

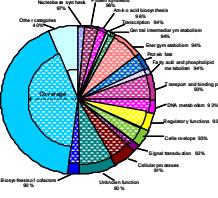
PNNL Protein Complex Characterization Efforts

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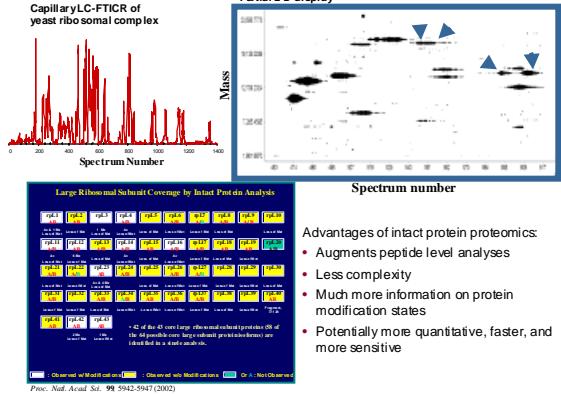
Identifying protein complexes using AMT tags

The database of *Shewanella* accurate mass and time (AMT) tags provides the basis for high throughput characterization of protein complexes at either the peptide level or intact protein level. The intact protein level analysis is enabled by the peptide level approach (by providing their initial identifications) and also complements the information obtainable at the peptide level with additional information on protein modifications (e.g., chemical modifications, protein truncation). Regardless, the use of UMT tags can greatly speed the analysis and potentially allows characterization times of <5 minutes per complex. Proteome coverage by AMT tags is indicated by the pie chart (right).



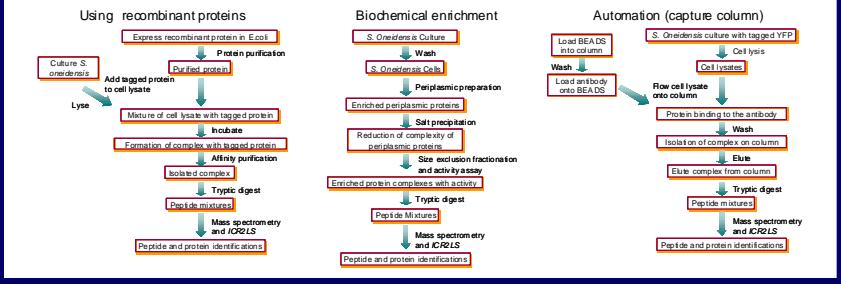
Value-added protein complex characterization at the intact protein level – initial demonstration

To evaluate our approach, we have initially studied the well characterized yeast large ribosomal subunit. The 43 proteins in this complex were previously identified at the peptide level using tandem MS, providing an expected set of tentative molecular weights. The constrained level of complexity associated with most protein complexes (ignoring obviously low-level contaminants) allows the detected masses to be assigned to the various proteins, as well as (in most cases) assignment of their modification states.

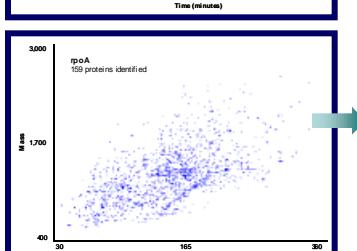
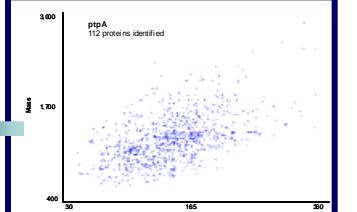


- Augments peptide level analyses
- Less complexity
- Much more information on protein modification states
- Potentially more quantitative, faster, and more sensitive

Complex generation and isolation



Initial results



Higher characterization throughput at lower cost

A component of the PNNL program is to develop an approach that provides both increased confidence, higher throughput, and a quantitative tool for characterizing protein complexes. We have initially explored the utility of characterizing protein complexes at the peptide level using AMT tags with Q-TOF instrumentation as an alternative to much less sensitive and lower throughput approaches based upon tandem MS (e.g., using ion trap mass spectrometers) or more expensive FTICR instrumentation that is needed for much more demanding "whole proteome" analyses.

This initial evaluation examined the highly active oxygen-evolving photosystem II (PSII) complex purified from the HT-3 strain of the cyanobacterium *Synechocystis* sp. PCC 6803. This initial study used high pressure capillary LC-Q-TOF instrumentation and used AMT tags that were generated from capillary LC-MS/MS analyses. The table below shows a partial listing of >120 proteins that were identified, along with a measure of their relative abundances based upon the integrated peak intensities for the corresponding peptides. The proteins highlighted in yellow were previously identified by Pakzad and coworkers (Kashino et al., *Biochemistry* 2002, 41, 8004 – 8012). All proteins that were assigned based upon more than one peptide are done so with very high confidence.

These results show:

- Q-TOF instrumentation augmented by the use of LC elution time information provides sufficient specificity for application of AMT tag approaches.
 - The AMT tag approach with LC-Q-TOF analysis provides sufficient specificity for protein complex characterization, along with high throughput, and preliminary quantitation.
 - The use of quantitative information and multiple analyses (e.g., using different wash conditions) will be needed with this approach to better qualify which proteins are part of the complex in contrast to being non-specifically associated.

Tagged proteins generated to date for pull-down studies at PNNL

Gene	Description	SGO#
a	periplasmic Fc hydrogengase small subunit	S00120
1A	periplasmic Fc hydrogengase large subunit	S00160
pA	periplasmic nitrate reductase	S00048
deoC	deoxycholate operon	S00179
dch	dechymate cytochrome C	S01178
Quinone-reducing NADH:quinone-oxidoreductase small subunit precursor	QDR	S02099
QDR	ubiquinol:cytochrome c reductase iron-sulfur protein	S00068
Baclovinocyte C-like subunit	BAC	S00301
Glyceraldehyde-3-phosphate dehydrogenase	GAPD	S03120
oxidative stress molybdate-dependent	MSOB	S00043
formate dehydrogenase	FDH	S00048
aliphatic phosphate propanoate phosphatase	S02098	
Tyrosine-specific protein phosphatase	TPP	S00124
Signal transduction histidine kinase	STK	S00476
methionine sulfoxide reductase (S-form A)	MsrA	S02337
methionine sulfoxide reductase (S-form B)	MsrB	S02588
RNA polymerase sigma factor	RpoS	S00140
ATP-dependent RNA helicase	DnaK	S00407
RNA polymerase sigma-70 factor	RpoD	S01284
Chaperone GroEL	GroEL	S00027
DNA-directed RNA polymerase alpha subunit	DnaG	S00250
DNA-directed RNA polymerase omega subunit	DnaG	S00360
RNA polymerase-associated protein	PNPB	S00056

Description	MW (kDa)	Average Abundance
rpoA	26	8.63%