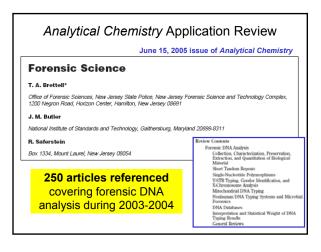


National Institute of Justice
 The Research, Development, and Evaluation Agency of the U.S. Department of Justice

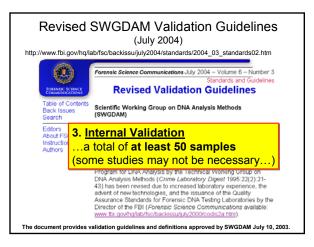
Current Areas of NIST Research Effort

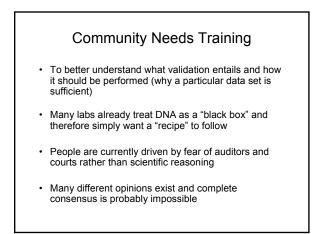
- Standard Information Resources (STRBase information, training materials/review articles, validation standardization, calibration datasets)
- Interlaboratory Studies (Real-time PCR, mixture interpretation)
- Resources for "Challenging Samples" (miniSTRs for degraded DNA)
- Information on New Loci (Y-Chromosome, new STRs, SNPs)



# Validation Project Purpose

- Review validation practices currently in use and available standards and guidelines (revised SWGDAM guidelines are too general)
- Help the community gain a better understanding of the validation process and how others have implemented validation in their labs so that validation in one's own lab may be performed more guickly
- Attempt to define a minimum number of samples that could be recommended for various validation scenarios
- Help with establishing uniformity throughout the field to aid auditors in their inspections





# Validation Definitions

### ISO 17025

5.4.5.1 Validation is the confirmation by examination and the provision of objective evidence that the particular requirements for a specific intended use are fulfilled

### DAB Quality Assurance Standards for Forensic DNA Testing Laboratories

2 (ff) Validation is a process by which a procedure is evaluated to determine its efficacy and reliability for forensic casework analysis and includes:

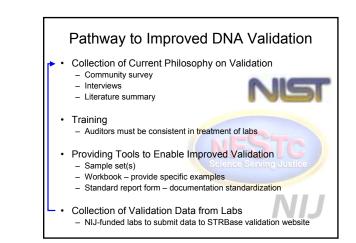
To demonstrate that a method is suitable for its intended purpose...

# DAB Quality Assurance Standards for Forensic DNA Testing Laboratories

# Manufacturer

- Developmental validation is the acquisition of test data and determination of conditions and limitations of a <u>new</u> or novel DNA methodology for use on forensic samples.
- (2) Internal validation is an accumulation of test data within the laboratory to demonstrate that <u>established</u> methods and procedures perform as expected in the laboratory.
  Forensic Lab

# Pathway to Improved DNA Validation Collection of Current Philosophy on Validation Community survey Interviews Literature summary Training Auditors must be consistent in treatment of labs Providing Tools to Enable Improved Validation Sample set(s) Workbook – provide specific examples Standard report form – documentation standardization Collection of Validation Data from Labs NIJ-funded labs to submit data to STRBase validation website



# Contacting the Community

- Validation Standardization Questionnaire handed out at NIJ DNA Grantees meeting (June 28-30, 2004)
- Emails sent to >200 scientists (July-Aug 2004)
  - Attendees from the NIJ DNA Grantees meeting
     Participants in NIST interlaboratory studies
  - Farticipants in NIST interlaboratory st
     Contacts through STRBase website
- Responses from <u>52 scientists</u> were compiled
  - Covering 27 states + Puerto Rico, 4 companies, 2 outside US
- Specific interviews were conducted to gain perspectives from a small lab, a large lab, a private lab, and court testimony experience

# Representative Labs Interviewed

- Montgomery County Crime Lab small lab, 3 analysts, ~180 cases/year; using PP16 and ABI 310
- Orchid Cellmark private contract lab, 40 analysts and technicians, ~5,000 cases/year; Profiler Plus/ COfiler and Identifiler with ABI 310 and ABI 3100; extensive court experience
- AFDIL large federal lab, ~120 analysts/technicians, remains identification rather than strictly forensic cases, >1,000 cases/year (mtDNA & STRs); Profiler Plus/COfiler and PP16 with ABI 377 and ABI 3100

Information from interviews is included in the written report of this project...

# Albany DNA Academy Workshop (Butler and McCord)

	Validation Standardization Questionnaire (conducted June-August 2004)
Validation Standardization Questionnaire Please return to John Butler (NIST) <u>John butler@nist.gov</u> or 301-975-8505 (fax)	Review of Survey Questions
Purpose of mustionnatre:       We are embarking on an effort to define the minimum number of samples needed to reliably validate DNA typing procedures. As part of this effort, we are conducting a survey of standard practices currently used by practitiones in forensic DNA laboratories. Your honest responses to the following quastions will help the entire commanity as we compile this information. Results will be summarized at the Promega meeting in October 2004 and made available on the NIST STRBase web site.         General Questions       What does the term validation mean to you? (define in a single semence if possible)         How do you know when you are finished validating a kit, instrument, software, or procedure?         What steps are needed in internal validation and how many samples should be run at a minimum?         Precision studies(indicate types of samples -i.e., ladders), # samples/run # runs         Misture studies	<ul> <li>What is validation?</li> <li>How do you know when you are finished validating a kit, instrument, software, or procedure?</li> <li>What steps are needed in internal validation and how many samples should be run at a minimum?</li> <li>How many total samples do you think it takes to internally "validate" a new forensic kit?</li> <li>How many different sets of samples are needed? Over what time period?</li> <li>What are some kits, software, instruments that you are considering for validation, raining, and proficiency testing related to one another?</li> <li>Do you think that the process of validation can be standardized?</li> <li>If a standard protocol or set of guidelines existed for validation, would you use it?</li> <li>Used to help define specific examples</li> </ul>



### Validation Standardization Questionnaire (conducted June-August 2004) How do you know when you are finished

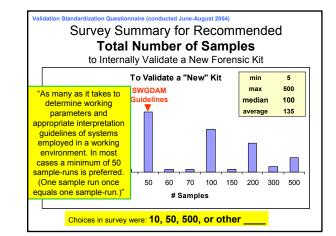
# with a validation study? (1)

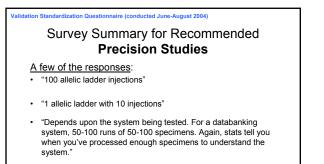
- "When you have demonstrated that it works as expected over a range of samples that is representative of what is seen in casework"
- "When repeat performance gave the same result"
- "When you pull the toothpick out and it is dry?... Meet at least minimum expectations and DAB guidelines"
- "You are very comfortable that you know how it works and your documentation will convince a reviewer you have put the kit thru a rigorous review/test."

### Validation Standardization Questionnaire (conducted June-August 2004)

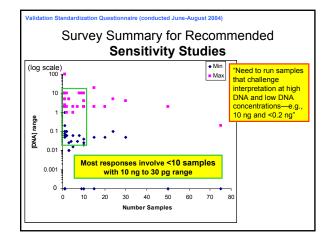
### How do you know when you are finished with a validation study? (2)

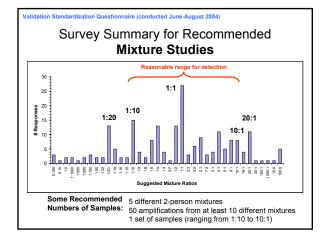
- "Once a reasonable body of data has been assembled and analyzed, quirks have been revealed, and the upper and lower limits of the system have been challenged using a range of samples that one could expect to encounter in the everyday operation of the system"
- "When you achieve accuracy and precision to the desired statistical level of certainty"
- "You can never know...but it is always nice to have more samples!"
- "Validation is never complete"





"Minimum: Run one sample at least 8 times. Recommended: Run at least two samples plus allelic ladder at least 8 times." (24 sample-runs)





### Validation Standardization Questionnaire (conducted June-August 2004)

### Survey Summary for Recommended **Non-Human Cases**

### A few of the responses:

- "10-20 food animals, companion animals, local wildlife, ferrets"
- "I don't believe this is necessary in internal validation if external results are published. This would not be expected to vary in different analysts' hands."
- "I've trusted system manufacturers to handle this. Should I have?"
- "Minimum: Include information from developmental studies. If performing developmental studies, include at least bacterial and yeast/fungal example, plus mammalian and non-mammalian examples.

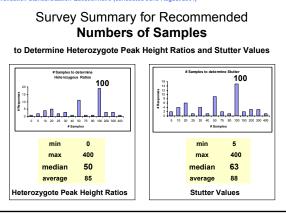
### Validation Standardization Questionnaire (conducted June-August 2004)

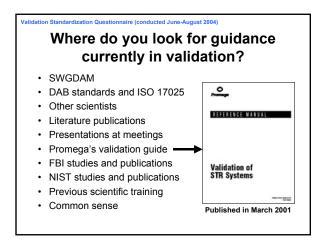
# Survey Summary for Recommended **Non-Probative Cases**

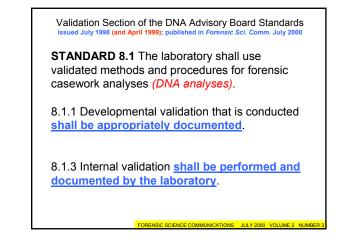
### A few of the responses:

- Most responses were between 5-10 cases (range 3-25)
- "More important than the number of cases is the range of forensic samples that are typed during validation."
- "Complete cases are not required to test a system." Recommended: Run at least 8 mock non-probative samples. Note: Non-probative samples are not guaranteed to provide complete profiles. They are needed only to show that false results are not generated. Lack of results or incomplete results do not affect the validity of a validation."

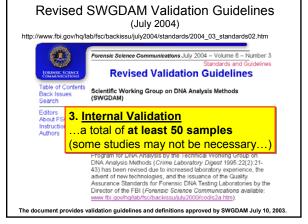
# Validation Standardization Questionnaire (conducted June-August 2004)







# SWGDAM Revised Validation Guidelines Section 1.1 Validation is the process by which the scientific community acquires the necessary information to (a) Assess the ability of a procedure to obtain reliable results. (b) Determine the conditions under which such results can be obtained. (c) Define the limitations of the procedure. The validation process identifies aspects of a procedure that are critical and must be carefully controlled and monitored. Reliability, Reproducibility, Robustness, Range



Validation Standardization Questionnaire (conducted June-August 2004)

# Can Validation be Standardized?

### Statements from survey responders...

### Over 86% (45/52) said yes

### Those who responded "no" said

- "to some degree it can be, however, validation is specific to the platform, kits,  $\ldots$  ",
- "a start-up lab should do much more than an experienced lab...",
- "validation builds on previous work by lab or published data",
- "parts of it can be standardized; I don't think the non-probative cases could be", and
- "only in a general way, as with the SWGDAM guidelines. The uniqueness of each new procedure would make standardization difficult."

### Our Conclusion...

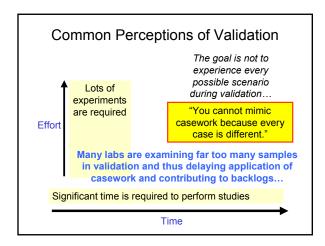
to a certain extent it can...but everyone will always have a different comfort level...and inflexible, absolute numbers for defined studies will not likely be widely accepted

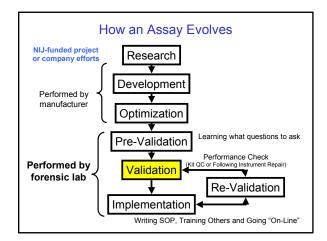
# A Thoughtful Comment from One Interviewee

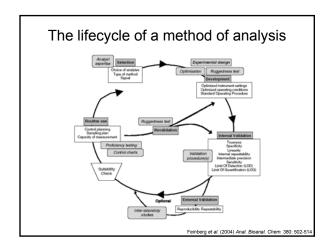
Before a set of validation experiments is performed ....

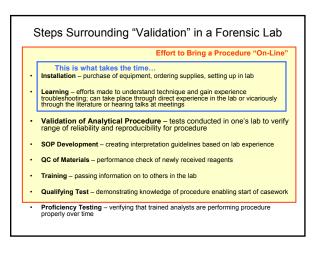
- The question should be asked "Do we already know the answer to this question from the literature or a previous study performed in-house?"
- If the answer is "yes" and we document how we know this answer, then there is no need to perform that set of validation experiments.

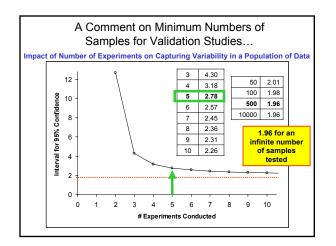
A good example of this scenario is non-human DNA studies.

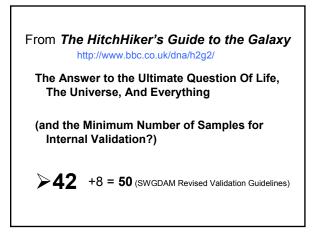


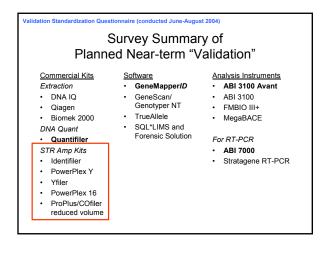












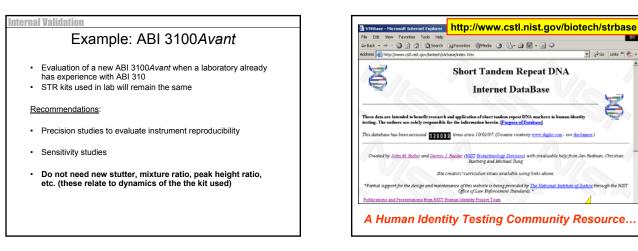


# Example: PowerPlex 16

Switch from ProfilerPlus/COfiler kits to PowerPlex 16 · Retaining same instrument platform of ABI 310

### Recommendations:

- Concordance study (somewhat, but better to review literature to see impact across a larger number of samples and which loci would be expected to exhibit allele dropout-e.g., D5S818)
- · Stutter quantities, heterozygote peak height ratio
- Some sensitivity studies and mixture ratios
- Do not need precision studies to evaluate instrument reproducibility



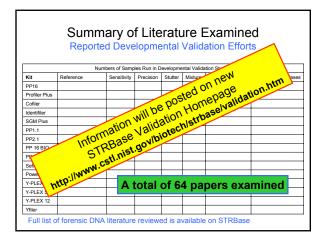
http://www	.cstl.	nist.gov/b	age on STRE iotech/strbase/v msic DNA Laborate	alidation.ht	t <mark>m </mark>	
Validation Summa			asic DNA Laborato	bries		
We are initiating an effort literature. The purpose of		Address () Mag (Sense call risk or	Addition studies that have been dated/itstat/albior/05_towPb:(7.0s	whiched in the	-	
tested, and the number of efforts by forensic DNA la SWGDAM Revised Valid documented and summar Below is listed a compila	samples boratorie ation Gui ized."	PowerPlex Y Validation Reference Poene et al (2001) State Completed Engle Source (Concerdence) Midure Ratio (mare female)	Description of Samples Tested Institution of Samples 1 Inter- 5 Later 2 Meter State Sector 2 Inter- 6 July 2 Meter Sector 2 Inter- 8 July 2 Meter Sector 2 Inter 8 July 2 Meter Sector 2 Inter 8 July 2 Meter Sector 2 In	1 100,1 300,1 1000,0 8 300,	#Ban. 40 112 112	
STR kits, in-house assays, instrum full reference bibliography is listed specific Validation Summary St		Mature Rato (male male) Sensitivity Non-Human ruti T (Hai	6 Jalos a 2 MM4 midures sames a 11 ratios (2.0,18.1,8.1,8.1,8.1,2.1,1.1,2.1.0,1.0,1.10,0.1) 7 Julio a 2 ierres a 8 amounts (10.50.250.1250.060.03) 24 animatis 6 components of DRe 2395			
Kit, Assay, or Instrument	Refer	Precision (481 3192 and 401 377) Non-Probative Cases Stutter	10 ledder replicates + 10 sample replicated + (0 lado 65 cases with 102 samples 412 makes used	How?	36 102 412	
PowerPiex Y	Brent	Peak Height Rate: Cycling Parameters	NAA (except for DY5385 but he studies were noted) 5 cycles (20/27/26/25/24) x 8 putch sizes x 2 sample		80	
Profiler Plus	Frank al. (2) Pawle	Annealing Temperature Reaction volume Thermal cyclar test Male-specificity	5 talis a 5 temperatures (34/580,082/64) a 1 sample 5 volumes (50/251512,56,26) a 35 emisurals - 5 con 4 models (40/240/460/960/9700) a 1 sample - D mo 2 females a 1 39/40/240/960/9700 a 1 sample - D Mo	deto x 3 sets x 12 samples) « 5 amounts	25 50 76 10	
COfiler	LaFo	Tagliold polymerase titration Primer pair titration Magnesium titration	5 amounts (1.392.06/2.75/3.444.13.0) x 4 quantities 5 amounts (2.5x8.25iartait.5x20 x 4 quantities (1/0.1 5 amounts (1/1.25/1.5x1.75/2.m41461 x 4 quantities (	V0 25/0 1 5 mg D1/40	20 20	
SGM Plus AmpFISTR Blue	Cotto			TOTAL SAMPLES EXAMPLES	1269	
AmpFISTR Orean I	Holt	Cammenta: Oth	er information and co	nclusions		

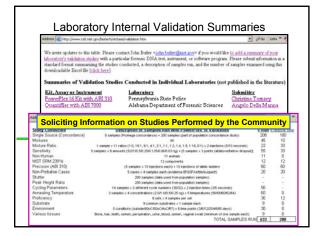
Validation	Summary Sheet for PowerPlex Y	
Study Completed (17 studies done)	Description of Samples Tested (performed in 7 labs and Promega)	# Run
Single Source (Concordance)	5 samples x 8 labs	40
Mixture Ratio (male:female)	6 labs x 2 M/F mixture series x 11 ratios (1:0,1:1,1:10,1:100,1:300,1:1000,0.5:300, 0.25:300,0.125:300, 0.0625:300, 0.03:300 ng M:F )	132
Mixture Ratio (male:male)	6 labs x 2 M/M mixtures series x 11 ratios (1:0, 19:1, 9:1, 5:1, 2:1, 1:1, 1:2, 1:5, 1:9, 1:19, 0:1)	132
Sensitivity	7 labs x 2 series x 6 amounts (1/0.5/0.25/0.125/0.06/0.03)	84
Non-Human	24 animals	24
NIST SRM	6 components of SRM 2395	6
Precision (ABI 3100 and ABI 377)	10 ladder replicates + 10 sample replicated + [8 ladders + 8 samples for 377]	36
Non-Probative Cases	65 cases with 102 samples	102
Stutter	412 males used	412
Peak Height Ratio	N/A (except for DYS385 but no studies were noted)	
Cycling Parameters	5 cycles (28/27/26/25/24) x 8 punch sizes x 2 samples	80
Annealing Temperature	5 labs x 5 temperatures (54/58/60/62/64) x 1 sample	25
Reaction volume	5 volumes (50/25/15/12.5/6.25) x [5 amounts + 5 concentrations]	50
Thermal cycler test	4 models (480/2400/9600/9700) x 1 sample + [3 models x 3 sets x 12 samples]	76
Male-specificity	2 females x 1 titration series (0-500 ng female DNA) x 5 amounts each	10
TaqGold polymerase titration	5 amounts (1.38/2.06/2.75/3.44/4.13 U) x 4 quantities (1/0.5/0.25/0.13 ng DNA)	20
Primer pair titration	5 amounts (0.5x/0.75x/1x/1.5x/2x) x 4 quantities (1/0.5/0.25/0.13 ng DNA)	20
Magnesium titration	5 amounts (1/1.25/1.5/1.75/2 mM Mg) x 4 quantities (1/0.5/0.25/0.13 ng DNA)	20
Krenke et al. (2005) Forensic	Sci. Int. 148:1-14 TOTAL SAMPLES EXAMINED	1269

24F

ble help from Jan Bedman, Christian

of Justice through the MIST

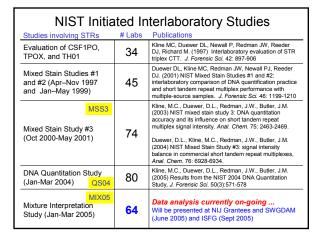


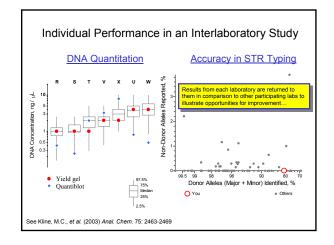


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# Interlaboratory Studies

DNA Quantitation (2004), Mixture Interpretation (2005)

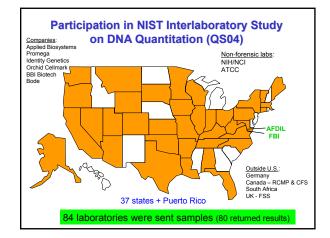


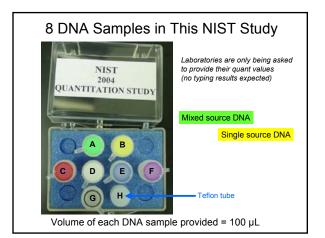


# NIST Quantitation Study 2004 (QS04) <u>Consisted of</u>: •8 DNA extracts labeled A – H •Shipped Dec 2003 – Jan 2004 to 84 laboratories for quantification; data received back by April 2004 •Labs were requested to use multiple methods / multiple analysts We received data from 80 Labs (95%) Total of 287 sets of data Portionets were figure to use tigents

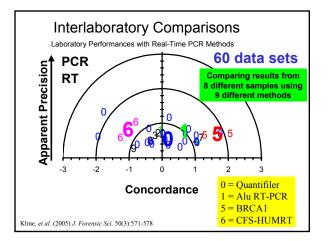
Participants used 19 different quantification methods (primarily variations on Quantiblot and Real-time PCR)

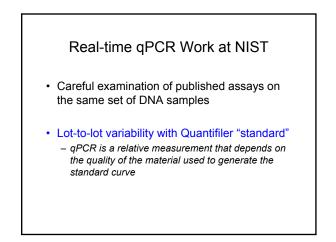
Information from this interlab study is being used to help construct SRM 2372 (Human DNA Quantitation Standard)





					Vantitat					
Target [D	NA] ng/µL	1.5	0.5	0.5	0.16	0.16	0.05	0.05	0.05	
Method	New	A	В	E	С	F	D	G	Н	
Quantifiler	37	100	100	100	100	100	100	100	100	
Other RT-PCR	23	100	100	100	100	100	100	100	100	
"ACES"	14	100	100	100	100	100	100	100	100	
AluQuant	13	100	100	100	100	100	100	100	100	
PicoGreen	12	100	100	92	100	100	92	83	83	
ECL	75	100	99	99	93	95	84	77	87	
TMB	98	100	100	99	93	94	59	62	63	
Yield gel	14	57	0	0	0	0	0	0	0	
	286									r performanc v level sampl
Quantitative result between contiguor standard if smaller calibration standar	is calibratio than the ta	n stand rget [D	ards, va NA], or	lues rej values	ralue: to porter in	o justii nstrum	fy purc	hase ( on and	of qPC	R ersion to





Sample (n = 4)	Standard Lot 1 (ng/mL)	Standard Lot 2 (ng/mL)			
1	4*	2.91 ± 0.04			
2	7.26 ± 0.79	4*			
3	2.93 ± 0.27	1.88 ± 0.09			
4	3.46 ± 0.30	2.22 ± 0.08			
5	2.99 ± 0.28	1.91 ± 0.08			
6	2.62 ± 0.22	1.70 ± 0.03			

# Mixture Interpretation Interlab Study (MIX05)

- Only involves interpretation of data
- 91 labs enrolled for participation (20 from overseas)
- 64 labs have returned results
- Four mock cases supplied with "victim" and "evidence" electropherograms (GeneScan .fsa files – that can be converted for Mac or GeneMapper; gel files made available to FMBIO labs)
- Data available with Profiler Plus, COffier, SGM Plus, PowerPlex 16, Identifiler, PowerPlex 16 BIO (FMBIO) kits
- Summary of results will involve training materials to illustrate various approaches to solving mixtures

