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ANALYSES

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Maps of Papillomavirus mRNA Transcripts

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Explanation of Maps

Maps showing the open reading frames and mRNA transcripts of seven papillomaviruses are presented on the following pages. Facing each map is a brief description of the transcripts, and following the maps is a list of references from which the maps were compiled. In each map the significant ORFs are shown in their proper reading frames as colored rectangles. At the upper left end of the rectangle are two numbers. The first corresponds to the nucleotide (nt) position of the ORF start, the first nucleotide following a stop codon. The second number records the nt position of the first ATG, which is also indicated by a dotted line within the rectangle. The position of the last nt in the stop codon of each ORF is printed at the lower right corner of the rectangles. The rectangle's colored fill begins at the ATG and extends to the end; thus it corresponds to the coding sequence, not the entire ORF. In the cases where no ATG exists in the ORF, only one number is present in the upper left corner and the rectangle is completely filled with color. Below the ORFs is a scale of the genome divided into thousands. On the scale are placed the positions of promotors (represented by arrows) and the poly(A) signals. The exact position of the poly(A) signal is printed below the scale line. Located below the genome scale are diagrams of mRNA species, most of which are spliced. The exons are illustrated by heavy black lines, while the introns are indicated by black hairlines between. The numbers printed below the lines indicate the 5' and 3' termini of the RNAs, and the 5' and 3' splice junction positions. The splice junction numbers give the position of the last nucleotide in the exon before the splice and the position of the first nucleotide of the exon following the splice. Splice junctions in parentheses were deduced from the genomic sequence and have not been confirmed by cDNA sequencing. Where 5' or 3' ends of the RNAs are uncertain, no nt position is given. Superimposed on the exons are colored rectangles representing the gene, or part thereof, coded by that portion of the exon. The coding potential of each transcript is also listed at the right. In that list a ^ symbol between two gene name (e.g. E1^E4) indicates a fusion product. The * symbol indicates different forms of the E6 product.

BPV-1 has served as the prototype for the genetic analysis of the papillomaviruses. The mRNAs from BPV-1 transformed C127 cells as well as productively infected bovine fibropapillomas have been exhaustively analyzed. The top of the BPV-1 map shows the genomic organization of BPV-1. The long control region is labelled and, in order to show the position of promoters, is repeated to the left of nt 0. The locations of known promoters [3,6,23,24,27] are indicated by arrows and labeled P_n where n is the approximate nucleotide position of the RNA start site for that promoter. P_{7185} , P_{89} , P_{890} , P_{2443} , and P_{3080} are also referred to as P_1 , P_2 , P_3 , P_4 , and P_5 [6,23], respectively. The late promoter (P_L) is the major promoter active in the fibropapilloma and is the only promoter not active in C127 cells transformed by BPV-1 [3]. The early and late polyadenylation signals are at nt 4180 and 7156, respectively. The structures of BPV-1 mRNAs from BPV-1 transformed mouse C127 cells (species A—Q) were determined by cDNA cloning as well as electron microscopy, nuclease protection, PCR, and primer extension. The 5' most ORF containing a translation initiation codon and a significant coding region is indicated at the right of each mRNA. Although an E6/E7 fusion ORF is the 5' most ORF for species I, the cDNA from which this structure was deduced has been shown to encode the E1 M protein [26].

Additional very rare mRNA species from cycloheximide treated BPV-1 transformed C127 cells have been characterized [6], but are not shown here. The structures of mRNAs unique to the BPV-1 fibropapilloma (species R–X) were determined by RT-PCR and cDNA cloning and sequencing [3,4]. Although the E2 and E4 ORFs are the first significant ORF for species W and X, these mRNAs may also encode the L2 protein. A more detailed discussion of BPV-1 transcription can be found elsewhere, including references for each mRNA [1,2].

BPV1R



HPV-16 mRNAs isolated from transfected cells and a variety of tumor cell lines and lesions containing both extrachromosomal and integrated HPV-16 genomes have been analyzed in multiple laboratories [8,15,18-20]. Viral/host chimeric mRNAs have been purposely omitted. All nucleotide positions correspond to the revised HPV-16 sequence published in Part I of this compendium. Most mRNA species were determined by RT-PCR, so the identity of the 5' and 3' ends are not known. However, only one promoter (P_{97}) has been definitively identified for HPV-16 [22]. Species A-K are most likely transcribed from this promoter. The promoter responsible for the transcription of the late mRNAs (species O-P) is not known. The early and late polyadenylation signals are located at nt 4215 and nt 7321, respectively [11,15].

HPV16R



HPV-31 mRNAs were investigated in CIN62 cells grown in monolayer cultures and in organotypic raft cultures [9,14]. CIN612 cells contain extrachromosomal HPV-31. Two promoters were identified by primer extension and RNAse protection experiments. P_{97} is active in both monolayer and raft cultures; P_{742} is active only in differentiated raft cultures [9]. No promoter could be identified in the E6 ORF [9]. The mRNA structures identified are shown in the facing map. Species A and B are presumably transcribed from P_{97} . Species C, D, and presumably E are transcribed from P_{742} . Additional L1 mRNAs appear to be transcribed from P_{97} , but the exact splice structure of these mRNAs is unknown. Early mRNAs were polyadenylated between nt 4099 and 4125 [9]. The exact sites of polyadenylation at the late poly(A) site were not determined in these studies, but a putative polyadenylation signal exists at nt 7227.



HPV-11 mRNAs, isolated from genital condyloma acuminata and experimental condylomatous cysts implanted under the renal capules of nude mice, have been analyzed by several laboratories [5,7,13,16,17]. The types of analyses include cDNA cloning (species A), electron microscopy analysis of R-loops (species A–G, J–K), and RT-PCR analysis (species C–F, H–J). The deduced mRNA structures are shown in the HPV-11 map. The 5' ends of individual mRNAs have not been mapped at the nucleotide level. However, three promoters have been mapped by Smotkin et al. [21] using nuclease S1 analysis: the E6 promoter initiates at nt 90, the E7 promoter at nt 264, and the E1 promoter at nts 674–714. The 5' end of the cDNA representing species A is located at nt 716 [13], so this mRNA is presumably transcribed from the E1 promoter. The early poly(A) site has been identified from analysis of species A cDNAs [13]. The late poly(A) site has not been identified experimentally, but the L1 and L2 mRNAs are presumably polyadenylated utilizing the poly(A) signal at nt 7457.



HPV-5 mRNAs isolated from benign skin lesions from patients with epidermodysplasia verruciformis were analyzed by RT-PCR (Haller, Stubenrauch, and Pfister, *Virology*, in press). The 5' and 3' ends of these mRNAs are unknown. However, in situ hybridization data as well as promoter mapping data for the related virus HPV-8 suggest that four promoters may exist, a late promoter in the LCR, an E6 promoter, an E7 promoter, and an E1 promoter similar to that identified for HPV-31 [9]. Species A–D are most likely transcribed by the late promoter and species E–F by the E6 promoter. The early and late poly(A) sites have not been experimentally determined. However, by analogy with the related virus HPV-47 [12], the early poly(A) signal is at nt 4438. Species A–B and E–G are most likely polyadenylated at the early poly(A) site, while species C–D and H–I at the late poly(A) site.



HPV-8 mRNAs have been investigated from both transfected cells and lesions from an epidermodysplasia vertuciformis patient. Analysis of RNA start sites suggest at least three promoters: a late promoter in the LCR with start site at nt 7535 (P_{7535}) [25], an E6 promoter with start sites at nt 175–179 (P_{175}) [10,25], and an E7 promoter with start site just upstream of the E7 ORF (not further mapped [10]). RT-PCR analysis of RNA from the EV lesion [25] identified two L1 mRNAs which presumably are transcribed from the late promoter. The exact 5' and 3' ends of these mRNAs have not been determined, however.



Analysis of mRNA from an HPV-47 transformed rat cell line using S1 and primer extension analysis revealed RNA start sites at nt 183–202 with a major start site at nt 198 [12]. The early poly(A) sites were mapped to nt 4444 and nt 4465 by cDNA cloning and sequencing. Three RNA structures were determined using cDNA cloning coupled with S1 analysis. Species A is the most abundant mRNA. Only one clone was identified for species B and C mRNAs. The 5' end of the cDNA insert for the species B clone was at an EcoRI site which was used for cloning the cDNA, so the actual cDNA may have extended further upstream.



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References

- Baker, C. C. 1990. Bovine papillomavirus type 1 transcription, p. 91-112. In H. Pfister (ed.), Papillomavirus and human cancer. CRC Press, Inc. Boca Raton, FL.
- \mathbf{N} K maps: locus maps of complex genomes . Cold Spring Harbor Laboratory Press, Cold Spring Harbor, Baker, C. C. 1993. The genomes of the papillomaviruses, p. 1.134-1.146. In S. O'Brien (ed.), Genetic
- ω Baker, C. C. and P. M. Howley. 1987. Differential promoter utilization by the bovine papillomavirus in transformed cells and productively infected wart tissues. EMBO J. 6:1027-1035.
- 4 papillomavirus type 1 P2443 and late promoters. J. Virol . 67:5605-5616. Barksdale, S. K. and C. C. Baker. 1993. Differentiation-specific expression from the bovine
- S papillomavirus type 11 is a sequence-specific transcription repressor. J. Virol. 65:3317-3329 Chiang, C.-M., T. R. Broker, and L. T. Chow. 1991. An E1M E2C fusion protein encoded by human
- 6 two forms of a transcriptional repressor: Structural and functional analysis of new viral cDNAs. J. Choe, J., P. Vaillancourt, A. Stenlund, and M. Botchan. 1989. Bovine papillomavirus type 1 encodes Virol. 63:1743–1755.
- -Chow, L. T., M. Nasseri, S. M. Wolinsky, and T. R. Broker. 1987. Human papillomavirus types 6 and 11 mRNAs from genital condylomata acuminata. *J. Virol*. **61**:2581–2588.
- ∞ of novel splicing patterns in a HPV16-containing keratinocyte cell line. Virology 178:254-262 Doorbar, J., A. Parton, K. Hartley, L. Banks, T. Crook, M. Stanley, and L. Crawford. 1990. Detection
- 9 **66**:6070-6080. transcription of human papillomavirus type 31b in cell lines containing viral episomes. J. Virol. Hummel, M., J. B. Hudson, and L. A. Laimins. 1992. Differentiation-induced and constitutive
- 10 Iftner, T., G. Sagner, H. Pfister, and F. O. Wettstein. 1990. The E7 protein of human papillomavirus 8 is a nonphosphorylated protein of 17 kDa and can be generated by two different mechanisms. Virology 179:428-436.
- 1 late mRNA 3' processing signals in vitro and in vivo. J. Virol . 64:1825-1829 Kennedy, I. M., J. K. Haddow, and J. B. Clements. 1990. Analysis of human papillomavirus type 16
- 12 Kiyono, T., K. Nagashima, and M. Ishibashi. 1989. The primary structure of major viral RNA in a cDNA and E6 gene. Virology 173:551-565 rat cell line transfected with type 47 human papillomavirus DNA and the transforming activity of its
- 13 Nasseri, M., R. Hirochika, T. R. Broker, and L. T. Chow. 1987. A human papilloma virus type 11 transcript encoding an E1-E4 protein. Virology 159:433-439.
- 14 Pray, T. R. and L. A. Laimins. 1995. Differentiation-dependent expression of E1 E4 proteins in cell lines maintaining episomes of human papillomavirus type 31b. Virology 206:679-685

- 5 Rohlfs, M., S. Winkenbach, S. Meyer, T. Rupp, and M. Durst. 1991. Viral transcription in human keratinocyte cell lines immortalized by human papillomavirus type-16. Virology 183:331–342.
- 16 Rotenberg, M. O., C.-M. Chiang, M. L. Ho, T. R. Broker, and L. T. Chow. 1989. Characterization of gene transfer. Virology 172:468–477. cDNAs of spliced HPV-11 E2 mRNA and other HPV mRNAs recovered via retrovirus-mediated
- 17 Rotenberg, M.O., L. T. Chow, and T. R. Broker. 1989. Characterization of rare human papillomavirus type 11 mRNAs coding for regulatory and structural proteins, using the polymerase chain reaction. Virology 172:489–497.
- 81 Sherman, L. and N. Alloul. 1992. Human papillomavirus type 16 expresses a variety of alternatively spliced mRNAs putatively encoding the E2 protein. Virology 191:953-959.
- 19 Sherman, L., N. Alloul, I. Golan, M. Durst, and A. Baram. 1992. Expression and splicing patterns of J. Cancer 50:356–364. human keratinocytes immortalized by HPV 16, and in cell lines established from cervical cancers. Int. human papillomavirus type-16 mRNAs in pre-cancerous lesions and carcinomas of the cervix, in
- 20 873. Smits, H. L., M. T. E. Cornelissen, M. F. Jebbink, J. G. Van den Tweel, A. P. H. B. Struyk, M. Briet, junctions and full-length viral copies in CaSki cells and in a cervical carcinoma. Virology 182:870and J. ter Schegget. 1991. Human papillomavirus type 16 transcripts expressed from viral-cellular
- 21 Smotkin, D., H. Prokoph, and F. O. Wettstein. 1989. Oncogenic and nononcogenic human genital papillomaviruses generate the E7 mRNA by different mechanisms. J. Virol . 63:1441-1447.
- 22 Smotkin, D. and F. O. Wettstein. 1986. Transcription of human papillomavirus type 16 early genes in a cervical cancer and a cancer-derived cell line and identification of the E7 protein. Proc. Natl. Acad. Sci. USA 83:4680-4684.
- 23 Stenlund, A., G. L. Bream, and M. R. Botchan. 1987. A promoter with an internal regulatory domain is part of the origin of replication in BPV-1. Science 236:1666-1671.
- 24 Stenlund, A., J. Zabielski, H. Ahola, J. Moreno-Lopez, and U. Pettersson. 1985. Messenger RNAs from the transforming region of bovine papilloma virus type I. J. Mol. Bio . 182:541-554
- 25 Stubenrauch, F., J. Malejczyk, P. G. Fuchs, and H. Pfister. 1992. Late promoter of human papillomavirus type 8 and its regulation. J. Virol. 66:3485-3493
- 26 Thorner, L., N. Bucay, J. Choe, and M. Botchan. 1988. The product of the bovine papillomavirus type 1 modulator gene (M) is a phosphoprotein. J. Virol . 62:2474-2482
- 27 Yang, Y. C., H. Okayama, and P. M. Howley. 1985. Bovine papillomavirus contains multiple transforming genes. Proc. Natl. Acad. Sci. USA 82:1030-1034.