

Abstract: Estimating the magnitude of ammonia (NH₃) volatilization loss under different conditions is important for proper fertilizer management. Both of the current field techniques used to trap volatilized NH₃ from soils amended with various N sources, and the techniques for the quantitative determination of NH₃. drain substantial, financial resources. This justifies re-evaluating existing popular methods, in order to develop a cost-effective field and laboratory method, to trap and determine volatilized NH₃ quantitatively and accurately. This study is designed to evaluate the effectiveness of (i) two absorbents (Phosphoric vs. Oxalic) in trapping volatilized NH., (ii) two extractants (KCl vs. deionized water) in removing absorbed NH., and (iii) two analytical procedures (Flow Injection Auto Analyzer vs. Ion Chromatography) in determining the concentration of NH, in extracts quantitatively, in most widely used two-passive flux methods. This evaluation was repeated under laboratory conditions, with two soils (light-textured Candler fine sand and medium-textured Ogeechee loamy sand) amended with various fertilizer N sources (Urea, Ammonium sulfate, Ammonium nitrate, Ammonium chloride along with an unamended control) and manure N sources (Chicken, Cattle, and Swine manures along with an unamended control) to identify the best combination of procedures to develop a cost effective method for determining volatilized NH₂. This presentation focuses on step-by-step evaluations of results obtained from several laboratory studies, so as to develop an effective method for field and laboratory scale determination of volatilized NH₃. Per our preliminary evaluation, the best combination of procedures would be to use 3% oxalic acid in acetone, to trap NH₃; deionized water, to extract the trapped NH₃; and Ion chromatography, to determine the concentration of NH₄. Nin extract. This promises to reduce chemicals, cost, and labor, without sacrificing accuracy of measurement of volatilized NH3.

Introduction:

- @ Estimating the magnitude of ammonia (NH₂) volatilization losses under different conditions is important for proper fertilizer management
- @ Current field techniques to trap volatilized NH, from soils amended with various N sources and techniques used for the quantitative determination of NH3 drain substantial financial resources
- @ This justifies the need to re-evaluate current methods, prior to the development of cost effective field and laboratory methods to trap and determine the volatilized NH2 quantitatively and accurately.
- @ Therefore, the current study was designed with the following objectives.

Objectives

Soils:

To evaluate the effectiveness of the most commonly used

(i) two absorbents (Phosphoric and Oxalic) in trapping volatilized NH.

(ii) two extractants (KCl vs. H₂O) in removing absorbed NH₃ and

(iii) two analytical techniques (Auto Analyzer vs. Ion chromatography) in determining the concentration of NH, in extracts quantitatively in most widely used passive flux methods

Materials and Methods Used:

1) Candler fine sand (pH = 6.8) - Lake Alfred, FL. [CFS]

2) Ogeechee loamy sand (pH = 5.2) - Savannah, GA. [OLS]

A. Treatment set up with inorganic fertilizer N sources nd rates (0 & 200 kg ha-1)

| N sources 1 | reatment ID | Amt of N (mg) |
|--|--|----------------------|
| Control | [T1] | 00.0 |
| Urea | [T2] | 57.0 |
| Ammonium sulfa | te [T3] | 42.0 |
| Ammonium nitra | te [T4] | 25.4 |
| Ammonium chlor | ide [T5] | 34.0 |
| B. Treatment set u rates (0, & 11.2 M | 1p with organic n g ha ⁻¹): | nanure N sources and |
| Control [T1]; Chic [T3]; and Swine m | cken manure [T2 anure [T4]. |]; Cattle manure |

Set up of Incubation Study to Measure NH₂ Emission:

Cumulative emission loss from CFS

12 15 18 21 24 0 3 6 9 12 15 18 21 24

9 12 15 18 21 24 0 3 6 9 12 15 18 21 24

Duration of experiment (days)

Duration of experiment (days)

KCl Extract

Water Extract



Incubation study was initiated by placing 100 g of Candler fine sand (pH = 6.8) from Lake Alfred, Florida, and Ogeechee loamy sand from Savannah, Georgia (pH =5.2), in 5cm diameter (1 L) mason jars. Soil samples in mason jars were amended on area basis with appropriate amounts various inorganic N sources to achieve amendments at the application rate of 0. 200 kg ha⁻¹ in experiment 1. Experiment 2 was conducted with animal manures (chicken, cattle, and swine) at the application rate of 0, 11.2 Mg ha⁻¹.

Soils and respective amendments were mixed thoroughly and incubated at their respective field capacity level moisture contents (7 and 14% for Florida and Georgia soil respectively) through-out the study period. All treatments were triplicated.

NILN NILC

9 12 15 18 21 24

Preparation of Acidified Sponge to Capture volatilized NH₃:

1a. Phosphoric-Glycerol: Sponges were made out by material used for seat-cushions, which were purchased from Jo Ann Fabrics in Savannah, GA, and cut into 2.5-cm diameter circles with an electric knife. The sponges were washed thoroughly in de-ionized water prior to transforming them into acidified sponges, to be used to capture emitted NH, from PL amended soils in mason jars. Acidified sponges were prepared by soaking and squeezing the sponges in various solutions in the following order: 1M H-PO., 1M KOH, de-ionized water. and a mixture of IM H,PO, and Glycerol. The sponges were treated with first three solutions and squeezed thoroughly prior to treating them with a mixture of 1M H₂PO₄ and Glycerol. During the final stage of sponge preparation, the sponges were allowed to retain at least 3-5 mL of mixture of 1M H.PO4 and Glycerol to facilitate the capture of emitted NH4 from the surface applied inorganic N sources.

1b. 3% Oxalic acid in Acetone: Acidified sponges were prepared by soaking and squeezing the sponges in various solutions in the following order: De-ionized water, and 3% oxalic acid in acetone. During the final stage of sponge preparation, the sponges were allowed to retain at least about 3-5 mL of 3% Oxalic-acetone solution to facilitate the capture of emitted NH, from the surface applied inorganic N sources

Extraction of NH₄-N from Sponge: Sponges were extracted individually with 40 mL of either 2M KCl or de-ionized water and stored for determination of NH4-N in the extract.

Measurement of NH₄-N in Extracts: Ammonium-N in KCl extract was measured by Flow Injection Analyzer (FIA) and Ammonium-N in de-ionized water by both Ion Chromatography and FIA.

