

Bone marrow stromal cell culture

Reagents:

PBS: PBS (Ca, Mg free) +2% P/S
Washing media: DMEM + glu +P/S + 2-mercaptoethanol (2-ME) (Gibco BRL cat#21985-023 0.5ml→500ml)
Culture Media: MEM α -media (Gibco Cat# 12571-063) + L-gln + 20% FBS+P/S + 2-mercaptoethanol (2-ME) (Gibco BRL cat#21985-023 0.5ml→500ml)

Procedure:

A. Preparation of bone marrow cells:

1. Euthanize mice, remove the legs of mice, and then put them in a 50ml conical tube in procedure room. Keep the tube on ice when take the legs to tissue culture room.
2. Aseptically remove femurs/tibia in hood.
3. Cut off ends of each bone to expose marrow. Flush out marrow with PBS into a clean petri dish with syringe.
4. Filter marrow soup through a cell strainer (Falcon, Cat# 352350) into a 50ml conical tube, and then put the tube on ice before counting and plating.

For CFU-F formation assay: Low density 1 or 2 million cells per flask (25cm²)

For bone marrow cell growth: Low density 10-15-20 million cells /100mm dish
Plate 5ml of cell suspension into each 100mm dish

B. Preparation of feeder cells:

1. Sacrificed 4-6 week old Hartley male guinea pig(s) (in building 14), remove the legs, and then put them in a 50ml conical tube in procedure room. Keep the tube on ice when take the legs to tissue culture room.
2. Aseptically remove femurs/tibia in hood.
3. Cut off ends of each bone to expose marrow. Flush out marrow with PBS into a clean petri dish with syringe.
4. Filter marrow soup through a cell strainer (Falcon, Cat# 352350) into a 50ml conical tube, and then put the tube on ice before counting and plating.
5. Resuspend pellet in culture media, α -irradiate (Caesium-137) for 7.5 min. with 6000 cGy by a Gemmacell irradiation. Count the cells.

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- 7. Aseptically remove femurs/tibia in hood.**
- 8. Cut off ends of each bone to expose marrow. Flush out marrow with PBS into a clean petri dish with syringe.**
- 9. Filter marrow soup through a cell strainer (Falcon, Cat# 352350) into a 50ml conical tube, and then put the tube on ice before counting and plating.**
- 10. Resuspend pellet in culture media, \square -irradiate (Caesium-137) for 7.5 min. with 6000 cGy by a Gemmacell irradiation.**
- 11. Count the cells.**

CFU-F formation assay:

Reagents:

**Culture Media: MEM α -media (Gibco cat# 12571-063) + L-gln + 20% FBS+P/S
+ 2-mercaptoethanol (2-ME) (Gibco BRL cat#21985-023
0.5ml \rightarrow 500ml)**

PBS: w/o Ca, Mg

Saturated methyl violet (sigma): Dissolve methyl violet into water and filter through Whatman paper filter.

- 1 Add 5 ml of mouse bone marrow stromal cells in each 3-5 of 25cm² tissue culture flasks; incubate for 3 hours to allow adherent cells to attach.**
- 2 Wash the cells twice with 5 ml PBS DMEM medium to remove non-adherent cells.**
- 3 Add 5 ml of feeder cells (10 million each flask) with or without TGF- α .**
- 4 Incubate for until colonies are formed (12-14 days).**
- 5 Wash the cells twice with 5 ml PBS w/o Ca and Mg⁺⁺.**
- 6 Fix the cells for 20min.with 3 ml of absolute methanol.**
- 7 Stain the cells with alkaline phosphatase (sigma) for 20 min.**
- 8 Wash with distilled water until water is clear.**
- 9 Count the number of colonies.**
- 10 Stain the cells with saturated methyl violet (sigma) for 20 min.**
- 11 Wash with distilled water until water is clear.**
- 12 Count the number of colonies.**

Passing stromal cells

Reagents:

Media: MEM-□ w/ 20%FBS, P/S L-glu + Dex (10^{-8})

Collagenase: 250u/ml in PBS w/o Ca⁺⁺ and Mg⁺⁺ + TLC

1. Aspirate media.
 2. Wash the cells with 5 ml PBS w/o Ca⁺⁺ and Mg⁺⁺ with EDTA.
 3. Add 5ml of collagenase, incubate for about 20-25 minutes, and then transfer media to a 50ml conical tube with 10ml media.
 4. Spin suspension for 5mins. at 1000 rpm, 4⁰C.
 5. Resuspend pellet in media, count
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1. Avg count per sample Add 4ml cell solution to 60mm dishes for pass the cells.
 2. Add 2ml cell solution to each well of a 12-well plate for bone formation assay
 3. Add 2ml cell solution to each well of a 6-well plate for western assay
 4. Add 300ul to each well of 8-well chamber slice for apoptosis assay.
 5. Add 300ul to each well of 8-well chamber slice for proliferation (BrdU) assay.
 6. Add 200ul to each well of 96-well plate for Ca²⁺⁺ deposition assay.
 7. Culture at 37⁰C 5% CO₂.

Proliferation assay (MTT)

Bone marrow Stromal cells

Media: MEM-□ w/ 20%FBS, P/S L-glu + Dex (10^{-8})

MTT kit: Boehringer Mannheim cat# 1465 007

- 1. Aspirate media.**
- 2. Wash the cells with 5 ml PBS w/o Ca⁺⁺ and Mg⁺⁺ with EDTA.**
- 3. Add 5ml of collagenase, incubate for about 20-25 minutes, and then transfer media to a 50ml conical tube with 10ml media.**
- 4. Spin suspension for 5mins. at 1000 rpm, 4⁰C.**
- 5. Resuspend pellet in media, count cells**
- 6. Add 100ul cell solution to wells of 96-well plate (Labeled 2000cells/well).**
- 7. Add 50ul media 50ul cell solution to wells that labeled 1000 cells/well).**
- 8. Culture for 1, 2, 3, 4, 5, 6, 7 days**
- 9. Add 10ul MTT labeling solution into each well, incubate for 4 hours, and add 100ul solubilization solution and incubate for overnight**
- 10. Measure OD at 560nm.**