

Charge-associated effects of fullerene derivatives on microbial structure integrity and central metabolism

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Abstract

The effects of four types of fullerene compounds (C₆₀, C₆₀-OH, C₆₀-COOH, C₆₀-NH₂) were examined on two model microorganisms (Escherichia coli W3110 and Shewanella oneidensis MR-1). Positively charged C60-NH2 at concentrations as low as 10 mg/L inhibited growth and reduced substrate uptake for both microorganisms. Scanning Electron Microscopy (SEM) revealed damage to cellular structures. Neutrally-charged C_{60} and C_{60} -OH had mild negative effects on S. oneidensis MR-1, whereas the negatively-charged C60-COOH did not affect either microorganism's growth. The effect of fullerene compounds on global metabolism was further investigated using [3-13C]L-lactate isotopic labeling, which tracks perturbations to metabolic reaction rates in bacteria by examining the change in the isotopic labeling pattern in the resulting metabolites. The 13C isotopomer analysis from all fullerene-exposed cultures revealed no significant differences in isotopomer distributions from unstressed cells. This result indicates that microbial central metabolism is robust to environmental stress inflicted by fullerene nanoparticles. In addition, although C60-NH2 compounds caused mechanical stress on the cell wall or membrane, both S. oneidensis MR-1 and E. coli W3110 can efficiently alleviate such stress by cell aggregation and precipitation of the toxic nanoparticles. The results presented here hypotheses that fullerenes cause more membrane stress than perturbation to energy metabolism.

Methods and materials

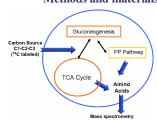


Figure 1.Schematics for the isotopic approach of investigating cellular global metabolism via 13C labeling pattern in amino acids. Labeling patterns of metabolites were used to evaluate the perturbation of central

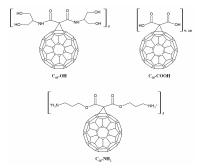


Figure 2, Co. fullerene derivatives used in this study. The fullerene-COOH-derivative carries negative charges in solution, the fullerene-NH3*-derivative is positively charged. The C60-serinol (-OH) is neutral.

Experimental Results and discussions

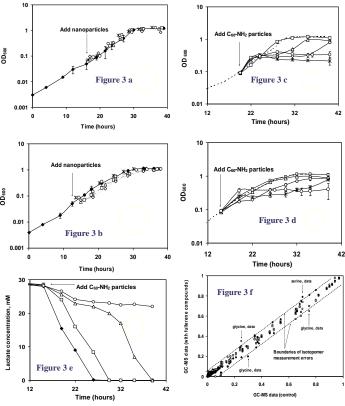


Figure 3. The effect of C₆₀, C₆₀-OH, C₆₀-COOH fullerene compounds (neutral or anionic charge) on *E. coli* W3110 (Figure 3 a) and S. oneidensis MR-1 (Figure 3 b) growth. •, control, 0 mg/l; \Box , C_{go} : 20 mg/l; Δ , C_{go} : OH, 80 mg/l; * , C_{go} : COOH, 20 mg/l; \circ , C_{go} : OOH, 80 mg/l; * , C_{go} : OGOH, 20 mg/l; \circ , C_{go} : OOH, 80 mg/l; \circ , O: OOH, 80 mg/l; \circ , 1 mg/l; Δ, 10 mg/l; ◊, 20 mg/l; *, 40 mg/l; ∘, 80 mg/l. (c) ♦, 0 mg/l; □, 20 mg/l; Δ, 40 mg/l; ∘, 80 mg/l. Isotopome distribution in proteogenic amino acids of S. oneidensis MR-1 cultured in [3-13C] L-lactate medium (Figure 3f). The GC-MS data include 14 amino acids ((M57)+ and (M159)+ mass values) for all three nanoparticle-stressed experiments (o 80 mg/L C₈₀-OH; □ 80 mg/L C₈₀-COOH; ♦ 20 mg/L C₈₀-NH₂).

Conclusion

- 1. Positive charged nanoparticles are more toxic.
- 2. Central metabolism is not altered by presence of nanoparticles.
- 3. Bacteria are able to remediate toxic nanoparticles.

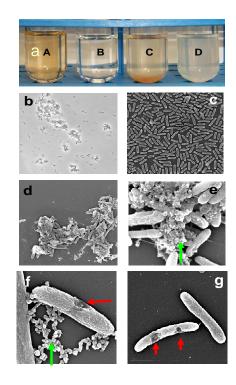


Figure 5. Images of S. oneidensis MR-1 exposed to C₆₀-NH₂. (a) Precipitation of S. oneidensis MR-1 with nanoparticles (A: no cells, C₆₀ NH₂; B: no cells, no nanoparticles; C: MR-1 cells at OD₆₀₀ 0.26, C₆₀ NH₂ added; D: MR-1 at OD₆₀₀ 0.26, no nanoparticles). (b) Light microscopy showed cell aggregation. (c) SEM of S. oneidensis MR-1 (no effect from NPs). (d) SEM of MR-1 aggregation in the presence of C₆₀-NH₂. (e) SEM of MR-1 in the presence of C₆₀-NH₂ (green arrow points to nanoparticles). (f) SEM of MR-1 in the presence of Co-NH_o aggregation (red arrow points to the damaged part of the cell), (g) SEM of individual MR-1 cells (red arrow points to the damaged part of the cell)

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