

## **Last time:**

- Pre-genetics studies of development
- Advantages of genetic approach
- Goals of a developmental geneticist
  1. Which genes
  2. Where and when are they expressed?
  3. How are they controlled?
  4. What do they do?
- Drosophila:
  - Hox genes
  - Segmentation genes

## **Today:**

1. Genetic screens and genetic analysis of development (*Drosophila* techniques)
2. Regulatory networks in development & cis-regulatory control of expression
3. Post-transcriptional regulation, pleiotropy & disease in development

**The goal of a genetic screen:** Identify genes specifically involved in a certain biological process.

**Developmental genetic screens:**

- Loss of function mutations are helpful because they often have a mutant phenotype in the cells where the WT gene is functioning. These are usually recessive.
- Gain of function mutations often hard to interpret but can be distinguished from loss of function mutations because they are usually dominant.

## **Components of a screen:**

1. Saturation mutagenesis
2. Isolate mutants and make stable lines
3. Perform complementation analysis on different lines with similar phenotypes
4. Further characterize

**Q:** After you mutagenize an individual, how do you isolate the specific kind of mutants you want and make stable lines of them?

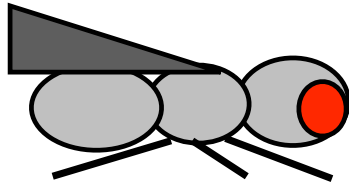
**Ans:** By being a clever geneticist or doing a lot of work and often both.

Each organism has its own set of visible and selectable markers as well as a host of other genetic tools to help one design a good screen.

## **Balancer chromosomes in Drosophila**

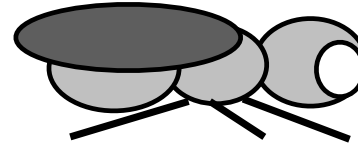
- been made for all chromosomes
- lots of large overlapping inversions prevent them from producing viable offspring if crossing over with a homologous chromosome occurs
- dominant visible marker (e.g. *Cy* curly wings)
- recessive lethal (e.g. *Cy*)

P



Bal/+

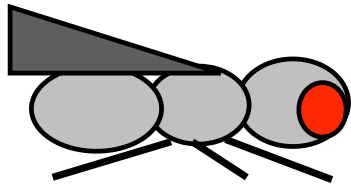
x



a w/a w

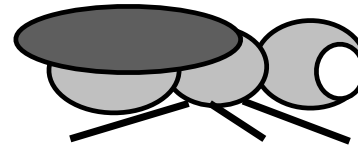
Chemical  
treatment of  
sperm

P



Bal/+

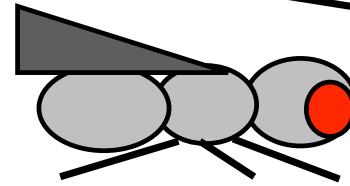
x



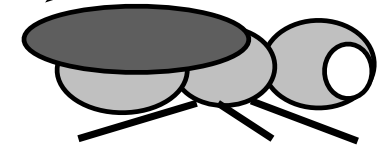
a w/a w

Chemical  
treatment of  
sperm

FI



a' w/Bal



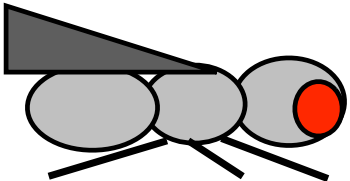
a' w/+





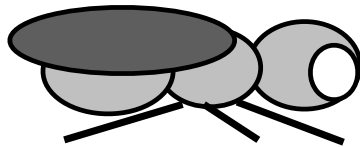
Chemical  
treatment of  
sperm

P



Bal/+

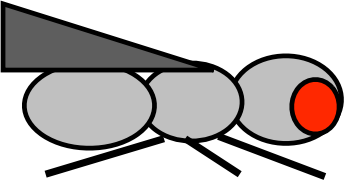
x



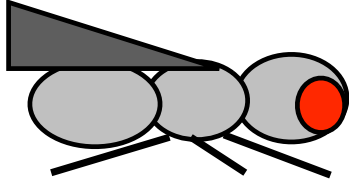
a w/a w

x

F1

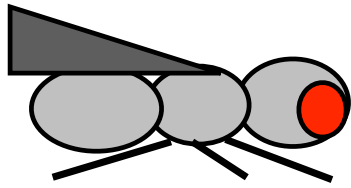


Bal/+



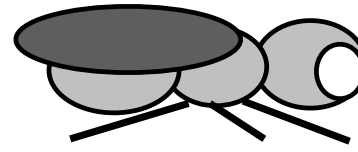
a' w/Bal

P



Bal/+

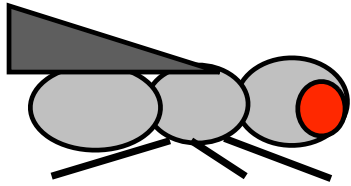
x



a w/a w

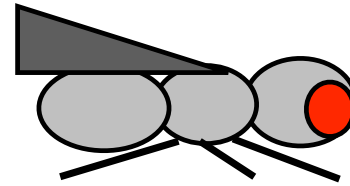
Chemical  
treatment of  
sperm

F1



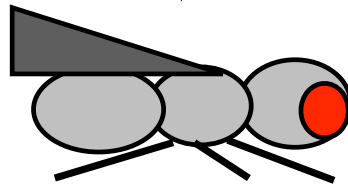
Bal/+

x



a' w/Bal

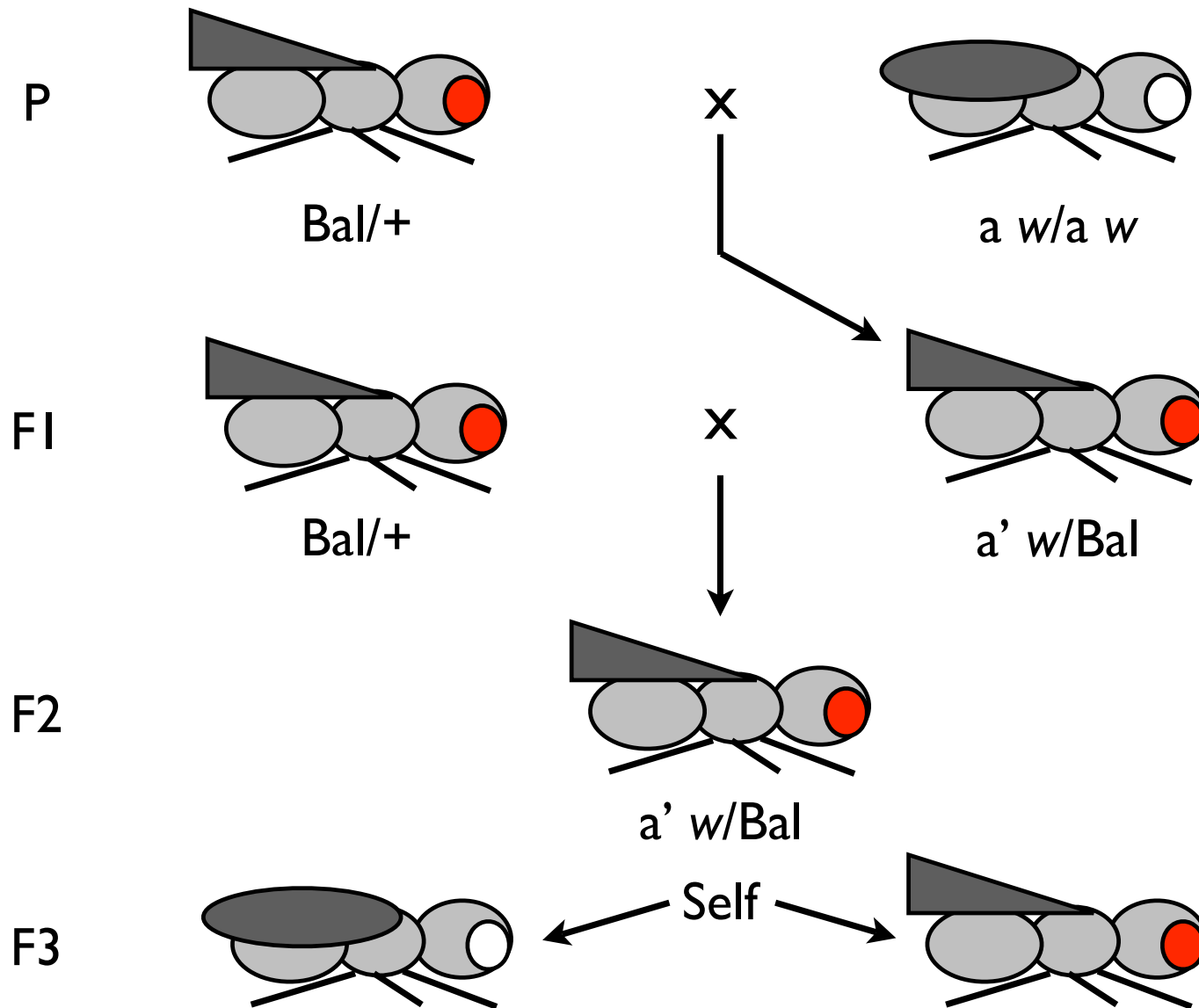
F2



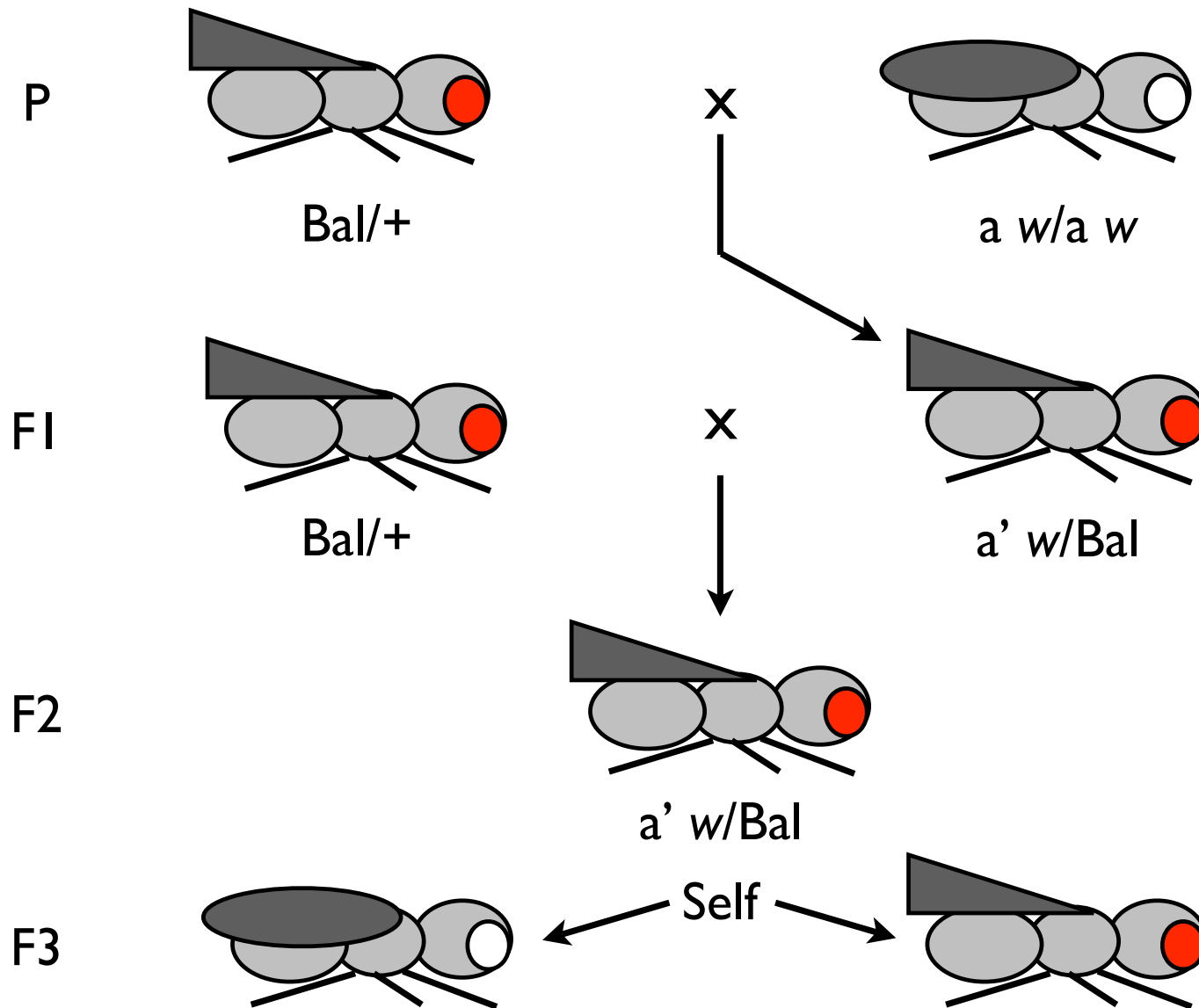
a' w/Bal

Balancer chrom has a recessive  
lethal gene so Bal/Bal always die

Chemical  
treatment of  
sperm

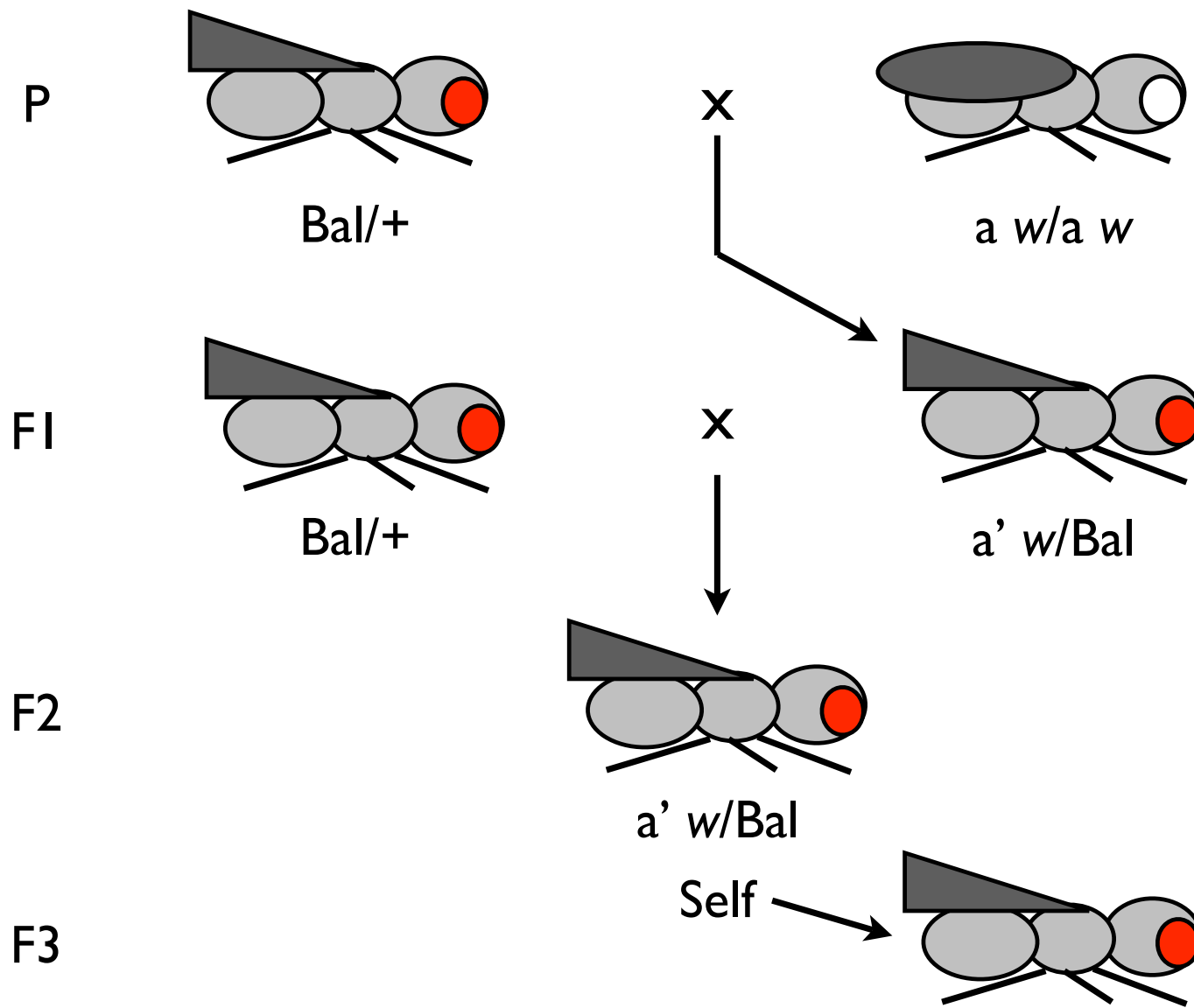


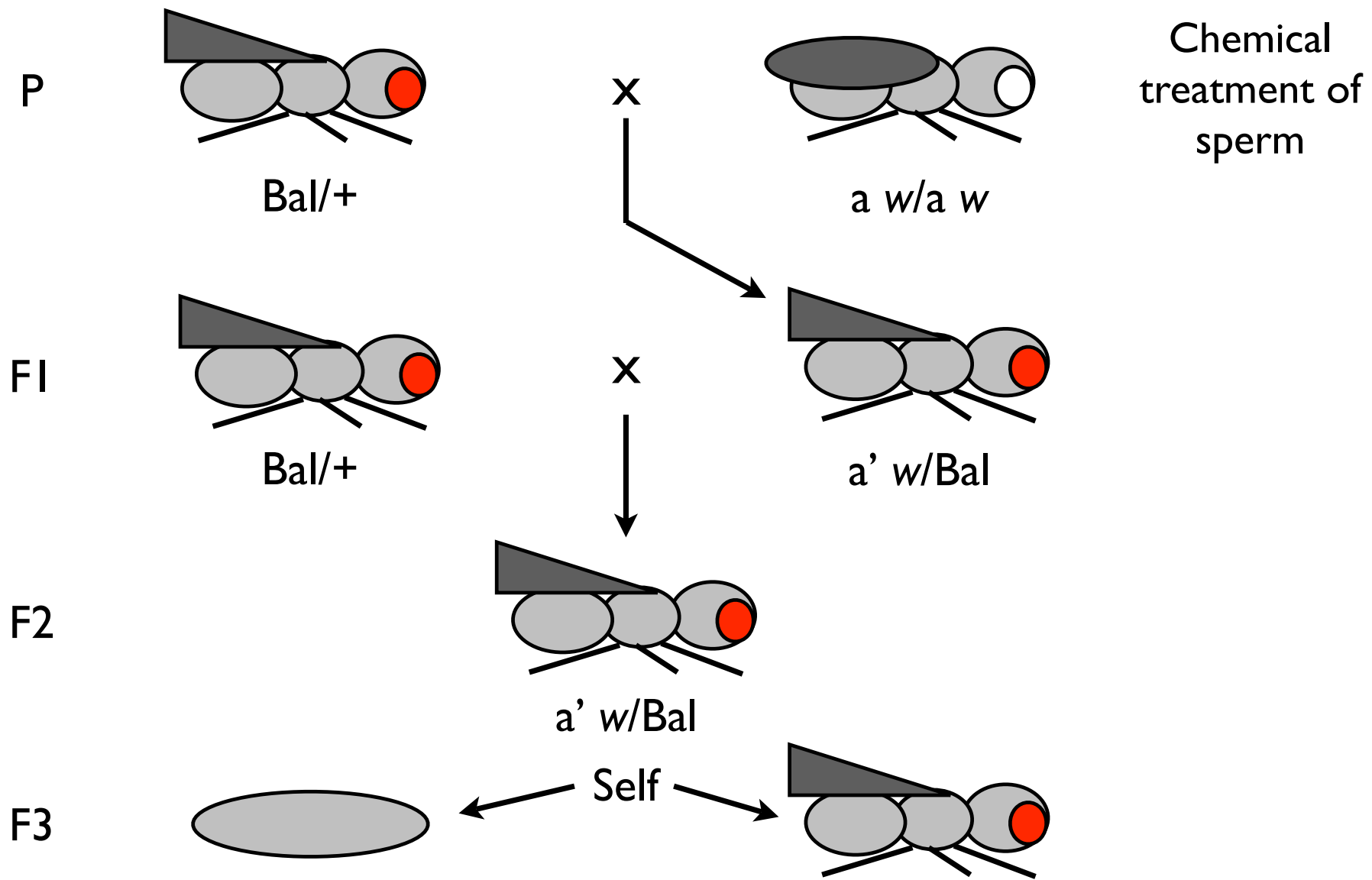
Chemical  
treatment of  
sperm



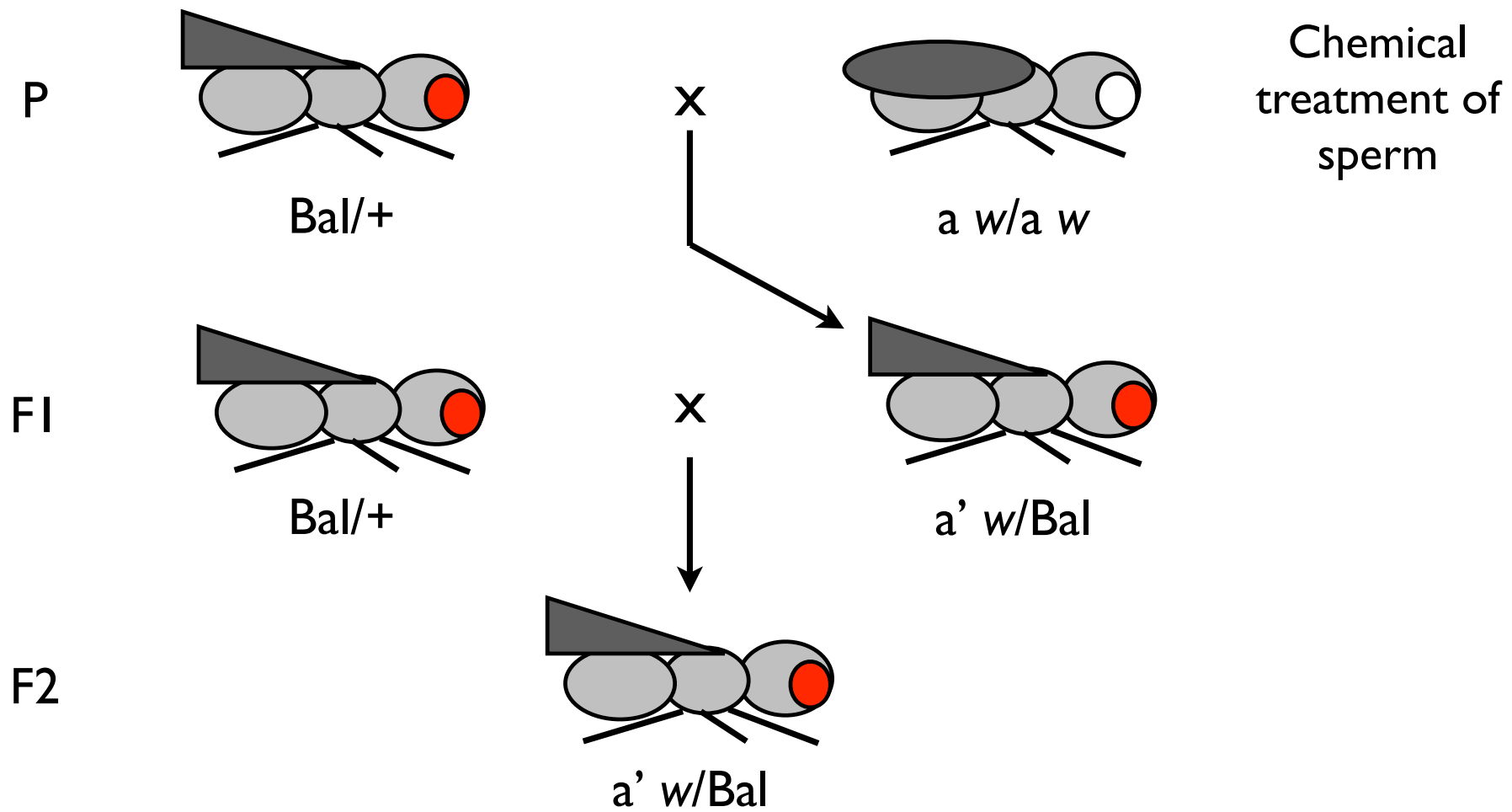
If you get any white eyed F3s it means  
no mutation or a' not recessive lethal

Chemical  
treatment of  
sperm





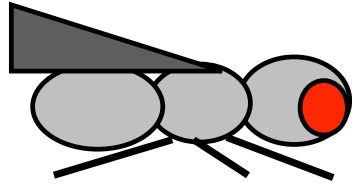
Only red eyed flies means you have a recessive lethal.  
 Next step is to examine embryos for phenotype.



Bonus is that a' w/Bal can be maintained as a stable non-recombining stock that will immediately reveal contamination and is suitable for many useful crosses.

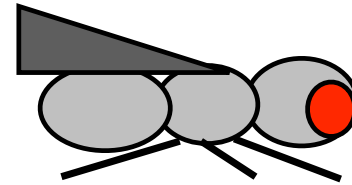
# Complementation test with balancers

P



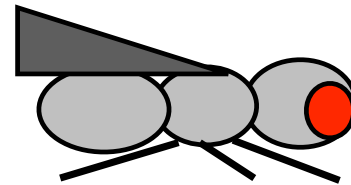
a' w/Bal

x



b' w/Bal

a' w/b' w?



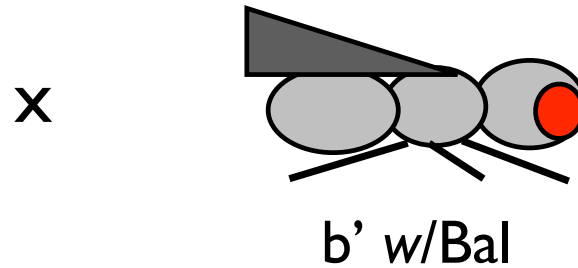
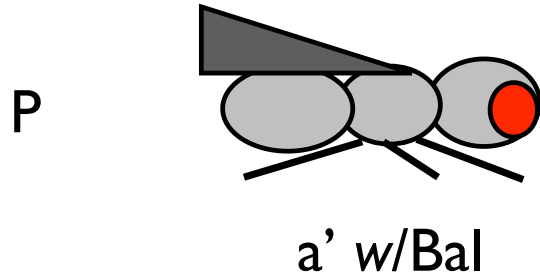
a' w/Bal or b' w/Bal

Dead

Bal/Bal



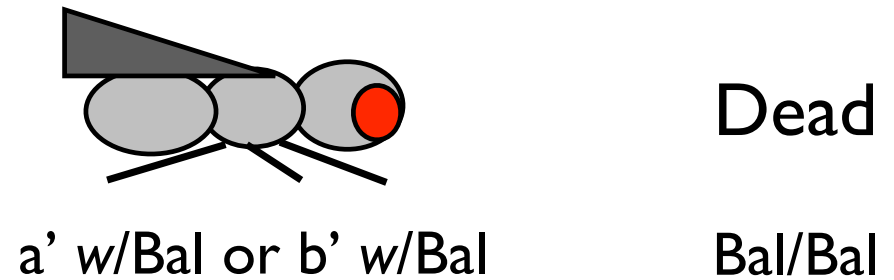
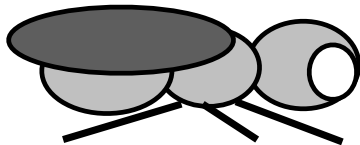
# Complementation test with balancers



a' w/b' w?

If allelic would die

If complement:



**Q:** Why might a genetic screen fail to find some genes involved in a given biological process?

**Q:** Why might a genetic screen fail to find some genes involved in a given biological process?

1. Lethality before time of selection (Christine Nusslein-Volhard & Eric Wieschaus' screen was really cool because it was 'looking' for embryonic lethality)
2. Redundancy masking mutant phenotype

## **Today:**

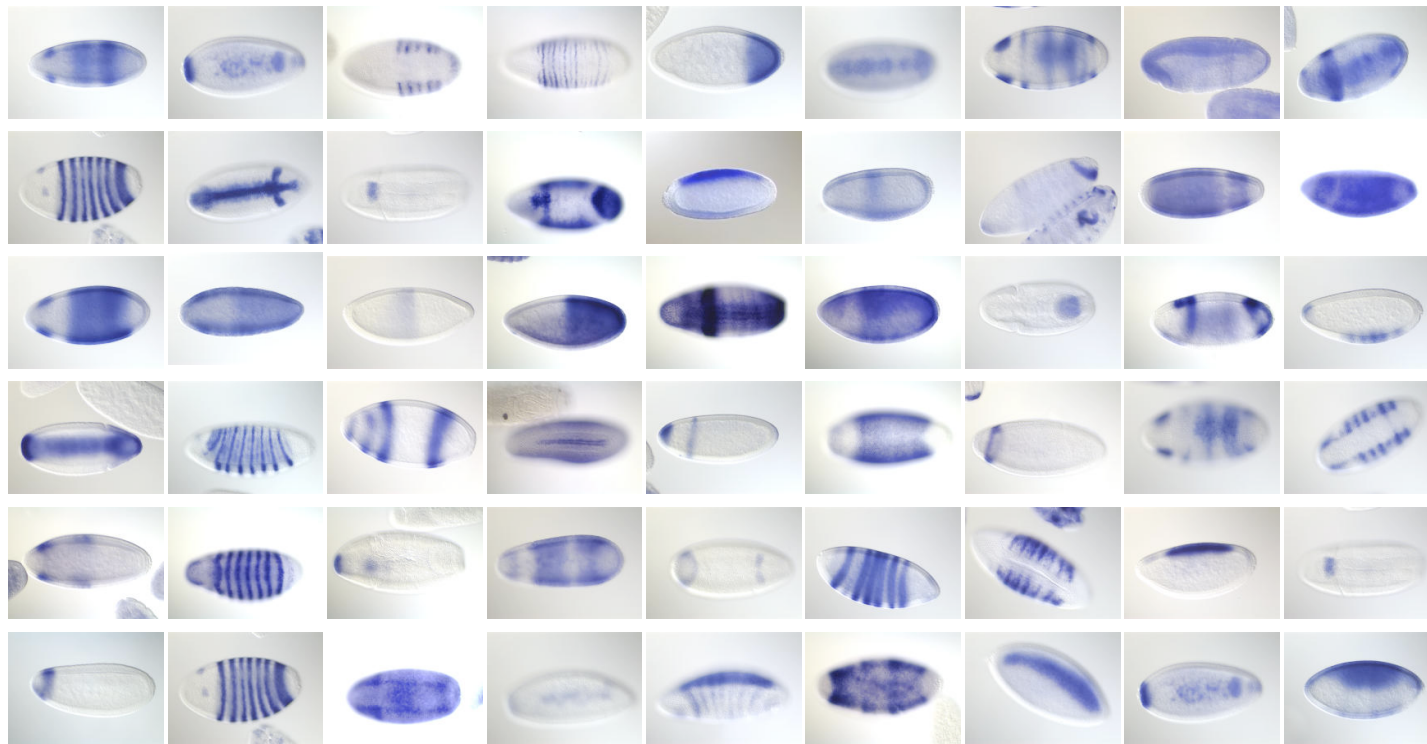
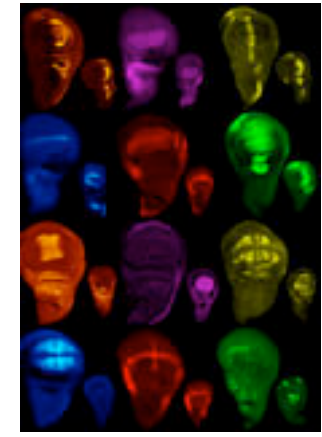
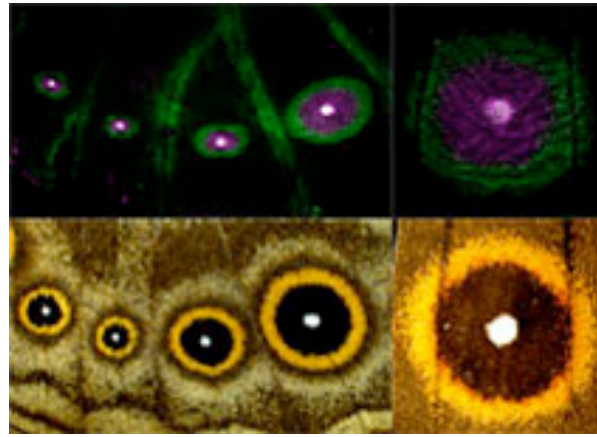
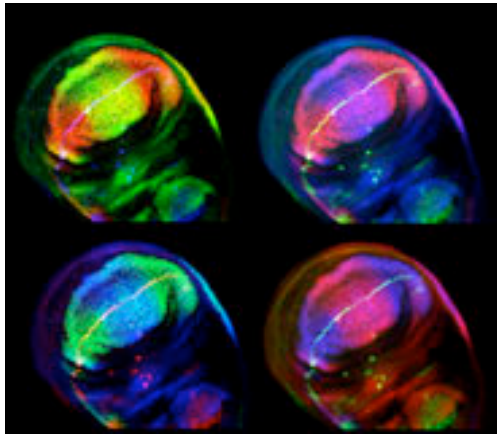
1. Genetic screens and genetic analysis of development (Drosophila techniques)
2. Regulatory networks in development & cis-regulatory control of expression
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**Review question:** How do you assay which cells in a developing organism are expressing a gene of interest?

Visualize using microscopy:

For mRNA use in situ hybridization

For protein use immunohistochemistry



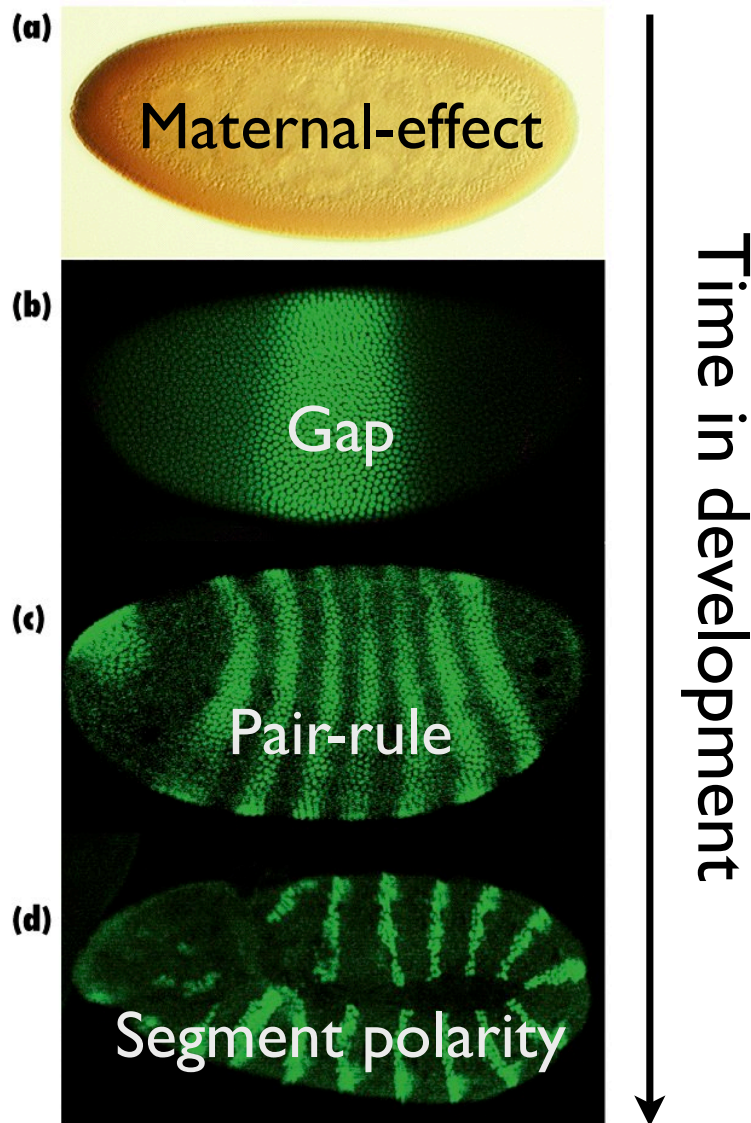


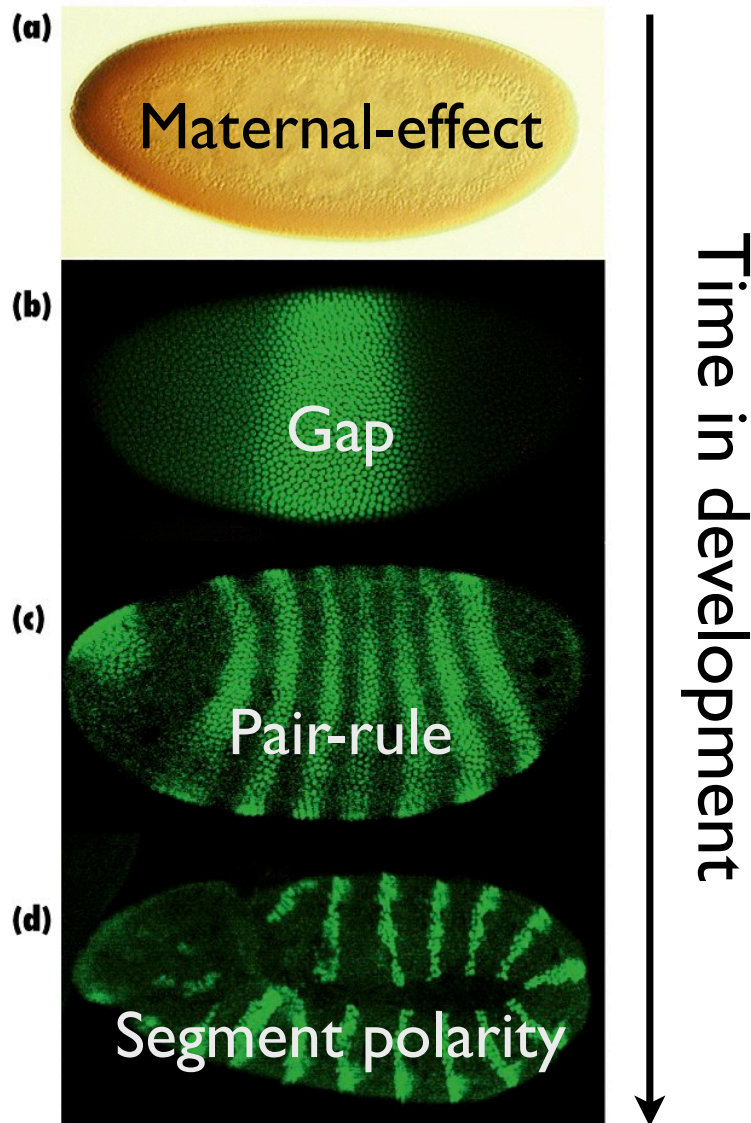
Figure 12-15  
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## From last time:

Expression patterns of four classes of genes found in screen for genes affecting segmentation

Most of these genes were found to be transcription factors

Appears to be a network where earlier genes affect later genes



How does a simple gradient lead to more complex expression patterns through transcriptional regulation?

Bicoid is the first expression pattern to appear with a gradient of high anterior to low posterior expression.

How can a downstream gene 'read' the bicoid gradient?

Figure 12-15  
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# Hunchback 'reads' the Bicoid gradient

The concentration dependent response of hunchback to the graded input of Bicoid gradient is achieved through multiple Bicoid binding sites in a 5' cis-acting regulatory element.

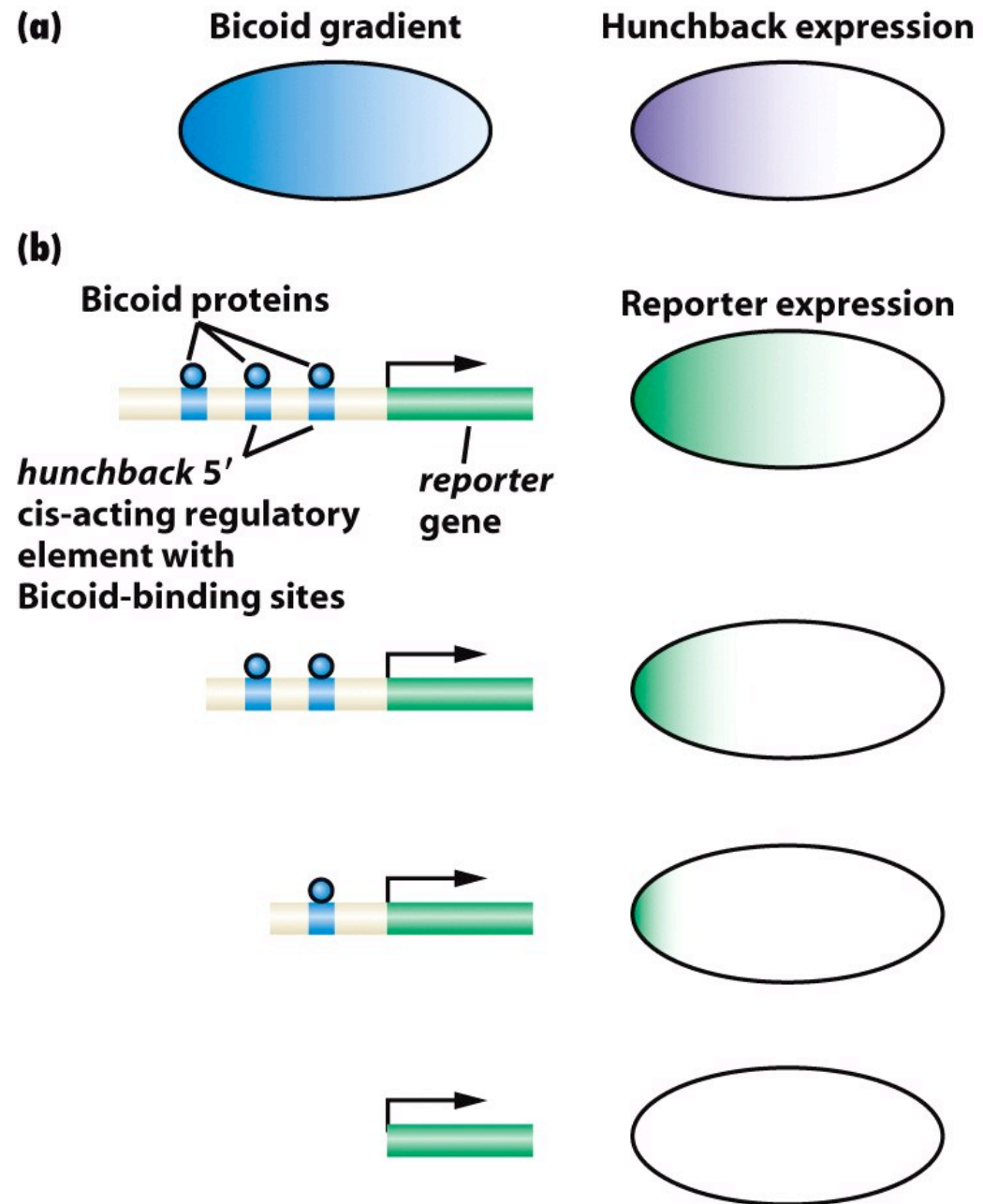


Figure 12-18  
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# Reporter gene assays allow dissection of independent regulator elements

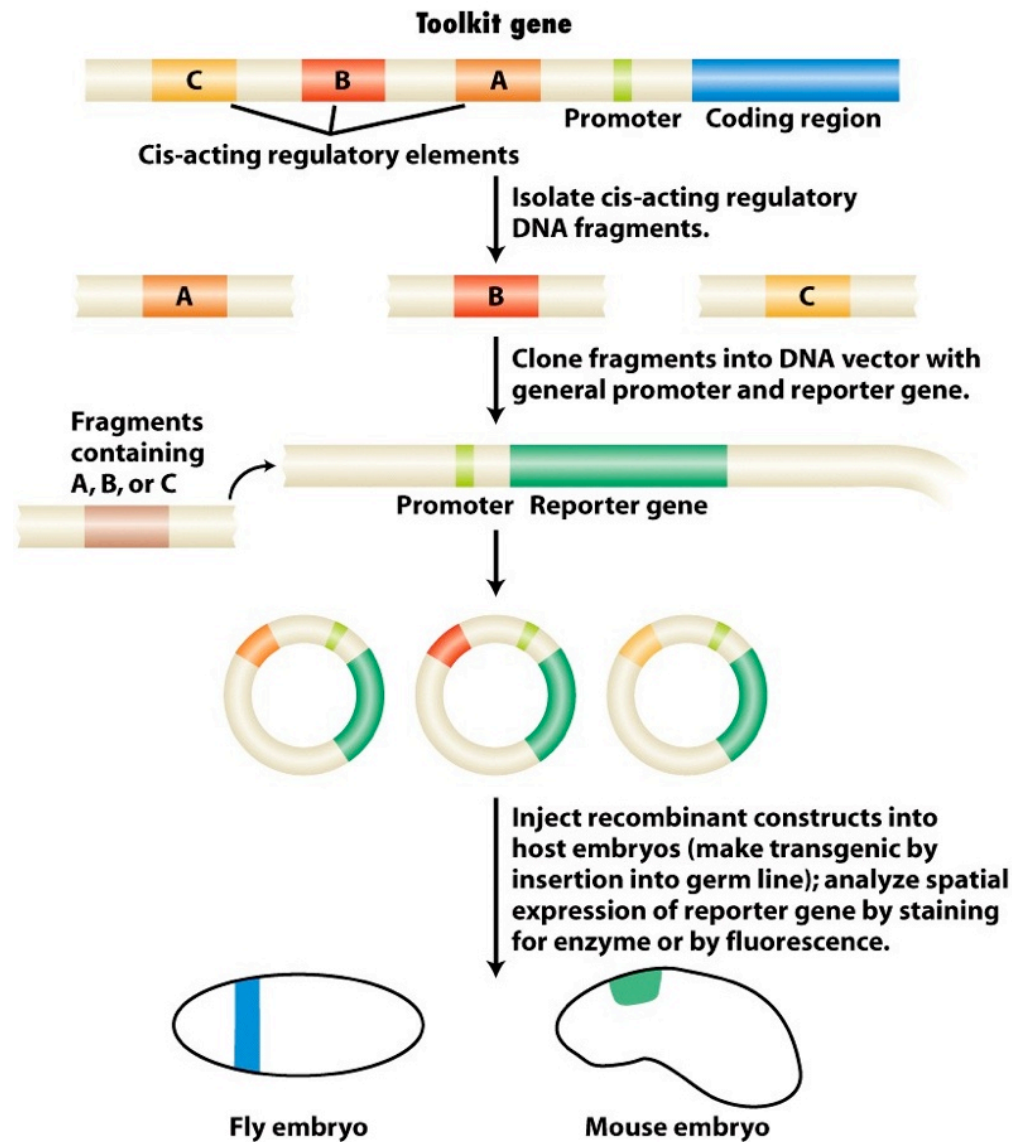
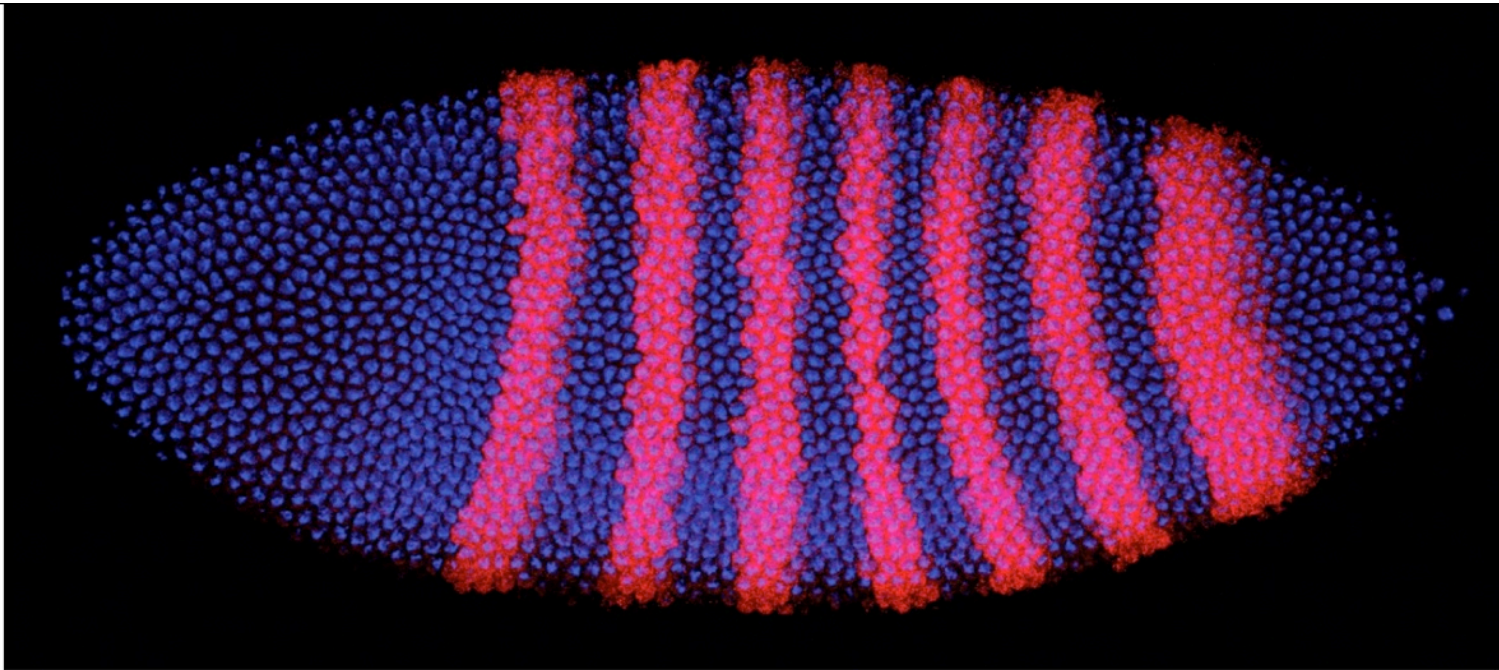


Figure 12-19  
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## **How do you find cis-regulatory sequences?**

1. 'Promoter bashing'. Break up and alter 5' upstream and test expression patterns in reporter gene assays.
2. Binding assays. Measure binding of a transcription factor to 5' upstream region. Various kinds with different resolutions and accuracies (DNase I footprinting, SELEX +bioinformatics & Chromatin IP).



Chapter 12 Opener  
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Eve stripe 2

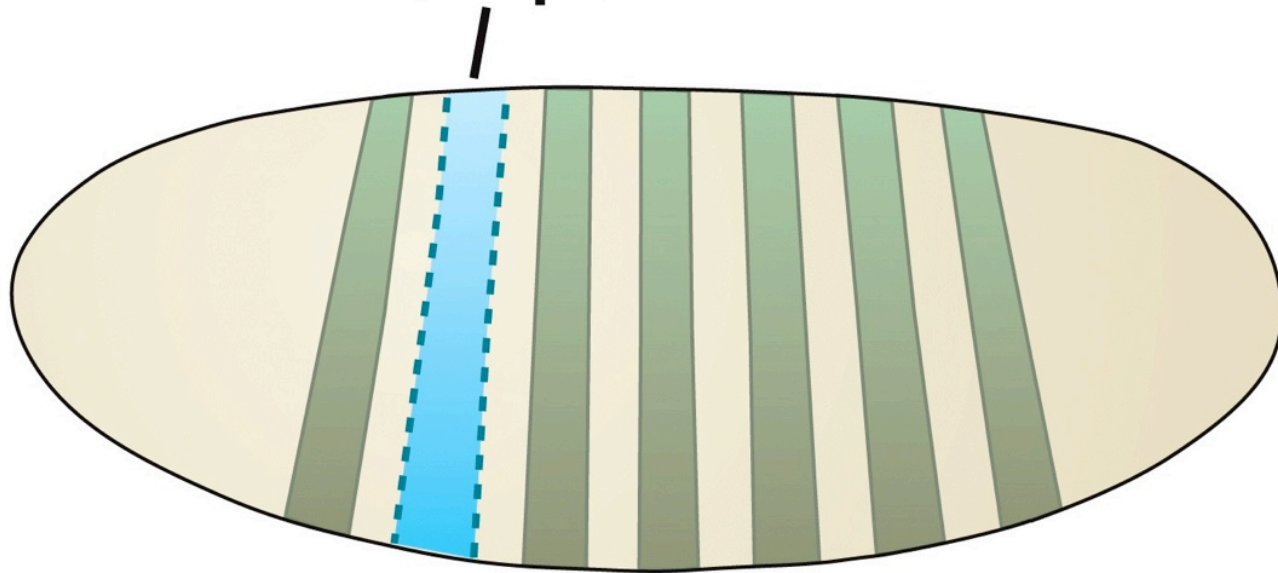


Figure 12-20a  
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Eve stripe 2

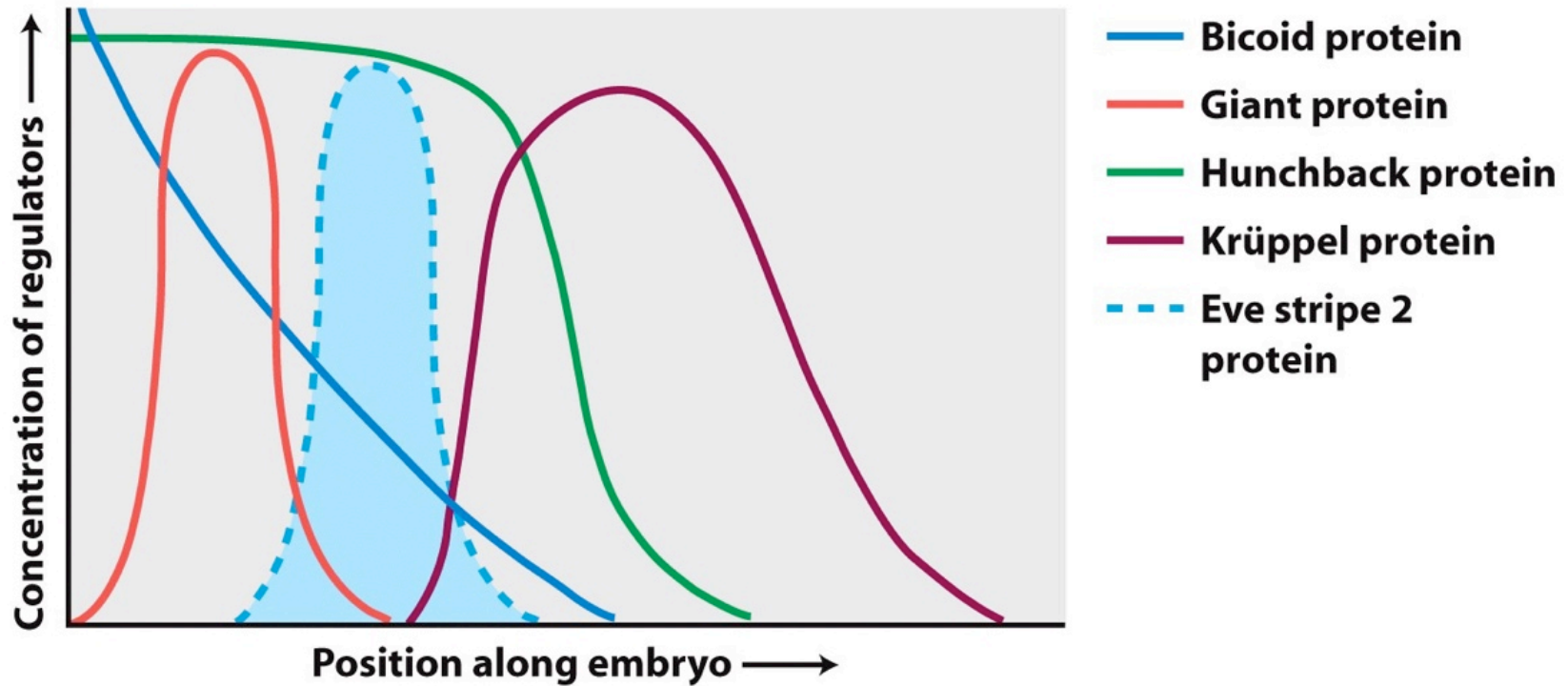
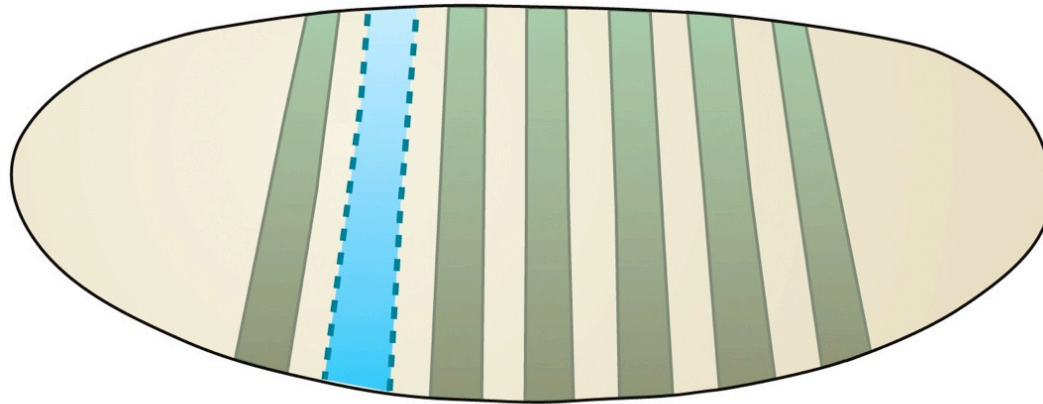


Figure 12-20b  
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## **Some principles:**

1. Cis-regulatory elements integrate combinations of activators and repressors in the spatial regulation of gene expression.
2. Complex patterns of inputs are integrated into simpler patterns through a single cis-regulatory element.
3. The full expression pattern of a gene is often derived from the patterns of many independent cis-regulatory elements.

# Back to Hox genes: how do you make segments different?

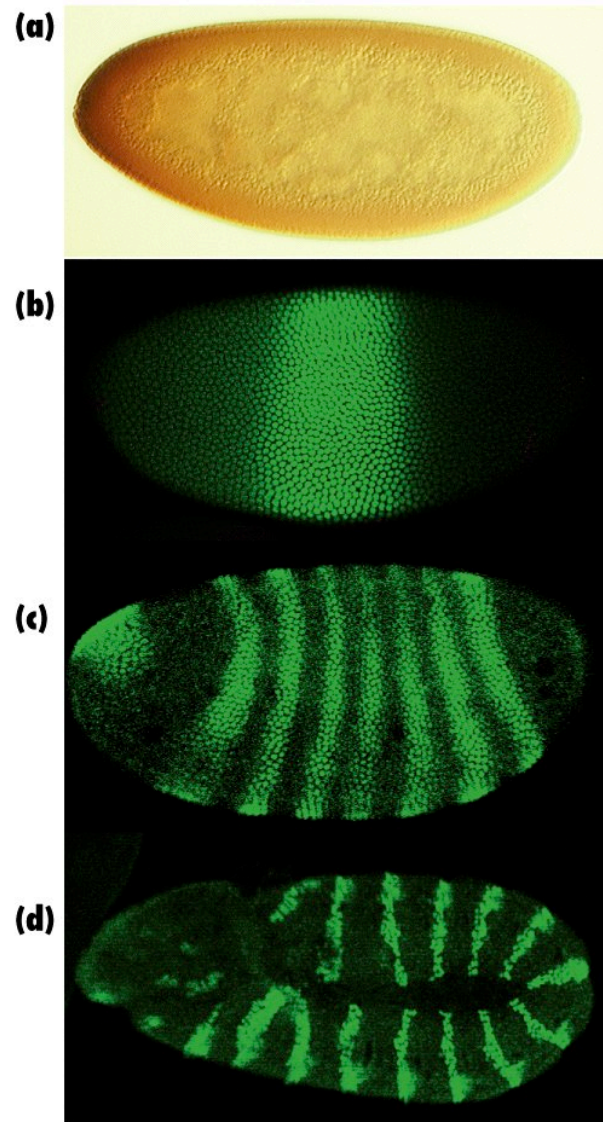


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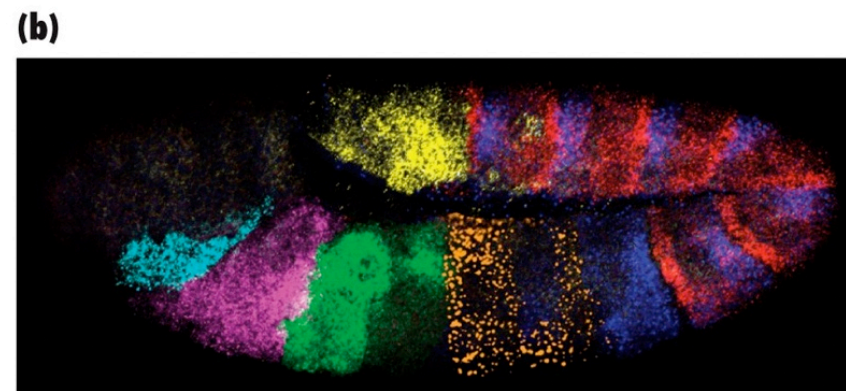
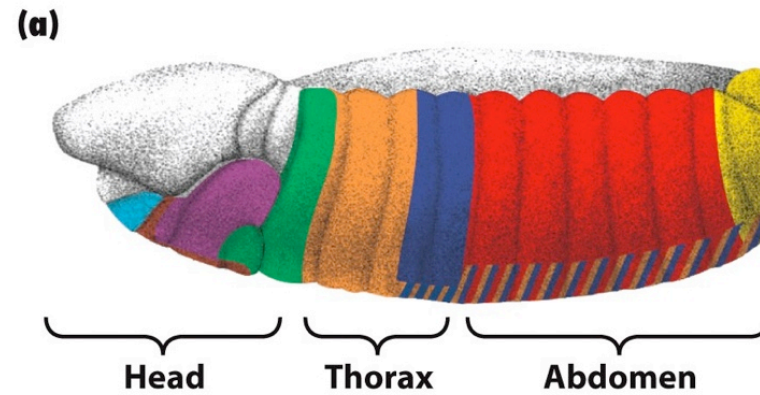
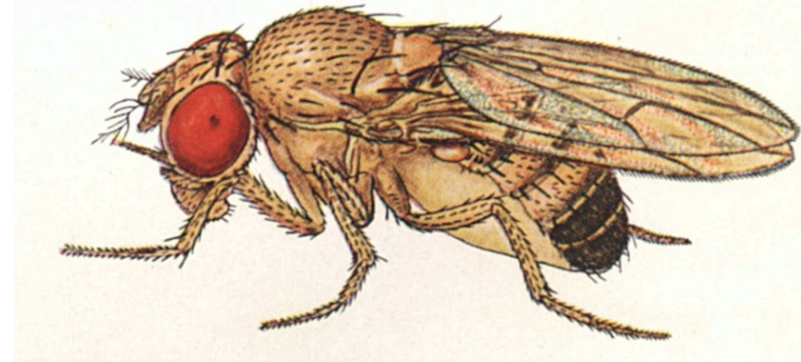


Figure 12-6  
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Distal-less (*Dll*) expression marks appendage primordia & is essential for limb development. How does the fly not develop limbs on its abdomen?

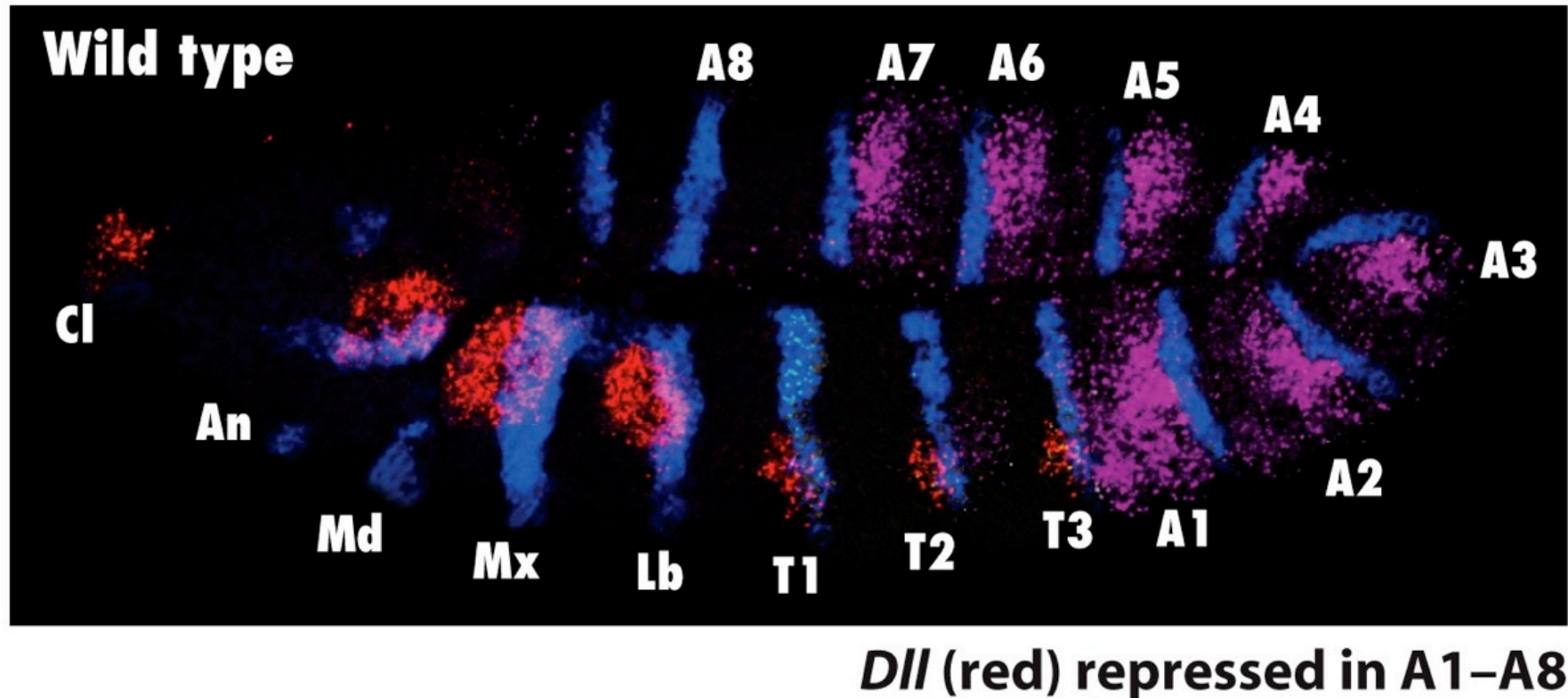


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# Ubx and abd-A (Hox genes) are necessary for repression of Dll in abdominal compartments A1-A7

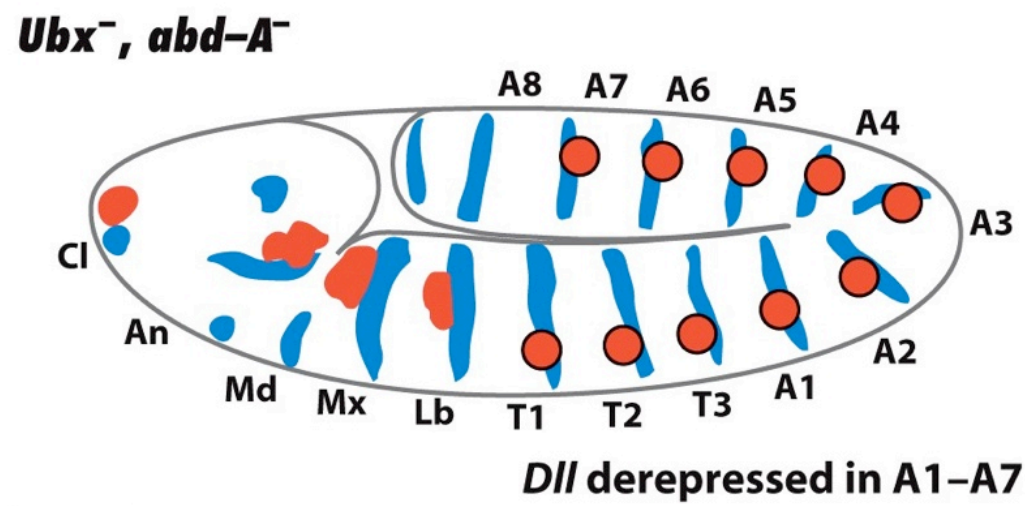
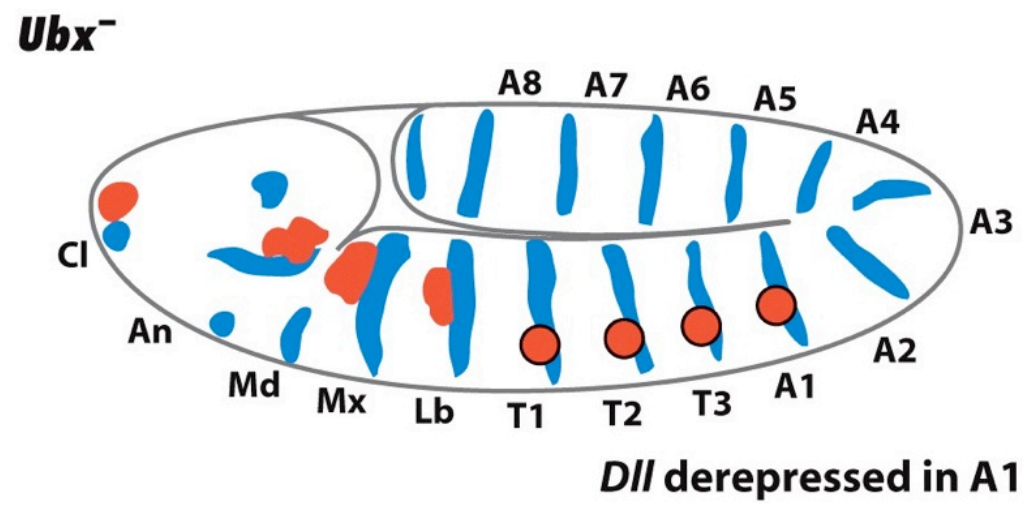


Figure 12-21bc  
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**CIS-ACTING REGULATORY ELEMENT**

**(a) Wild type**



**(b) Hox mutations**



**(c) Slp mutation**



**(d) En mutation**



**(e) Slp, En mutations**



**(f) Exd, Hth mutations**



**REPORTER-GENE EXPRESSION**

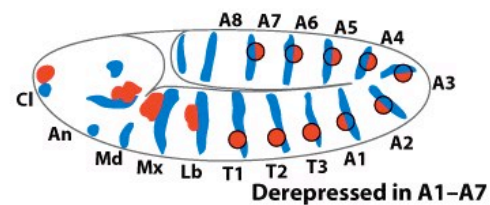
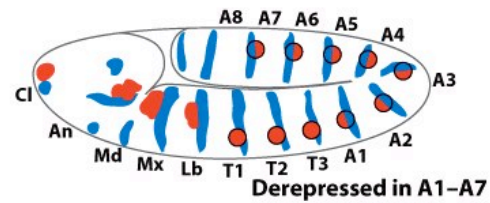
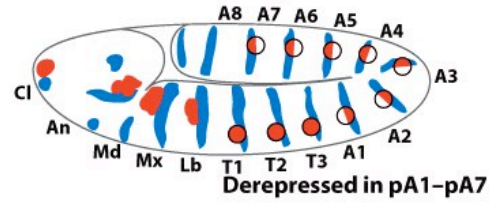
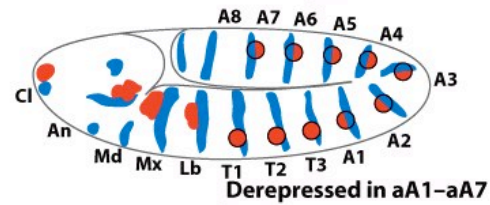
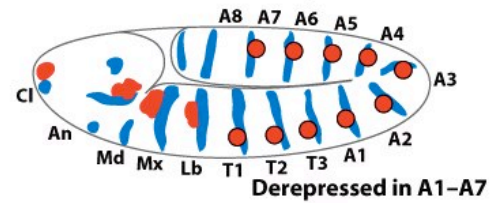
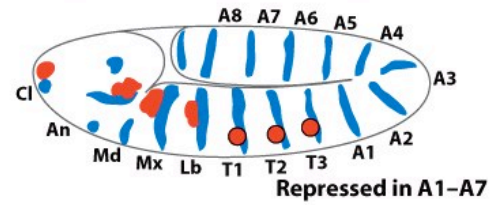


Figure 12-22

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## **Some principles:**

1. Cis-regulatory elements integrate combinations of activators and repressors in the spatial regulation of gene expression.
2. Complex patterns of inputs are integrated into simpler patterns through a single cis-regulatory element.
3. The full expression pattern of a gene is often derived from the patterns of many independent cis-regulatory elements.
4. Combinatorial and cooperative regulation imposes greater specificity on spatial patterns of gene expression.

## **Today:**

1. Genetic screens and genetic analysis of development (*Drosophila* techniques)
2. Regulatory networks in development & cis-regulatory control of expression
3. Post-transcriptional regulation, pleiotropy & disease in development

# Alternative splicing as a means of gene expression regulation

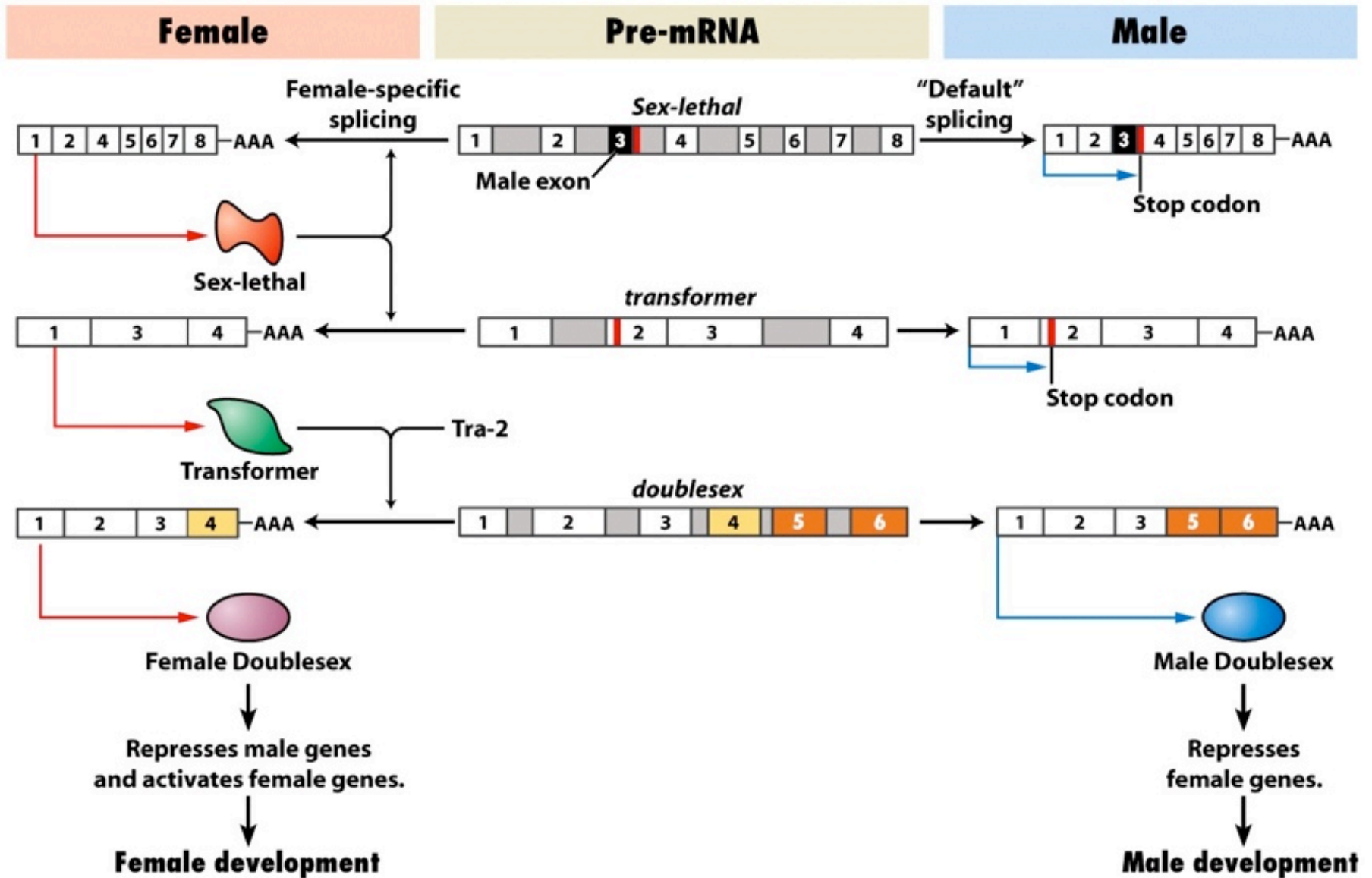


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# Regulation of mRNA translation in *C. elegans*:

How is differential expression of GLP achieved at the four cell stage?

GLD-1 binds 3' UTR and represses translation in posterior cells

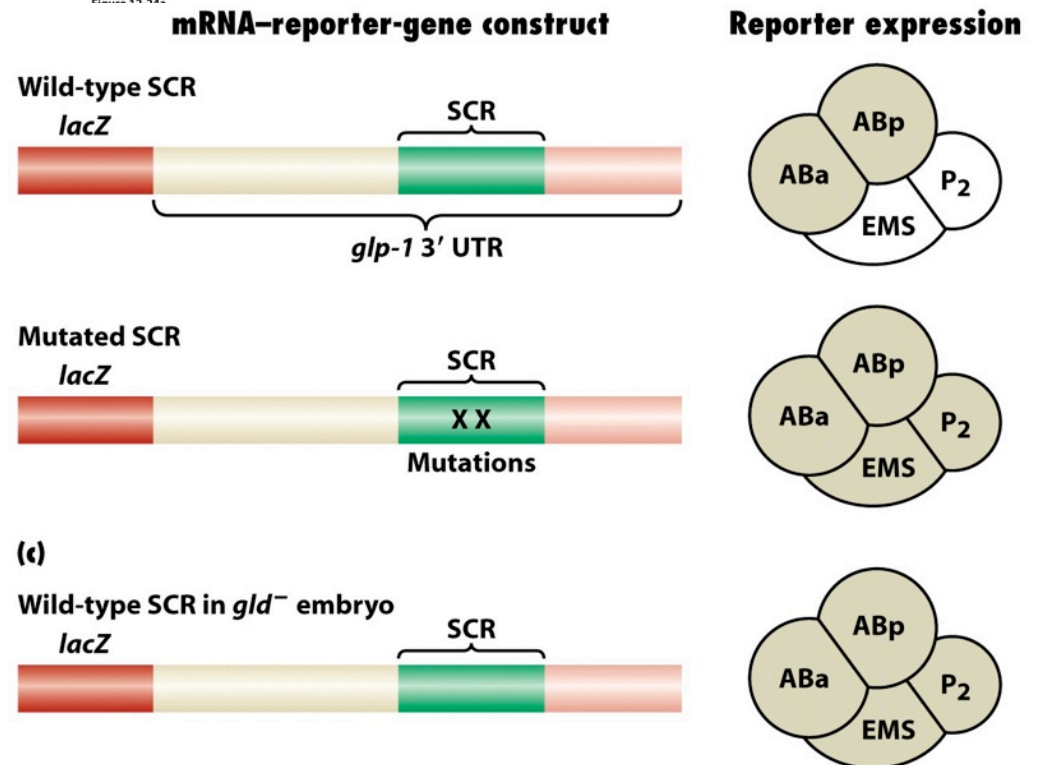
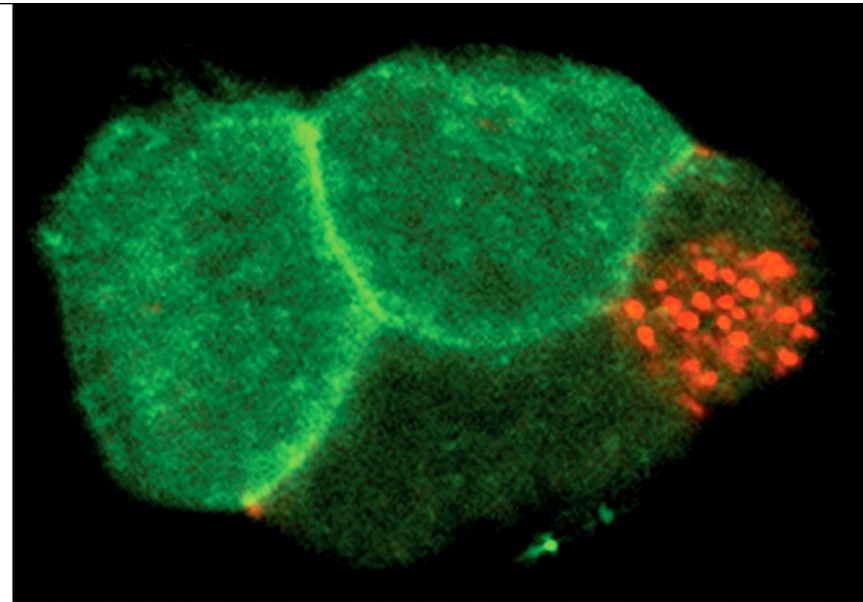
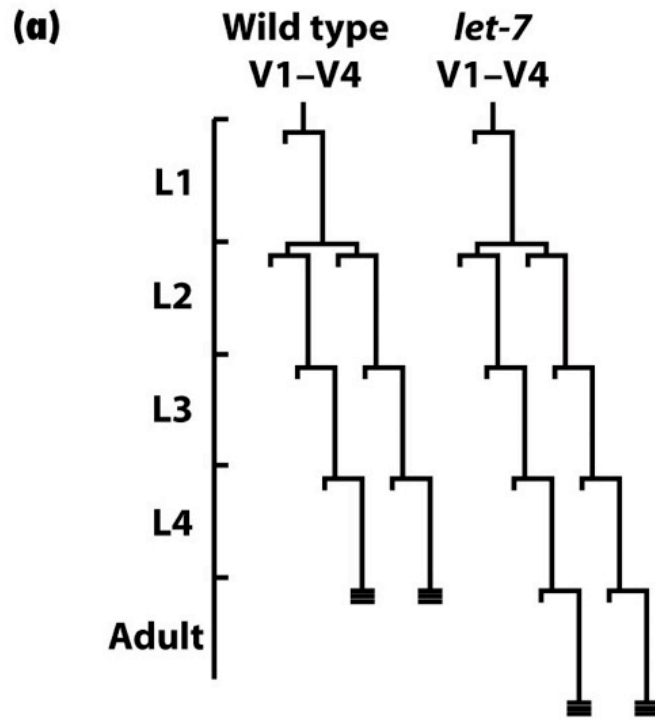


Figure 12-24b  
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# miRNA control of developmental timing in *C. elegans*:



*let-7* represses translation of target genes leading to adult cell fates

*let-7* encodes 70 bp RNA precursor that is processed into a 22 nucleotide miRNA

miRNA binding 3' UTRs of target genes and repress translation

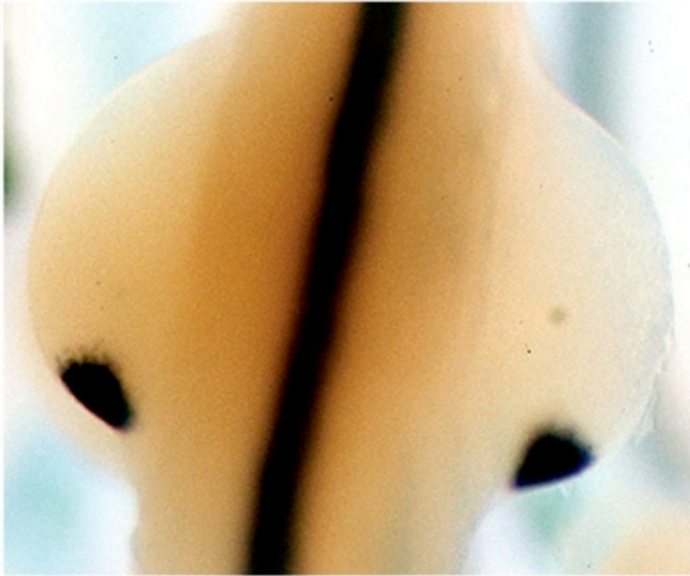
(b)



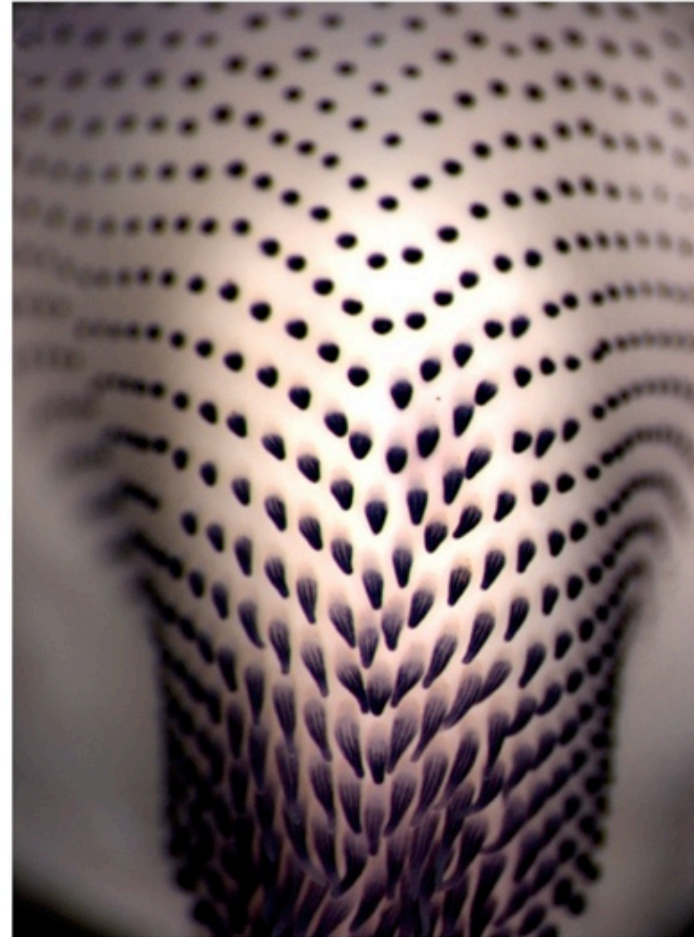


Developmental genes are often used many times throughout development and adult life

(a)



(b)



Sonic hedgehog: segment polarity gene, developing limb bud polarizing zone, neural tube & feather buds

# Developmental genes and disease: from model organisms to human health

Polydactyly is caused by mis-expression of Shh in developing limb bud as a result of mutations in cis-regulatory elements.



Figure 12-27  
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Note that cis-regulatory mutations are often dominant and will only affect one aspect of a genes function.

Holoprosencephaly is a developmental disease leading to abnormalities in brain and midline development caused by a loss of function mutation in the coding region of Shh. Disease can molecularly be understood because of research in model organisms like *Drosophila*.

# Developmental genes discovered in *Drosophila* associated with Cancer

**Table 12-2** Some Toolkit Genes Having Roles in Cancer

|   | Fly gene             | Mammalian gene                 | Cancer type               |                                       |
|---|----------------------|--------------------------------|---------------------------|---------------------------------------|
| <b>Signaling-Pathway Components</b>           | <b>Wingless</b>      | <i>armadillo</i>               | Colon and skin            |                                       |
|   |                      | <i>D.TCF</i>                   | Colon                     |                                       |
|   | <b>Hedgehog</b>      | <i>cubitus interruptus</i>     | <i>Gli1</i>               | Basal cell carcinoma                  |
|   |                      | <i>patched</i>                 | <i>patched</i>            | Basal cell carcinoma, medulloblastoma |
|   |                      | <i>smoothened</i>              | <i>smoothened</i>         | Basal cell carcinoma                  |
|   | <b>Notch</b>         | <i>Notch</i>                   | <i>hNotch1</i>            | Leukemia, lymphoma                    |
|   | <b>EGF receptor</b>  | <i>torpedo</i>                 | <i>C-erbB-2</i>           | Breast and colon                      |
| <b>Decapentaplegic/TFG-<math>\beta</math></b> | <i>Medea</i>         | <i>DPC4</i>                    | Pancreatic and colon      |                                       |
| <b>Toll</b>                                   | <i>dorsal</i>        | <i>NF-<math>\kappa</math>B</i> | Lymphoma                  |                                       |
| <b>Other</b>                                  | <i>extradenticle</i> | <i>Pbx1</i>                    | Acute pre-B-cell leukemia |                                       |