW = hard red winter wheat

HWSW = hard white spring wheat

HWWW = hard white winter wheat

kD = kilodalton

LMW = low molecular weight (glutenins)

MAS = marker-assisted selection

NILs = near-isogenic lines

NIR = near infrared

NSW = New South Wales, region of Australia

PAGE = polyacrylamide gel electrophoresis

PCR = polymerase chain reaction

PFGE = pulsed-field gel electrophoresis

PMCs = pollen mother cells

PNW = Pacific Northwest (a region of North America including the states of Oregon and Washington in the U.S. and the province of Vancouver in Canada)

PPO = polyphenol oxidase

QTL = quantitative trait loci

RAPD = random amplified polymorphic DNA

RFLP = restriction fragment length polymorphism

RILs = recombinant inbred lines

SAMPL = selective amplification of microsatellite polymorphic loci

SAUDPC = standardized area under the disease progress curve

SCAR = sequence-characterized amplified region

SDS-PAGE = sodium dodecyl sulphate polyacrylamide gel electrophoresis

SH = synthetic hexaploid

SNP = single nucleotide polymorphism

SRPN = Southern Regional Performance Nursery

SRWW = soft red winter wheat

SRSW = soft red spring wheat

STMA = sequence tagges microsatellite site

SWWW = soft white winter wheat

SSD = single-seed descent

SSR = single-sequence repeat

STS = sequence-tagged site

TKW = 1,000-kernel weight

UESRWWN = Uniform Experimental Soft Red Winter Wheat Nursery

VII. ADDRESSES OF CONTRIBUTORS.

The E-mail addresses of contributors denoted with a '*' are included in section IX.

GRAINGENES.

David Matthews: matthews@greenengenes.cit.cornell.edu. Department of Biometry and Plant Breeding, Cornell University, Ithaca, NY 14853 USA, (607) 255-9951.

Olin Anderson: oandersn@pw.usda.gov. USDA-ARS, WRRC, 800 Buchanan Street, Albany, CA 94710 USA, (510) 559-5773.

Gerard Lazo: lazo@pw.usda.gov. USDA-ARS, WRRC, 800 Buchanan Street, Albany, CA 94710 USA, (510) 559-5777.

AGRIPRO WHEAT 806 N. Second Street, P.O. Box 30, Berthoud, CO 80513, USA. Rollie G. Sears*, D. Worrall, R. Bruns*.

Southern Plains Hard Winter Wheat, AgriPro Wheat, 12167 Hwy 70, P.O. Box 1739, Vernon, TX 76385, USA. 940-552-8881 (Tel), 940-552-0061 (FAX). David Worrall, David Graf, and Bradley Burkett.

Central Plains Hard Winter Wheat, AgriPro Wheat, 6515 Ascher Road, Junction City, KS 66441, USA. 785-210-0218 (Tel), 785-210-0231 (FAX). R.G. Sears*, Charles Johnson, John Robbins, Jon Rich, and Harold Erichsen.

Northern Plains Hard Red Spring Wheat, AgriPro Wheat, 806 N. Second Street, P.O. Box 30, Berthoud, CO 80513, USA. 970-532-3721 (Tel), 970-532-2035 (FAX). R. Bruns*, Joe A. Smith, Dennis Tweed, Scott Seifert, Linda Sizemore, and Bill Schabinger.

Canadian Hard Red Spring Wheat, Proven Research Farm, Morden, Manitoba, and Saskatoon, Saskatchewan. Kevin McCallum and Jim Dyck.

Northern Soft Red Winter Wheat, AgriPro Wheat, P.O. Box 411, 520 East 1050, South Brookston, IN 47923, USA. 765-563-311 (Tel), 765-563-6848 (FAX). Curtis Beazer*, Don Eckoff, Dayna Scruggs, and Eugene Glover.

Southeastern Soft Red Winter Wheat, AgriPro Wheat, P.O. Box 2365, Jonesboro, AR 72402, USA. 870-935-3941 (Tel), 870-935-3941 (*51) (FAX). Barton Fogelman, Gary Moore, Michael Montgomery, and Christopher DeArmond.

Pacific Northwest, AgriPro Wheat, 806 N. Second Street, P.O. Box 30, Berthoud, CO 80513, USA. 970-532-3721 (Tel), 970-532-2035 (FAX). John Moffatt*, Bob Knudson, and Jim Hemerick.

OR SEED BREEDING COMPANY Rua João Battisti, 71 – Passo Fundo, RS-CEP, 99050-380, Brazil. O.S. Rosa* and O. Rosa-Filho. 55-54-311-7499 (Tel/FAX).

STOLLER ARGENTINA S.A. Av. Malagueño s/n-Complejo Industrial U. CO. MA. Ferreyra, C.P. X5020CST, Córdoba, Argentina. W. Londero, L.E. Torres*, and Ricardo H. Maich*.

ARGENTINA

CÓRDOBA NATIONAL UNIVERSITY College of Agriculture, P.O. Box 509, Casilla de Correo 509, 5000 Córdoba, Argentina. (051) 334116/7 (Tel), (051) 334118 (FAX). María E. Dubois*, M.M. Cerana*, S.P. Gil, and R.H. Maich*.

INSTITUTO DE GÉNETICA ‘EWALD A FAVRET’ – CRN INTA Castelar, C.C. 25 (1712) Pcia. de Buenos Aires, Argentina. 54-11-4450-1876 (Tel), 54-11-4550-0805 (FAX). Cristina A. Kamlofski, Ruben Marrero, and Antonio D. Paleo*.

INSTITUTO DE RECURSOS BIOLÓGICOS – INTA Castelar, Las Cabañas y Los Reseros s/n, (1712) Villa Udaondo, Pcia. de Buenos Aires, Argentina. 54-1-621-5663/0125/1684 (Tel), 54-11-4621-6903 (FAX). M.M. Manifesto*, E.Y. Suárez, Gabriela Tranquilli*, and Alberto Acevedo*.

UNIVERSIDAD NACIONAL DE QUILMES Departamento de Ciencia y Tecnología, Roque Sáenz Peña 180, B1876BXD Bernal, Buenos Aires, Argentina. 54-11-4365-7132 (FAX). Adrián E. Bossio and Cecilia Bender.

AUSTRALIA
NEW SOUTH WALES

THE UNIVERSITY OF ADELAIDE Waite Campus, Department of Plant Science, Glen Osmond, 5064, SA, Australia. Daryl J. Mares*, Anna Campbell, and Kolumnina Mrva.

VICTORIA

VICTORIAN INSTITUTE FOR DRYLAND AGRICULTURE Department of Natural Resources and Environment, Private Bag 260, Horsham, VIC 3401, Australia. F.C. Ogbonnaya*, F. Dreccer*, R.F. Eastwood*, P.R. Hearnden*, E. Martin*, J. Oman*, D. Rodríguez*. and J.S. Brown*.

AUSTRIA

INTERUNIVERSITY RESEARCH INSTITUTE FOR AGRICULTURAL BIOTECHNOLOGY Department of Plant Biotechnology, Konrad Lorenz Str. 20, A-3430, Tulln, Austria. 43-2272-66280-205/203 (Tel), 43-2272-66280-77/203 (FAX). Hermann Buerstmayr*, M. Lemmens, U. Scholz, P. Ruckenbauer, M. Steiner, M. Mardi, M. Stierschneider, and M. Griesser.

BRAZIL

NATIONAL RESEARCH CENTER FOR WHEAT – EMBRAPA Centro Nacional de Pesquisa de Trigo, Rodovia BR 285, km 174, Caixa Postal 569, 99001-970, Passo Fundo, Rio Grande do Sul, Brazil. (54) 312-3444 (Tel), (54) 311 3617 (FAX). Cantído N.A. de Sousa*, Leo de J.A. Del Duca*, Amarilis L. Barcellos, Eliana M. Guarienti, Leila M. Costamilan, Márcio Só e Silva, Pedro L. Scheeren, Osmar Rodrigues, Gilberto R. da Cunha, R.S. Fontaneli, J. Almeida, N. Antoniazzi, R. Molin, F. Franco, S.R. Dotto, Sandra P. Brammer*, Daniela S. Boscardin, and Caren R.C. Lamb.

CANADA**MANITOBA**

AGRICULTURE AND AGRI-FOOD CANADA Cereal Research Centre, 195 Dafoe Road, Winnipeg, Manitoba R3T 2M9, Canada. (204) 983-3031 (Tel), (204) 983-4604 (FAX). Steve Haber*, Ron DePauw, Julian Thomas, and John Noll*.

CROATIA

BC INSTITUTE FOR BREEDING AND PRODUCTION OF FIELD CROPS d.d. Zageb, Marulicev trg 5/I, 10 000 Zagreb, Croatia. 385-1-65-45-576 (Tel), 385-1-65-45-579 (FAX). <http://www.bc.institut.hr> S. Tomasovic*, R. Mlinar, S. Gasperov, I. Ikic, and K. Puskaric.

CZECH REPUBLIC

RESEARCH INSTITUTE OF CROP PRODUCTION Drnovská 507, 161 06 Prague 6 – Ruzyne, Czech Republic. 420-2-33022-364 (Tel); 420-2-33022-286 (FAX). Zdenek Stehno*, L. Cejka, I. Faberova, J. Chrpova, M. Skorpik, V. Síp, L. Bobková, S. Sykorová, and L. Papousková.

ESTONIA

ESTONIAN AGRICULTURAL UNIVERSITY Institute of Experimental Biology, Department of Plant Genetics, 76902, Harku, Estonia. T. Enno, H. Peusha, Oskar Priilinn*, and Maimu Tohver*.

JÖGEVA PLANT BREEDING INSTITUTE EE 48309 Jõgeva, Estonia. R. Koppel.

TALLINN TECHNICAL UNIVERSITY Ehitajate tee 5, EE 19086 Tallinn, Estonia. A. Kann, A. Mihhalevski, I. Rahnu, and R. Täht.

ETHIOPIA

INSTITUTE OF BIODIVERSITY CONSERVATION AND RESEARCH P.O. Box 30726, Addis Ababa, Ethiopia. Ysmane Tsehay*.

GERMANY

INSTITUT FÜR PFLANZENGENETIK UND KULTURPFLANZENFORSCHUNG (IPK) Corrensstraße 3, 06466 Gatersleben, Germany. (049) 39482 5229 (Tel), (049) 39482 280/5139 (FAX). Andreas Börner*, U. Freytag, U. Sperling, K.F.M. Salem, and E.K. Khlestkina.

HUNGARY

AGRICULTURAL RESEARCH INSTITUTE OF THE HUNGARIAN ACADEMY OF SCIENCES Brunszvik u. 2, Martonvásár, H-2462, Hungary. 36(22)569-500 (Tel), 36(22)460-213 (FAX). Z. Bedö*, L. Szunic, L. Láng*, O. Veisz, I. Karsai*, A. Juhász, M. Rakszegi, Gy. Vida, P. Szűcs, Cs. Kuti, M. Megyeri, M. Gál, J. Sutka, G. Galiba, M. Molnár-Láng*, G. Kocsy, G. Kóvacs, G. Linc, A. Vágújfalvi, E.D. Nagy, A.F. Bálint, B. Tóth, and I. Molnár.

INDIA

BHABHA ATOMIC RESEARCH CENTRE Nuclear Agriculture and Biotechnology and Molecular Biology and Agriculture Divisions, Mumbai-400085, India. B.K. Das, S.G. Bhagwat*, N. Eswaran, A. Saini, N. Jawali, J.K. Sainis, S.P. Shouche, and R. Rastogi.

BHARATHIAR UNIVERSITY Cytogenetics Laboratory, Department of Botany, Coimbatore-641 046, Tamil Nadu, India. 091-422222 Ext. 359 (Tel), 091-422-422387 (FAX). V. Rama Koti Reddy, K.M. Gothandam*, and G. Kalaiselvi.

CH. CHARAN SINGH UNIVERSITY Wheat Biotechnology Project, Department of Agricultural Botany, Meerut-250 004 (U.P.), India. 91-121-768195/770335 (Tel), 91-121-767018/760577/764070 (FAX). P.K. Gupta*, H.S. Balyan, S. Kumar, Manoj Prasad, Joy K. Roy, N. Kumar, S. Sharma, P.L. Kulwal, S. Rustgi, and R. Singh.

INDIAN AGRICULTURAL RESEARCH INSTITUTE REGIONAL STATION Wellington-643 231, The Nilgiris, Tamilnadu, India. M. Sivasamy, A.J. Prabakaran, K.A. Nayeem, R.N. Brama, and M.K. Menon.

INDIAN AGRICULTURAL RESEARCH INSTITUTE REGIONAL STATION Genetics Division, New Delhi–110012, India. (011)-5783077, 5781481 (Tel). S.S. Singh*, G.P. Singh*, J.B. Sharma, Nanak Chand, D.N. Sharma, J.B. Singh*, and S.M.S. Tomar.

INDIAN AGRICULTURAL RESEARCH INSTITUTE REGIONAL STATION National Research Centre on Plant Biotechnology, New Delhi–110012, India. T. Mohapatra and K.P. Singh.

INDIAN AGRICULTURAL RESEARCH INSTITUTE REGIONAL STATION Nuclear Research Laboratory, New Delhi–110012, India. P.C. Pandey and M.R.S. Kaim.

ITALY

EXPERIMENTAL INSTITUTE FOR CEREAL RESEARCH Via Cassia 176, 00191, Rome, Italy. 06/36307716 (Tel), 06/36308761 (FAX). V. Vallega*.

EXPERIMENTAL INSTITUTE FOR CEREAL RESEARCH Via Mulino 3, 26866 S. Angelo Lodigiano, Italy. G. Boggini*, M. Perenzin, M. Crobellini, M. Cattaneo, P. Vaccino, S. Empilli, N.E. Pogna*, L. Gazza, and A. Brandolini.

ISTITUTO DE PATHOLOGIA VEGETALE Dipartimento di Scienze e Technologie Agroalimentari (DiSTA), Via Filippo Re 8, 40126 Bologna, Italy. 051-2091436 (Tel). C. Rubies-Autonell* and C. Ratti*.

JAPAN

GIFU UNIVERSITY Faculty of Agriculture, 1-1 Yanagido, Gifu 501-11, Japan. Nobuyoshi Watanabe*.

TOHOKU NATIONAL AGRICULTURAL EXPERIMENT STATION Ministry of Agriculture, Forestry and Fisheries, Morioka, Iwate 020-0198, Japan. 81-19-643-3512 (Tel), 81-19-643-3510 (FAX). Hiro Nakamura*.

MEXICO

INTERNATIONAL MAIZE AND WHEAT IMPROVEMENT CENTER (CIMMYT INT.) Lisboa 27, Colonia Juárez, Apdo. Postal 6-641, 06600 México, D.F., México. (52-5) 726-9091 (Tel), (52-5) 726 75-58/9 (FAX). A. Mujeeb-Kazi*, R. Delgado, S. Cano, A.A. Vahidy, T. Razzaki, J.L. Diaz-de-León, A. Cortés, V. Rosas, F. Shafiq, Lucy Gilchrist, M. Zaharieva, and M. William.

PAKISTAN

REGIONAL AGRICULTURAL RESEARCH INSTITUTE Model Town-A, Bahawalpur, Punjab, Pakistan.

Muhammad Sarwar Cheema, Liaquat Ali, and Muhammad Akhtar.

ROMANIA

S.C.A.—AGRICULTURAL RESEARCH STATION Turda, 3350, str. Agriculturii 27 Jud Cluj, Romania. 00-40-64-311134 (Tel/FAX). Vasile Moldovan*, Maria Moldovan, and Rozalia Kadar.

RUSSIAN FEDERATION

AGRICULTURAL RESEARCH INSTITUTE FOR SOUTH-EAST REGIONS – ARISER Toulaikov Str., 7, Saratov, 410020, Russian Federation. 8452-64-76-88 (FAX). S.A. Voronina, N. Sibikeev, V.A. Krupnov*, A.E. Druzhin, A.Yu. Buyenkov, and T.I. Djatchouk.

INSTITUTE OF COMPLEX ANALYSIS OF REGIONAL PROBLEMS Karl Marx str., 105 A, kb. 167, Khabarovsk, 680009, Russian Federation. Ivan M. Shindin.

RUSSIAN RESEARCH INSTITUTE OF PHYTOPATHOLOGY Department of Mycology and Immunity of Agricultural Plants, 143050, Moscow region, B. Vyazemy, Russian Federation. E.D. Kovalenko*, A.I. Zhemchuzina, and N.N. Kryazheva.

RUSSIAN UNIVERSITY OF PEOPLES' FRIENDSHIP ul. Miklukho-Maklaya 6, Moskow, 117918, Russian Federation. A.K. Federov*.

SARATOV STATE AGRARIAN UNIVERSITY named after N.I. Vavilov, Department of Biotechnology, Plant Breeding and Genetics, 1 Teatralnaya Sg., Saratov 410600, Russia. Yu.V. Lobachev, M.V. Noritsina, and O.V. Tkachenko*.

SIBERIAN INSTITUTE OF PLANT PHYSIOLOGY AND BIOCHEMISTRY Lermontov str., 132, 664033, Irkutsk-33, P.O. Box 1243, Russian Federation. A.K. Glyanko*, G.G. Vasileva, A.V. Kolesnichenko*, E.L. Tauson, V.V. Zykova, E.S. Klimenko, O.I. Grabelnych, T.P. Pobezhimova, V.V. Tourchaninova, V.K. Voinikov, S.V. Osipova*, V.A. Trufanov, T.A. Pshenichnikova, S.V. Lankevich*, V.M. Sumtsova, R.K. Salyaev, L.V. Dudareva, S.V. Lankevich, V.M. Sumtsova, N.I. Rekoslavskaya*, O.V. Yurieva, B.A. Shainyan, T.V. Kopytina, R.K. Salyaev, V.A. Davydov, A.V. Permyakov*, S.L. Didenko, L.V. Pomazkina*, and L.G. Kotova.

VAVILOV INSTITUTE OF GENERAL GENETICS Gubkin str. 3, 117809 Moscow, Russian Federation. 7-095-3304022 (Tel), 7-095-3307301 (FAX). Sergei Martynov* and T.V. Dobrotvorskaya.

SOUTH AFRICA

SMALL GRAIN INSTITUTE Private Bag X29, Bethlehem 9700, Republic of South Africa. 27 58 3073444 (Tel), 27 58 3033952 (FAX). J.C. Aucamp, D.J. Exley, F. Middleton, P. Delpot, W.H.P. Boshoff, A.D. Barnard*, M.V. van Wyk, R. Prins, V.P. Ramburan, C.W. Miles, K.B. Majola, M.L.T. Moloi, M.M. Radebe, N.E.M. Mtjale, C.N. Matla, M.M. Mofokeng, M.L. Dhalmini, N.N. Mtshali, J.H. Hatting, L. Visser, W. Otto, V.L. Tolmay, Karen Naudé,

UNIVERSITY OF FREE STATE Plant Pathology Department, Republic of South Africa. Z.A. Pretorius.

UNIVERSITY OF STELLENBOSCH Department of Genetics, Private Bag X1, Matieland 7602, Republic of South Africa. 27-21-8085829 (Tel), 27-21-8085833 (FAX). G.F. Marais*, H.S. Roux, A.S. Marais, W.C. Botes, and J.H. Louw.

SPAIN

UNIVERSITY OF LLEIDA AND INSTITUTE FOR FOOD AND AGRICULTURAL RESEARCH AND TECHNOLOGY (UdL-IRTA) Center of R&D, Alcalde Rovira Roure 177, 25198 Lleida, Spain. 34-973-702569 (Tel), 34-973-238301 (FAX). J.A. Martín-Sánchez*, E. Sin, C. Martínez, and A. Michelena.

UNIVERSIDAD POLITÉCNICA DE MADRID Departamento de Biotecnología, E.T.S. Ingenieros Agrónomos, Ciudad Universitaria, 28040 Madrid, Spain. A. Delibes*, I. López-Braña, M.J. Montes, M. Gómez Colmenarejo, C. González-Belinchón.

CONSEJO SUPERIOR DE INVESTIGACIONES CIENTÍFICAS Serrano, 115, 28006 Madrid, Spain. D. Romero and M.F. Andrés.

JUNTA DE EXTREMADURA Servicio de Investigación Agraria, Finca La Orden, 06187 Guadajira, Badajoz, Spain. J. del Moral, F. Pérez Rojas, and F.J. Espinal.

THE UKRAINE

INSTITUTE OF PLANT PRODUCTION N.A. V. YA. YURJEV National Centre for Plant Genetic Resources of Ukraine, Yurjev Plant Production Institute, Moskovsky prospekt, 142, 310060 Kharkov, Ukraine. 00380 (0572) 920354 (Tel/FAX). S.V. Rabinovich*, T.Y. Markova, R.L. Boguslavsky, I.N. Chernyaeva, Yury G. Krasilovets*, Z.V. Usova, A.E. Litvinov, V.V. Sotnikov, N.V. Kouzmenko, V.A. Wlasenko, O.Ju. Leonov, I.A. Panchenko, S.Ju. Didenko, and R.G. Parkhomenko.

UNITED KINGDOM

JOHN INNES CENTRE Norwich Research Park, Colney, Norwich NR4 7UH, United Kingdom. 44-1603-450611 (Tel), 44-1603-450023/450045 (FAX). Robert M.D. Koebner*, James Brown, Lia Arraiano, Penny Brading, Clare Ellerbrook, Elaine Foster, Tony Worland, Lesley Boyd, Phil Smith, Peter Minchin, Clare Ellenbrook, Jacqueline Garrood, Elizabeth Chandler, Richard Draeger, Nick Gosman, Martha Thomsett, Duncan Simpson, Andy Steed, Lorenzo Covarelli, Paul Nicholson, Simon Griffiths, Steve Reader, Tracie Foote, Graham Moore, John Flintham, Manoel Bassoi, Rachael Adlam, and Mike Gale.

THE UNITED STATES

COLORADO

COLORADO STATE UNIVERSITY Department of Agronomy, Ft. Collins, CO 80523, USA. S. Haley*, J. Stromberger, B. Clifford, S. Clayschulte, T. Mulat, E. Ball, S. Slough, and A. Brown.

GEORGIA / FLORIDA

UNIVERSITY OF GEORGIA Department of Agronomy, Griffin, GA 30212-1197, USA. 770-228 7321 (Tel), 770-229-3215 (FAX). J.W. Johnson*, R.D. Barnett, B.M. Cunfer, and G.D. Buntin.

IDAHO

UNIVERSITY OF IDAHO Plant and Soil Science Department, Moscow, ID 83343, USA and the Agricultural Experiment Station, P.O. Box AA, Aberdeen, ID 83210, USA. R. Zemetra*, E. Souza, S. Guy, L. Robertson, B. Brown, N. Bosque-Pérez, J. Hansen, K. O'Brien, M. Guttieri, D. Schotzko, Y. Wu, T. Koehler, L. Sorensen, and J. Clayton.

USDA-ARS NATIONAL SMALL GRAINS GERMPLASM RESEARCH FACILITY P.O. Box 307, Aberdeen, ID 83210, USA. H.E. Bockelman*, D.M. Wesenberg, C.A. Erickson, and B.J. Goates.

INDIANA

PURDUE UNIVERSITY Departments of Agronomy, Entomology, and Botany and Plant Pathology, and USDA-ARS, West Lafayette, IN 47907, USA. 317-494-8072 (Tel), 317-496-2926 (FAX). J.M. Anderson*, H.W. Ohm*, F.L. Patterson, H.C. Sharma*, J. Uphaus, G. Buechley, S. Goodwin*, D.M. Huber*, G. Shaner*, X.R. Xu, R.H. Ratcliffe*, R. Shukle, C.E. Williams*, S. Cambron, C. Collier, and J.J. Stewart*.

KANSAS

KANSAS DEPARTMENT OF AGRICULTURE U.S. Department of Agriculture, 632 SW Van Buren, Rm. 200, P.O. Box 3534, Topeka, KS 66601-3534, USA. 913-233-2230 (Tel). <http://www.nass.usda.gov/ks/>. E.J. Thiessen, Sherri Hand, and Ron Sitzman.

KANSAS STATE UNIVERSITY

Environmental Physics Group Department of Agronomy, Kansas State University, Waters Hall, Manhattan, KS 66502, USA. 913-532-5731 (Tel), 913-532-6094 (FAX). M.S. Liphadzi and M.B. Kirkham*.

The Wheat Genetics Resource Center Departments of Plant Pathology and Agronomy and the USDA-ARS, Throckmorton Hall, Manhattan, KS 66506-5502, USA. 913-532-6176 (Tel), 913 532-5692 (FAX). W.J. Raupp*, B.S. Gill*, B. Friebel*, P. Zhang*, L.L. Qi*, S. Muthukrishnan, and W.L. Li*.

U.S. GRAIN MARKETING AND PRODUCTION RESEARCH CENTER — USDA, Agricultural Research Service, Manhattan, KS 66502, USA. O.K. Chung*, F.E. Dowell*, S.R. Bean*, G.L. Lookhart*, J.B. Ohm*, M. Tilley*, L.M. Seitz*, M.S. Ram*, D.B. Bechtel*, M.E. Casada*, S.H. Park*, J.D. Hubbard*, B.W. Seabourn*, M.S. Caley*, J.D. Wilson*, R.E. Dempster*, J.M. Downing*, J.E. Throne*, J.E. Baker*, and C.S. Chang.

MINNESOTA

CEREAL DISEASE LABORATORY, USDA-ARS University of Minnesota, 1551 Lindig, St. Paul, MN 55108, USA. 612-625-6299 (Tel), 612-649-5054 (FAX). <http://www.edl.umn.edu> D.L. Long, D.V. McVey*, J.A. Kolmer, M.E. Hughes*, and L.A. Wanschura.

NEBRASKA

UNIVERSITY OF NEBRASKA AND THE USDA-ARS Department of Agronomy and the USDA-ARS Wheat, Sorghum and Forage Unit, Keim Hall, Lincoln, NE 68583, USA. 402-472-1563 (Tel), 402-472-4020 (FAX). P.S. Baenziger*, R.A. Graybosch*, T. Clemente, S. Sato, M. Dickman, A. Mitra, S. Mitra, J. Watkins, J. Schimelfenig, H. Budak, T. Campbell, M. Erayman, Y. Mater, K.S. Gill, K. Eskridge, I. Dweikat, S. Dere, D. Baltensperger, and B. Beecher.

NORTH DAKOTA

USDA-ARS CEREAL CROPS RESEARCH UNIT Northern Crop Science Laboratory, Fargo, ND 58078-5051, USA. Justin D. Faris*, James Miller, Daryl Klindworth, Leonard Joppa, Phil Meyer, Mary Johnshoy, David Birdsall, Karri Haen, Kristin Simons, and Erik Doebler.

OKLAHOMA

OKLAHOMA STATE UNIVERSITY

Department of Entomology and Plant Pathology, 123 Noble Research Center, Stillwater, OK 74078, USA. Robert Hunger, Brian Olson, Larry L. Singleton, Jeanmarie Verchot, and Mark E. Payton (Department of Statistics).

Department of Plant and Soil Sciences, 368 Ag Hall, Stillwater, OK 74078-6028, USA. B.F. Carver*, G. Bai, A.K. Klatt, and Ray Sidwell.

SOUTH DAKOTA

SOUTH DAKOTA STATE UNIVERSITY AND THE USDA-ARS Plant Science Department and the Northern Grain Insect Research Laboratory (NGIRL), Brookings, SD 57007, USA. <http://triticum.sdsstate.edu> 605-688-4453 (Tel), 605-688-4452 (FAX). A.M.H. Ibrahim*, M.A.C. Langham, S.A. Kalsbeck, R.S. Little, F. Hakizimana, D. Gustafson, Yang Yen*, Denghui Xing, Lanfang Bai, Lieceng Zhu, Yue Jin*, Howard J. Woodard, Anthony Bly, Dwayne Winther, L. Hesler, W. Riedel, Cynthia I. Bergman, and Marie A.C. Langham.

VIRGINIA

VIRGINIA POLYTECHNIC INSTITUTE AND STATE UNIVERSITY Department of Crop and Soil Environmental Sciences and Department of Plant Pathology, Physiology and Weed Science, 419A Smyth Hall, Blacksburg, VA 24061-0404, USA. 540-231-7624 (Tel), 540-231-3431 (FAX). Wendy L. Rohrer*, Carl A. Griffey*, Daniel E. Brann, Linda Whitcher, Jianli Chen, M.A. Saghai Maroof, W. Zhao, J. Wilson, D. Nabati, Thomas H. Pridgen, and B. Robinson.

Matthew Chappell, E.L. Stromberg, R.M. Biyashev, and W. Xie.

WASHINGTON

USDA-ARS WHEAT GENETICS, QUALITY, PHYSIOLOGY AND DISEASE RESEARCH Department of Crop & Soil Sciences and Plant Pathology, Washington State University, P.O. Box 646420, 209 Johnson Hall, Pullman, WA 99164, USA. 509-335-3632 (Tel), 509-335-2553 (FAX). R.E. Allan*, X.M. Chen, D.A. Wood, M.K. Moore, Guiping Yan, and R.F. Line*.

WASHINGTON STATE UNIVERSITY Spring Wheat Breeding and Genetics Program, Department of Crop and Soil Sciences, 201 Johnson Hall, Pullman, WA 99164-6420, USA. 509-335-7247 (Tel). K.G. Kidwell*, G. Shelton, V. DeMacon, M. McClendon, J. Smith, J. Baley, R. Higgonbotham, and T. Paulitz.

YUGOSLAVIA

INSTITUTE OF AGRICULTURAL RESEARCH ‘SERBIA’ Center for Small Grains Kragujevac, S. Kovacevica 31, Kragujevac 34000, Yugoslavia. 381-34-65-808/33-663 (Tel), 381-36-343-333 (FAX). D. Knezevic*, V. Zecevic, N. Djukic, D. Micanovic, D. Urosevic, and B. Dimitrijevic.

VIII. E-MAIL DIRECTORY OF SMALL GRAINS WORKERS.

Acevedo, Alberto	aacevedo@unq.edu.ar, aacevedo@inta.gov.ar	INTA, Castelar, Argentina
Aldana, Fernando	fernando@pronet.net.gt	ICTA, Guatemala
Allan, Robert E	allanre@mail.wsu.edu	USDA-ARS, Pullman, WA
Altenbach, Susan	altnbach@pw.usda.gov	USDA-WRRE, Albany, CA
Altman, David	dwa1@cornell.edu	ISAAA-Cornell University, Ithaca, NY
Alvarez, Juan B	alvarez@unitus.it	University of Córdoba, Argentina
Anderson, Jim M	ander319@tc.umn.edu	University of Minnesota, St. Paul
Anderson, Joseph M	janderson@purdue.edu	Purdue University, W. Lafayette, IN
Anderson, Olin	oandersn@pw.usda.gov	USDA-WRRE, Albany, CA
Appels, Rudi	rudi@pican.pi.csiro.au	CSIRO-Division of Plant Industry
Armstrong, Ken	armstrongkc@em.agr.ca	AAFC-Ottawa, Ontario
Aung, T	taung@mbrswi.agr.ca	AAFC-Winnipeg
Babaoglu, Metin	metin_babaoglu@edirne.tagem.gov.tr	Thrace Ag Research Institute, Turkey
Bacon, Robert	rb27412@uafsysb.uark.edu	University of Arkansas, Fayetteville
Baenziger, P Stephen	agro104@unlnotes.unl.edu	University of Nebraska, Lincoln
Baker, Cheryl A	cbaker@pswcr.ars.usda.gov	USDA-ARS, Stillwater, OK
Baker, JE	baker@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Barnard, Anri D	anri@kgs1.agric.za	Small Grain Institute, South Africa
Barreto, D	dbarreto@cnia.inta.gov.ar	INTA, Buenos Aires, Argentina
Barker, Susan	sbarker@waite.adelaide.edu.au	Waite, University Adelaide
Bariana, Harbans	harbansb@camden.usyd.edu.au	PBI Cobbitty, Australia
Barkworth, Mary	uf7107@cc.usu.edu	USDA-ARS, Pullman, WA
Bartos, Pavel	bartos@hb.vrvu.cv	RICP, Prague, Czech Republic
Bhagwat, SG	sbhagwat@apsara.barc.ernet.in	Bhabha Atomic Res Cen, Mumbai, India
Buerstmayr, Hermann	buerst@ifa-tulln.ac.at	IFA-Tulln, Austria
Bean, Scott R	scott@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Beazer, Curtis	cbeazer@dcwi.com	AgriPro Seeds, Inc., Lafayette, IN
Bechtel DB	don@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Bedö, Zoltan	bedoz@buza.mgki.hu	Martonvásár, Hungary
Bentley, Stephen	bentleys@phibred.com	Pioneer Hi-Bred-Frouville, France
Bergstrom, Gary	gcb3@cornell.edu	Cornell University, Ithaca, NY
Berzonsky, William A	berzonsk@badlands.nodak.edu	North Dakota State University, Fargo
Bhagwat, SG	sbhagwat@apsara.barc.ernet.in	Bhabha Atomic Res Center, India
Bhatta, MR	rwp@nwrp.mos.com.np	Natl Wheat Research Program, Nepal
Blake, Nancy	nblake@montana.edu	Montana State University, Bozeman
Blake, Tom	isstb@montana.edu	Montana State University, Bozeman
Blanco, Antonia	blanco@afr.umiba.it	Institue of Plant Breeding, Bari, Italy
Blum, Abraham	vcablm@volcani.agri.gov.il	Volcani Center, Israel
Bockelman, Harold E	hbockelman@ars-grin.gov	USDA-ARS, Aberdeen, ID
Boggini, Gaetano	cerealicoltura@iscsal.it	Exp Inst Cereal Research, Italy
Boguslavskiy, Roman L	bogus@ncpgru.relcom.kharkov.ua	Kharkov Inst Plant Protection, Ukraine
Börner, Andreas	boerner@ipk-gatersleben.de	IPK, Gatersleben, Germany
Borovskii, Genadii	gena@sifibr.irk.ru	Siberian Inst Plant Physiology, Irkutsk
Bowden, Robert	rbowden@plantpath.ksu.edu	Kansas State University, Manhattan
Boyko, Elena	olena@plantpath.ksu.edu	Kansas State University, Manhattan
Brown, John S	john.brown@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Brammer, Sandra P	sandra@cnpt.embrapa.br	EMBRAPA, Passo Fundo, Brazil
Bradová, Jane	bradova@hb.vrv.cz	RICP, Prague, Czech Republic
Braun, Hans J	H.J.Braun@cgiar.org	CIMMYT-Turkey
Brennan, Paul	paulb@qdpit.sth.dpi.qld.gov.au	Queensland Wheat Research Institute
Brown, Douglas	dbrown@em.agr.ca	AAFC-Winnipeg, Manitoba
Brown, James	jbrown@bbsrc.ac.uk	JI Centre, Norwich, UK
Brown-Guedira, Gina	gbg@ksu.edu	USDA-ARS, Manhattan, KS
Bruckner, Phil	bruckner@montana.edu	Montana State University, Bozeman
Bruns, Rob	rbruns@frii.com	AgriPro Seeds, Inc., Berthoud, CO

Buerstmayr, Hermann	buerst@ifa-tulln.ac.at	IFA Tulln, Austria
Burd, John D	jdburd@pswcrl.ars.usda.gov	USDA-ARS, Stillwater, OK
Busch, Robert	Robert.H.Busch-1@umn.edu	USDA-ARS, St. Paul, MN
Caley, MS	margo@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Cambron, Sue	sue_cambron@entm.purdue.edu	Purdue University
Campbell, Kim	campbell.210@osu.edu	Ohio State University, Wooster
Carmona, M	mcarmona@sion.com.ar	University of Buenos Aires, Argentina
Carver, Brett F	bfc@soilwater.agr.okstate.edu	Oklahoma State University, Stillwater
Cerana, María M	macerana@agro.uncor.edu	Córdoba National University, Argentina
Casada, ME	casada@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Chapin, Jay	jchapin@clust1.clemson.edu	Clemson University
Chung, OK	okchung@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Cisar, Gordon L	glcisa@ccmail.monsanto.com	Hybritech-Lafayette, IN
Clark, Dale R	dclark@westbred.com	Western Plant Breeders, Bozeman, MT
Corke, Harold	harold@hkuxa.hku.hk	Hong Kong University
Czarnecki, E	eczarnecki@mbrswi.agr.ca	AAFC-Winnipeg, Manitoba
Davydov, VA	gluten@sifibr.irk.ru	Siberian Inst Plant Physiology, Russia
Das, BK	bkdas@magnum.barc.ernet.in	Bhaba Atomic Res Cen, Mumbai, India
Del Duca, Leo JA	delduca@cnpt.embrapa.br	EMBRAPA, Brazil
Delibes, A	adelibes@bit.etsia.upm.es	Universidad Politécnica de Madrid
del Moral, J.	moral@inia.es	Junta de Extremadura Servicio, Spain
Dempster, RE	rdempster@aibonline.org	Amer Inst Baking, Manhattan, KS
de Sousa, Cantido NA	cantidio@cnpt.embrapa.br	EMBRAPA, Brazil
DePauw, Ron	depauw@em.agr.ca	AAFC-Swift Current
Devos, Katrien	devos@bbsrc.ac.uk	JI Centre, Norwich UK
Dill-Macky, Ruth	ruthdm@puccini.crl.umn.edu	University Of Minnesota, St. Paul
Dotlacil, Ladislav	dotlacil@hb.vurv.cz	RICP, Prague, Czech Republic
Dorlencourt, Guy	dorlencourt@phibred.com	Pioneer Hi-bred-Frouville France
Dowell, FE	fdowell@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Downing, JM	jdowning@atrxlab.frii.com	
Dreccer, F	fernanda.dreccer@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Dubcovsky, Jorge	jdubcovsky@ucdavis.edu	University of California, Davis
Dubin, Jesse	JDubin@cimmyt.mx	CIMMYT, Mexico
Dubois, María E	mdubois@agro.uncor.edu	Córdoba National University, Argentina
Dundas, Ian	idundas@waite.adelaide.edu.au	University of Adelaide, Australia
Dunphy, Dennis	dennis.j.dunphy@monsanto.com	Monsanto Corp., Lafayette, IN
Dvorak, Jan	jdvorak@ucdavis.edu	University of California, Davis
Eastwood, Russell	russell.eastwood@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Edge, Benjamin	edgeben@phibred.com	Pioneer Hi-Bred-St. Matthews, SC
Edwards, Ian	iane@biotest.com.au	Grain Biotech Australia, Joondalup
Egorov, Tsezi	egorov@imb.ac.ru	Englehardt Institute, Moscow, Russia
Elias, Elias	elias@prairie.nodak.edu	North Dakota State University, Fargo
Elliott, Norman C	nelliott@ag.gov	USDA-ARS, Stillwater, OK
Endo, Takashi R	endo@kais.kyoto-u.ac.jp	Kyoto University, Japan
Faberova, Iva	faberova@genbank.vurv.cz	RICP, Prague, Czech Republic
Fahima, Tzion	rabi310@haifaumv.bitnet	University of Haifa, Israel
Faris, Justin D	Justin_Faris@ndsu.nodak.edu	UDSA-ARS, Fargo, ND
Fedak, George	fedakga@em.agr.ca	AAFC-Ottawa, Ontario
Fedorov, A.K.	meraserv@mega.ru	Russian Univ People Friend, Moscow
Feldman, Moshe	lpfeld@weizmann.weizmann.ac.il	Weizmann Institute, Rehovot, Israel
Fellers, John P	jpf@alfalfa.ksu.edu	USDA-ARS, Manhattan, KS
Fox, Paul	pfox@alphac.cimmyt.mx	CIMMYT-Mexico
Fogelman Jr, J Barton	jbarton@ipa.net	AgriPro Seeds, Inc., Jonesboro, AK
Frank, Robert W	frankr@idea.ag.uiuc.edu	University of Illinois, Urbana
Fritz, Alan K	akf@ksu.edu	Kansas State University, Manhattan
Friebe, Bernd	friebe@ksu.edu	Kansas State University, Manhattan
Gale, Mike	gale@bbsrc.ac.uk	JI Centre, Norwich, UK

Giese, Henriette	h.giese@risoe.dk	Risoe National Lab, DK
Gilbert, Jeannie	jgilbert@mbrswi.agr.ca	Agriculture Canada–Winnipeg
Gill, Bikram	bsg@ksu.edu	Kansas State University, Manhattan
Giroux, Mike	mgiroux@montana.edu	Montana State University, Bozeman
Gitt, Michael	mgitt@pw.usda.gov	USDA–ARS–WRRC, Albany, CA
Glyanko, AK	usttaft@sifibr.irk.ru	Siberian Inst PI Physio Biochem, Russia
Gonzalez-de-Leon, Diego	dgdeleon@alphac.cimmyt.mx	CIMMYT–Mexico
Gooding, Rob	rgooding@magnus.acs.ohio-state.edu	Ohio State University, Wooster
Goodwin, Steve	goodwin@btny.purdue.edu	Purdue University, W. Lafayette, IN
Gothandam, KM	gothandam@yahoo.com	Bharathiar University, Coimbatore, India
Grausgruber, Heinrich	grausgruber@ipp.boku.ac.at	Univ of Agriculture Sciences, Vienna
Graham, W Doyce	dgraham@clust1.clemson.edu	Clemson University
Graybosch, Bob	rag@unlserve.unl.edu	USDA–ARS, Lincoln, NE
Greenstone Matthew H	mgreenstone@pswcr.ars.usda.gov	USDA–ARS, Stillwater, OK
Grienenger, Jean M	grienenger@medoc.u-strasbg.fr	University of Strasberg, France
Griffey, Carl	cgriffey@vt.edu	Virginia Tech, Blacksburg
Griffin, Bill	griffinw@lincoln.cri.nz	DSIR, New Zealand
Groeger, Sabine	probstdorfer.saatzucht@netway.at	Probstdorfer Saatzucht, Austria
Guenzi, Arron	acg@soilwater.agr.okstate.edu	Oklahoma State University, Stillwater
Guidobaldi, Héctor A.	guidobaldi@uol.com.ar	Univrsity of Córdoba, Argentina
Gupta, PK	pkgupta@ndf.vsn1.net.in	Ch. Charan Singh Univ, Meerut, India
Gustafson, Perry	pgus@showme.missouri.edu	USDA–ARS, University of Missouri
Haber, Steve	shaber@agr.gc.ca	Winnipeg, Manitoba, Canada
Haley, Scott	shaley@lamar.colostate.edu	Colorado State University, Ft. Collins
Hancock, June	june.hancock@seeds.Novartis.com	Novartis Seeds Inc., Bay, AR
Harrison, Steve	sharris@lsuvm.sncc.lsu.edu	Louisiana State University, Baton Rouge
Harder, Don	dharder@mbrswi.agr.ca	Winnipeg, Manitoba, Canada
Hart, Gary E	ghart@acs.tamu.edu	Texas A & M Univ, College Station
Hays, Dirk B	dhays@ag.gov	USDA–ARS, Stillwater, OK
Hayes, Pat	hayesp@css.orst.edu	Oregon State University, Corvallis
Hearnden, PR	phillippa.hearden@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Henzell, Bob	bobh@qdpit.sth.dpi.qld.gov.au	Warwick, Queensland, AU
Hershman, Don	dhershman@ca.uky.edu	University Of Kentucky, Lexington
Heslop-Harrison, JS (Pat)	hharrison@bbsrc.ac.uk	JI Centre, Norwich, UK
Hoffman, David	A03dhoffman@attmail.com	USDA–ARS, Aberdeen, ID
Hohmann, Uwe	uhemail@botanik.biologie.unimuenchen.de	Botanical Institute, Munich, Germany
Hoisington, David	dhoisington@cimmyt.mx	CIMMYT–Mexico
Hole, David	dhole@mendel.usu.edu	Utah State University, Logan
Howes, Neil	nhowes@mbrswi.agr.ca	Winnipeg, Manitoba, Canada
Hubbard, JD	john@gmprc.ksu.edu	USDA–ARS–GMPRC, Manhattan, KS
Huber, Don M	huber@btny.purdue.edu	Purdue University, W. Lafayette, IN
Hucl, Pierre	hucl@sask.usask.ca	University of Saskatchewan
Hughes, Mark E	markh@puccini.crl.umn.edu	USDA–Univ. of Minnesota, St. Paul
Hulbert, Scot	shulbrt@plantpath.ksu.edu	Kansas State University, Manhattan
Ibrahim, Amir	amir_ibrahim@sdstate.edu	South Dakota State Univ, Brookings
Isaac, Peter G	mbnis@seqnet.dl.ac.uk	Nickerson Biocem, UK
Jacquemin, Jean	stamel@fsagx.ac.be	Cra-Gembloix, Belgium
Jelic, Miodrag	miодраг@knez.uis.kg.ac.yu	ARI Center Small Grains, Yugoslavia
Jiang, Guo-Liang	dzx@njau.edu.cn	Nanjing Agricultural University, China
Jin, Yue	jiny@ur.sdstate.edu	South Dakota State Univ, Brookings
Johnson, Doug	djohnson@ca.uky.edu	University of Kentucky, Lexington
Johnson, Jerry	jjohnso@gaes.griffin.peachnet.edu	University of Georgia, Griffin
Johnston, Paul	paulj@qdpit.sth.dpi.qld.gov.au	Warwick, Queensland, AU
Jones, Steven S	jones@wsuvml.csc.wsu.edu	Washington State University, Pullman
Joppa, Leonard	joppa@badlands.nodak.edu	USDA–ARS, Fargo, ND
Karabayev, Muratbek	mkarabayev@astel.kz	CIMMYT, Kazakhstan
Karow, Russell S	Russell.S.Karow@orst.edu	Oregon State University, Corvallis

Karsai, Ildiko	karsai@buza.mgki.hu	ARI, Martonvasar, Hungary
Kasha, Ken	kkasha@crop.uoguelph.ca	University of Guelph, Canada
Keefer, Peg	peg_keefer@entm.purdue.edu	Purdue University, West Lafayette, IN
Keller, Beat	bkeller@botinst.unizh.ch	University of Zurich, Switzerland
Kidwell, Kim	kidwell@mail.wsu.edu	Washington State University, Pullman
Kindler, S Dean	sdkindler@pswcr1.ars.usda.gov	USDA-ARS, Stillwater, OK
Kirkham, MB	mbk@ksu.edu	Kansas State University, Manhattan
Kisha, Theodore	tkisha@dept.agry.purdue.edu	Purdue University, W. Lafayette, IN
Kleinhofs, Andy	coleco@bobcat.csc.wsu.edu	Washington State University, Pullman
Knezevic, Desimir	deskok@knezuis.kg.ac.yu	ARI Center Small Grains, Yugoslavia
Koebner, Robert	koebner@bbsrc.ac.uk	JI Centre, Norwich, UK
Koemel, John Butch	jbk@soilwater.agr.okstate.edu	Oklahoma State University, Stillwater
Kokhmetova, Alma	kalma@ippgb.academ.alma-ata.su	Kazakh Research Institute of Agriculture
Kolb, Fred	fkolb@ux1.cso.uiuc.edu	University Of Illinois, Urbana
Kolesnichenko, AV	akol@sifibr.irk.ru	Siberian Inst Plant Physiology, Irkutsk
Koppel, R	Reine.Koppel@jpbi.ee	Jõgeva Plant Breeding Institute, Estonia
Korol, Abraham	rabi309@haifaumv.bitnet	University of Haifa
Kovalenko, E.D.	kovalenko@vniif.rosmail.com	Russian Res Inst Phytopath, Moscow
Krasilovets, Yuri G	ncpgru@kharkov.ukrtel.net	Inst Plant Production, Karkiv, Ukraine
Kronstad, Warren E	kronstaw@css.orst.edu	Oregon State University, Corvallis
Krupnov, V	imu@ssau.saratov.ru	Agric Res Inst SE Reg, Saratov, Russia
Kuhr, Steven L	slkuhr@ccmail.monsanto.com	Hybritech-Mt. Hope, KS
Lafferty, Julia	lafferty@edv1.boku.ac.at	Saatzucht Donau, Austria
Lagudah, Evans	e.lagudah@pi.csiro.au	CSIRO, Australia
Lankevich, SV	laser@sifibr.irk.ru	Siberian Inst Plant Physiology, Russia
Láng, László	langl@buza.mgki.hu	Martonvásár, Hungary
Langridge, Peter	plangridge@waite.adelaide.edu.au	University of Adelaide, Australia
Lapitan, Nora LV	nlapitan@lamar.colostate.edu	Colorado State University, Ft. Collins
Lapochkina, Inna F	lapochkina@chat.ru	Research Inst of Agric, Moscow, Russia
Laskar, Bill	laskarb@phibred.com	Pioneer Hi-Bred-Windfall, IN
Leach, Jan E	jeleach@ksu.edu	Kansas State University, Manhattan
Leath, Steve	steven_leath@ncsu.edu	USDA-ARS, Raleigh, NC
Leonard, Kurt J	kurtl@puccini.crl.umn.edu	USDA-ARS, St. Paul, MN
Leroy, Philippe	leroy@valmont.clermont.inra.fr	INRA, Clermont
Lewis, Hal A	halewi@ccmail.monsanto.com	Hybritech-Corvallis OR
Lewis, Silvina	slewis@cirn.inta.gov.ar	CNIA-INTA, Buenos Aires, Argentina
Li, Wanlong	wli@plantpath.ksu.edu	Kansas State University, Manhattan
Liang, GH	ghliang@ksu.edu	Kansas State University, Manhattan
Line, RF	rline@wsu.edu	USDA-ARS, Pullman, WA
Liu, Dajun	djliu@public1.ptt.js.cn	Nanjing Agricultural University, China
Lively, Kyle	livelyk@phibred.com	Pioneer Hi-Bred-Windfall, IN
Long, David	davidl@puccini.crl.umn.edu	USDA St. Paul, MN
Lookhart, George	george@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Luckow, Odean	alvkow@em.agr.ca	AAFC-Winnipeg, Manitoba
Lukaszewski, Adam	ajoel@ucrac1.ucr.edu	University of California-Riverside
Maas, Fred	fred_maas@entm.purdue.edu	Purdue University, West Lafayette, IN
Mackay, Michael	mackaym@quord.agric.nsw.gov.au	AWEE, Tamworth, NSW
Maich, Ricardo H.	rimaich@agro.uncor.edu	University of Córdoba, Argentina
Manifesto, María M	mmanifes@cicv.intgov.ar	INTA Castelar, Argentina
Marais, G. Frans	gfm@akad.sun.ac.za	University of Stellenbosch, R.S.A.
Mares, Daryl J	dmares@adelaid.edu.au	University of Adelaide, Australia
Marshall, David	d-marshall@tamu.edu	Texas A & M University, Dallas
Marshall, Gregory C	marshallg@phibred.com	Pioneer Hi-Bred-Windfall, IN
Martin, Erica	erica.martin@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Martín-Sánchez, JA	JuanAntonio.Martin@irta.es	IRTA, Lleida, Spain
Martynov, Sergei	sergej_martynov@mail.ru	Vavilov Inst Plant Prod, St. Petersburg
Mather, Diane	indm@musicb.mcgill.ca	McGill University, Canada

Matthews, Dave	matthews@greengenes.cit.cornell.edu	Cornell University, Ithaca, NY
McCallum, John	mccallumj@lan.lincoln.cri.nz	Crop & Food Res. Ltd, NZ
McGuire, Pat	pemcguire@ucdavis.edu	University of California, Davis
McIntosh, Robert A	bobm@camden.usyd.edu.au	PBI Cobbitty, Australia
McKendry, Anne L	mckendrya@missouri.edu	University of Missouri, Columbia
McKenzie, RIH	rmckenzie@em.agr.ca	AAFC-Winnipeg, Manitoba
McVey, Donald	domv@puccini.crl.umn.edu	USDA-ARS, St. Paul, MN
Mi, Q.L.	qlm@ksu.edu	Kansas State University, Manhattan
Milach, Sandra	mila0001@student.tc.umn.edu	University of Minnesota, St. Paul
Miller, James	millerid@fargo.ars.usda.gov	USDA-ARS, Fargo, ND
Milovanovic, Milivoje	mikim@knez.uis.kg.ac.yu	ARI Center Small Grains, Yugoslavia
Milus, Gene	gmilus@comp.uark.edu	University of Arkansas, Fayetteville
Miskin, Koy E	miskin@dcwi.com	Agipro Seeds, Inc.
Mochini, RC	rmoschini@inta.gov.ar	INTA, Castelar, Argentina
Moffat, John	apwheat@frii.com	AgriPro Wheat, Berthoud, CO
Moldovan, Vasile	scaturda@rdslink.ro	Agric Research Station, Turda, Romania
Molnár-Láng, Marta	molnarm@fsnew.mgki.hu	Martonvásár, Hungary
Moore, Paul	ejh@uhccvx.uhcc.hawaii.edu	University of Hawaii
Moreira, João C.S.	moreira@cnpt.embrapa.br	EMBRAPA, Passo Fundo, Brazil
Morgounov, Alexei	amorgounov@astel.kz	CIMMYT, Kasakhstan
Morino-Sevilla, Ben	bmoreno-sevilla@westbred.com	Western Plant Breeders, Lafayette, IN
Mornhinweg, Dolores W	dmornhin@ag.gov	USDA-ARS, Stillwater, OK
Morris, Craig	wwql@wsuax.csc.wsu.edu	USDA Pullman, WA
Morrison, Laura	alura@peak.org	Oregon State University, Corvallis
Moser, Hal	hsmoser@iastate.edu	Iowa State University, Ames
Mujeeb-Kazi, A	mkazi@cimmyt.mx	CIMMYT, Mexico
Mukai, Yasuhiko	ymukai@cc.osaka-kyoiku.ac.jp	Osaka Kyoiku University, Japan
Murphy, Paul	njpm@unity.ncsu.edu	North Carolina State University
Murray, Tim	tim_murray@wsu.edu	Washington State University, Pullman
Muthukrishnan, S	smk@ksu.edu	Kansas State University, Manhattan
Nakamura, Hiro	hiro@affrc.go.jp	Tohoku Nat Ag Exp Sta, Japan
Nass, Hans	nassh@em.agr.ca	AAFC-Prince Edward Island, Canada
Nelson, Lloyd R	lr-nelson@tamu.edu	Texas A & M University
Nevo, Eviatar	rabi301@haifaunivm.bitnet	University of Haifa, Israel
Nicol, Julie	j.nicol@cgiar.org	CIMMYT, Mexico
Nguyen, Henry T	bwlab@ttacs1.ttu.edu	Texas Tech University, Lubbock
Noll, John S.	jnoll@em.agr.ca	AAFC-Winnipeg, Canada
Nyachiyo, Joseph	jnyachir@gpu.srv.ualberta.ca	University of Alberta
O'Donoughue, Louise	em220cyto@nccot2.agr.ca	AAFC-Canada
Ogbonnaya, Francis C	fc.ogbonnaya@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Ohm, Herb	hohm@purdue.edu	Purdue University, West Lafayette, IN
Ohm, Jay B	jay@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Oman, Jason	jason.oman@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Osipova, AV	gluten@sifibr.irk.ru	Siberian Inst Plant Physiology, Russia
Paelo, Antonio D	adiazpaleo@cnia.inta.gov.ar	CRN INTA Castelar, Argentina
Park, SH	seokho@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Payne, Thomas	t.payne-t@cgiar.org	CIMMYT, Addis Ababa, Ethiopia
Penix, Susan	agsusan@mizzou1.missouri.edu	University of Missouri, Columbia
Permyakov, AV	gluten@sifibr.irk.ru	Siberian Inst Plant Physiology, Russia
Perry, Keith	perry@btny.purdue.edu	Purdue University, W. Lafayette, IN
Perry, Sid	sidgsr@southwind.com	Goertzen Seed Research, Haven, KS
Pérez, Beatriz A	baperez@inta.gov.ar	INTA, Castelar, Argentina
Peterson, CJ	cjp@orst.edu	Oregon State University, Corvallis
Pickering, Richard	pickeringr@crop.cri.nz	Christchurch, NZ
Pomazkina, L	agroeco@sifibr.irk.ru	Siberian Inst Plant Physiology, Russia
Pogna, Norberto	isc.gen@iol.it	Inst Exper Cereal, Rome, Italy
Porter, David	dporter@pswcrl.ars.usda.gov	USDA-ARS, Stillwater, OK

Poulsen, David	davep@qdpit.sth.dpi.qld.gov.au	Warwick, Queensland AU
Priillin, Oskar	ebi@ebi.ee	Estonian Agricultural University, Harku
Pukhalskiy, VA	pukhalsk@iogen.msk.su	N.I. Vavilov Institute, Moscow
Qi, Lili	qilili@plantpath.ksu.edu	Kansas State University, Manhattan
Qualset, Cal	coqualset@ucdavis.edu	University of California–Davis
Quick, Jim	jquick@ceres.agsci.colostate.edu	Colorado State University, Ft. Collins
Rabinovich, Svetlana	svr@svr.kharkiv.com	Inst Plant Production, Karkiv, Ukraine
Rajaram, Sanjaya	srajaram@cimmyt.mx	CIMMYT, Mexico
Ram, MS	ramms@gmprc.ksu.edu	USDA–ARS–GMPRC, Manhattan, KS
Ratcliffe, Roger H	roger_ratcliffe@entm.purdue.edu	USDA–ARS, W. Lafayette IN
Ratti, C	cratte@tin.it	Istit Patologia Vegetale, Bologna, Italy
Raupp, WJ	raupp@ksu.edu	Kansas State University, Manhattan
Rayapati, John	nanster@iastate.edu	Iowa State University, Ames
Reddy, VRK	botany@bharathi.ernet.in	Bharathiar University, Coimbatore, India
Reisner, Alex	reisner@angis.su.oz.au	Australia
Rekoslavskaya, Natalya I	phytolab@sifibr.irk.ru	Siberian Inst Plant Physiology, Russia
Riera-Lizarazu, Oscar	oscar.rierd@orst.edu	Oregon State University, Corvallis
Roberts, John	jrobert@gaes.griffin.peachnet.edu	USDA–ARS, Griffin, GA
Rodríguez, Daniel	daniel.rodriguez@nre.vic.gov.au	Victorian Inst Dryland Agric, Australia
Rohrere, Wendy	wrohrer@vt.edu	Virginia Tech, Blacksburg
Romig, Robert W	bobromig@aol	Trigen Seed Services LLC, MN
Rosa, OS	ottoni@ginet.com.br	OR Seed Breeding Co., Brazil
Rudd, Jackie	ruddj@mg.sdstate.edu	South Dakota State Univ, Brookings
Rubies-Autonell, C	crubies@agrsci.unibo.it	Istit Patologia Vegetale, Bologna, Italy
Safranski, Greg	greg_safranski@entm.purdue.edu	Purdue University, W. Lafayette, IN
Săulescu, Nicolae	saulescu@valhalla.racai.ro	Fundulea Institute, Romania
Schwarzacher, Trude	schwarzta@bbsrc.ac.uk	JI Centre, Norwich, UK
Seabourn, BW	brad@gmprc.ksu.edu	USDA–ARS–GMPRC, Manhattan, KS
Sears, Rollie	rsears@flinthsills.com	AgriPro Wheat, Junction City, KS
Seitz, LM	larry@gmprc.ksu.edu	USDA–ARS–GMPRC, Manhattan, KS
Shaner, Greg	shaner@btny.purdue.edu	Purdue University, W. Lafayette, IN
Sharma, Hari	hsharma@purdue.edu	Purdue University, W. Lafayette, IN
Sharp, Peter	peters@camden.usyd.edu.au	PBI Cobbitty, Australia
Sheppard, Ken	ksheppard@waite.adelaide.edu.au	University of Adelaide, Australia
Shields, Phil	shieldsp@phibred.com	Pioneer Hi-Bred, St. Matthews, SC
Shroyer, Jim	jshroyr@ksuvn.edu	Kansas State University, Manhattan
Shufran, Kevin A	kashufran@pswcrl.ars.usda.gov	USDA–ARS, Stillwater, OK
Shukle, Rich	rich_shukle@entm.purdue.edu	Purdue University, West Lafayette, IN
Singh, GP	dwrpratap@rediffmail.com	IARI, New Delhi, India
Singh, JB	jbsingh1@rediffmail.com	IARI, New Delhi, India
Singh, S.S.	singhss@rediffmail.ocm	IARI, New Delhi, India
Sinnott, Quinn	quinn@prime.ars-grin.gov	USDA–ARS, Beltsville, MD
Síp, Vaclav	sip@hb.vurv.cz	RICP, Prague, Czech Republic
Skovmand, Bent	bskovmand@cimmyt.mx	CIMMYT–Mexico
Smith, Joe A	jasmith@frii.com	AgriPro Seeds, Inc., Berthoud, CO
Snape, John	john.snape@bbsrc.ac.uk	JI Centre, Norwich, UK
Sorrells, Mark	mark_sorrells@qmrelay.mail.cornell.edu	Cornell University, Ithaca, NY
Spetsov, Penko	iws@eos.dobrich.acad.bg	Inst Wheat and Sunflower, Bulgaria
Steffenson, Brian	bsteffen@badlands.nodak.edu	North Dakota State University, Fargo
Stehno, IZ	stehno@vurv.cz	RICP, Prague, Czech Republic
Stift, G.	stift@ifa-tulln.ac.at	IFA-Tulln, Austria
Stoddard, Fred	stoddard@extro.ucc.edu.oz.ua	University of Sydney, Australia
Stuart, Jeffery J	jeff_stuart@entm.purdue.edu	Purdue University, W. Lafayette, IN
Stupnikova, IV	irina@sifibr.irk.ru	Siberian Inst Plant Physiology, Irkutsk
Suchy, Jerry	isuchy@em.arg.ca	AAFC–Winnipeg, Manitoba
Sun, Mei	meisun@hkucc.hku.hk	Hong Kong University
Szabo, Les	lszabo@puccini.crl.umn.edu	USDA–ARS, University of Minnesota

Talbert, Luther	usslt@montana.edu	Montana State University, Bozeman
Therrien, Mario C	therrien@mbrsbr.agr.ca	AAFC-Manitoba
Throne, JE	throne@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Tilley, M	mtilley@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Tinker, Nick	cznt@agradm.lan.mcgill.ca	McGill University
Tkachenko, OV	agm@ssau.saratov.ru	Saratov State Agrarian Univ, Russia
Tohver, Maimu	maimu.tohver@mail.ee	Estonian Agricultural University, Harku
Tomasovic, Slobodan	bc-botinec-hr@bc-institut.hr	Bc Institute, Zagreb, Croatia
Townley-Smith, TF	tsmith@em.agr.ca	AAFC-Winnipeg, Manitoba
Trottet, Maxime	mtrottet@rennes.inra.fr	INRA, Le Rheu Cedex, France
Torres, Lorena	letorres_k@yahoo.com.ar	University of Córdoba, Argentina
Tranquilli, Gabriela	granqui@cirn.inta.gov.ar	INTA Castelar, Argentina
Tsehaye, Yemane	yemtse@yahoo.com	Inst Biodiversity Conservation, Ethiopia
Tsujimoto, Hisashi	tsujimot@yokohama-cu.ac.jp	Kihara Institute, Japan
Urbano, Jose Maria	urbano@phibred.com	Pioneer Hi-Bred, Sevilla, Spain
D'utra Vaz, Fernando B	ferbdvaz@pira.cena.usp.br	University De Sao Paulo, Brazil
Vallega, Victor	vallegavictor@mclink.it	Exp Inst Cerealicoltura, Rome, Italy
Vassiltchouk, NS	ariser@mail.saratov.ru	ARISER, Saratov, Russia
Van Sanford, Dave	agr38@pop.uky.edu	University of Kentucky, Lexington
Varughese, George	g.varughese@cernet.com	CIMMYT, Mexico
Verhoeven, Mary C	Mary.C.Verhoeven@orst.edu	Oregon State University, Corvallis
Vida, Gyula	h8607vid@ella.hu	Hungary
Von Allmen, Jean-Marc	bvonal@abru.cg.com	Ciba-Geigy, Basel, Switzerland
Voss, Márcio	voss@cnpt.embrapa.br	EMBRAPA, Passo Fundo, Brazil
Waines, Giles	waines@ucrac1.ucr.edu	University of California, Riverside
Walker-Simmons, MK	ksimmons@wsu.edu	USDA-ARS, Pullman, WA
Wang, Richard	rrcwang@cc.usu.edu	Utah State University, Logan
Ward, Richard	wadri@pilot.msu.edu	Michigan State University, East Lansing
Watanabe, Nobuvoshi	watnb@cc.gifu-u.ac.jp	Gifu University, Japan
Webster, James A	jwebster@pswcr.ars.usda.gov	USDA-ARS, Stillwater, OK
Wesley, Annie	awesley@rm.agr.ca	AAFC-Winnipeg, Manitoba
Wildermuth, Graham	wilderg@prose.dpi.gld.gov.au	Leslie Research Centre, Australia
Williams, Christie	christie_williams@entm.purdue.edu	Purdue University, W. Lafayette, IN
Wilson, Dean	trio@feist.com	Trio Research, Wichita, KS
Wilson, Duane L	dlwil@ksu.edu	Kansas State University, Manhattan
Wilson, James A	trio@feist.com	Trio Research, Wichita, KS
Wilson, Jeff D	jdw@gmprc.ksu.edu	USDA-ARS-GMPRC, Manhattan, KS
Wilson, Paul	wilsonp@phibred.com	Pioneer Hi-bred, Northants, UK
Wilson, Peter	hwauast@mpx.com.au	Hybrid Wheat Australia, Tamworth
Yen, Yang	yeny@ur.sdstate.edu	South Dakota State Univ, Brookings
Zeller, Frederich	zeller@mm.pbz.agrar.tu-muenchen.de	Technical Univ.of Munich, Germany
Zemetra, Robert	rzemetra@uidaho.edu	University of Idaho, Moscow
Zhang, Peng	pzhang@plantpath.ksu.edu	Kansas State University, Manhattan
Zhu, Yu Cheng	zhuyuc@ag.gov	USDA-ARS, Stillwater, OK

IX. ANNUAL WHEAT NEWSLETTER FUND.

Financial Statement on account #7768480 at the Home National Bank, 4th and Duck, Stillwater, OK 74074, USA, Brett C. Carver, Treasurer, Annual Wheat Newsletter.

Five corporate sponsors and 24 individuals have contributed to Volume 48.

(Contributions over \$100)

Okkyung Kim Chung, USDA-ARS, BMPRC, USGMRL, 1515 College Ave., Manhattan, KS 66502, USA.
Dale R. Clark, Western Plant Breeders, Inc., 8111 Timberline Drive, Bozeman, MT 59715 USA.
James A. Wilson, 6414 N. Sheridan, Trio Research Inc., Wichita, KS 67204-6606, USA.

(Contributions \$50 to \$99)

Robert E. Allan, 3202 Old Moscow Road, Washington State University, USDA-ARS, Pullman, WA, USA.
Grain Marketing Production and Research Center, USDA-ARS-NPA, 1515 College Ave., Manhattan, KS 66502, USA.
M.B. Kirkhan, Department of Agronomy, Kansas State University, Manhattan, ID 66506-5501, USA.
Robert Koebner, John Innes Centre, Norwich Research Park, Colney, Norwich NR4 7UH, UNITED KINGDOM.
G.F. Marais, University of Stellenbosch, Department of Genetics, Private Bag X1, Matieland, 7602, SOUTH AFRICA.
J.A. Martín-Sánchez, Alcaide Rovira Roure, UdL-IRTA, Lelida 25198, SPAIN.
Patsy Sperry, 209 Johnson Hall, USDA-ARS Washington State University, Pullman, 99164-6420 WA, USA.

(Contributions to \$50)

Cereal Research Centre, Agriculture and Agri-Food Canada, 195 Dafoe Rd., Winnipeg, Manitoba R3T 2M9, CANADA.
Harold Bockelman, USDA-ARS, P.O. Box 307, Aberdeen, ID 83210, USA.
Gaetano Boggini, Via Mulino 3, Instituto sperimentale per la Cerealicoltura, S. Angelo Lodigiano, LO 26866, ITALY.
Xianming Chen, 361 Johnson Hall, P.O. Box 646430, Washington State University, Pullman, WA 99164-6430 USA.
Byrd C. Curtis, 1904 Sequoia Street, Ft. Collins, CO 80525-1540, USA.
Cereal Disease Lab, USDA-ARS, 1551 Lindig St., St. Paul, MN 55108, USA.
Ricardo Hector Maich, Faculty of Ciencias Agropecuarias, Universidad Nacional Casillia de Correro, 509-C Central,
5000 Córdoba, ARGENTINA.
D. Mares, Plant Science, Waite Campus, University of Adelaide, Glen Osmond 5064, AUSTRALIA.
Robert A. McIntosh, Plant Breeding Institute, Cobbitty, Private Bag 1, Camden NSW 2570, AUSTRALIA.
Hiro Nakamura, Shimokuriagava, Tohoku National Agricultural Experiment Station, Morioka Iwate, 020-0198, JAPAN.
Herbert W. Ohm, Purdue University, Department of Agronomy, 105 Lilly Hall, W. Lafayette, IN 47907, USA.
Fred L. Patterson, Purdue University, Department of Agronomy, 105 Lilly Hall, W. Lafayette, IN 47907, USA.
James S. Quick, Soil and Crop Sciences Department, Colorado State University, Ft. Collins, CO 80525, USA.
Ottoni de Sousa Rosa, OR Seed Breeding Company, Rua João Battisti, 71-Passo Fundo, 99050-380, BRAZIL.
Ralf Schachschneider, Nordsaat Saatzucht GMBH, Hauptstrasse 1, D-38895 Bohnshausen, GERMANY.
Bent Skovmand, Apdo Postal 6-641, CIMMYT, México D.F. 06600, MÉXICO.
F. du Toit, Pannar Ltd. (PTY), P.O. Box 17164, Bainsvlei 9338, SOUTH AFRICA.
Wayne E. Vian, 1207 Harrison Street, Grand Island, NE 68803-6352, USA.
Nobuyoshi Watanabe, Faculty of Agriculture, Gifu Univ., 1-1 Yanagido, Gifu 501-11, JAPAN.

X. VOLUME 49 MANUSCRIPT GUIDELINES.

Manuscript guidelines for the *Annual Wheat Newsletter*, volume 49. The required format for Volume 49 of the *Annual Wheat Newsletter* will be similar to Volume 48 and previous editions.

CONTRIBUTIONS MAY INCLUDE:

- Current activities on your projects.
- New cultivars and germ plasm released.
- Special reports of particular interest, new ideas, etc., normally not acceptable for scientific journals.
- A list of recent publications.
- News: new positions, advancements, retirements, necrology.
- Wheat stocks; lines for distribution, special equipment, computer software, breeding procedures, techniques, etc.

FORMATTING & SUBMITTING MANUSCRIPTS:

Follow the format in volume 44–48 of the *Newsletter* in coordinating and preparing your contribution, particularly for state, station, contributor names, and headings. Limited editing is done. Use the WordPerfect or Word programs, or send an RTF file that can be converted. Use Times 12 CPI and 1.0" (2.5 cm) margins. DO NOT use the table settings or column setting functions, create tables with tabs and spaces. Double-space the text of your contribution if you must use a typewriter.

All text will be entered in computer files; therefore, please submit manuscript in any of the above formats. Mail a file on a disk to W. John Raupp, Department of Plant Pathology, Throckmorton Hall, Kansas State University, Manhattan KS 66506-5502. If submitting by E-mail, send to raupp@ksu.edu.

DISTRIBUTION:

The primary method of distribution of Volume 49 will be CD-ROM in HTML format. These files can be read with any internet browser. A hard copy will be sent only if requested by 1 March, 2003, and will cost \$40.

The *Annual Wheat Newsletter* will continue to be available (Vol. 37–48) through the Internet on GrainGenes, the USDA–ARS Wheat Database at <http://wheat.pw.usda.gov/ggpages/awn/> and Internet gopher access at "greengenes.cit.cornell.edu".

COST:

The cost of publishing the *Annual Wheat Newsletter* is financed by voluntary contributions from individuals, commercial companies, international programs, and organizations with a direct or indirect interest in wheat. Funds on hand and contributions have been insufficient to pay for hard copies.

In the interest of remaining solvent, the NWIC has approved future distribution primarily by computer diskette. We are asking that you renew your contribution or, if you have not contributed in the past, to join the growing list of contributors. Contributions from individuals in the range of \$15 to \$30 play a significant role in financing the Newsletter. An increase in the number of individual contributors is very important, and we are confident that, with continued corporate support, we will be able to meet our financial obligations in 2003. The address for contributions is Dr. Brett Carver, Department of Agronomy, Oklahoma State University, Stillwater, OK 74078, U.S.A.